

SARS-CoV-2 and COVID-19: An Evolving Review of Diagnostics and Therapeutics

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1 Pathogenesis, Symptomatology, and Transmission of SARS-CoV-2 through Analysis of Viral Genomics and Structure

1.1 Abstract

The novel coronavirus SARS-CoV-2, which emerged in late 2019, has since spread around the world and infected hundreds of millions of people with coronavirus disease 2019 (COVID-19). While this viral species was unknown prior to January 2020, its similarity to other coronaviruses that infect humans has allowed for rapid insight into the mechanisms that it uses to infect human hosts, as well as the ways in which the human immune system can respond. Here, we contextualize SARS-CoV-2 among other coronaviruses and identify what is known and what can be inferred about its behavior once inside a human host. Because the genomic content of coronaviruses, which specifies the virus's structure, is highly conserved, early genomic analysis provided a significant head start in predicting viral pathogenesis and in understanding potential differences among variants. The pathogenesis of the virus offers insights into symptomatology, transmission, and individual susceptibility. Additionally, prior research into interactions between the human immune system and coronaviruses has identified how these viruses

can evade the immune system's protective mechanisms. We also explore systems-level research into the regulatory and proteomic effects of SARS-CoV-2 infection and the immune response. Understanding the structure and behavior of the virus serves to contextualize the many facets of the COVID-19 pandemic and can influence efforts to control the virus and treat the disease.

1.2 Importance

COVID-19 involves a number of organ systems and can present with a wide range of symptoms. From how the virus infects cells to how it spreads between people, the available research suggests that these patterns are very similar to those seen in the closely related viruses SARS-CoV-1 and possibly MERS-CoV. Understanding the pathogenesis of the SARS-CoV-2 virus also contextualizes how the different biological systems affected by COVID-19 connect. Exploring the structure, phylogeny, and pathogenesis of the virus therefore helps to guide interpretation of the broader impacts of the virus on the human body and on human populations. For this reason, an in-depth exploration of viral mechanisms is critical to a robust understanding of SARS-CoV-2 and, potentially, future emergent HCoV.

1.3 Introduction

The current coronavirus disease 2019 (COVID-19) pandemic, caused by the *Severe acute respiratory syndrome-related coronavirus 2* (SARS-CoV-2) virus, represents an acute global health crisis. Symptoms of the disease can range from mild to severe or fatal (8) and can affect a variety of organs and systems (9). Outcomes of infection can include acute respiratory distress (ARDS) and acute lung injury, as well as damage to other organ systems (9, 10). Understanding the progression of the disease, including these diverse symptoms, depends on understanding how the virus interacts with the host. Additionally, the fundamental biology of the virus can provide insights into how it is transmitted among people, which can, in turn, inform efforts to control its spread. As a result, a thorough understanding of the pathogenesis of SARS-CoV-2 is a critical foundation on which to build an understanding of COVID-19 and the pandemic as a whole.

The rapid identification and release of the genomic sequence of the virus in January 2020 (11) provided early insight into the virus in a comparative genomic context. The viral genomic sequence clusters with known coronaviruses (order *Nidovirales*, family *Coronaviridae*, subfamily *Orthocoronavirinae*). Phylogenetic analysis of the coronaviruses reveals four major subclades, each corresponding to a genus: the alpha, beta, gamma, and delta coronaviruses. Among them, alpha and beta coronaviruses infect mammalian species, gamma coronaviruses infect avian species, and delta coronaviruses infect both mammalian and avian species (12). The novel virus now known as SARS-CoV-2 was identified as a beta coronavirus belonging to the B lineage based on phylogenetic analysis of a polymerase chain reaction (PCR) amplicon fragment from five patients along with the full genomic sequence (13). This lineage also includes the *Severe acute respiratory syndrome-related coronavirus* (SARS-CoV-1) that caused the 2002-2003 outbreak of Severe Acute Respiratory Syndrome (SARS) in humans (13). (Note

that these subclades are not to be confused with variants of concern within SARS-CoV-2 labeled with Greek letters; i.e., the Delta variant of SARS-CoV-2 is still a beta coronavirus.)

Because viral structure and mechanisms of pathogenicity are highly conserved within the order, this phylogenetic analysis provided a basis for forming hypotheses about how the virus interacts with hosts, including which tissues, organs, and systems would be most susceptible to SARS-CoV-2 infection. Coronaviruses that infect humans (HCoV) are not common, but prior research into other HCoV such as SARS-CoV-1 and *Middle East respiratory syndrome-related coronavirus* (MERS-CoV), as well as other viruses infecting humans such as a variety of influenza species, established a strong foundation that accelerated the pace of SARS-CoV-2 research.

Coronaviruses are large viruses that can be identified by their distinctive “crown-like” shape (Figure 1). Their spherical virions are made from lipid envelopes ranging from 100 to 160 nanometers in which peplomers (protruding structures) of two to three spike (S) glycoproteins are anchored, creating the crown (14, 15). These spikes, which are critical to both viral pathogenesis and to the response by the host immune response, have been visualized using cryo-electron microscopy (16). Because they induce the human immune response, they are also the target of many proposed therapeutic agents (2, 3). Viral pathogenesis is typically broken down into three major components: entry, replication, and spread (17). However, in order to draw a more complete picture of pathogenesis, it is also necessary to examine how infection manifests clinically, identify systems-level interactions between the virus and the human body, and consider the possible effects of variation or evolutionary change on pathogenesis and virulence. Thus, clinical medicine and traditional biology are both important pieces of the puzzle of SARS-CoV-2 presentation and pathogenesis.

1.4 Coronavirus Structure and Pathogenesis

1.4.1 Structure of Coronaviruses

Genome structure is highly conserved among coronaviruses, meaning that the relationship between the SARS-CoV-2 genome and its pathogenesis can be inferred from prior research in related viral species. The genomes of viruses in the *Nidovirales* order share several fundamental characteristics. They are non-segmented, which means the viral genome is a single continuous strand of RNA, and are enveloped, which means that the genome and capsid are encased by a lipid bilayer. Coronaviruses have large positive-sense RNA (ssRNA+) genomes ranging from 27 to 32 kilobases in length (18, 19). The SARS-CoV-2 genome lies in the middle of this range at 29,903 bp (19). Genome organization is highly conserved within the order (18). There are three major genomic regions: one containing the replicase gene, one containing the genes encoding structural proteins, and interspersed accessory genes (18) (Figure 1). The replicase gene comprises about two-thirds of the genome and consists of two open reading frames that are translated with ribosomal frameshifting (18). This polypeptide is then translated into 16 non-structural proteins (nsp), except in gammacoronaviruses where nsp1 is absent, that form the replication

machinery used to synthesize viral RNA (20). The remaining third of the genome encodes structural proteins, including the spike (S), membrane, envelope, and nucleocapsid proteins. Additional accessory genes are sometimes present between these two regions, depending on the species or strain. Much attention has been focused on the S protein, which is a critical structure involved in cell entry.

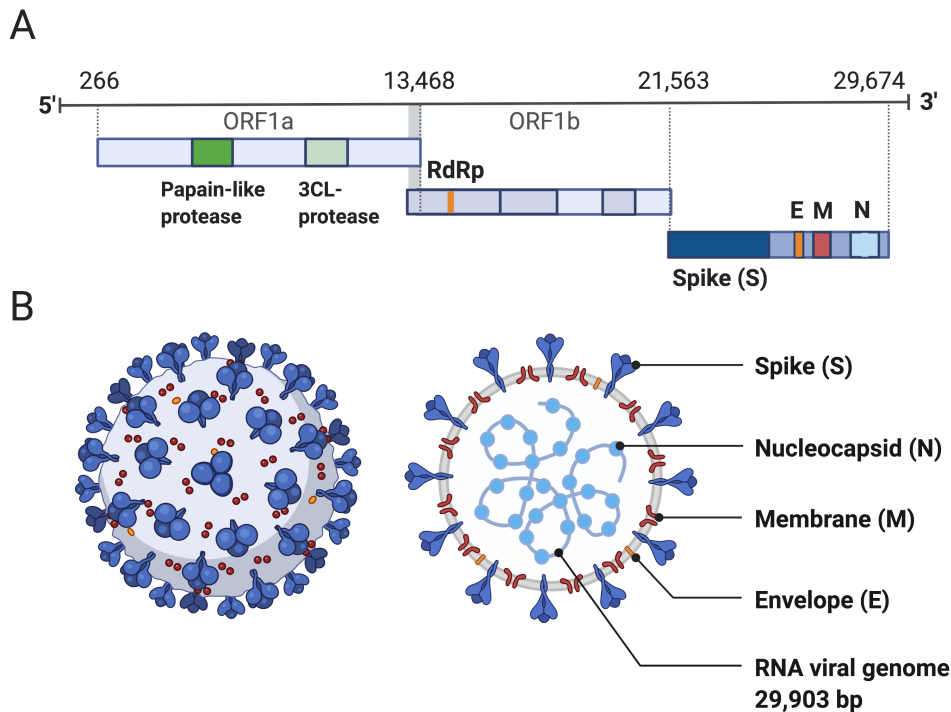


Figure 1: Structure of SARS-CoV-2 capsid and genome. A) The genomic structure of coronaviruses is highly conserved and includes three main regions. Open reading frames (ORF) 1a and 1b contain two polyproteins that encode the non-structural proteins (nsp). The nsp include enzymes such as RNA-dependent RNA Polymerase (RdRp). The last third of the genome encodes structural proteins, including the spike (S), envelope (E), membrane (M) and nucleocapsid (N) proteins. Accessory genes can also be interspersed throughout the genome (18). B) The physical structure of the coronavirus virion, including the components determined by the conserved structural proteins S, E, M and N. This figure was adapted from “Human Coronavirus Structure”, by BioRender.com (2020), retrieved from <https://app.biorender.com/biorender-templates>.

1.4.2 Pathogenic Mechanisms of Coronaviruses

While it is possible that SARS-CoV-1 and SARS-CoV-2, like most viruses, enter cells through endocytosis, a process conserved among coronaviruses enables them to target cells for entry through fusion with the plasma membrane (21, 22). Cell entry proceeds in three steps: binding, cleavage, and fusion. First, the viral spike protein binds to a host cell via a recognized receptor or entry point. Coronaviruses can bind to a range of host receptors (23, 24), with binding conserved only at the genus level (12). Viruses in the beta coronavirus genus, to which SARS-CoV-2 belongs, are known to bind to the CEACAM1 protein, 5-N-acetyl-9-O-acetyl neuraminic acid, and to angiotensin-converting enzyme 2 (ACE2) (23). This recognition is driven by domains in the S1 subunit (25). SARS-CoV-2 has a high affinity for human ACE2, which is expressed in the vascular epithelium, other epithelial cells, and cardiovascular and renal tissues (26, 27), as well as many others (28). The binding process is guided by the molecular structure of the spike protein, which is structured in three segments: an ectodomain, a transmembrane

anchor, and an intracellular tail (29). The ectodomain forms the crown-like structures on the viral membrane and contains two subdomains known as the S1 and S2 subunits (30). The S1 (N-terminal) domain forms the head of the crown and contains the receptor binding motif, and the S2 (C-terminal) domain forms the stalk that supports the head (30). The S1 subunit guides the binding of the virus to the host cell, and the S2 subunit guides the fusion process (29).

After the binding of the S1 subunit to an entry point, the spike protein of coronaviruses is often cleaved at the S1/S2 boundary into the S1 and S2 subunits by a host protease (25, 31, 32). This proteolytic priming is important because it prepares the S protein for fusion (31, 32). The two subunits remain bound by van der Waals forces, with the S1 subunit stabilizing the S2 subunit throughout the membrane fusion process (25). Cleavage at a second site within S2 (S2') activates S for fusion by inducing conformational changes (25). Similar to SARS-CoV-1, SARS-CoV-2 exhibits redundancy in which host proteases can cleave the S protein (33). Both transmembrane protease serine protease-2 (TMPRSS-2) and cathepsins B/L have been shown to mediate SARS-CoV-2 S protein proteolytic priming, and small molecule inhibition of these enzymes fully inhibited viral entry *in vitro* (33, 34). Other proteases known to cleave the S1/S2 boundary in coronaviruses include TMPRSS-4, trypsin, furin, cathepsins, and human airway trypsin-like protease (HAT) (34).

Unlike in SARS-CoV-1, a second cleavage site featuring a furin-like binding motif is also present near the S1/S2 boundary in SARS-CoV-2 (35). This site is found in HCoV belonging to the A and C lineages of beta coronavirus, including MERS-CoV, but not in the other known members of the B lineage of beta coronavirus that contains SARS-CoV-1 and SARS-CoV-2 (35). It is associated with increased virulence in other viral species (35) and may facilitate membrane fusion of SARS-CoV-2 in the absence of other proteases that prime the S1/S2 site (36). However, given that proteases such as HAT are likely to be present in targets like the human airway, the extent to which this site has had a real-world effect on the spread of SARS-CoV-2 was initially unclear (36). Subsequent research has supported this site as an important contributor to pathogenesis: *in vitro* analyses have reported that it bolsters pathogenicity specifically in cell lines derived from human airway cells (Calu3 cell line) (37–39) and that furin inhibitors reduced pathogenic effects in VeroE6 cells (40).

Electron microscopy suggests that in some coronaviruses, including SARS-CoV-1 and MERS-CoV, a six-helix bundle separates the two subunits in the postfusion conformation, and the unusual length of this bundle facilitates membrane fusion through the release of additional energy (12). The viral membrane can then fuse with the endosomal membrane to release the viral genome into the host cytoplasm. Once the virus enters a host cell, the replicase gene is translated and assembled into the viral replicase complex. This complex then synthesizes the double-stranded RNA (dsRNA) genome from the genomic ssRNA(+). The dsRNA genome is transcribed and replicated to create viral mRNAs and new ssRNA(+) genomes (18, 41). From there, the virus can spread into other cells. In SARS-CoV-2, the insertion of the furin-like binding site near the S1/S2 boundary is also thought to increase cell-cell adhesion, making it possible for the viral genome to spread directly from cell

to cell rather than needing to propagate the virion itself (42). In this way, the genome of SARS-CoV-2 provides insight into the pathogenic behavior of the virus.

Evidence also suggests that SARS-CoV-2 may take advantage of the specific structure of endothelial cells to enter the circulatory system. Endothelial cells are specialized epithelial cells (43) that form a barrier between the bloodstream and surrounding tissues. The endothelium facilitates nutrient, oxygen, and cellular exchange between the blood and vascularized tissues (44). The luminal (interior) surface of the endothelium is lined with glycocalyx, a network of both membrane-bound and soluble proteins and carbohydrates, primarily proteoglycans and glycoproteins (45, 46). The glycocalyx varies in thickness from 0.5 microns in the capillaries to 4.5 microns in the carotid arteries and forms a meshwork that localizes both endothelial- and plasma-derived signals to the inner vessel wall (45). Heparan sulfate is the dominant proteoglycan in the glycocalyx, representing 50-90% of glycocalyx proteoglycan content (47). The SARS-CoV-2 spike protein can bind directly to heparan sulfate, which serves in part as a scaffolding molecule to facilitate ACE2 binding and entry into endothelial cells (46). A heparan sulfate binding site has also been identified near the ACE2 binding site on the viral receptor binding domain (RBD), and modeling has suggested that heparan sulfate binding yields an open conformation that facilitates binding to ACE2 on the cell surface (46). Degrading or removing heparan sulfate was associated with decreased binding (46). Heparan sulfate may also interact with the S1/S2 proteolytic cleavage site and other binding sites to promote binding affinity (48). Notably, treatment with soluble heparan sulfate or even heparin (a commonly used anti-coagulant and vasodilator that is similar in structure to heparan sulfate (49)) potentially blocked spike protein binding and viral infection (46). This finding is particularly interesting because degradation of heparan sulfate in the glycocalyx has previously been identified as an important contributor to ARDS and sepsis (50), two common and severe outcomes of COVID-19, and suggests that heparan sulfate could be a target for pharmaceutical inhibition of cell entry by SARS-CoV-2 (51–55). Together, this evidence suggests that heparan sulfate can serve as an important adhesion molecule for SARS-CoV-2 cell entry. It may represent a therapeutic target but has not been pursued as much as other candidate targets (3).

1.4.3 Immune Evasion Strategies

Research in other HCoV provides some indication of how SARS-CoV-2 infection can proceed despite human immune defenses. Infecting the epithelium can help viruses such as SARS-CoV-1 bypass the physical barriers, such as mucus, that comprise the immune system's first line of defense (56). Once the virus infiltrates host cells, it is adept at evading detection. CD163+ and CD68+ macrophage cells are especially crucial for the establishment of SARS-CoV-1 in the body (56). These cells most likely serve as viral reservoirs that help shield SARS-CoV-1 from the innate immune response. According to a study on the viral dissemination of SARS-CoV-1 in Chinese macaques, viral RNA could be detected in some monocytes throughout the process of differentiation into dendritic cells (56). This lack of active viral replication allows SARS-CoV-1 to escape the innate immune response because reduced levels of detectable viral RNA allow the virus to avoid both natural killer cells

and Toll-like receptors (56). Even during replication, SARS-CoV-1 is able to mask its dsRNA genome from detection by the immune system. Although dsRNA is a pathogen-associated molecular pattern that would typically initiate a response from the innate immune system (57), *in vitro* analysis of nidoviruses including SARS-CoV-1 suggests that these viruses can induce the development of double-membrane vesicles that protect the dsRNA signature from being detected by the host immune system (58). This protective envelope can therefore insulate these coronaviruses from the innate immune system's detection mechanism (59).

HCoVs are also known to interfere with the host immune response, rather than just evade it. For example, the virulence of SARS-CoV-2 is increased by nsp1, which can suppress host gene expression by stalling mRNA translation and inducing endonucleolytic cleavage and mRNA degradation (60). SARS-CoV-1 also evades the immune response by interfering with type I IFN induction signaling, which is a mechanism that leads to cellular resistance to viral infections. SARS-CoV-1 employs methods such as ubiquitination and degradation of RNA sensor adaptor molecules MAVS and TRAF3/6 (61). Also, MERS-CoV downregulates antigen presentation via MHC class I and MHC class II, which leads to a reduction in T cell activation (61). These evasion mechanisms, in turn, may facilitate systemic infection. Coronaviruses such as SARS-CoV-1 are also able to evade the humoral immune response through other mechanisms, such as inhibiting certain cytokine pathways or down-regulating antigen presentation by the cells (58).

1.4.4 Host Cell Susceptibility

ACE2 and TMPRSS-2 have been identified as the primary entry portal and as a critical protease, respectively, in facilitating the entry of SARS-CoV-1 and SARS-CoV-2 into a target cell (16, 33, 62–64). This finding has led to a hypothesized role for the expression of these molecules in determining which cells, tissues, and organs are most susceptible to SARS-CoV-2 infection. ACE2 is expressed in numerous organs, such as the heart, kidney, and intestine, but it is most prominently expressed in alveolar epithelial cells; this pattern of expression is expected to contribute to the virus' association with lung pathology (26, 65, 66) as well as that of SARS (67). A retrospective observational study reported indirect evidence that certain antineoplastic therapies, such as the chemotherapy drug gemcitabine, may reduce risk of SARS-CoV-2 infection in patients with cancer, possibly via decreased ACE2 expression (68). Additionally, the addition of the furin site insertion at the S1/S2 boundary means that SARS-CoV-2 does not require TMPRSS-2 when furin, an ubiquitously expressed endoprotease (69), is present, enabling cell-cell fusion independent of TMPRSS-2 availability (70).

Clinical investigations of COVID-19 patients have detected SARS-CoV-2 transcripts in bronchoalveolar lavage fluid (BALF) (93% of specimens), sputum (72%), nasal swabs (63%), fibrobronchoscopy brush biopsies (46%), pharyngeal swabs (32%), feces (29%), and blood (1%) (71). Two studies reported that SARS-CoV-2 could not be detected in urine specimens (71, 72); however, a third study identified four urine samples (out of 58) that were positive for SARS-CoV-2 nucleic acids (73). Although respiratory failure remains the leading cause of death for COVID-19 patients (74), SARS-CoV-2 infection can damage many other organ systems including the heart (75),

kidneys ([76](#), [77](#)), liver ([78](#)), and gastrointestinal tract ([79](#), [80](#)). As it becomes clear that SARS-CoV-2 infection can damage multiple organs, the scientific community is pursuing multiple avenues of investigation in order to build a consensus about how the virus affects the human body.

1.5 Clinical Presentation of COVID-19

SARS-CoV-2 pathogenesis is closely linked with the clinical presentation of the COVID-19 disease. Reports have described diverse symptom profiles associated with COVID-19, with a great deal of variability both within and between institutions and regions. Definitions for non-severe, severe, and critical COVID-19, along with treatment recommendations, are available from the World Health Organization living guidelines ([81](#)). A large study from Wuhan, China conducted early in the pandemic identified fever and cough as the two most common symptoms that patients reported at hospital admission ([82](#)), while a retrospective study in China described the clinical presentations of patients infected with SARS-CoV-2 as including lower respiratory tract infection with fever, dry cough, and dyspnea (shortness of breath) ([83](#)). This study ([83](#)) noted that upper respiratory tract symptoms were less common, suggesting that the virus preferentially targets cells located in the lower respiratory tract. However, data from the New York City region ([84](#), [85](#)) showed variable rates of fever as a presenting symptom, suggesting that symptoms may not be consistent across individuals. For example, even within New York City, one study ([84](#)) identified low oxygen saturation (<90% without the use of supplemental oxygen or ventilation support) in 20.4% of patients upon presentation, with fever being present in 30.7%, while another study ([85](#)) reported cough (79.4%), fever (77.1%), and dyspnea (56.5%) as the most common presenting symptoms; both of these studies considered only hospitalized patients. A later study reported radiographic findings such as ground-glass opacity and bilateral patchy shadowing in the lungs of many hospitalized patients, with most COVID-19 patients having lymphocytopenia, or low levels of lymphocytes (a type of white blood cell) ([82](#)). Patients may also experience loss of smell, myalgias (muscle aches), fatigue, or headache. Gastrointestinal symptoms can also present ([86](#)), and the CDC includes nausea and vomiting, as well as congestion and runny nose, on its list of symptoms consistent with COVID-19 ([8](#)). An analysis of an app-based survey of 500,000 individuals in the U.S. found that among those tested for SARS-CoV-2, a loss of taste or smell, fever, and a cough were significant predictors of a positive test result ([87](#)). It is important to note that in this study, the predictive value of symptoms may be underestimated if they are not specific to COVID-19. This underestimation could occur because the outcome measured was a positive, as opposed to a negative, COVID-19 test result, meaning an association would be more easily identified for symptoms that were primarily or exclusively found with COVID-19. At the time the surveys were conducted, due to limits in U.S. testing infrastructure, respondents typically needed to have some symptoms known to be specific to COVID-19 in order to qualify for testing. Widespread testing of asymptomatic individuals may therefore provide additional insight into the range of symptoms associated with COVID-19.

Consistent with the wide range of symptoms observed and the pathogenic mechanisms described above, COVID-19 can affect a variety of systems within the body in addition to causing respiratory problems ([88](#)). For

example, COVID-19 can lead to acute kidney injury, especially in patients with severe respiratory symptoms or certain preexisting conditions (89). Some patients are at risk for collapsing glomerulopathy (90).

COVID-19 can also cause neurological complications (91–93), potentially including stroke, seizures or meningitis (94, 95). One study on autopsy samples suggested that SARS-CoV-2 may be able to enter the central nervous system via the neural–mucosal interface (96). However, a study of 41 autopsied brains (97) found no evidence that the virus can actually infect the central nervous system. Although there was viral RNA in some brain samples, it was only found in very small amounts, and no viral protein was found. The RNA may have been in the blood vessels or blood components and not in the brain tissue itself. Instead, the neuropathological effects of COVID-19 are more likely to be caused indirectly by hypoxia, coagulopathy, or inflammatory processes rather than by infection in the brain (97). COVID-19 has been associated with an increased incidence of large vessel stroke, particularly in patients under the age of 40 (98), and other thrombotic events including pulmonary embolism and deep vein thrombosis (99). The mechanism behind these complications has been suggested to be related to coagulopathy, with reports indicating the presence of antiphospholipid antibodies (100) and elevated levels of d-dimer and fibrinogen degradation products in deceased patients (101). Other viral infections have been associated with coagulation defects and changes to the coagulation cascade; notably, SARS was also found to lead to disseminated intravascular coagulation and was associated with both pulmonary embolism and deep vein thrombosis (102). The mechanism behind these insults has been suggested to be related to inflammation-induced increases in the von Willebrand factor clotting protein, leading to a pro-coagulative state (102). Abnormal clotting (thromboinflammation or coagulopathy) has been increasingly discussed recently as a possible key mechanism in many cases of severe COVID-19, and may be associated with the high d-dimer levels often observed in severe cases (103–105). This excessive clotting in lung capillaries has been suggested to be related to a dysregulated activation of the complement system, part of the innate immune system (106, 107).

Finally, concerns have been raised about long-term sequelae of COVID-19. Some COVID-19 patients have reported that various somatic symptoms (such as shortness of breath, fatigue, chest pain) and psychological (depression, anxiety or mild cognitive impairment) symptoms can last for months after infection (108). Such long-term effects occur both in adults (109) and children (110). Sustained symptoms affecting a variety of biological systems have been reported across many studies (e.g., (108, 111, 112)). The phenomenon of “long COVID” is not fully understood although various possible explanations have been proposed, including damage caused by immune response to infection as well as by the infection itself, in addition to negative consequences of the experience of lengthy illness and hospitalization. However, a lack of consistency among definitions used in different studies makes it difficult to develop precise definitions or identify specific symptoms associated with long-term effects of COVID-19 (113, 114). Patient and family support groups for “long haulers” have been formed online, and patient-driven efforts to collect data about post-acute COVID-19 provide valuable

sources of information (e.g., [\(111\)](#)). The specific relationship between viral pathogenesis and these reported sequelae remains to be uncovered, however.

1.5.1 Pediatric Presentation

The presentation of COVID-19 infection can vary greatly among pediatric patients and, in some cases, manifests in distinct ways from COVID-19 in adults. Evidence suggests that children and adolescents tend to have mostly asymptomatic infections and that those who are symptomatic typically exhibit mild illness ([\(115–118\)](#)). One review examined symptoms reported in 17 studies of children infected with COVID-19 during the early months of the COVID-19 epidemic in China and one study from Singapore ([\(119\)](#)). In the more than a thousand cases described, the most common reports were for mild symptoms such as fever, dry cough, fatigue, nasal congestion and/or runny nose, while three children were reported to be asymptomatic. Severe lower respiratory infection was described in only one of the pediatric cases reviewed. Gastrointestinal symptoms such as vomiting or diarrhea were occasionally reported. Radiologic findings were not always reported in the case studies reviewed, but when they were mentioned they included bronchial thickening, ground-glass opacities, and/or inflammatory lesions ([\(119\)](#)). Neurological symptoms have also been reported ([\(120\)](#)).

These analyses indicate that most pediatric cases of COVID-19 are not severe. Indeed, it is estimated that less than 1% of pediatric cases result in critical illness ([\(117, 121\)](#)), although reporting suggests that pediatric hospitalizations may be greater with the emergence of the Delta variant of concern (VOC) ([\(122–124\)](#)). Serious complications and, in relatively rare cases, deaths have occurred ([\(125\)](#)). Of particular interest, children have occasionally experienced a serious inflammatory syndrome, multisystem inflammatory syndrome in children (MIS-C), following COVID-19 infection ([\(126\)](#)). This syndrome is similar in some respects to Kawasaki disease, including Kawasaki disease shock syndrome ([\(127–129\)](#)), and is thought to be a distinct clinical manifestation of SARS-CoV-2 due to its distinct cytokine profile and the presence of burr cells in peripheral blood smears ([\(130, 131\)](#)). MIS-C has been associated with heart failure in some cases ([\(132\)](#)). A small number of case studies have identified presentations similar to MIS-C in adults associated with SARS-CoV-2 ([\(133–136\)](#)). However, not all cases of severe COVID-19 in children are characterizable as MIS-C. A recent study ([\(137\)](#)) described demographic and clinical variables associated with MIS-C in comparison with non-MIS-C severe acute COVID-19 in young people in the United States. Efforts to characterize long-term sequelae of SARS-CoV-2 infection in children face the same challenges as in adults, but long-term effects remain a concern in pediatric patients ([\(110, 138, 139\)](#)), although some early studies have suggested that they may be less of a concern than in adults ([\(140–142\)](#)). Research is ongoing into the differences between the pediatric and adult immune responses to SARS-CoV-2, and future research may shed light on the factors that lead to MIS-C; it is also unknown whether the relative advantages of children against severe COVID-19 will remain in the face of current and future variants ([\(143\)](#)).

1.5.2 Cytokine Release Syndrome

The inflammatory response was identified early on as a potential driver of COVID-19 outcomes due to existing research in SARS and emerging research in COVID-19. While too low of an inflammatory response is a concern because it will fail to eliminate the immune threat (144), excessive pro-inflammatory cytokine activity can cascade (145) and cause cell damage, among other problems (146). A dysregulated immune response can cause significant damage to the host (147–149) including pathogenesis associated with sepsis. Sepsis, which can lead to multi-organ failure and death (150, 151), is traditionally associated with bacterial infections. However, sepsis associated with viral infections may be underidentified (152), and sepsis has emerged as a major concern associated with SARS-CoV-2 infection (153). Hyperactivity of the pro-inflammatory response due to lung infection is commonly associated with acute lung injury and more rarely with the more severe manifestation, ARDS, which can arise from pneumonia, SARS, and COVID-19 (145, 150). Damage to the capillary endothelium can cause leaks that disrupt the balance between pro-inflammatory cytokines and their regulators (154), and heightened inflammation in the lungs can also serve as a source for systemic inflammation, or sepsis, and potentially multi-organ failure (150). The shift from local to systemic inflammation is a phenomenon often referred to broadly as a cytokine storm (150) or, more precisely, as cytokine release syndrome (155).

Cytokine dysregulation is therefore a significant concern in the context of COVID-19. In addition to the known role of cytokines in ARDS and lung infection more broadly, immunohistological analysis at autopsy of deceased SARS patients revealed that ACE2-expressing cells that were infected by SARS-CoV-1 showed elevated expression of the cytokines IL-6, IL-1 β , and TNF- α (156). Similarly, the introduction of the S protein from SARS-CoV-1 to mouse macrophages was found to increase production of IL-6 and TNF- α (157). For SARS-CoV-2 infection leading to COVID-19, early reports described a cytokine storm syndrome-like response in patients with particularly severe infections (65, 158, 159). Sepsis has been identified as a major contributor to COVID-19-related death. Among patients hospitalized with COVID-19 in Wuhan, China, 112 out of 191 (59%) developed sepsis, including all 54 of the non-survivors (83).

While IL-6 is sometimes used as a biomarker for cytokine storm activity in sepsis (150), the relationship between cytokine profiles and the risks associated with sepsis may be more complex. One study of patients with and at risk for ARDS, specifically those who were intubated for medical ventilation, found that shortly after the onset of ARDS, anti-inflammatory cytokine concentration in BALF increased relative to the concentration of pro-inflammatory cytokines (154). The results suggest that an increase in pro-inflammatory cytokines such as IL-6 may signal the onset of ARDS, but recovery depends on an increased anti-inflammatory response (154). However, patients with severe ARDS were excluded from this study. Another analysis of over 1,400 pneumonia patients in the United States reported that IL-6, tumor necrosis factor (TNF), and IL-10 were elevated at intake in patients who developed severe sepsis and/or ultimately died (160). However, unlike the study analyzing pro- and anti-inflammatory cytokines in ARDS patients (154), this study reported that unbalanced pro-/anti-inflammatory cytokine profiles were rare. This discrepancy could be related to the fact that the sepsis study measured only three cytokines. Although IL-6 has traditionally

been considered pro-inflammatory, its pleiotropic effects via both classical and trans-signaling allow it to play an integral role in both the inflammatory and anti-inflammatory responses (161), leading it to be associated with both healthy and pathological responses to viral threat (162). While the cytokine levels observed in COVID-19 patients fall outside of the normal range, they are not as high as typically found in patients with ARDS (163). Regardless of variation in the anti-inflammatory response, prior work has therefore made it clear that pulmonary infection and injury are associated with systemic inflammation and with sepsis. Inflammation has received significant interest both in regards to the pathology of COVID-19 as well as potential avenues for treatment, as the relationship between the cytokine storm and the pathophysiology of COVID-19 has led to the suggestion that a number of immunomodulatory pharmaceutical interventions could hold therapeutic value for the treatment of COVID-19 (3, 164).

1.6 Insights from Systems Biology

Systems biology provides a cross-disciplinary analytical paradigm through which the host response to an infection can be analyzed. This field integrates the “omics” fields (genomics, transcriptomics, proteomics, metabolomics, etc.) using bioinformatics and other computational approaches. Over the last decade, systems biology approaches have been used widely to study the pathogenesis of diverse types of life-threatening acute and chronic infectious diseases (165). Omics-based studies have also provided meaningful information regarding host immune responses and surrogate protein markers in several viral, bacterial and protozoan infections (166). Though the complex pathogenesis and clinical manifestations of SARS-CoV-2 infection are not yet fully understood, omics technologies offer the opportunity for discovery-driven analysis of biological changes associated with SARS-CoV-2 infection.

1.6.1 Transcriptomics

Through transcriptomic analysis, the effect of a viral infection on gene expression can be assessed. Transcriptomic analyses, whether *in vivo* or *in situ*, can potentially reveal insights into viral pathogenesis by elucidating the host response to the virus. For example, infection by some viruses, including by the coronaviruses SARS-CoV-2, SARS-CoV-1, and MERS-CoV, is associated with the upregulation of ACE2 in human embryonic kidney cells and human airway epithelial cells (65). This finding suggests that SARS-CoV-2 facilitates the positive regulation of its own transmission between host cells (65). The host immune response also likely plays a key role in mediating infection-associated pathologies. Therefore, transcriptomics is one critical tool for characterizing the host response in order to gain insight into viral pathogenesis. For this reason, the application of omics technologies to the process of characterizing the host response is expected to provide novel insights into how hosts respond to SARS-CoV-2 infection and how these changes might influence COVID-19 outcomes.

Several studies have examined the cellular response to SARS-CoV-2 *in vitro* in comparison to other viruses. One study (167) compared the transcriptional responses of three human cell lines to SARS-CoV-2 and to other respiratory

viruses, including MERS-CoV, SARS-CoV-1, *Human parainfluenza virus 3*, *Respiratory syncytial virus*, and *Influenza A virus*. The transcriptional response differed between the SARS-CoV-1 infected cells and the cells infected by other viruses, with changes in differential expression specific to each infection type. Where SARS-CoV-2 was able to replicate efficiently, differential expression analysis revealed that the transcriptional response was significantly different from the response to all of the other viruses tested. A unique pro-inflammatory cytokine signature associated with SARS-CoV-2 was present in cells exposed to both high and low doses of the virus, with the cytokines IL-6 and IL1RA uniquely elevated in response to SARS-CoV-2 relative to other viruses. However, one cell line showed significant IFN-I or IFN-III expression when exposed to high, but not low, doses of SARS-CoV-2, suggesting that IFN induction is dependent on the extent of exposure. These results suggest that SARS-CoV-2 induces a limited antiviral state with low IFN-I or IFN-III expression and a moderate IFN-stimulated gene response, in contrast to other viruses. Other respiratory viruses have been found to encode antagonists to the IFN response ([168](#), [169](#)), including SARS-CoV-1 ([170](#)) and MERS-CoV ([171](#)).

The analysis of SARS-CoV-2 suggested that this transcriptional state was specific to cells expressing ACE2, as it was not observed in cells lacking expression of this protein except with ACE2 supplementation and at very high (10-fold increase) level of SARS-CoV-2 exposure ([167](#)). In another study, direct stimulation with inflammatory cytokines such as type I interferons (e.g., IFN β) was also associated with the upregulation of ACE2 in human bronchial epithelial cells, with treated groups showing four-fold higher ACE2 expression than control groups at 18 hours post-treatment ([172](#)). This hypothesis was further supported by studies showing that several nsp in SARS-CoV-2 suppress interferon activity ([173](#)) and that the SARS-CoV-2 *ORF3b* gene suppresses IFNB1 promoter activity (IFN-I induction) more efficiently than the SARS-CoV-1 *ORF3b* gene ([174](#)). Taken together, these findings suggest that a unique cytokine profile is associated with the response to the SARS-CoV-2 virus, and that this response differs depending on the magnitude of exposure.

Susceptibility and IFN induction may also vary by cell type. Using poly(A) bulk RNA-seq to analyze dynamic transcriptional responses to SARS-CoV-2 and SARS-CoV-1 revealed negligible susceptibility of cells from the H1299 line (< 0.08 viral read percentage of total reads) compared to those from the Caco-2 and Calu-3 lines (>10% of viral reads) ([175](#)). This finding suggests that the risk of infection varies among cell types, and that cell type could influence which hosts are more or less susceptible. Based on visual inspection of microscopy images alongside transcriptional profiling, the authors also showed distinct responses among the host cell lines evaluated ([175](#)). In contrast to Caco-2, Calu-3 cells infected with SARS-CoV-2 showed signs of impaired growth and cell death at 24 hours post infection, as well as moderate IFN induction with a strong up-regulation of IFN-stimulated genes. Interestingly, the results were similar to those reported in Calu-3 cells exposed to much higher levels of SARS-CoV-2 ([167](#)), as described above. This finding suggests that IFN induction in Calu-3 cells is not dependent on the level of exposure, in contrast to A549-ACE2 cells. The discrepancy could be explained by the observations that Calu-3 cells are highly susceptible to SARS-CoV-2 and show rapid viral replication ([34](#)), whereas A549 cells are incompatible with SARS-

CoV-2 infection ([176](#)). This discrepancy raises the concern that *in vitro* models may vary in their similarity to the human response, underscoring the importance of follow-up studies in additional models.

As a result, transcriptional analysis of patient tissue is an important application of omics technology to understanding COVID-19. Several studies have collected blood samples from COVID-19 patients and analyzed them using RNA-Seq ([177–182](#)). Analyzing gene expression in the blood is valuable to understanding host-pathogen interactions because of the potential to identify alterations associated with the immune response and to gain insights into inflammation, among other potential insights ([177](#)). One study compared gene expression in 39 COVID-19 inpatients admitted with community-acquired pneumonia to that of control donors using whole blood cell transcriptomes ([177](#)). They also evaluated the effect of mild versus severe disease. A greater number of differentially expressed genes were found in severe patients compared to controls than in mild patients compared to controls. They also identified that the transcriptional profiles clustered into five groups and that the groups could not be explained by disease severity. Most severe cases fell into two clusters associated with increased inflammation and granulocyte and neutrophil activation. The presence of these clusters suggests the possibility that personalized medicine could be useful in the treatment of COVID-19 ([177](#)). Longitudinal analysis of granulocytes from patients with mild versus severe COVID-19 revealed that granulocyte activation-associated factors differentiated the disease states, with greater numbers of differentially expressed genes early in disease course ([177](#)). This study therefore revealed distinct patterns associated with COVID-19 and identified genes and pathways associated with each cluster.

Many other studies have also identified transcriptomic signatures associated with the immune response and inflammation. Other studies have profiled the transcriptome of BALF ([179](#)) and the nasopharynx ([183](#)). One study used single-cell transcriptomics techniques to investigate cell types including brain and choroid plexus cells compared to healthy controls and controls with influenza; among other signals of neuroinflammation, this study reported cortical T cells only in COVID-19 patients ([184](#)). Transcriptomic analysis can thus provide insight into the pathogenesis of SARS-CoV-2 and may also be useful in identifying candidate therapeutics ([177](#)).

1.6.2 Proteomics

Proteomics analysis offers an opportunity to characterize the response to a pathogen at a level above transcriptomics. Especially early on, this primarily involved evaluating the effect of the virus on cell lines. One early proteomics study investigated changes associated with *in vitro* SARS-CoV-2 infection using Caco-2 cells ([185](#)). This study reported that SARS-CoV-2 induced alterations in multiple vital physiological pathways, including translation, splicing, carbon metabolism and nucleic acid metabolism in the host cells. Another area of interest is whether SARS-CoV-2 is likely to induce similar changes to other HCoV. For example, because of the high level of sequence homology between SARS-CoV-2 and SARS-CoV-1, it has been hypothesized that sera from convalescent SARS-CoV-1 patients might show some efficacy in cross-neutralizing SARS-CoV-2-S-driven entry ([33](#)). However, despite the high level of sequence homology, certain protein structures might be

immunologically distinct, which would be likely to prohibit effective cross-neutralization across different SARS species (186). Consequently, proteomic analyses of SARS-CoV-1 might also provide some essential information regarding the new pathogen (187, 188).

Proteomics research has been able to get ahead of the timeline for development of omics-level big data sets specific to SARS-CoV-2 by adopting a comparative bioinformatics approach. Data hubs such as UniProt (189), NCBI Genome Database (190), The Immune Epitope Database and Analysis Resource (191), and The Virus Pathogen Resource (192) contain a wealth of data from studies in other viruses and even HCoV. Such databases facilitate the systems-level reconstruction of protein-protein interaction networks, providing opportunities to generate hypotheses about the mechanism of action of SARS-CoV-2 and identify potential drug targets. In an initial study (193), 26 of the 29 SARS-CoV-2 proteins were cloned and expressed in HEK293T kidney cells, allowing for the identification of 332 high-confidence human proteins interacting with them. Notably, this study suggested that SARS-CoV-2 interacts with innate immunity pathways. Ranking pathogens by the similarity between their interactomes and that of SARS-CoV-2 suggested *West Nile virus*, *Mycobacterium tuberculosis*, and *human papillomavirus* infections as the top three hits. The fact that the host-pathogen interactome of the bacterium *Mycobacterium tuberculosis* was found to be similar to that of SARS-CoV-2 suggests that changes related to lung pathology might comprise a significant contributor to these expression profiles. Additionally, it was suggested that the envelope protein, E, could disrupt host bromodomain-containing proteins, i.e., BRD2 and BRD4, that bind to histones, and the spike protein could likely intervene in viral fusion by modulating the GOLGA7-ZDHHC5 acyl-transferase complex to increase palmitoylation, which is a post-translational modification that affects how proteins interact with membranes (194).

An example of an application of this *in silico* approach comes from another study (195), which used patient-derived peripheral blood mononuclear cells to identify 251 host proteins targeted by SARS-CoV-2. This study also reported that more than 200 host proteins were disrupted following infection. In particular, a network analysis showed that nsp9 and nsp10 interacted with NF-Kappa-B-Repressing Factor, which encodes a transcriptional repressor that mediates repression of genes responsive to Nuclear Factor kappa-light-chain-enhancer of activated B-cells. These genes are important to pro-, and potentially also anti-, inflammatory signaling (196). This finding could explain the exacerbation of the immune response that shapes the pathology and the high cytokine levels characteristic of COVID-19, possibly due to the chemotaxis of neutrophils mediated by IL-8 and IL-6. Finally, it was suggested (197) that the E protein of both SARS-CoV-1 and SARS-CoV-2 has a conserved Bcl-2 Homology 3-like motif, which could inhibit anti-apoptosis proteins, e.g., BCL2, and trigger the apoptosis of T cells. Several compounds are known to disrupt the host-pathogen protein interactome, largely through the inhibition of host proteins. Therefore, this research identifies candidate targets for intervention and suggests that drugs modulating protein-level interactions between virus and host could be relevant to treating COVID-19.

As with other approaches, analyzing the patterns found in infected versus healthy human subjects is also important. COVID-19 infection has been associated with quantitative changes in transcripts, proteins, metabolites, and lipids in patient blood samples ([198](#)). One longitudinal study ([199](#)) compared COVID-19 patients to symptomatic controls who were PCR-negative for SARS-CoV-2. The longitudinal nature of this study allowed it to account for differences in the scale of inter- versus intraindividual changes. At the time of first sampling, common functions of proteins upregulated in COVID-19 patients relative to controls were related to immune system mediation, coagulation, lipid homeostasis, and protease inhibition. They compared these data to the patient-specific timepoints associated with the highest levels of SARS-CoV-2 antibodies and found that the actin-binding protein gelsolin, which is involved in recovery from disease, showed the steepest decline between those two time points. Immunoglobulins comprised the only proteins that were significantly different between the COVID-19 and control patients at both of these timepoints. The most significantly downregulated proteins between these time points were related to inflammation, while the most significantly upregulated proteins were immunoglobulins. Proteins related to coagulation also increased between the two timepoints. The selection of a symptomatic control cohort rather than healthy comparisons also suggests that the results are more likely to highlight the response to SARS-CoV-2 and COVID-19 specifically, rather than to disease more broadly. This study also compared the disease course in patients who ultimately survived to those who died and found that ITIH4, a protein associated with the inflammatory response to trauma, may be a biomarker useful to identifying patients at risk of death. Thus, these results indicate the value of studying patients in a longitudinal manner over the disease course. By revealing which genes are perturbed during SARS-CoV-2 infection, proteomics-based analyses can thus provide novel insights into host-virus interaction and serve to generate new avenues of investigation for therapeutics.

1.7 Viral Virulence

Like that of SARS-CoV-1, the entry of SARS-CoV-2 into host cells is mediated by interactions between the viral spike glycoprotein, S, and human ACE2 (hACE2) ([25](#), [33](#), [200–205](#)). Differences in how the S proteins of the two viruses interact with hACE2 could partially account for the increased transmissibility of SARS-CoV-2. Studies have reported conflicting binding constants for the S-hACE2 interaction, though they have agreed that the SARS-CoV-2 S protein binds with equal, if not greater, affinity than the SARS-CoV-1 S protein does ([16](#), [25](#), [203](#)). The C-terminal domain of the SARS-CoV-2 S protein in particular was identified as the key region of the virus that interacts with hACE2, and the crystal structure of the C-terminal domain of the SARS-CoV-2 S protein in complex with hACE2 reveals stronger interaction and a higher affinity for receptor binding than that of SARS-CoV-1 ([204](#)). Among the 14 key binding residues identified in the SARS-CoV-1 S protein, eight are conserved in SARS-CoV-2, and the remaining six are semi-conservatively substituted, potentially explaining variation in binding affinity ([25](#), [203](#)). Studies of crystal structure have shown that the RBD of the SARS-CoV-2 S protein, like that of other coronaviruses, undergoes stochastic hinge-like movement that flips it from a “closed” conformation, in which key binding residues are hidden at the interface between protomers, to an “open” one

(16, 25). Spike proteins cleaved at the furin-like binding site are substantially more likely to take an open conformation (66%) than those that are uncleaved (17%) (206). Because the RBD plays such a critical role in viral entry, blocking its interaction with ACE2 could represent a promising therapeutic approach. Nevertheless, despite the high structural homology between the SARS-CoV-2 RBD and that of SARS-CoV-1, monoclonal antibodies targeting SARS-CoV-1 RBD failed to bind to SARS-CoV-2-RBD (16). However, in early research, sera from convalescent SARS patients were found to inhibit SARS-CoV-2 viral entry *in vitro*, albeit with lower efficiency than it inhibited SARS-CoV-1 (33).

Comparative genomic analysis reveals that several regions of the coronavirus genome are likely critical to virulence. The S1 domain of the spike protein, which contains the receptor binding motif, evolves more rapidly than the S2 domain (23, 24). However, even within the S1 domain, some regions are more conserved than others, with the receptors in S1's N-terminal domain (S1-NTD) evolving more rapidly than those in its C-terminal domain (S1-CTD) (24). Both S1-NTD and S1-CTD are involved in receptor binding and can function as RBDs to bind proteins and sugars (23), but RBDs in the S1-NTD typically bind to sugars, while those in the S1-CTD recognize protein receptors (12). Viral receptors show higher affinity with protein receptors than sugar receptors (12), which suggests that positive selection on or relaxed conservation of the S1-NTD might reduce the risk of a deleterious mutation that would prevent binding. The SARS-CoV-2 S protein also contains an RRAR furin recognition site at the S1/S2 junction (16, 25), setting it apart from both bat coronavirus RaTG13, with which it shares 96% genome sequence identity, and SARS-CoV-1 (207). Such furin cleavage sites are commonly found in highly virulent influenza viruses (208, 209). The furin recognition site at the S1/S2 junction is likely to increase pathogenicity via destabilization of the spike protein during fusion to ACE2 and the facilitation of cell-cell adhesion (16, 25, 42, 206, 208, 209). These factors may influence the virulence of SARS-CoV-2 relative to other beta coronaviruses. Additionally, a major concern has been the emergence of SARS-CoV-2 variants with increased virulence. The extent to which evolution within SARS-CoV-2 may affect pathogenesis is reviewed below.

1.8 Molecular Signatures, Transmission, and Variants of Concern

Genetic variation in SARS-CoV-2 has been used to elucidate patterns over time and space. Many mutations are neutral in their effect and can be used to trace transmission patterns. Such signatures within SARS-CoV-2 have provided insights during outbreak investigations (210–212). Similar mutations observed in several patients may indicate that the patients belong to the same transmission group. The tracking of SARS-CoV-2 mutations is recognized as an essential tool for controlling future outbreaks and tracing the path of the spread of SARS-CoV-2. In the first months of the pandemic in early 2020, early genomic surveillance efforts in Guangdong, China revealed that local transmission rates were low and that most cases arising in the province were imported (213). Since then, efforts have varied widely among countries: for example, the U.K. has coordinated a national database of viral genomes (214), but efforts to collect this type of data in the United States

have been more limited ([215](#)). Studies have applied phylogenetic analyses of viral genomes to determine the source of local COVID-19 outbreaks in Connecticut (USA), ([216](#)), the New York City area (USA) ([217](#)), and Iceland ([218](#)). There has been an ongoing effort to collect SARS-CoV-2 genomes throughout the COVID-19 outbreak, and as of summer 2021, millions of genome sequences have been collected from patients. The sequencing data can be found at GISAID ([219](#)), NCBI ([220](#)), and COVID-19 data portal ([221](#)).

Ongoing evolution can be observed in genomic data collected through molecular surveillance efforts. In some cases, mutations can produce functional changes that can impact pathogenesis. One early example is the spike protein mutation D614G, which appeared in March 2020 and became dominant worldwide by the end of May 2020 ([222](#), [223](#)). This variant was associated with increased infectivity and increased viral load, but not with more severe disease outcomes ([222](#), [224](#)). This increased virulence is likely achieved by altering the conformation of the S1 domain to facilitate binding to ACE2 ([224](#)). Similarly, the N439K mutation within the RBD of the spike protein is likely associated with increased transmissibility and enhanced binding affinity for hACE2, although it is also not thought to affect disease outcomes ([225](#)). In contrast, a mutation in ORF8 that was identified in Singapore in the early months of 2020 was associated with cases of COVID-19 that were less likely to require treatment with supplemental oxygen ([226](#)), and a deletion surrounding the furin site insertion at the S1/S2 boundary has been identified only rarely in clinical settings ([227](#)), suggesting that these mutations may disadvantage viral pathogenesis in human hosts. Thus, mutations have been associated with both virological and clinical differences in pathogenesis.

Several VOCs have also been identified and designated through molecular surveillance efforts ([228](#)). The Alpha variant (lineage B.1.1.7) was first observed in the U.K. in October 2020 before it quickly spread around the world ([229](#)). Other variants meriting further investigation have also been identified, including the Beta variant (B.1.351 lineage) first identified in South Africa and the Gamma variant (P.1 lineage) initially associated with outbreaks in Brazil. These lineages share independently acquired mutations that may affect pathogenicity ([230–234](#)). For example, they are all associated with a greater binding affinity for hACE2 than that of the wildtype variant ([232](#), [235](#), [236](#)), but they were not found to have more efficient cell entry than the wildtype virus ([237](#)). A fourth VOC, the Delta variant (B.1.617.2 and AY.1, AY.2, and AY.3 lineages), was identified in India in late 2020 ([238](#)). Some of the mutations associated with this lineage may alter fusogenicity and enhance furin cleavage, among other effects associated with increased pathogenicity ([239](#)). The changes in these VOC demonstrate how ongoing evolution in SARS-CoV-2 can drive changes in how the virus interacts with host cells.

1.9 Quantifying Viral Presence

Assessing whether a virus is present in a sample is a more complex task than it initially seems. Many diagnostic tests rely on real-time polymerase chain reaction (RT-PCR) to test for the presence versus absence of a virus ([7](#)). They may report the cycle threshold (C_t) indicating the number of doubling cycles required for the target (in this case, SARS-CoV-2) to become detectable. A

lower C_t therefore corresponds to a higher viral load. The C_t that corresponds to a positive can vary widely, but is often around 35. This information is sufficient to answer many questions, since an amplicon must be present in order to be duplicated in RT-PCR. For example, if a patient is presenting with COVID-19 symptoms, a positive RT-PCR test can confirm the diagnosis.

However, RT-PCR analysis alone cannot provide the information needed to determine whether a virus is present at sufficient levels to be infectious (240). Some studies have therefore taken the additional step of cultivating samples *in vitro* in order to observe whether cells become infected with SARS-CoV-2. One study collected upper respiratory tract samples from COVID-19 patients, analyzed them with RT-PCR to determine the cycle threshold, and then attempted to cultivate the SARS-CoV-2 virus in VeroE6 cells (240). This study found that out of 246 samples, less than half (103) produced a positive culture. Moreover, at a C_t of 35, only 5 out of 60 samples grew *in vitro*. Therefore, the RT-PCR-confirmed presence of SARS-CoV-2 in a sample does not necessarily indicate that the virus is present at a high enough concentration to grow and/or spread.

1.10 Mechanisms of Transmission

When a human host is infected with a virus and is contagious, person-to-person viral transmission can occur through several possible mechanisms. When a contagious individual sneezes, coughs, or exhales, they produce respiratory droplets that can contain a large number of viral particles (241). Viral particles can enter the body of a new host when they then come in contact with the oral, nasal, eye, or other mucus membranes (241). The primary terms typically used to discuss the transmission of viruses via respiratory droplets are droplet, aerosol, and contact transmission (242). The distinction between droplet and aerosol transmission is typically anchored on whether a particle containing the virus is larger or smaller than 5 micrometers (μm) (243, 244). Droplet transmission typically refers to contact with large droplets that fall quickly to the ground at close range, such as breathing in droplets produced by a sneeze (241, 243). Aerosol transmission typically refers to much smaller particles (less than 5 μm) produced by sneezing, coughing, or exhaling (241, 242) that can remain suspended over a longer period of time and potentially to be moved by air currents (241). It is also possible that viral particles deposited on surfaces via large respiratory droplets could later be aerosolized (241). The transmission of viral particles that have settled on a surface is typically referred to as contact or fomite transmission (241, 245). Any respiratory droplets that settle on a surface could contribute to fomite transmission (241). Droplet and contact transmission are both well-accepted modes of transmission for many viruses associated with common human illnesses, including influenza and rhinovirus (241).

The extent to which aerosol transmission contributes to the spread of respiratory viruses is more widely debated. In influenza A, for example, viral particles can be detected in aerosols produced by infected individuals, but it is not clear to what extent these particles drive the spread of influenza A infection (241, 242, 246–248). Regardless of its role in the spread of influenza

A, however, aerosol transmission likely played a role in outbreaks such as the 1918 Spanish Influenza (H1N1) and 2009 “swine flu” (pH1N1) (248). All three of these mechanisms have been identified as possible contributors to the transmission of HCoVs (241), including the highly pathogenic coronaviruses SARS-CoV-1 and MERS-CoV (249, 250). Transmission of SARS-CoV-1 is thought to proceed primarily through droplet transmission, but aerosol transmission is also considered possible (241, 251, 252), and fomite transmission may have also played an important role in some outbreaks (253). Similarly, the primary mechanism of MERS transmission is thought to be droplets because inter-individual transmission appears to be associated with close interpersonal contact (e.g., household or healthcare settings), but aerosolized particles of the MERS virus have been reported to persist much more robustly than influenza A under a range of environmental conditions (254, 255). However, few of these analyses have sought to grow positive samples in culture and thus to confirm their potential to infect new hosts.

Contact, droplet, and aerosol transmission are therefore all worth evaluating when considering possible modes of transmission for a respiratory virus like SARS-CoV-2. The stability of the SARS-CoV-2 virus both in aerosols and on a variety of surfaces was found to be similar to that of SARS-CoV-1 (256). Droplet-based and contact transmission were initially put forward as the greatest concern for the spread of SARS-CoV-2 (257), with droplet transmission considered the dominant mechanism driving the spread of the virus (258) because the risk of fomite transmission under real-world conditions is likely to be substantially lower than the conditions used for experimental analyses (259). The COVID-19 pandemic has, however, exposed significant discrepancies in how terms pertaining to airborne viral particles are interpreted in different contexts (243). The 5- μ m distinction between “droplets” and “aerosols” is typical in the biological literature but is likely an artifact of historical science rather than a meaningful boundary in biology or physics (244). Additionally, various ambient conditions such as air flow can influence how particles of different sizes fall or spread (243). Despite initial skepticism about airborne transmission of SARS-CoV-2 through small particles (244), evidence now suggests that small particles can contribute to SARS-CoV-2 transmission (256, 260–262). For example, one early study detected SARS-CoV-2 viral particles in air samples taken from hospitals treating COVID-19 patients, although the infectivity of these samples was not assessed (263). Subsequently, other studies have been successful in growing SARS-CoV-2 in culture with samples taken from the air (264, 265) while others have not (266, 267) (see (268) for a systematic review of available findings as of July 2020). The fact that viable SARS-CoV-2 may exist in aerosolized particles calls into question whether some axioms of COVID-19 prevention, such as 2-meter social distancing, are sufficient (244, 264, 269).

1.10.1 Symptoms and Viral Spread

Other aspects of pathogenesis are also important to understanding how the virus spreads, especially the relationship between symptoms, viral shedding, and contagiousness. Symptoms associated with reported cases of COVID-19 range from mild to severe (8), but some individuals who contract COVID-19 remain asymptomatic throughout the duration of the illness (270). The incubation period, or the time period between exposure and the onset of symptoms, has been estimated at five to eight days, with means of 4.91 (95%

confidence interval (CI) 4.35-5.69) and 7.54 (95% CI 6.76-8.56) reported in two different Asian cities and a median of 5 (IQR 1 to 6) reported in a small number of patients in a Beijing hospital ([271](#), [272](#)).

However, the exact relationship between contagiousness and viral shedding remains unclear. Estimates suggest that viral shedding can, in some cases, begin as early as 12.3 days (95% CI 5.9-17.0) before the onset of symptoms, although this was found to be very rare, with less than 0.1% of transmission events occurring 7 or more days before symptom onset ([273](#)).

Transmissibility appeared to peak around the onset of symptoms (95% CI -0.9 - 0.9 days), and only 44% (95% CI 30–57%) of transmission events were estimated to occur from presymptomatic contacts ([273](#)). A peak in viral load corresponding to the onset of symptoms was also confirmed by another study ([240](#)). As these trends became apparent, concerns arose due to the potential for individuals who did not yet show symptoms to transmit the virus ([274](#)). Recovered individuals may also be able to transmit the virus after their symptoms cease. A study of the communicable period based on twenty-four individuals who tested positive for SARS-CoV-2 prior to or without developing symptoms estimated that individuals may be contagious for one to twenty-one days, but they note that this estimate may be low ([270](#)). In an early study, viral nucleic acids were reported to remain at observable levels in the respiratory specimens of recovering hospitalized COVID-19 patients for a median of 20 days and with a maximum observed duration through 37 days, when data collection for the study ceased ([83](#)).

As more estimates of the duration of viral shedding were released, they converged around approximately three weeks from first positive PCR test and/or onset of symptoms (which, if present, are usually identified within three days of the initial PCR test). For example, in some studies, viral shedding was reported for up to 28 days following symptom onset ([275](#)) and for one to 24 days from first positive PCR test, with a median of 12 days ([72](#)). On the other hand, almost 70% of patients were reported to still have symptoms at the time that viral shedding ceased, although all symptoms reduced in prevalence between onset and cessation of viral shedding ([276](#)). The median time that elapsed between the onset of symptoms and cessation of viral RNA shedding was 23 days and between first positive PCR test and cessation of viral shedding was 17 days ([276](#)). The fact that this study reported symptom onset to predate the first positive PCR test by an average of three days, however, suggests that there may be some methodological differences between it and related studies. Furthermore, an analysis of residents of a nursing home with a known SARS-CoV-2 case measured similar viral load in residents who were asymptomatic regardless of whether they later developed symptoms, and the load in the asymptomatic residents was comparable to that of residents who displayed either typical or atypical symptoms ([277](#)). Taken together, these results suggest that the presence or absence of symptoms are not reliable predictors of viral shedding or of SARS-CoV-2 status (e.g., ([278](#))). However, it should be noted that viral shedding is not necessarily a robust indicator of contagiousness. The risk of spreading the infection was low after ten days from the onset of symptoms, as viral load in sputum was found to be unlikely to pose a significant risk based on efforts to culture samples *in vitro* ([275](#)). The relationship between symptoms, detectable levels of the virus, and risk of viral spread is therefore complex.

The extent to which asymptomatic or presymptomatic individuals are able to transmit SARS-CoV-2 has been a question of high scientific and community interest. Early reports (February and March 2020) described transmission from presymptomatic SARS-CoV-2-positive individuals to close family contacts ([279](#), [280](#)). One of these reports ([280](#)) also included a description of an individual who tested positive for SARS-CoV-2 but never developed symptoms. Later analyses also sought to estimate the proportion of infections that could be traced back to a presymptomatic or asymptomatic individual (e.g., ([281](#))). Estimates of the proportion of individuals with asymptomatic infections have varied widely. The proportion of asymptomatic individuals on board the Diamond Princess cruise ship, which was the site of an early COVID-19 outbreak, was estimated at 17.9% ([282](#)). In contrast, a model using the prevalence of antibodies among residents of Wuhan, China estimated a much higher rate of asymptomatic cases, at approximately 7 in 8, or 87.5% ([283](#)). An analysis of the populations of care homes in London found that, among the residents (median age 85), the rate of asymptomatic infection was 43.8%, and among the caretakers (median age 47), the rate was 49.1% ([284](#)). The duration of viral shedding may also be longer in individuals with asymptomatic cases of COVID-19 compared to those who do show symptoms ([285](#)). As a result, the potential for individuals who do not know they have COVID-19 to spread the virus raises significant concerns. In Singapore and Tianjin, two cities studied to estimate incubation period, an estimated 40-50% and 60-80% of cases, respectively, were considered to be caused by contact with asymptomatic individuals ([271](#)). An analysis of viral spread in the Italian town of Vo', which was the site of an early COVID-19 outbreak, revealed that 42.5% of cases were asymptomatic and that the rate was similar across age groups ([286](#)). The argument was thus made that the town's lockdown was imperative for controlling the spread of COVID-19 because it isolated asymptomatic individuals. While more models are likely to emerge to better explore the effect of asymptomatic individuals on SARS-CoV-2 transmission, these results suggest that strategies for identifying and containing asymptomatic but contagious individuals are important for managing community spread.

1.10.2 Estimating the Fatality Rate

Estimating the occurrence of asymptomatic and mild COVID-19 cases is important to identifying the mortality rate associated with COVID-19. The mortality rate of greatest interest would be the total number of fatalities as a fraction of the total number of people infected. One commonly reported metric is the case fatality rate (CFR), which compares the number of COVID-19 related deaths to the number of confirmed or suspected cases. However, in locations without universal testing protocols, it is impossible to identify all infected individuals because so many asymptomatic or mild cases go undetected. Therefore, a more informative metric is the infection fatality rate (IFR), which compares the known deaths to the estimated number of cases. It thus requires the same numerator as CFR, but divides by an approximation of the total number of cases rather than only the observed/suspected cases. IFR varies regionally, with some locations observed to have IFRs as low as 0.17% while others are as high as 1.7% ([287](#)). Estimates of CFR at the national and continental level and IFR at the continent level is maintained by the Centre for Evidence-Based Medicine ([288](#)). Several meta-analyses have also sought to estimate IFR at the global scale. These estimates have varied; one

peer-reviewed study aggregated data from 24 other studies and estimated IFR at 0.68% (95% CI 0.53%–0.82%), but a preprint that aggregated data from 139 countries calculated a global IFR of 1.04% (95% CI 0.77%–1.38%) when false negatives were considered in the model ([287](#), [289](#)). A similar prevalence estimate was identified through a repeated cross-sectional serosurvey conducted in New York City that estimated the IFR as 0.97% ([290](#)). Examination of serosurvey-based estimates of IFR identified convergence on a global IFR estimate of 0.60% (95% CI 0.42%–0.77%) ([287](#)). All of these studies note that IFR varies widely by location, and it is also expected to vary with demographic and health-related variables such as age, sex, prevalence of comorbidities, and access to healthcare and testing ([291](#)). Estimates of infection rates are becoming more feasible as more data becomes available for modeling and will be bolstered as serological testing becomes more common and more widely available. However, this research may be complicated due to the emergence of variants over time, as well as the varying availability and acceptance of vaccines in different communities and locations.

1.11 Dynamics of Transmission

Disease spread dynamics can be estimated using R_0 , the basic reproduction number, and R_t , the effective reproduction number. Accurate estimates of both are crucial to understanding the dynamics of infection and to predicting the effects of different interventions. R_0 is the average number of new (secondary) infections caused by one infected person, assuming a wholly susceptible population ([292](#)), and is one of the most important epidemiological parameters ([293](#)). A simple mechanistic model used to describe infectious disease dynamics is a susceptible-infected-recovered compartmental model ([294](#), [295](#)). In this model, individuals move through three states: susceptible, infected, and recovered; two parameters, γ and β , specify the rate at which the infectious recover, and the infection transmission rate, respectively, and R_0 is estimated as the ratio of β and γ ([293](#), [296](#)). A pathogen can invade a susceptible population only if $R_0 > 1$ ([293](#), [297](#)). The spread of an infectious disease at a particular time t can be quantified by R_t , the effective reproduction number, which assumes that part of the population has already recovered (and thus gained immunity to reinfection) or that mitigating interventions have been put into place. For example, if only a fraction S_t of the population is still susceptible, $R_t = S_t \times R_0$. When R_t is greater than 1, an epidemic grows (i.e., the proportion of the population that is infectious increases); when R_t is less than 1, the proportion of the population that is infectious decreases. R_0 and R_t can be estimated directly from epidemiological data or inferred using susceptible-infected-recovered-type models. To capture the dynamics of SARS-CoV-2 accurately, the addition of a fourth compartment, i.e. a susceptible-exposed-infectious-recovered model, may be appropriate because such models account for the relative lengths of incubation and infectious periods ([298](#)).

Original estimates of R_0 for COVID-19 lie in the range $R_0=1.4$ – 6.5 ([299](#)–[301](#)). Variation in R_0 is expected between different populations, and the estimated values of R_0 discussed below are for specific populations in specific environments. The different estimates of R_0 should not necessarily be interpreted as a range of estimates of the same underlying parameter. In one

study of international cases, the predicted value was $R_0=1.7$ (302). In China (both Hubei province and nationwide), the value was predicted to lie in the range $R_0=2.0-3.6$ (299, 303, 304). Another estimate based on a cruise ship where an outbreak occurred predicted $R_0=2.28$ (305). Susceptible-exposed-infectious-recovered model-derived estimates of R_0 range from 2.0 - 6.5 in China (306–309) to $R_0=4.8$ in France (310). Using the same model as for the French population, a study estimated $R_0=2.6$ in South Korea (310), which is consistent with other studies (311). From a meta-analysis of studies estimating R_0 , (300) the median R_0 was estimated to be 2.79 (IQR 1.16) based on twelve studies published between January 1 and February 7, 2020.

Inference of the effective reproduction number can provide insight into how populations respond to an infection and the effectiveness of interventions. In China, R_t was predicted to lie in the range 1.6-2.6 in January 2020, before travel restrictions (312). R_t decreased from 2.35 one week before travel restrictions were imposed (January 23, 2020), to 1.05 one week after. Using their model, the authors also estimated the probability of new outbreaks occurring. Assuming individual-level variation in transmission comparable to that of MERS or SARS, the probability of a single individual exporting the virus and causing a large outbreak is 17-25%, and assuming variation like that of SARS and transmission patterns like those observed for COVID-19 in Wuhan, the probability of a large outbreak occurring after ≥ 4 infections exist at a new location is greater than 50%. An independent study came to similar conclusions, finding $R_t=2.38$ in the two-week period before January 23 with a decrease to $R_t = 1.34$ (using data from January 24 to February 3) or $R_t=0.98$ (using data from January 24 to February 8) (301). In South Korea, R_t was inferred for February through March 2020 in two cities, Daegu (the center of the outbreak) and Seoul (311). Metro data was also analyzed to estimate the effects of social distancing measures. R_t decreased in Daegu from around 3 to <1 over the period that social distancing measures were introduced. In Seoul, R_t decreased slightly, but remained close to 1 (and larger than R_t in Daegu). These findings indicate that social distancing measures appeared to be effective in containing the infection in Daegu, but in Seoul, R_t remained above 1, meaning secondary outbreaks remained possible. The study also shows the importance of region-specific analysis: the large decline in case load nationwide was mainly due to the Daegu region and could mask persistence of the epidemic in other regions, such as Seoul and Gyeonggi-do. In Iran, estimates of R_t declined from 4.86 in the first week to 2.1 by the fourth week after the first cases were reported (313). In Europe, analysis of 11 countries inferred the dynamics of R_t over a time range from the beginning of the outbreak until March 28, 2020, by which point most countries had implemented major interventions (such as stay-at-home orders, public gathering bans, and school closures) (314). Across all countries, the mean R_t before interventions began was estimated as 3.87; R_t varied considerably, from below 3 in Norway to above 4.5 in Spain. After interventions, R_t decreased by an average of 64% across all countries, with mean $R_t=1.43$. The lowest predicted value was 0.97 for Norway and the highest was 2.64 for Sweden, which could be related to the fact that Sweden did not implement social distancing measures on the same scale as other countries. The study concludes that while large changes in R_t are observed, it is too early to tell whether the interventions put into place are sufficient to decrease R_t below 1.

Evolution within SARS-CoV-2 has also driven changes in the estimated reproduction number for different populations at different times. As of June 2021, the reproduction number had increased globally relative to 2020, and increased transmissibility over the wildtype variant was observed for the Alpha, Beta, Gamma, and Delta VOC (315). In the U.S. between December 2020 and January 2021, B.1.1.7 (Alpha) was estimated to have an increased transmission of 35 to 45% relative to common SARS-CoV-2 variants at the time, with B.1.1.7 the dominant SARS-CoV-2 variant in some places at some timepoints (316). This lineage was estimated to have increased transmissibility of 43 to 90% in the U.K. (317). An estimate of the reproduction number of B.1.1.7 in the U.K. from September to December 2020 yielded 1.59 overall and between 1.56 and 1.95 in different regions of the country (234). The Delta variant is particularly transmissible, and it has been estimated to be twice as transmissible than the wildtype variant of SARS-CoV-2 (315). A review of the literature describing the Delta variant identified a mean estimated R_0 of 5.08 (318). Such differences can affect fitness and therefore influence the relative contributions of different lineages to a given viral gene pool over time (319). Therefore, the evolution of the virus can result in shifts in the reproduction rate.

More generally, population-level epidemic dynamics can be both observed and modeled (296). Data and empirically determined biological mechanisms inform models, while models can be used to try to understand data and systems of interest or to make predictions about possible future dynamics, such as the estimation of capacity needs (320) or the comparison of predicted outcomes among prevention and control strategies (321, 322). Many current efforts to model R_t have also led to tools that assist the visualization of estimates in real time or over recent intervals (323, 324). These are valuable resources, yet it is also important to note that the estimates arise from models containing many assumptions and are dependent on the quality of the data they use, which varies widely by region.

1.12 Effect of Vaccines on Pathogenesis and Community Spread

The vaccine clinical trial data demonstrate a significant reduction in the likelihood of contracting symptomatic COVID-19, thereby succeeding in the primary goal of vaccination. The mRNA vaccines in particular were initially so effective in preventing disease that they were also assumed to have an effect on the likelihood of transmission (e.g., venues requiring proof of vaccination). However, in light of the reduced efficacy in response to VOC, it is especially important to consider whether this assumption is supported by the available evidence.

This question is made up of several components. The crux is whether vaccinated individuals with a SARS-CoV-2 infection, regardless of symptom status, are as contagious as unvaccinated, infected individuals. Additionally, as outlined above, an important qualification is that the variants of SARS-CoV-2 circulating at the time of each study must be considered in light of the effect of evolution on vaccine efficacy.

The phase II/III clinical trials evaluating the mRNA vaccines assessed vaccine efficacy based on COVID-19 diagnosis, thereby detecting only patients who received a diagnosis. In order to identify patients infected with SARS-CoV-2 who did not receive a diagnosis, for example, potentially those who did not develop symptoms, it would be necessary to conduct routine PCR testing even in the absence of symptoms. Prior to the development of vaccines, the evidence suggested that asymptomatic individuals could spread SARS-CoV-2. Investigation of viral dynamics of asymptomatic infection in early 2020 indicated that asymptomatic patients continued to shed the virus for a duration similar to that of symptomatic patients ([325](#)) (although viral shedding should not be conflated with contagiousness without further investigation). Another study found viral load to be higher in the nasopharyngeal/oropharyngeal samples of asymptomatic patients compared to symptomatic patients hospitalized due to symptoms and/or known exposure ([326](#)). However, the sample size in both of these studies was small, and a larger study found higher viral load in symptomatic than asymptomatic cases ([327](#)) along with a systematic review finding a reduced probability of asymptomatic transmission ([328](#)). While far from conclusive, these studies suggest that asymptomatic cases still carry a risk of transmitting SARS-CoV-2.

One important consideration is therefore how likely vaccinated individuals are to develop asymptomatic SARS-CoV-2. Considering asymptomatic cases is necessary to establish a more complete picture of efficacy with respect to spread. Routine testing of healthcare workers in California who had received an mRNA vaccine revealed slightly higher rates of absolute risk for testing positive than those identified in the phase II/III trials, although the extent to which asymptomatic infection influenced these numbers was not investigated ([329](#)). Another study analyzed the results of COVID-19 screening tests administered to asymptomatic individuals prior to receiving certain medical services at the Mayo Clinic in several locations across the United States. This study found patients who had received two doses of an mRNA vaccine to be 73% less likely to have asymptomatic COVID-19 than patients who had received zero doses ([330](#)). Because this study began on December 17, 2020, a date selected to coincide with the first day vaccines were available at the Mayo Clinic, this number may underestimate the efficacy of the vaccines given that many people eligible for early vaccination were at increased risk for exposure (e.g., healthcare workers and residents of long-term care facilities) ([330](#)). In Israel, a longitudinal study of nearly 12,000 healthcare workers found that of the 5,372 fully vaccinated people with Pfizer/BioNTech BNT162b2, 8 developed symptomatic COVID-19 (0.15%) and 19 developed asymptomatic COVID-19 (0.35%) ([331](#)). While the study itself analyzed the efficacy of the vaccine based on person-days, these findings also suggest that many or even the majority of SARS-CoV-2 infections in vaccinated individuals are likely to be asymptomatic. Therefore, in addition to the symptomatic cases reported by the vaccine clinical trials, these findings suggest that asymptomatic cases can also occur in vaccinated people. In the absence of symptoms, individuals are less likely to know to self-isolate, and therefore evaluating the effect of the vaccine on viral load is critical to understanding the role vaccinated individuals can play in spreading SARS-CoV-2.

Another question of interest is therefore whether vaccinated individuals positive for SARS-CoV-2 carry a similar viral load to unvaccinated individuals. Viral load is often approximated by cycle threshold (C_t), or the cycle at which viral presence is detected during RT-qPCR, with a lower C_t corresponding to a greater viral load. A prospective cohort study that evaluated front-line workers in six U.S. states from December 2020 to April 2021 reported a 40% reduction in viral load even with just a single dose of an mRNA vaccine (332). The vaccine also appeared to influence the time to viral clearance: the risk of having detectable levels of SARS-CoV-2 for more than one week was reduced by 66% in participants who had received at least one dose (332). However, this study compared the mean viral load across the two groups, meaning that these findings cannot be extrapolated across all points in the disease course. Similarly, between December 2020 and February 2021, positive RT-qPCR tests were analyzed for almost 5,000 Israeli patients (333). C_t was analyzed relative to when each patient received the first dose of the Pfizer mRNA vaccine. A sharp increase in C_t (corresponding to reduced viral load) was observed between days 11 and 12, consistent with what is known about the onset of immunity following vaccination. This pattern therefore suggested a direct effect of vaccination on viral load.

Other studies, however, have not offered support for a reduced viral load in breakthrough cases. In Singapore, which has strict protocols for screening individuals with potential COVID-19 exposure, a retrospective cohort of patients who tested positive for SARS-CoV-2 between April and June 2021 was analyzed to compare viral kinetics and symptom course between vaccinated and unvaccinated cases. Vaccinated individuals who tested positive experienced fewer symptoms than unvaccinated, SARS-CoV-2-positive individuals and were more likely to be asymptomatic (334). Additionally, this study analyzed C_t over time and found that, though the median values were similar between the two groups at disease onset, viral load appeared to decrease more rapidly in vaccinated cases (334). This study is likely to have evaluated a more accurate representation of all COVID-19 outcomes than has been feasible in most studies, but one limitation was that the RT-PCR reactions were conducted in many different facilities. A third study investigated viral load (as approximated by C_t) using samples processed in a single laboratory during the summer of 2021 (335). This study identified no significant differences in C_t between fully vaccinated and unvaccinated cases, but this study used samples sent for diagnosis and was not longitudinal. It offered the additional benefit of culturing samples to assess whether their C_t threshold was likely to represent contagiousness and found that SARS-CoV-2 could be cultured from 51 of 55 samples with C_t less than 25 (the cut-off used in many studies). Another study of samples collected at two sites in San Francisco, one of which tested only asymptomatic individuals, reported no difference in C_t between asymptomatic and symptomatic cases regardless of whether vaccination status was included in the model (336). Though each of these three studies offers distinct strengths and weaknesses, taken together, they suggest that viral load is likely to be similar in vaccinated and unvaccinated individuals, but that vaccinated individuals clear the virus more rapidly, meaning that the average viral load is lower over time.

Given the emergence of VOC, especially the Delta and Omicron variants, for which breakthrough infections are more common, the potential for vaccinated individuals to spread SARS-CoV-2 is not static over time. In fact, studies reporting reduced viral load in vaccinated individuals collected samples, for the most part, prior to the emergence of the Delta variant's dominance. The emergence of this variant may partially account for why more recent studies tend to find no difference between viral load in vaccinated and unvaccinated cases.

Taken together, these findings can provide some insight into how vaccines influence community spread. While vaccinated individuals may be more likely to experience asymptomatic infection, current evidence about viral load in asymptomatic versus symptomatic cases is ambiguous. Similarly, no conclusions can be drawn about whether viral load is different in vaccinated versus unvaccinated cases. Therefore, at present, the evidence suggests that vaccinated individuals who are infected can still contribute to community spread. The one potential mitigating factor supported at present is that differences in the viral kinetics may result in vaccinated cases infecting fewer individuals over time due to a more rapid decrease in viral load ([334](#)), although this study did not examine patterns in secondary transmission. Thus, the virological evidence suggests that public health measures such as masking and distancing remain important even in areas with high vaccination rates.

1.13 Conclusions

The novel coronavirus SARS-CoV-2 is the third HCoV to emerge in the 21st century, and research into previous HCoVs has provided a strong foundation for characterizing the pathogenesis and transmission of SARS-CoV-2. Critical insights into how the virus interacts with human cells have been gained from previous research into HCoVs and other viral infections. With the emergence of three devastating HCoV over the past twenty years, emergent viruses are likely to represent an ongoing threat. Contextualizing SARS-CoV-2 alongside other viruses serves not only to provide insights that can be immediately useful for combating this virus itself but may also prove valuable in the face of future viral threats.

Host-pathogen interactions provide a basis not only for understanding COVID-19, but also for developing a response. As with other HCoVs, the immune response to SARS-CoV-2 is likely driven by detection of its spike protein, which allows it to enter cells through ACE2. Epithelial cells have also emerged as the major cellular target of the virus, contextualizing the respiratory and gastrointestinal symptoms that are frequently observed in COVID-19. Many of the mechanisms that facilitate the pathogenesis of SARS-CoV-2 are currently under consideration as possible targets for the treatment or prevention of COVID-19 ([2](#), [3](#)). Research in other viruses also provides a foundation for understanding the transmission of SARS-CoV-2 among people and can therefore inform efforts to control the virus's spread. Airborne forms of transmission (droplet and aerosol transmission) have emerged as the primary modes by which the virus spreads to new hosts. Asymptomatic transmission was also a concern in the SARS outbreak of 2002-03 and, as in

the current pandemic, presented challenges for estimating rates of infection (337). These insights are important for developing a public health response, such as the CDC's shift in its recommendations surrounding masking (338).

Even with the background obtained from research in SARS and MERS, COVID-19 has revealed itself to be a complex and difficult-to-characterize disease that has many possible presentations that vary with age. Variability in presentation, including cases with no respiratory symptoms or with no symptoms altogether, were also reported during the SARS epidemic at the beginning of the 21st century (337). The variability of both which symptoms present and their severity have presented challenges for public health agencies seeking to provide clear recommendations regarding which symptoms indicate SARS-CoV-2 infection and should prompt isolation. Asymptomatic cases add complexity both to efforts to estimate statistics such as R_0 and R_t , which are critical to understanding the transmission of the virus, and IFR, which is an important component of understanding its impact on a given population. The development of diagnostic technologies over the course of the pandemic has facilitated more accurate identification, including of asymptomatic cases (7). As more cases have been diagnosed, the health conditions and patient characteristics associated with more severe infection have also become more clear, although there are likely to be significant sociocultural elements that also influence these outcomes (339). While many efforts have focused on adults, and especially older adults because of the susceptibility of this demographic, additional research is needed to understand the presentation of COVID-19 and MIS-C in pediatric patients. As more information is uncovered about the pathogenesis of HCoV and SARS-CoV-2 specifically, the diverse symptomatology of COVID-19 has and likely will continue to conform with the ever-broadening understanding of how SARS-CoV-2 functions within a human host.

While the SARS-CoV-2 virus is very similar to other HCoV in several ways, including in its genomic structure and the structure of the virus itself, there are also some differences that may account for differences in the COVID-19 pandemic compared to the SARS and MERS epidemics of the past two decades. The R_0 of SARS-CoV-2 has been estimated to be similar to SARS-CoV-1 but much higher than that of MERS-CoV (340), although a higher R_0 has been estimated for some VOC. While the structures of the viruses are very similar, evolution among these species may account for differences in their transmissibility and virulence. For example, the acquisition of a furin cleavage site the S1/S2 boundary within the SARS-CoV-2 S protein may be associated with increased virulence. Additionally, concerns have been raised about the accumulation of mutations within the SARS-CoV-2 species itself, and whether these could influence virulence (341). These novel variants may be resistant to vaccines and antibody treatments such as Bamlanivimab that were designed based on the wildtype spike protein (3, 6). As a consequence of reliance on targeting the SARS-CoV-2 spike protein for many therapeutic and prophylactic strategies, increased surveillance is required to rapidly identify and prevent the spread of novel SARS-CoV-2 variants with alterations to the spike protein. The coming of age of genomic technologies has made these types of analyses feasible, and genomics research characterizing changes in SARS-CoV-2 along with temporal and spatial movement is likely to provide additional insights into whether within-species evolution influences the effect of the virus on the human host. Additionally, the rapid

development of sequencing technologies over the past decade has made it possible to rapidly characterize the host response to the virus. For example, proteomics analysis of patient-derived cells revealed candidate genes whose regulation is altered by SARS-CoV-2 infection, suggesting possible approaches for pharmaceutical invention and providing insight into which systems are likely to be disrupted in COVID-19 ([195](#)). As more patient data becomes available, the biotechnological advances of the 2000s are expected to allow for more rapid identification of potential drug targets than was feasible during the SARS, or even MERS, pandemic.

Thus, the COVID-19 crisis continues to evolve, but the insights acquired over the past 20 years of HCoV research have provided a solid foundation for understanding the SARS-CoV-2 virus and the disease it causes. As the scientific community continues to respond to COVID-19 and to elucidate more of the relationships between pathogenesis, transmission, host regulatory responses, and symptomatology, this understanding will no doubt continue to evolve and to reveal additional connections among virology, pathogenesis, and health. This review represents a collaboration between scientists from diverse backgrounds to contextualize this virus at the union of many different biological disciplines ([4](#)). At present, understanding the SARS-CoV-2 virus and its pathogenesis is critical to a holistic understanding of the COVID-19 pandemic. In the future, interdisciplinary work on SARS-CoV-2 and COVID-19 may guide a response to a new viral threat.

2 Evolutionary Perspectives on SARS-CoV-2

2.1 Abstract

2.2 Importance

2.3 Introduction

The emergence of what is now known to be the pathogen SARS-CoV-2 has dramatically reshaped modern life for the past two years. The genomic revolution provided the tools needed to understand the virus in ways that were not feasible during previous pandemics. For example, the first genome sequence of the pathogen was released on January 3, 2020, providing valuable information about the pathogen within a month and a half of the first known cases. As the pandemic has unfolded, evolutionary questions and methods of investigation have framed the scientific approach to understanding the virus. These questions have evolved along with the pandemic. Thus far, five major evolutionary questions have emerged. The first was “what is it?”, the second “where did it come from?”, the third and fourth address “whom does it affect?”, the fifth “how is it changing?” and the sixth “what is next?” Evolutionary biology provides a framework through which these questions can be evaluated and explored.

2.4 Question 1: What Is It?

What is now known as SARS-CoV-2 emerged in November 2019 as an unknown pathogen causing a cluster of pneumonia cases in Wuhan, China. The initial genome sequence, which was released in early January 2020, revealed the pathogen to be a novel coronavirus ([11](#)). Although most coronaviruses show little transmission in humans, several human coronaviruses (HCoV) have been identified since the 1960s. Therefore, in the early days of the pandemic, many strategies to understand or manage the emergent viral threat focused on contextualizing it amongst better-studied coronaviruses.

Many people have previously been infected by an HCoV. Approximately one-third of common cold infections are thought to be caused by four seasonal HCoV: *Human coronavirus 229E* (HCoV-229E), *Human coronavirus NL63* (HCoV-NL63), *Human coronavirus OC43* (HCoV-OC43), and *Human coronavirus HKU1* (HCoV-HKU1) ([342–344](#)). The first HCoV were identified in the 1960s: HCoV-229E in 1965 ([345](#)) and HCoV-OC43 in 1967 ([346](#)). Both of these viruses typically cause cold-like symptoms, including upper and lower respiratory infections ([347–349](#)), but they have also been associated with gastrointestinal symptoms ([350](#)). Two additional HCoV were subsequently identified ([351, 352](#)). In 2003, HCoV-NL63 ([351](#)) was first identified in a 7-month-old infant and then in clinical specimens collected from seven additional patients, five of whom were infants younger than 1 year old and the remainder of whom were adults. CoV-HKU1 was identified in samples collected from a 71-year-old pneumonia patient in 2004 and then found in samples collected from a second adult patient ([352](#)). These viruses are associated with respiratory diseases of varying severity, ranging from common cold to severe pneumonia, with severe symptoms mostly observed in immunocompromised individuals ([353](#)), and also have gastrointestinal involvement in some cases ([350](#)).

In addition to these relatively mild HCoV, however, highly pathogenic human coronaviruses have been identified, including *Severe acute respiratory syndrome-related coronavirus* (SARS-CoV or SARS-CoV-1) and *Middle East respiratory syndrome-related coronavirus* (MERS-CoV) ([250, 342, 354](#)). At the time that SARS-CoV-1 emerged in the early 2000s, no HCoV had been identified in almost 40 years ([250](#)). The first case of SARS was reported in November 2002 in the Guangdong Province of China, and over the following month, the disease spread more widely within China and then into several countries across multiple continents ([250, 340](#)). Unlike previously identified HCoV, SARS was much more severe, with an estimated death rate of 9.5% ([340](#)). It was also highly contagious via droplet transmission, with a basic reproduction number (R_0) of 4 (i.e., each person infected was estimated to infect four other people) ([340](#)).

However, the identity of the virus behind the infection remained unknown until April of 2003, when the SARS-CoV-1 virus was identified through a worldwide scientific effort spearheaded by the WHO ([250](#)). SARS-CoV-1 belonged to a distinct lineage from the two other HCoV known at the time ([340](#)). By July 2003, the SARS outbreak was officially determined to be under control, with the success credited to infection management practices ([250](#)). A decade later, a second outbreak of severe respiratory illness associated with a coronavirus emerged, this time in the Arabian Peninsula. This disease,

known as Middle East respiratory syndrome (MERS), was linked to another novel coronavirus, MERS-CoV. The fatality rate associated with MERS is much higher than that of SARS, at almost 35%, but the disease is much less easily transmitted, with an R_0 of 1 (340). Although MERS is still circulating, its low reproduction number has allowed for its spread to be contained (340). The COVID-19 pandemic is thus associated with the seventh HCoV to be identified and the fifth since the turn of the millennium, though additional HCoVs may be in circulation but remain undetected (e.g., (355)).

Following the release of the SARS-CoV-2 genome sequence, multiple research groups sequenced the genomes of SARS-CoV-2 specimens identified in clinical samples. These samples were primarily collected from patients' lower respiratory tract, namely bronchoalveolar lavage fluid (BALF), and the upper respiratory tract, in the form of throat and nasopharyngeal swabs (19, 207, 356). Integration of these sequences allowed for a more complete picture of the viral genome. Analysis of the viral genome revealed significant sequence homology with two known HCoV: the novel coronavirus shared about 79% sequence identity with SARS-CoV-1 and 50% with MERS-CoV (19). Therefore, this early phylogenetic analysis of the novel coronavirus allow its similarity to other, known viruses to be established. SARS-CoV-1 and MERS-CoV were ultimately managed largely through infection management practices (e.g., mask wearing) and properties of the virus itself (i.e., low rate of transmission), respectively (250, 340). Research in response to prior outbreaks of HCoV-borne infections, such as SARS and MERS, provided a strong foundation for hypotheses about the pathogenesis of SARS-CoV-2 as well as potential diagnostic and therapeutic approaches, as we review elsewhere (1, 3, 5, 6). Therefore, this phylogenetic information was valuable for gaining an understanding of the pathogen and identifying strategies to manage it.

2.5 Question 2: Where Did It Come From?

Despite the high degree of similarity to SARS-CoV-1, even greater sequence identity was observed between SARS-CoV-2 and zoonotic coronaviruses. A 2001 literature review estimated that 61% of human pathogens have a zoonotic origin (357). A zoonotic disease, or zoonosis, arises when a pathogen can both a) infect and b) cause a disease in humans (358). As a result, the risk of zoonotic disease increases when there is substantial interaction between humans and wildlife (358). Many factors can influence this human/wildlife interface and therefore the risk of zoonotic transmission events (358, 359).

In the SARS epidemic, SARS-CoV-1 was also thought to have emerged in a live animal market. A survey of a market in Shenzhen, China revealed that individuals from two carnivore species, namely several masked palm civets (*Paguma larvata*) and one raccoon dog (*Nyctereutes procyonoides*), were likely carriers of SARS-CoV-1, despite presenting as healthy (360). However, further analysis suggested that these species might be only intermediate hosts who were exposed in the market setting (361). A closely related virus was identified in Chinese horseshoe bats (*Rhinolophus sinicus*), but the sequence identity was only 88% with SARS-CoV-1 (362). Therefore, the species of origin for SARS-CoV-1 remains unresolved.

In the case of SARS-CoV-2, early interest for the emergence of the pathogen turned to live-animal markets in Wuhan ([363](#), [364](#)–add-to-Wuhan-riddle), where it would later emerge that many animals were sold suffering from poor health and hygiene ([365](#)). A large percentage of early patients had visited the Huanan seafood market in Wuhan, and next-generation sequencing of samples collected from nine patients, eight of whom had visited the market, revealed extremely high sequence identity (99.98%), indicative of rapid spread ([19](#)). The sequence of the viral pathogen collected from these patients was also compared to known zoonotic pathogens. In particular, genomic research quickly highlighted significant similarity (about 88% sequence identity) between SARS-CoV-2 and bat-derived SARS-like coronaviruses, namely bat-SL-CoVZC45 and bat-SL-CoVZXC21 ([19](#)). Other analyses have reported even greater similarity between SARS-CoV-2 and the bat coronavirus BatCoV-RaTG13, with shared sequence identity as high as 96.2% ([207](#), [211](#)). Bats are well-established as a disease reservoir, including for RNA viruses ([366–368](#)). This evidence therefore suggested that the virus may have emerged as a result of zoonotic transfer of a virus from bats to humans, with the wildlife trade considered a potential source of exposure.

Nevertheless, some fragments of the genome differ between SARS-CoV-2 and RATG13 by up to 17%, suggesting a complex natural selection process. Additionally, SARS-CoV-2 is closely related (91.02%) to a novel coronavirus identified in Malayan pangolins (*Manis javanica*) infected with a respiratory disease in October 2019 ([369](#)). Although the genome-wide sequence identity was lower between SARS-CoV-2 and this pangolin virus than BatCoV-RaTG13, its particularly high similarity in the receptor binding domain (RBD) of the spike (*S*) gene with SARS-CoV-2 drew further attention ([369](#), [370](#)). The SARS-CoV-2 RBD differs from the pangolin coronavirus RBD by only one amino acid change ([369](#)), and the sequence identity between the regions is 97.4% ([370](#)). Pangolins were therefore identified as a potential intermediate host of SARS-CoV-2 between bats and humans.

However, data collected from May 2017 to November 2019 by a research team interested in tick-borne illnesses identified no bats or pangolins sold at these markets leading up to the emergence of COVID-19 ([365](#)). Additionally, endemic bat species are typically in hibernation at the time that SARS-CoV-2 emerged ([19](#)). Therefore, it is possible that animals associated with these markets were infected by bats, but it is not clear whether the disease emerged in a different location and/or whether it is associated with a different species. There were 38 species observed at the market in the 2.5 years leading up to the emergence of SARS-CoV-2, indicating significant diversity in the animals with which humans were interacting ([365](#)). As with SARS-CoV-1, the species of origin for SARS-CoV-2 therefore remains unresolved.

Genomic analyses and comparisons to other known coronaviruses suggest that SARS-CoV-2 is unlikely to have originated in a laboratory – either purposely engineered and released, or escaped – and instead evolved naturally in an animal host ([371](#)). However, potentially due to public misunderstanding about recombination and complex evolutionary processes like coevolution, the similarity to pangolin *S* has resulted in popular conspiracy theories that the virus did not arise naturally. The similarity of *S* to that of pangolin viruses could arise from either recombination or coevolution

([211](#), [372](#)), rather than requiring human intervention. Such suspicions may also have been fueled, in part, by the lack of well-characterized bat coronaviruses which means that SARS-CoV-2 is still relatively derived from documented coronaviruses surveyed in bats ([373](#)). While it has been suggested that more thorough investigation of the origins of COVID-19 may have some value ([374](#)), in many cases, support for the “lab-leak” theory is politically motivated ([375](#)). A more robust panel of zoonotic viruses against which to compare SARS-CoV-2 would allow for conclusive dismissal of these politicized claims, underscoring another potential benefit of more thorough monitoring of zoonotic diseases. More importantly, it would allow researchers to have a better understanding of and to community concerns about potential emerging viral threats.

2.6 Question 3: Which Species Are Susceptible?

Given the strong evidence for a zoonotic origin of SARS-CoV-2, another evolutionary question that received significant attention, especially early on, was whether humans could infect other species with SARS-CoV-2. In the modern age, opportunities for human-to-animal transmission events could arise in interactions with companion animals, zoo animals, house pests, hunting, urbanized wildlife, and livestock. Outbreaks of zoonotic diseases have been known to originate in environments such as zoos, farms, and petting zoos ([376](#)), indicating that disease transmission is likely to be possible in these contexts. Additionally, many coronaviruses infect animals and have been the subject of veterinary medical investigations and vaccine development efforts due to their effect on the health of companion and agricultural animals ([377](#)). Concerns about anthroponotic (human-to-animal) transmission focused on a few issues. First, if animal species were susceptible to COVID-19-like infection, in addition to concerns about animal health, infections in livestock could have significant effects on food supply chains. Additionally, even if pathology in these species was limited, if they could serve as viral reservoirs, then they would pose additional risk to humans. The breadth of species susceptible to infection by a pathogen is known as the pathogen’s host range ([378](#)). Understanding the host-pathogen relationship throughout SARS-CoV-2’s host range can therefore offer valuable information for managing the spread of SARS-CoV-2.

The phylogeny of the species implicated in the origination of COVID-19 suggested that the host range of SARS-CoV-2 could encompass many species with a high level of interaction with humans. Humans last shared an ancestor with bats and pangolins almost 100 million years ago ([379](#)). Bats belong to the order Chiroptera and pangolins to Pholidota, which both belong to the clade *Pegasoferae* ([380–382](#)). They are closely related to many other species that have close relationships with humans, namely odd-toed ungulates (Euungulata) and carnivores (Carnivora) ([380–383](#)). The part of the evolutionary tree that includes both humans and the *Pegasoferae* encompasses many species of significant social and economic importance. Therefore, concerns were raised that the species with which humans have close interactions, many of which are much more closely related to bats and pangolins than humans are, could also be infected. It seemed plausible that the host range could include both livestock, many of which are odd-toed ungulates, and companion animals, many of which are carnivores. Infection of these animals was identified as a major concern ([384](#)).

Genomic analyses seeking to identify which species were likely to be susceptible focused largely on the comparative genetics of angiotensin-converting enzyme 2 (ACE2). ACE2 is the primary protein used by SARS-CoV-2 to enter the cell (see [\(1\)](#)). Recognition of this protein is largely determined by domains in the S1 subunit of the RBD ([\(25\)](#)). Alignment of the ACE2 sequence from 19 species revealed high conservation among mammals ([\(385\)](#)). This analysis suggested that non-human primates (three monkey and two ape species), companion animals (dogs and cats), and livestock (both odd- and even-toed ungulates) may all be susceptible to SARS-CoV-2 ([\(385\)](#)). Similarly, another study conducted an *in silico* analysis of ACE2 protein structures and their predicted binding to SARS-CoV-2 for 410 vertebrate species ([\(386\)](#)). The species identified as having the highest predicted binding affinities were all primates, including humans. Other taxa with high predicted affinities included other primates, rodents, even-toed ungulates (namely, several species of cetaceans and deer), and anteaters. Reindeer were the only domesticated species predicted to belong to either of these groups, but many common zoo animal species with threatened or worse IUCN risk status were identified as at risk.

Considering the evidence generated by *in silico* studies, it may not be surprising that many cases of reverse zoonotic, or anthroponotic, SARS-CoV-2 transmission have been reported. Ferrets (*Mustela furo*) as well as cats and dogs were reported to be susceptible to SARS-CoV-2 in an experimental infection study ([\(387\)](#)). The earliest reported anthroponotic transmission events were observed in house pets, primarily cats (*Felis catus*) ([\(388–390\)](#)). Similarly, cases of SARS-CoV-2 infection have been reported in dogs (*Canis familiaris*): two of fifteen dogs monitored for SARS-CoV-2 by the Hong Kong Agriculture, Fisheries, and Conservation Department during the owners' quarantine in March 2020 were found to be positive for SARS-CoV-2 ([\(389\)](#)). Comparing estimates in studies where cats (*Felis catus*) living with SARS-CoV-2-positive humans were tested for SARS-CoV-2 suggest that 6 to 15% of house cats may become infected ([\(388\)](#)), and a large-scale study of pet dogs and cats in Italy suggested that 4.5% of cats and 12.8% of dogs from known COVID-19-positive households had developed antibodies to the virus ([\(391\)](#)). Some of these SARS-CoV-2-positive domestic carnivores have also shown clinical symptoms ([\(392\)](#)), and a pilot study of seven cats and three dogs found that cats, but not dogs, shed SARS-CoV-2 virus for several days after viral challenge, although none of the animals were symptomatic ([\(393\)](#)). A few dogs and cats have reportedly died after becoming infected with SARS-CoV-2, although in most cases whether the virus is causally related to the death is unclear ([\(394–396, 397/?sh=4b653381275e, 398, 399\)](#)).

Domestic pests, on the other hand, seem to be less susceptible to SARS-CoV-2. In the comparative genomic analysis of ACE2, the two rodent species analyzed, despite being the most phylogenetically similar to humans aside from the other primates, showed the most sequence divergence in ACE2 ([\(385\)](#)). This finding was supported by experimental evidence that SARS-CoV-2 cannot use mouse (*Mus musculus*) ACE2 for cell entry ([\(207\)](#)). In fact, research using murine models to study SARS and COVID-19 therefore uses transgenic mice designed to be sensitive to the virus (as summarized in [\(400\)](#)).

Similarly, SARS-CoV-2 in livestock also raised concern because of the potential effect on food supply. However, studies using *in vivo* viral challenge reported that livestock species in general do not develop clinical manifestations of SARS-CoV-2 and do not shed infectious virus ([387](#), [401](#)). *In vitro* exposure to SARS-CoV-2 suggested that sheep (*Ovis aries*), but not cattle (*Bos taurus*), might be susceptible to infection, but *in vivo* viral challenge suggested that sheep did not show notable susceptibility to infection ([402](#)). Similarly, analyses of antibody response ([403](#)) suggested that sheep exposed to a high level of human interaction did not appear to have developed infections. Following viral challenge of several species, including cattle, sheep, and horses (*Equus ferus caballus*), none were found to shed culturable levels of virus ([401](#)). As a note, despite the low risk posed by livestock themselves, the working conditions of the meat industry itself were associated with a very high risk of SARS-CoV-2 infection for workers that did cause disruptions to food supply chains ([404](#), [405](#)).

However, one species of domesticated agricultural animal severely affected by SARS-CoV-2 was the mink (*Neovison vison*). While fur farming has declined significantly since the twentieth century, mink farming is still common in China and some European countries, and mink farms continue to exist in the United States. Mink belong to the Mustelidae family within Carnivora. SARS-CoV-2 was first reported on mink farms in the Netherlands and Denmark in 2020 ([406](#), [407](#)). Mink were observed to show symptoms of respiratory infection, with varied severity among individuals ([406](#)). Dissection revealed lung pathology consistent with interstitial pneumonia ([406](#)). An analysis of five farms in the United States reported mortality rates between 35 and 55% of adult minks ([408](#)). Subsequently, mink farms worldwide reported outbreaks of SARS-CoV-2. Concerns were amplified when novel variants of SARS-CoV-2 were identified as having emerged on Danish mink farms and spread into the human population ([407](#), [407](#), [409–411](#)). The fact that these variants appeared in mink populations before being observed in humans suggests that mink can indeed serve as a viral reservoir ([407](#)). Concerns about mink-to-human transmission led to the mass destruction of domesticated mink populations in Europe ([412](#), [413](#)). Introgression from fur farms into wild populations (i.e., feralization) may have also resulted in the spread of SARS-CoV-2 into wild mink populations ([414](#), [415](#)). Therefore, while the specific zoonotic origin of SARS-CoV-2 may still not be clear, the potential for the virus to take hold in species other than humans has been clearly demonstrated by the mink outbreak.

Finally, some species of zoo animals were also monitored to determine whether they were at risk. Several species closely related to humans (i.e., the Great Apes) are threatened with extinction and had been identified through *in silico* studies as likely to be susceptible to SARS-CoV-2 ([386](#)), and therefore the potential for a virus to target these close relatives presented a major concern. In early 2021, three gorillas (*Gorilla beringei beringei*) at the San Diego Zoo Safari Park developed respiratory symptoms that were confirmed to be associated with SARS-CoV-2 ([416](#)). Gorillas at other zoos have also been infected ([417–419](#)). Additionally, given the susceptibility of house cats, it is not so surprising that other felids are also susceptible to SARS-CoV-2. Infections of several “big cats” including Malayan tigers (*Panthera tigris jacksoni*), Amur tigers (*Panthera tigris altaica*), and African lions (*Panthera leo krugeri*) were reported at New York City’s Bronx Zoo in March 2020 ([420](#)). In

late 2020, four lions (*Panthera leo bleyenberghi*) at the Barcelona Zoo also developed respiratory symptoms that were found to be caused by SARS-CoV-2 (411). Several captive snow leopards (*Panthera uncia*) in the United States have died from COVID-19 (421, 422).

While discussions of zoonoses often focus on the risk that animal diseases carry for human populations, the COVID-19 pandemic has also underscored the risks that human diseases pose for animals. COVID-19 precautions may have reduced the spread of other respiratory illnesses to wild mountain gorilla (*Gorilla beringei beringei*) populations (423), reducing one of the most significant threats to this endangered species (424). In the case of gorillas, the potential for cross-species application of pharmaceutical advances has also become clear: captive gorillas with COVID-19 received monoclonal antibodies (425). Additionally, several companies are developing veterinary vaccines against SARS-CoV-2. The most visible has been Zoetis, a veterinary pharmaceutical company, that has developed vaccines that have been administered to several species, including felids in zoos, minks, and gorillas (426–429). Russian researchers have also developed a COVID-19 vaccine for carnivores (430).

Therefore, the host range of SARS-CoV-2 is broad, including primates, bats, and carnivores. The most severe infections have been observed in humans, felids, and mustelids (426). In the United States, as of late 2021, dogs and cats made up the majority of non-human SARS-CoV-2 infections (431), but the most severe infections were observed in felids and mustelids (in addition to humans) (426). Interestingly, comparing ACE2 binding activity across species (432, 433) revealed that it did not always align with which species known to be susceptible to SARS-CoV-2 infection, suggesting other binding sites might also be important. While the specific zoonotic origins of SARS-CoV-2 remain unknown, pharmaceutical developments in the treatment of COVID-19 have included non-human species. The complex relationship between animals, humans, and disease highlights the importance of a broad perspective on health that extends beyond a single species.

2.7 Question 4: Do Genes Influence Who is Affected?

Throughout the pandemic, many hypotheses have been raised about factors that might influence individuals' susceptibility to COVID-19 or to severe disease. Many risk factors, such as underlying health conditions, are related to the body's inflammatory response, as we review elsewhere (339). Here, we focus narrowly on genetic bases of differences in susceptibility or outcomes. Historically, the identification of genetic risk factors for a disease typically utilized a candidate gene approach, where a gene of interest was evaluated to identify variants that showed an association with the outcome of interest. While economical in terms of sequencing, this approach is prone to spurious results when applied to complex traits (434). Today, in the age of next-generation sequencing (NGS), alternative approaches have emerged. NGS makes it possible to conduct genome-wide scans where a large number of single-nucleotide polymorphisms (SNPs) or variants are evaluated to identify regions of the genome associated with variation in a phenotype. Genome-

wide association studies (GWAS) in particular are a popular approach that employs this strategy. During COVID-19, both of these paradigms have been applied to the problem of identifying genetic correlates of disease severity.

2.7.1 Candidate-Gene Approaches

Many candidate genes have been investigated throughout the pandemic. Here, we review three examples of candidate gene studies in COVID-19. First, an early study (published in April 2020) investigated a known variant in interferon-induced transmembrane protein 3 (*IFITM3*) among hospitalized patients in Beijing (435). This gene and variant were selected because of a prior candidate gene study by some of the same authors that found an association with influenza severity among Chinese patients during the 2009 influenza A H1N1/09 pandemic (436). Here, they evaluated a small number (n=80) hospitalized COVID-19 patients to determine whether homozygosity for the previously identified risk allele was associated with mild versus severe disease (435). They stated that they found an association between homozygosity for the SNP of interest and the severity of COVID-19. A follow-up study demonstrated worldwide variation in the frequency of these SNPs (437), and subsequent studies claimed to support this result by comparing the frequency of the SNP in different groups to the COVID-19 case fatality rate in those groups; they examined SNPs in several candidate genes and identified an association with another SNP in *IFITM3* (438). However, in the original study, the population-level frequency of the risk allele was consistent with its frequency in the mild population (436). A similar analysis examined both SNPs in Britons of different ancestral backgrounds and also reported a correlation (439). While this gene has been investigated for functions potentially relevant to COVID-19 pathogenesis by other groups as well (e.g., (440, 441), a follow-up analysis in Germany evaluated the effect of in 239 cases and 252 controls and reported non-significant effects (439). The narrative surrounding *IFITM3* therefore reflects a broad methodological critique about candidate gene studies, where results often fail to replicate (442). The region associated with this gene was not identified in the large-scale GWAS conducted by the COVID-19 Host Genetics Initiative (COVID-19 HGI) (443), which is described in more detail below.

A second source of genetic variability that was hypothesized to have an effect on COVID-19 outcomes were human leukocyte antigens (HLA), or the major histocompatibility complex (MHC). Both MHC classes I and II play a critical role in both the innate and adaptive immune system because they are a pivotal component of antigen presentation. HLA classes I and II are also the most polymorphic loci in the human genome (444). Additionally, because HLA polymorphisms are associated with geographic ancestry, study location and participant background offers important context (445). Given the important role of the HLA complex in the immune response and the standing variation in the human population, HLA variation has been investigated for potential associations with COVID-19 outcomes.

Several approaches have been taken to evaluate a potential role of HLA in COVID-19. *In silico* analysis suggested one particular HLA locus that could affect binding of SARS-CoV-2 peptides to MHC class II (446). Other studies evaluated outcomes using retrospective cohort analyses. An analysis of 95 South Asian COVID-19 patients found that HLA genotype was not significant

in differentiating case severity when the necessary statistical corrections were applied (447). Another study in a European population (n =147) did identify HLA alleles associated with severity (448). In St. Louis, MO (USA), another study enrolled 234 COVID-19 cases, who were genotyped for HLA alleles and compared to a control population of 20,000 individuals from the National Marrow Donor Program (449). They compared cases and controls on the basis of four “race/ethnic” populations and reported alleles showing a statistical association within each group (449). However, because of this stratification, two of the demographic categories had less than ten cases. Across all of these studies, there was minimal overlap in the risk alleles identified, and the small sample sizes raise concerns about the possibility for spurious hits. The hypervariability of this region means that statistical power will necessarily be reduced, with much higher recruitment needed than for studies of biallelic loci. A much larger analysis of 72,912 Israelis, 8.8% of whom tested positive for COVID-19, found no association between HLA genotype and infection or hospitalization (450). Therefore, while MHC is functionally important to the immune response to COVID-19, it is not clear whether HLA genotypes are predictive of COVID-19 severity, and certainly such studies face exacerbated versions of the typical challenges of candidate gene studies. Because of the challenges associated with analyzing such a variable region, it was excluded from the large-scale COVID-19 HGI GWAS analysis (443).

Finally, significant attention has been paid to the question of whether ABO blood type is associated with COVID-19 outcomes. ABO blood type has been found to modulate susceptibility to other pathogens (451). While ABO blood type is a genetic trait, it is more easily evaluated than the genetic regions discussed above because of the simple relationship between genetic variants and phenotype. The possibility for an association between blood type and COVID-19 infection was raised early in the pandemic in a preprint that reported associations in 2,173 patients in Wuhan and Shenzhen, China (452). The protective effect of O and increased risk associated with A blood types that they reported was subsequently investigated by many studies that returned varied results (e.g., (453–456); see (457) for a literature review). Observations of higher and lower risk, respectively, of SARS-CoV-2 infection with A and O blood types was supported by a meta-analysis (458). While the support for the association was independent of a mechanism, a possible relationship between ACE activity and blood type has been proposed (459) as has an effect on carbohydrate-carbohydrate interactions relevant to ACE2 binding (460). This is the only candidate gene described that has received additional support from GWAS, as is discussed below.

The COVID-19 literature related to candidate gene investigations demonstrates relatively low inter-study consistency in findings. In particular, sample size is a major challenge in designing these studies. However, for many traits, the relationships between genes and phenotypes are complex, and selecting which variants to sequence is not always straightforward. As a result, in the age of next-generation sequencing, discovery-driven studies have emerged as an alternative approach.

2.7.2 Genome-Wide Association Studies

Genome-wide association studies (GWAS) offers a discovery-driven approach that provides a different perspective than candidate gene studies. Instead of selecting a gene or variant *a priori*, in GWAS, a large number of SNPs (usually several million) are evaluated at once to identify those most likely to vary in correlation with a trait of interest. Because of the large number of statistical tests, statistical power and multiple hypothesis testing are both very important considerations in executing GWAS, which have also struggled with issues related to replicability ([461](#)). In cases such as COVID-19 where outcomes can differ among ancestry groups (likely for non-genetic reasons, as reviewed in ([339](#))), it is especially important that GWAS samples be selected with attention paid to ancestry, as incorrect or misleading associations can otherwise be identified with neutral markers indicative of ancestry itself ([462](#)).

Over the past two years, many GWAS have been undertaken with the aim of identifying variants associated with COVID-19 outcomes. In some cases, the results have been consistent with hypothesized genetic correlates of susceptibility to COVID-19. One study conducted a GWAS on a total of 435 COVID-19 patients from four countries and identified another HLA allele to be associated with an increased risk of intubation ([463](#)). Other GWAS have identified an association with the ABO blood group locus. One conducted a case/control GWAS in two populations, Italians and Spaniards, with 1980 cases and 2205 controls. They reported two loci that met the genome-wide significance threshold, one on chromosome 3 and one on chromosome 9 ([464](#)). The hit on chromosome 9 fell on the ABO locus and the alleles identified suggested a protective association with blood group O and a risk association with blood group A ([464](#)).

As the pandemic has progressed, large-scale efforts have been assembled to conduct GWAS on massive scales. In March 2020, COVID-19 HGI was established as a world-wide consortium that combines data to conduct meta-analyses ([465](#)). One year later, COVID-19 HGI released a meta-analysis of data from 46 studies, comprising 49,562 cases and 1,770,206 controls ([443](#)). They identified 13 loci, seven of which were significant at the genome-wide level when considering all data available, that were associated with one or more phenotypes related to COVID-19 infection or severity. Notably, strong signals were identified for both of the loci suggested by previous medium-scale GWAS in association with COVID-19 infection ([464](#)). Additionally, several other loci could be mapped onto hypotheses about genetic contributors to immune function, lung function and disease. This world-wide GWAS study made an effort towards strategic incorporation of genetic information from different ancestral groups. Interestingly, the risk variant on chromosome 3 is likely to be inherited from Neanderthal introgression, meaning it is likely to be more prevalent in certain populations, especially non-African populations ([466](#), [467](#)). The potential functional relationship between this region of the genome and COVID-19 is unknown, but phenome-wide association study has suggested blood cell traits as a potential trait regulated by this region ([468](#)).

Identifying genetic variants associated with a complex disease is always complicated. In COVID-19 studies, the results of candidate gene analyses have in general been difficult to replicate. However, large-scale collaboration on GWAS has made it possible to detect at least two loci that do appear to replicate across studies and potentially even across ancestral backgrounds.

2.8 Question 5: How is it Changing?

Evolution in SARS-CoV-2 has also been observed over a short timescale. After zoonotic transfer, SARS-CoV-2 continued evolving in the human population (210). The SARS-CoV-2 mutation rate is moderate compared to other RNA viruses (212), which likely restricts the pace of evolution in SARS-CoV-2. Nevertheless, genomic analyses have yielded statistical evidence of ongoing evolution. Initially, two known variants of the spike protein emerged that differed by a single amino acid at position 614 (G614 and D614), and there is evidence that G614 had become more prevalent than D614 by June 2020 (222). While there is a hypothesis that this genomic change increased the SARS-CoV-2 infectivity and virulence, this hypothesis has not yet been tested due to a lack of data (469). Another study (212) identified 198 recurrent mutations in a dataset of 7,666 curated sequences, all of which defined non-synonymous protein-level modifications. This pattern of convergent evolution at some sites could indicate that certain mutations confer an adaptive advantage. While it is evident that SARS-CoV-2 exhibits moderate potential for ongoing and future evolution, the relationship between mutations and pathogenicity is not yet known. Additional data is needed in order to understand patterns of evolutionary change and the mechanisms by which they might affect virulence.

Several factors could promote the evolution of SARS-CoV-2, including host immunodeficiency and transient exposure to antibodies directed against SARS-CoV-2 proteins. A single case study of SARS-CoV-2 infection in an immunocompromised female with chronic lymphocytic leukemia and hypogammaglobulinemia (470) suggested that an accelerated evolution of the virus could occur in conditions of immunodeficiency. A first administration of convalescent plasma did not clear the virus, and an ensuing increase in the genomic diversity in the samples was observed, suggesting an accelerated evolution due to selection pressure. A second administration of convalescent plasma cleared the virus from the host 105 days after the initial diagnosis. However, throughout the duration of infection, the patient was asymptomatic but contagious. A second single case study in a 45-year old male with antiphospholipid syndrome (471) confirmed the earlier results, providing evidence of persistent COVID-19 symptoms in an immunocompromised patient for 154 days following diagnosis, ultimately leading to the death of patient. The treatments administered included remdesivir and the Regeneron anti-spike protein antibody cocktail. Genomic analyses of the patient's nasopharyngeal swabs confirmed an accelerated evolution of the virus through mutations in the spike gene and the receptor-binding domain. In summary, these two case studies suggested an accelerated evolution and persistent shedding of the virus in conditions of immunodeficiency. In particular, the first case highlighted the role of convalescent plasma in creating escape variants. In fact, one study (472) exposed the SARS-CoV-2 virus to convalescent plasma *in vitro* repeatedly to see how much plasma was required to neutralize the virus. The results of the first six exposures were similar, but they reported that after the seventh exposure (on day 45), the amount of plasma required began to increase. In analyzing the viral variants present, they found that this viral escape was promoted by the sudden accumulation of mutations, especially in the receptor-binding domain (RBD) and N-terminal domain (NTD), that quickly rose in frequency. By the thirteenth exposure (day 85), the virus had evolved

three mutations and could no longer be neutralized by the plasma used, even though the plasma was comprised of polyclonal serum that targeted a variety of epitopes. Taken together, these observations suggest that evolutionary analyses of SARS-CoV-2 can provide crucial information about the conditions that promote resistance in SARS-CoV-2 and the kinetics of how resistance develops, information which will be important for understanding the implications of how vaccine regimens are designed and whether/when next-generation vaccines will be needed.

When variants occur, they can rise in frequency by chance or through an adaptive process that confers a competitive advantage to the virus. Variants that had the D614G mutation in the spike glycoprotein seemed to spread faster. However, it has been suggested that the mutation rose in frequency due to early chance events rather than by adaptive events ([473](#)). Another mutation, Y453F, that occurred in the receptor binding domain of *S*, was first detected in mink; however, the transmission to humans has been established. In mink, this mutation conferred an advantage by increasing the affinity towards ACE2 ([474](#)). Similarly, N501Y mutation induces an increased affinity towards human ACE2 and has been involved in the dominance of B.1.1.7 by outcompeting other variants ([475](#)). Therefore, genomic surveillance is essential to prevent the emergence of super-spreaders ([476](#)).

Emerging methods are being applied to this problem in an effort to understand which mutations are most likely to be of significant concern. Novel machine learning methods were developed to predict the mutations in the sequence that promote viral escape. While they preserve the pathogenicity of the virus, escape mutations change the virus's sequence to evade detection by the immune system. By using tools from natural language processing (NLP), viral escape was modeled as an NLP problem ([477](#)) where a modification makes a sentence grammatically correct but semantically different. Therefore, language models of viruses could predict mutations that change the presentation of the virus to the immune system but preserve its infectivity.

2.8.1 Variants of Concern and Variants under Surveillance

Viral replication naturally leads to the occurrence of mutations, and thus to genetic variation ([478](#)). However, due to an intrinsic RNA proof-reading process in the SARS-CoV-2 virus, the pace of evolution of SARS-CoV-2 is moderate in comparison to other viruses ([479](#)). The declaration of the first SARS-CoV-2 variant of concern (VOC) B.1.1.7 in December 2020 has attracted significant media attention. While the B.1.1.7 lineage garnered attention in November 2020, two genomes of the lineage were detected as early as September 20th, 2020 from routine genomic data sampled in Kent (U.K.) by the COVID-19 Genomics UK Consortium (COG-UK). The following day, a second B.1.1.7 genome was reported in greater London ([234](#), [473](#), [480](#), [481](#)). Since then, B.1.1.7 has spread across the UK and internationally, and it has now been detected in at least 62 countries ([482](#)), despite several countries imposing travel restrictions on travelers from the UK. Of the twenty-three mutations that define B.1.1.7 from the original strain isolated in Wuhan (lineage A), fourteen are lineage-specific and three appear to be biologically consequential mutations associated with the spike protein, namely N501Y,

P681H, and 69-70del ([480](#), [481](#)). The latter is a 6-bp deletion that leads to the loss of two amino acids and has consequences for immune recognition; it may, in conjunction with N501Y, be responsible for the increased transmissibility of the B.1.1.7 VOC due to changes in the RBD that increase binding affinity with ACE2 ([230](#), [480](#)). B.1.1.7 has increased transmissibility by up to 56%, leading to an R_0 of approximately 1.4. Additionally, this VOC has been shown to be associated with increased disease severity and increased mortality ([483](#)). Other variants also express the 69-70del mutation ([484](#), [485](#)), and public health officials in the United States and the UK have been able to use RT-PCR-based assays (ThermoFisher TaqPath COVID-19 assay) to identify sequences with this deletion because it occurs where the qPCR probe binds ([234](#)). In the UK, B.1.1.7 is present in more than 97% of diagnostic tests that return negative for S-gene targets and positive for the other targets; thus, the frequency of S-gene target failure can be used as a proxy for the detection of B.1.1.7 ([480](#), [486](#)). The FDA has highlighted that the performance of three diagnostic tests may be affected by the B.1.1.7 lineage because it could cause false negative tests ([487](#)).

While B.1.1.7 is currently the main VOC, other genetic variants also currently designated as VOCs have been detected, including B.1.351 and P.1, both of which emerged independently ([488](#), [489](#)). B.1.351 was first detected in October 2020 in South Africa, was later detected in the EU on December 28th, 2020 and has now spread to at least 26 countries ([231](#), [490](#), [491](#)). B.1.351 contains several mutations at the RBD including K417N, E484K, and N501Y. While the biological significance of these mutations are still under investigation, it does appear that this lineage may be associated with increased transmissibility ([492](#)) due to the N501Y mutation ([230](#), [481](#)). Additionally, an analysis of a pseudovirus expressing the 501Y.V2 spike protein (B.1.351) showed that this variant demonstrates increased resistance to neutralization by convalescent plasma, even though total binding activity remained mostly intact ([493](#)). Further, using a live virus neutralization assay (LVNA), it was shown that 501Y.V2 (B.1.351) is poorly neutralized by convalescent plasma obtained from individuals who responded to non-501Y.V2 variants ([494](#)). However, 501Y.V2 infection-elicited plasma was able to cross-neutralize earlier non-501Y.V2 variants, suggesting that vaccines targeting VOCs may be effective against other mutant lineages ([494](#)).

The P.1 variant is a sublineage of the B.1.1.28 lineage that was first detected in Japan in samples obtained from four travelers from Brazil during a screening at a Tokyo airport on January 10, 2021 ([495](#)). Shortly thereafter, it was established that there was a concentration of cases of the P.1 variant in Manaus, Brazil. In a small number of samples (n=31) sequenced in Manaus, 42% were identified as the P.1 variant as early as mid-December, but the variant seemed to be absent in genome surveillance testing prior to December ([496](#)). To date, at least eight countries have detected the P.1 lineage ([497](#)). While the majority of P.1 cases detected internationally have been linked to travel originating from Brazil, the UK has also reported evidence of community transmission detected via routine community sequencing ([497](#), [498](#)).

P.1 has eight lineage-specific mutations along with three concerning spike protein mutations in the RBD, including K417T, E484K, and N501Y ([492](#)).

There have been multiple different SARS-CoV-2 lineages detected that have mostly been of no more clinical concern than the original devastating lineage originating in Wuhan (499). However, the spotlight has been cast on other variants of unknown clinical relevance due to the increase of cases observed that have been associated with B.1.1.7 in particular.

Although early in its ascendancy, B.1.427/429 are SARS-CoV-2 variants that was detected in California, USA and also known as CAL.20C (500). It was first detected in July 2020 but was not detected again until October 2020. In December 2020, B.1.427/429 accounted for ~24% of the total cases in Southern California and ~36% of total cases in the Los Angeles area.

B.1.427/429 have now been detected in several U.S. states and at least 38 countries worldwide (500, 501). This variant is characterized by five key lineage-specific mutations (ORF1a: I4205V, ORF1b:D1183Y, S: S13I;W152C;L452R). The latter spike mutation, L452R, is found in an area of the RBD known to resist monoclonal antibodies to the spike protein (502), and it is hypothesized that this mutation may resist polyclonal sera in convalescent patients or in individuals post-vaccination (500, 503).

B.1.427/429 are now designated VOCs (489); however, further research is still required to determine the implications of the mutations encoded in this genetic variant.

Another notable variant has recently been discovered in 35 patients in a Bavarian hospital in Germany; however, the sequencing data has not been published to date and it remains to be determined whether this variant is of any further concern (504).

There are several shared mutations and deletions between the three lineages, P.1, B.1.1.7, and B.1.315 and indeed other variants of SARS-CoV-2 that are under investigation (496). For example, N501Y, which appears to have occurred independently in each of the three lineages.

E484K is present in both B.1.351 and P.1 (505). The mutations N501Y and E484K are found in the RBD within the receptor-binding motif responsible for forming an interface with the ACE2 receptor, which seems to be consequential for ACE2 binding affinity (506). Indeed, N501Y is associated with increased virulence and infectivity in mouse models (507). E484K has also been associated with evasion from neutralizing antibodies (472, 503, 508). The del69-70 (del:11288:9) is also shared between P.1 and B.1.1.7 and happens to be a common deletion found in the N terminal mutation of the spike protein. This deletion has also been associated with several RBD mutations (230, 481, 509). There is concern that mutations in the spike protein of variants may lead to clinical consequences for transmissibility, disease severity, re-infection, therapeutics, and vaccinations (472, 503, 510–514).

Vaccine producers are working to determine whether the vaccines are still effective against the novel genetic variants. Moderna recently published data for their mRNA-1273 vaccine that showed no significant impact of neutralization against the B.1.1.7 variant upon vaccination in humans and non-human primates. On the other hand, Moderna reported a reduced but significant neutralization against the B.1.351 variant upon vaccination (515). Indeed, Pfizer–BioNTech reported that sera from twenty participants vaccinated with the BNT162b COVID-19 vaccine in previous clinical trials (516, 517) elicited equivalent neutralizing titers against isogenic Y501 SARS-CoV-2 on an N501Y genetic background *in vitro* (518). Another study has reported

that the plasma neutralizing activity against SARS-CoV-2 variants encoding the combination of K417N:E484K:N501Y or E484K or N501Y was variably and significantly reduced in the sera of twenty participants who received either the Pfizer–BioNTech BNT162b (n = 6) vaccine or the Moderna’s mRNA-1273 vaccine (n =14) ([519](#)). In a study focusing on serum samples from a combination of convalescent individuals, those who obtained the mRNA-1273 vaccine, and those who obtained Novavax, in comparison to the D614G variant, the B.1.419 variant was 2-3 times less sensitive to neutralization while the B.1.351 variant was 9-14 times less sensitive ([520](#)). Indeed, the E484K substitution seen in the P.1 and B.1.315 variants of the B.1.1.7 lineage are broadly reported to substantially reduce the efficacy of mRNA-based vaccines ([520–522](#)). For now, the consensus appears to be that the FDA-approved vaccines still seem to be generally effective against the genetic variants of SARS-CoV-2 and their accompanying mutations, albeit with a lower neutralizing capacity ([515](#), [518](#), [519](#), [523](#)), though select VOCs may present challenges. Further research is required to discern the clinical, prophylactic, and therapeutic consequences of these genetic SARS-CoV-2 variants as the pandemic evolves.

2.9 SARS-CoV-2 Evolution and Vaccine Efficacy

With these vaccines in place, one concern is how the virus’s continued evolution will affect their efficacy. Since the start of this pandemic, we have already seen multiple variants emerge: B.1.1.7, which emerged in the UK, B.1.351, which emerged in South Africa, and P.1, which emerged in Brazil.

Viruses evolve or mutate at different rates. Mutation rate is measured as the number of substitutions per nucleotide per cell infected ($\mu_{s/n/c}$) ([524](#)). RNA viruses tend to have mutation rates between 10^{-6} to 10^{-4} ([524](#)). As a reference, influenza A virus has a mutation rate of 10^{-5} , whereas the mutation rate of SARS-CoV-2 is lower, with the mutation rate estimated at 10^{-6} ([525](#)). The accumulation of mutations allows the virus to escape recognition by the immune system ([526](#)).

The efficacy of vaccines depends on their ability to train the immune system to recognize the virus. Therefore, viruses can develop resistance to vaccines through the accumulation of mutations that affect recognition. The lower mutation rate of SARS-CoV-2 suggests the possibility of SARS-CoV-2 vaccines having a more long-lasting effect compared to vaccines targeting the influenza A virus.

2.9.1 Alpha and Beta Variants

The current SARS-CoV-2 vaccines in distribution have been reported to provide similar efficacy against the B.1.1.7 variant compared to the variants common at the time they were developed but reduced efficacy against the B.1.351 variant ([527](#)). Pfizer and Moderna announced that they are working on developing a booster shot to improve efficacy against the B.1.351 variant ([528](#)). The WHO continues to monitor the emergence of variants and their impact on vaccine efficacy ([529](#)). Previous research in the computational prediction of the efficacy of vaccines targeting the influenza A virus might

complement efforts to monitor these types of viral outbreaks ([530](#)). To adapt, future vaccines may need to account for multiple variants and strains of SARS-CoV-2, and booster shots may be required ([531](#)).

2.9.2 Delta Variant and C_t

One preprint ([334](#)) analyzed a retrospective cohort of patients in Singapore who contracted COVID-19 from April to June of 2021. This study focused on those who were confirmed or inferred to have been infected by the Delta variant of concern, and its aim was to analyze virological kinetics. They identified 218 cases, 71 (33%) of whom were fully vaccinated with either the Pfizer/BioNTech or Moderna mRNA vaccines, 13 (6%) of whom had received only one dose or had received the second dose less than two weeks prior to infection, and four (2%) of whom had received a vaccine developed with another technology. Unvaccinated patients were more likely to be symptomatic or to progress to severe COVID-19 and showed more symptoms than vaccinated patients, despite the higher age of the vaccinated cohort. C_t was assessed over disease course, although the specific procedures for when additional RT-PCR was conducted is not clear; however, it is stated that the data was smoothed based on day of illness. There was no significant difference in median C_t in the initial samples taken from fully vaccinated and unvaccinated patients, but C_t increased (signifying reduced viral load) more rapidly in fully vaccinated patients. Like most analyses analyzing C_t ([1](#)), this study does not provide the data to make conclusions about contagiousness, as the samples were not cultured. All the same, these findings do suggest that vaccinated individuals may be able to clear the infection more quickly.

A second analysis was based in Dane County in Wisconsin, USA during summer 2021, when the Delta variant was known to be the dominant variant in the region ([335](#)). According to Our World in Data ([532](#)), at the beginning of the study, 49.3% of residents of Dane County were fully vaccinated, with this number rising to 51.4% by the end of the study, although an earlier version of the preprint reported the vaccination rate in Dane County as 67.4%. The authors identified no significant differences in C_t among fully vaccinated and unvaccinated cases. The C_t thresholds reported were consistent with contagiousness as evaluated in other studies, and in the present study, SARS-CoV-2 could be cultured from 51 of 55 samples with C_t less than 25. This study was not longitudinal, but the timing of testing relative to symptom onset between symptomatic vaccinated and unvaccinated patients. The findings of this study are therefore consistent with the idea that vaccinated people are less likely to contract symptomatic or severe COVID-19, but in cases of breakthrough infection, are still likely to be able to transmit SARS-CoV-2 to others.

2.10 Question 6: What is Next?

The SARS-CoV-2 pandemic has presented many unprecedented scientific opportunities. The rapid identification of the genomic sequence of the virus allowed for early contextualization of SARS-CoV-2 among other known respiratory viruses, and the scientific community has continued to collect, analyze, and disseminate information about the SARS-CoV-2 virus and the associated illness, COVID-19 at previously unimaginable rates ([4](#)). The

accessibility of genome sequencing technology has allowed for deep sequencing of the virus to establish a level of viral surveillance that had never before been achieved ([214](#), [533](#), [534](#)). The information obtained from genetic, bioinformatics, and evolutionary analysis has played a significant role in shaping the global pandemic response ([533](#), [535](#), [536](#)). For example, wastewater surveillance has emerged as a potential epidemiological tool to monitor SARS-CoV-2 spread over large regions, complementing clinical surveillance ([537–539](#)). Humans shed SARS-CoV-2 viral RNA in feces ([540](#)) that can be detected in wastewater. Protocols have been developed to safely and reproducibly isolate and quantify SARS-CoV-2 in samples obtained from wastewater processing plants ([539](#), [541](#)). To date, studies show that wastewater surveillance is an effective tool to monitor SARS-CoV-2 spread over large sewersheds ([537–539](#), [542](#)). Indeed, data from a study in New York City indicated that wastewater SARS-CoV-2 detection correlated with clinical detection of infection ([542](#)). Similar studies have been conducted in Nevada ([543](#)) and Boston ([544](#)). To date, studies have shown that factors such as temperature, the travel time of wastewater, and diurnal variability may affect detection of SARS-CoV-2 ([537](#), [543](#)). Additionally, wastewater surveillance provides a tool to monitor fluctuations in the viral strains present in a community ([545](#), [546](#)). Due to its demonstrated utility so far, the United States CDC established the National Wastewater Surveillance System (NWSS), which has emerged as an important surveillance tool for SARS-CoV-2 spread ([547](#)).

Knowledge of the evolution of SARS-CoV-2 is imperative to managing it moving forward ([533](#), [548](#)).

The evolutionary questions highlighted here all point back to the fact that efforts to prevent future epidemics and pandemics will benefit greatly from long-term, sustainable efforts to monitor disease. Beyond understanding the status and evolution of known pathogens via genomic surveillance, greater preparedness for novel viral threats would also result from monitoring zoonotic disease. If not addressed, economic and environmental stressors are likely to cause future zoonotic transfer of diseases in the future ([549](#)). The COVID-19 pandemic has highlighted both the incredible insights available with modern evolutionary and genomic methodologies, but has also revealed the reluctance of political actors to commit resources to these efforts outside of periods of acute need. The One Health framework has emerged from collaborations by many prominent non-governmental organizations such as the World Health Organization to promote scientific goals supportive of pandemic preparedness ([534](#)). Genomic surveillance of human pathogens and of pathogens at the human-wildlife interface is an important component needed to meet the goals of One Health ([534](#)). These efforts are especially important as anthropogenic alterations to the landscape such as climate change and urbanization increase the risk of zoonotic disease transmission ([550](#), [551](#)). With the COVID-19 pandemic serving as a clear illustration of why this surveillance is imperative and of its feasibility, wider awareness and adoption of the One Health paradigm is the last piece needed to develop practices that will prevent the next pandemic.

3 Molecular and Serologic Diagnostic Technologies for SARS-CoV-2

3.1 Abstract

The COVID-19 pandemic has presented many challenges that have spurred biotechnological research to address specific problems. Diagnostics is one area where biotechnology has been critical. Diagnostic tests play a vital role in managing a viral threat by facilitating the detection of infected and/or recovered individuals. From the perspective of what information is provided, these tests fall into two major categories, molecular and serological.

Molecular diagnostic techniques assay whether a virus is present in a biological sample, thus making it possible to identify individuals who are currently infected. Additionally, when the immune system is exposed to a virus, it responds by producing antibodies specific to the virus. Serological tests make it possible to identify individuals who have mounted an immune response to a virus of interest and therefore facilitate the identification of individuals who have previously encountered the virus. These two categories of tests provide different perspectives valuable to understanding the spread of SARS-CoV-2. Within these categories, different biotechnological approaches offer specific advantages and disadvantages. Here we review the categories of tests developed for the detection of the SARS-CoV-2 virus or antibodies against SARS-CoV-2 and discuss the role of diagnostics in the COVID-19 pandemic.

3.2 Importance

Testing is critical to pandemic management. Among molecular tests, messaging about testing strategies has varied widely between countries, with the United States in particular emphasizing the higher sensitivity of polymerase chain reaction tests above immunoassays. However, these tests offer different advantages, and a holistic view of the testing landscape is needed to identify the information provided by each test and its relevance to addressing different questions. Another important consideration is the ease of use and ability to scale for each test, which determines how widely they can be deployed. Here we describe the different diagnostic technologies available as well as the information they provide about SARS-CoV-2 and COVID-19.

3.3 Introduction

Since the emergence of *Severe acute respiratory syndrome-like coronavirus 2* (SARS-CoV-2) in late 2019, significant international efforts have focused on managing the spread of the virus. Identifying individuals who have contracted coronavirus disease 2019 (COVID-19) and may be contagious is crucial to reducing the spread of the virus. Given the high transmissibility of SARS-CoV-2 and the potential for asymptomatic or presymptomatic

individuals to be contagious ([1](#)), the development of rapid, reliable, and affordable methods to detect SARS-CoV-2 infection is and was vitally important for understanding and controlling spread. For instance, test-trace-isolate procedures were an early cornerstone of many nations' efforts to control the outbreak ([552–554](#)). Such efforts depend on diagnostic testing.

The genetic sequence of the SARS-CoV-2 virus was first released by Chinese officials on January 10, 2020 ([555](#)), and the first test to detect the virus was released about 13 days later ([556](#)). The genomic information was critical to the development of diagnostic approaches. There are two main classes of diagnostic tests: molecular tests, which can diagnose an active infection by identifying the presence of SARS-CoV-2, and serological tests, which can assess whether an individual was infected in the past via the presence or absence of antibodies against SARS-CoV-2. Over the course of the COVID-19 pandemic, a variety of tests have emerged within these two categories.

Molecular tests detect either viral RNA or protein in a patient sample. They are essential to identifying infected individuals, which can be important for determining courses of action related to treatment, quarantine, and contact tracing. Tests for viral RNA are done by reverse transcription (RT) of viral RNA to DNA followed by DNA amplification, usually with polymerase chain reaction (PCR) ([557](#)). Tests for viral proteins typically use an antibody pair for detection as implemented in techniques such as lateral flow tests (LFTs) and enzyme-linked immunosorbent assays (ELISAs) ([558](#), [559](#)). Molecular tests require the viral genome sequence in order to develop DNA primers for viral RNA detection or to express a viral protein for use as an antigen in antibody production.

Serological tests, on the other hand, detect the presence of antibodies in blood plasma samples or other biological samples, providing insight into whether an individual has acquired immunity against SARS-CoV-2. Assays that can detect the presence of antibodies in blood plasma samples include ELISA, lateral flow immunoassay, and chemiluminescence immunoassay (CLIA) ([560](#)). To distinguish past infection from vaccination, serological tests detect antibodies that bind the nucleocapsid protein of the SARS-CoV-2 virus ([561](#)). They are useful for collecting population-level information for epidemiological analysis, as they can be used to estimate the extent of the infection in a given area. Thus, serological tests may be useful to address population-level questions, such as the percent of cases that manifest as severe versus mild and for guiding public health and economic decisions regarding resource allocation and counter-disease measures.

Molecular and serological tests therefore offer distinct, complementary perspectives on COVID-19 infections. Some of the same technologies are useful to both strategies, and different technologies have been employed to varying extents throughout the world since the start of the COVID-19 pandemic. Two of the primary metrics used to evaluate these tests are sensitivity and specificity. Sensitivity refers to a test's ability to correctly identify a true positive; for example, a test with 50% sensitivity would identify SARS-CoV-2 in only one of every two positive samples. On the other hand, specificity refers to how well a test is able to identify a negative sample as negative. This metric can be relevant both in terms of understanding the risk of false positives and in discussing whether a test is susceptible to identifying

other coronaviruses. Here, we review the different types of tests within each category that have been developed and provide perspective on their applications.

3.4 Molecular Tests to Identify SARS-CoV-2

Molecular tests are used to identify distinct genomic subsequences of a viral molecule in a sample or the presence of viral protein, and they thus can be used to diagnose an active viral infection. An important first step is identifying which biospecimens are likely to contain the virus in infected individuals and then acquiring these samples from the patient(s) to be tested. Common sampling sources for molecular tests include nasopharyngeal cavity samples, such as throat washes, throat swabs, and saliva ([562](#)), and stool samples ([563](#)). Once a sample is acquired from a patient, molecular tests detect SARS-CoV-2 based on the presence of either viral nucleic acids or viral proteins.

3.4.1 PCR-Based Tests

When testing for RNA from viruses like SARS-CoV-2, the first step involves pre-processing in order to create complementary DNA (cDNA) from the RNA sample using RT. The second step involves the amplification of a region of interest in the cDNA using successive cycles of heating and cooling. Depending on the application, this amplification is achieved using variations of PCR. Reverse transcription polymerase chain reaction (RT-PCR) tests determine whether a target is present by amplifying a region of interest of cDNA ([564](#)). Some tests use the results of the PCR itself (e.g., a band on a gel) to determine whether the pathogen is present. However, this approach has not been employed widely in diagnostic testing, and instead most PCR-based tests are quantitative.

3.4.1.1 Quantitative Real-Time PCR

In contrast to RT-PCR, quantitative, real-time PCR uses fluorescent dyes that bind to the amplified DNA, thereby allowing a real time assessment of the amplification procedure ([564](#)) (in this manuscript we refer to quantitative real-time PCR as qPCR, following the Minimum Information for Publication of Quantitative Real-Time PCR Experiments guidelines ([565](#)), and when combined with reverse transcriptase steps, as is required for the evaluation of RNA, it is known as RT-qPCR.) The time resolution provided by qPCR and RT-qPCR is useful because the amount of fluorescence emitted by the sample is proportional to the amount of DNA amplified, and therefore the amount of virus present can be indirectly measured using the cycle threshold (C_t) determined by qPCR.

The first test developed and validated for the detection of SARS-CoV-2 used RT-qPCR to detect several regions of the viral genome: the *ORF1b* of the RNA-dependent RNA polymerase (RdRP), the envelope protein gene (*E*), and the nucleocapsid protein gene (*N*) ([556](#)). The publication reporting this test was released on January 23, 2020, less than two weeks after the sequence of the virus was first reported ([556](#)). In collaboration with several other labs in Europe and in China, the researchers confirmed the specificity of this test

with respect to other coronaviruses against specimens from 297 patients infected with a broad range of respiratory agents. Specifically, this test uses two probes against RdRP, one of which is specific to SARS-CoV-2 (556). Importantly, this assay was not found to return false positive results.

In January 2020, Chinese researchers developed a test that used RT-qPCR to identify two gene regions of the viral genome, *ORF1b* and *N* (566). This assay was tested on samples from two COVID-19 patients and a panel of positive and negative controls consisting of RNA extracted from several cultured viruses. The assay uses the *N* gene to screen patients, while the *ORF1b* gene region is used to confirm the infection (566). The test was designed to detect sequences conserved across sarbecoviruses, or viruses within the same subgenus as SARS-CoV-2. Considering that *Severe acute respiratory syndrome-related coronavirus 1* (SARS-CoV-1) and SARS-CoV-2 are the only sarbecoviruses currently known to infect humans, a positive test can be assumed to indicate that the patient is infected with SARS-CoV-2, although this test is not able to discriminate the genetics of viruses within the sarbecovirus clade. The fact that the targets are so conserved offers the advantage of reduced concern about sensitivity in light of the evolution of SARS-CoV-2.

qPCR tests have played an important role in diagnostics during the COVID-19 pandemic. For SARS-CoV-2, studies have typically considered a patient to be infectious if the C_t is below 33 or sometimes 35 (1, 567, 568). A lower C_t corresponds to fewer qPCR cycles needed to reach a detectable level, indicating that higher amounts of virus were present in the initial reaction. Interpretations of the C_t values obtained from these tests have raised some interesting questions related to viral load and contagiousness. Lower C_t values correspond to a higher probability of a positive viral culture, but no threshold could discriminate all positive from all negative cultures (240). Additionally, because of the variability introduced by sample collection and clinical components of testing, C_t is not a proxy for viral load (569). Positive PCR results have also been reported for extended periods of time from symptom onset and/or the first positive PCR test (275), meaning that in some cases, a positive PCR may not indicate that someone is contagious (1).

In addition to the nuance required to interpret PCR results, there are also factors that influence their accuracy. The specificity of these tests is very high (570), meaning that a positive RT-PCR result is very likely to indicate SARS-CoV-2 infection. The weight given to these tests as an indicator of SARS-CoV-2 infection regardless of other clinical considerations is not typical (571). In fact, while the analytical specificity of the assay is extremely high, the challenges of implementing testing can introduce variability that results in a lower clinical specificity (571). Several factors may influence the sensitivity and specificity, with sample collection being a critically important factor in the reliability of RT-PCR results. The most reliable results were found to come from nasopharyngeal swabs and from pooled nasal and throat swabs, with lower accuracies produced by saliva or by throat or nasal swabs alone (570, 572). Differences in experimental parameters such as the use of primers more specific to SARS-CoV-2 has been found to improve sensitivity in these specimens (573). Additionally, the impact of viral evolution on RT-PCR sensitivity is a concern (574, 575). Using a panel that includes multiple targets can mitigate these effects (576). Additionally, a test designed to incorporate

genomic differences with SARS-CoV-1 was found to offer improved sensitivity and specificity ([573](#)). Thus, while various factors can influence the exact parameters of testing accuracy, RT-PCR is known to have very high specificity and lower, but still high, sensitivity.

3.4.1.2 Digital PCR

Digital PCR (dPCR) is a new generation of PCR technologies offering an alternative to traditional qPCR ([577](#)). In dPCR, a sample is partitioned into thousands of compartments, such as nanodroplets (droplet dPCR or ddPCR) or nanowells, and a PCR reaction takes place in each compartment. This design allows for a digital read-out where each partition is either positive or negative for the nucleic acid sequence being tested for, allowing for absolute target quantification through Poisson statistics. While dPCR equipment is not yet as common as that for qPCR, dPCR for DNA targets generally achieves higher sensitivity than other PCR technologies while maintaining high specificity, though sensitivity is slightly lower for RNA targets ([578](#)).

High sensitivity is particularly relevant for SARS-CoV-2 detection, since low viral load in clinical samples can lead to false negatives. In one study, Suo et al. ([579](#)) performed a double-blind evaluation of ddPCR for SARS-CoV-2 detection. They evaluated on 63 samples collected from suspected positive outpatients and 14 from supposed convalescent patients. Of the 63 outpatients, only 21 (33%) were identified as positive for SARS-CoV-2 with qPCR. However, ddPCR identified 49 (78%) as positive, 10 (16%) as negative, and 4 (6%) as suspected/borderline for SARS-CoV-2 infection. While both qPCR and ddPCR were found to have very high specificity (100%), this analysis reported that the sensitivity was 40% with qPCR compared to 94% with ddPCR. Analysis of serial dilutions of a linear DNA standard suggested that ddPCR was approximately 500 times more sensitive than qPCR ([579](#)). Thus, this study suggests that ddPCR provides an extremely sensitive molecular test that is able to detect SARS-CoV-2 even at very low viral loads.

A second study ([580](#)) confirmed that RT-ddPCR is able to detect SARS-CoV-2 at a lower threshold for viral load relative to RT-PCR. This study analyzed 196 samples, including 103 samples from suspected patients, 77 from contacts and close contacts, and 16 from suspected convalescents, using both RT-qPCR and RT-ddPCR. First, the authors evaluated samples from the 103 suspected cases. Using RT-qPCR, 29 (28%) were identified as positive, 25 (24%) as negative, and 49 (48%) as borderline, i.e., the C_t value was higher than the positive threshold of 35 but lower than the negative threshold of 40. When the 61 negative and borderline samples were reanalyzed with ddPCR, 19 (31%) of the negative and 42 (69%) of the borderline samples were identified as positive. All of the suspected cases were later confirmed to be COVID-19 through a combination of symptom development and RT-qPCR resampling, indicating that ddPCR improved the overall detection rate compared to RT-qPCR from 28.2% to 87.4%.

They repeated this analysis in patient samples from contacts and close contacts. Patients who tested negative with both methods ($n = 48$) were observed to remain healthy over a period of 14 days. Among the remaining 29 samples from contacts, RT-qPCR identified 12 as positive, 1 as negative, and 16 as borderline. All of the samples that tested positive using RT-qPCR

also tested positive using ddPCR. In contrast, the negative result and all but one of the borderline results were identified as positive by RT-ddPCR, and these patients were later determined to be SARS-CoV-2 positive based on clinical evaluation and repeated molecular sampling. Similarly, in the final group, 16 convalescent patients, RT-qPCR identified 12 as positive, three as suspect, and one as negative, but RT-dPCR identified all as positive. The evidence from this study therefore supports a lower limit of detection with ddPCR. Overall, these studies suggest that ddPCR is a promising tool for overcoming the problem of false negatives in SARS-CoV-2 RNA testing, but this method is unlikely to affect the current pandemic due to its lack of availability.

3.4.1.3 Sequencing

In some cases, the DNA amplified with PCR is sequenced. Sequencing requires an additional sample pre-processing step called library preparation. Library preparation is the process of preparing the sample for sequencing, typically by fragmenting the sequences and adding adapters ([581](#)). In some cases, library preparation can involve other modifications of the sample, such as adding barcodes to identify a particular sample within the sequence data. Barcoding can therefore be used to pool samples from multiple sources. There are different reagents used for library preparation that are specific to identifying one or more target sections with PCR ([582](#)). Sequential pattern matching is then used to identify unique subsequences of the virus, and if sufficient subsequences are found, the test is considered positive. Therefore, tests that use sequencing require a number of additional molecular and analytical steps relative to tests that use PCR alone.

Sequencing has been an important strategy for discovery of SARS-CoV-2 variants (e.g., see ([500](#))). Sequencing elucidates any genetic variants located between the PCR primers. For this reason, it is critical to genomic surveillance efforts. Genomic surveillance is an important complement to epidemiological surveillance efforts ([583](#)), as described below. Through genomic surveillance, it has become possible to monitor the emergence of variants of interest and variants of concern (VOC) that may pose additional threats due to increased contagiousness, virulence, or immune escape ([583](#), [584](#)). Sequencing also allows for analysis of the dominant strains in an area at a given time. Worldwide, the extent of genomic surveillance varies widely, with higher-income countries typically able to sequence a higher percentage of cases ([585](#)). Sequencing efforts are important for identifying variants containing mutations that might affect the reliability of molecular diagnostic tests, as well as mitigation measures such as therapeutics and prophylactics ([574](#), [575](#)). Therefore, sequencing is an important component of diagnostics: while it is not necessary for diagnosing an individual case, it is critical to monitoring trends in the variants affecting a population and to staying aware of emerging variants that may pose additional challenges.

3.4.1.4 Pooled and Automated PCR Testing

Due to limited supplies and the need for more tests, several labs have found ways to pool or otherwise strategically design tests to increase throughput. The first such result came from Yelin et al. ([586](#)), who reported that they could pool up to 32 samples in a single qPCR run. This was followed by

larger-scale pooling with slightly different methods ([587](#)). Although these approaches are also PCR based, they allow for more rapid scaling and higher efficiency for testing than the initial PCR-based methods developed. Conceptually, pooling could also be employed in analysis with RT-qPCR ([588](#)), and this strategy has been evaluated in settings such as schools ([589](#)) and hospitals ([590](#)).

3.4.2 RT-LAMP

RT-PCR remains the gold standard for detection of SARS-CoV-2 RNA from infected patients, but the traditional method requires special equipment and reagents, including a thermocycler. Loop-mediated isothermal amplification (LAMP) is an alternative to PCR that does not require specialized equipment ([591](#)). In this method, nucleic acids are amplified in a 25 μ L reaction that is incubated and chilled on ice ([591](#)). It uses primers designed to facilitate auto-cycling strand displacement DNA synthesis ([591](#)). LAMP can be combined with reverse transcription (RT-LAMP) to enable the detection of RNA.

One study showed that RT-LAMP is effective for detection of SARS-CoV-2 with excellent specificity and sensitivity and that this method can be applied to unprocessed saliva samples ([592](#)). This method was benchmarked against RT-PCR using 177 human nasopharyngeal RNA samples, of which 126 were COVID positive. The authors break down the sensitivity of their test according to the C_t value from RT-PCR of the same samples; RT-LAMP performs at 100% sensitivity for samples with a C_t from RT-PCR of 32 or less. The performance is worse when considering all RT-PCR positive samples (including those with C_t values between 32-40). However, there is some evidence suggesting that samples obtained from individuals that achieve C_t values >30 measured using RT-PCR tend to be less infective than those that record a C_t value <30 ([593–595](#)), so RT-LAMP is still a useful diagnostic tool. Various combinations of reagents are available, but one example is the WarmStart Colorimetric LAMP 2X Master Mix with a set of six primers developed previously by Zhang et al. ([596](#)). To determine assay sensitivity, serial tenfold dilutions of *in vitro* transcribed *N*-gene RNA standard were tested using quantities from 10^5 copies down to 10 copies. The assay readout is the color of the dye changing from pink to yellow due to binding to the DNA product over 30 minutes. The RT-LAMP assay was then applied to clinical nasopharyngeal samples. For viral loads above 100 copies of genomic RNA, the RT-LAMP assay had a sensitivity of 100% and a specificity of 96.1% from purified RNA. The sensitivity of the direct assay of saliva by RT-LAMP was 85%. Sensitivity and specificity metrics were obtained by comparison with results from RT-PCR. RT-LAMP pilot studies for detection of SARS-CoV-2 were reviewed in a meta-analysis ([597](#)). In the meta-analysis of all 2,112 samples, the cumulative sensitivity of RT-LAMP was calculated at 95.5%, and the cumulative specificity was 99.5%.

This test aims to bring the sensitivity of nucleic acid detection to the point of care or home testing setting. It could be applied for screening, diagnostics, or as a definitive test for people who are positive based on LFTs (see below). The estimated cost per test is about 2 euros when RNA extraction is included. The main strength of this test over RT-PCR is that it can be done isothermally, but the main drawback is that it is about 10-fold less sensitive than RT-PCR. The low cost, excellent sensitivity/specificity, and quick readout of RT-LAMP

makes this an attractive alternative to RT-PCR. Alternative strategies like RT-LAMP are needed to bring widespread testing away from the lab and into under-resourced areas.

3.4.3 CRISPR-based Detection

Technology based on CRISPR (clustered regularly interspaced short palindromic repeats) ([598](#)) has also been instrumental in scaling up testing protocols. Two CRISPR-associated nucleases, Cas12 and Cas13, have been used for nucleic acid detection. Multiple assays exploiting these nucleases have emerged as potential diagnostic tools for the rapid detection of SARS-CoV-2 genetic material and therefore SARS-CoV-2 infection. The SHERLOCK method (Specific High-sensitivity Enzymatic Reporter unLOCKing) from Sherlock Biosciences relies on Cas13a to discriminate between inputs that differ by a single nucleotide at very low concentrations ([599](#)). The target RNA is amplified by real-time recombinase polymerase amplification (RT-RPA) and T7 transcription, and the amplified product activates Cas13a. The nuclease then cleaves a reporter RNA, which liberates a fluorescent dye from a quencher. Several groups have used the SHERLOCK method to detect SARS-CoV-2 viral RNA. An early study reported that the method could detect 7.5 copies of viral RNA in all 10 replicates, 2.5 copies in 6 out of 10, and 1.25 copies in 2 out of 10 runs ([600](#)). It also reported 100% specificity and sensitivity on 114 RNA samples from clinical respiratory samples (61 suspected cases, among which 52 were confirmed and nine were ruled out by metagenomic next-generation sequencing, 17 SARS-CoV-2-negative but human coronavirus (HCoV)-positive cases, and 36 samples from healthy subjects) and a reaction turnaround time of 40 minutes. A separate study screened four designs of SHERLOCK and extensively tested the best-performing assay. They determined the limit of detection to be 10 copies/ μ l using both fluorescent and lateral flow detection ([601](#)).

LFT strips are simple to use and read, but there are limitations in terms of availability and cost per test. Another group therefore proposed the CREST (Cas13-based, Rugged, Equitable, Scalable Testing) protocol, which uses a P51 cardboard fluorescence visualizer, powered by a 9-volt battery, for the detection of Cas13 activity instead of immunochromatography ([602](#)). CREST can be run, from RNA sample to result, with no need for AC power or a dedicated facility, with minimal handling in approximately 2 hours. Testing was performed on 14 nasopharyngeal swabs. CREST picked up the same positives as the CDC-recommended TaqMan assay with the exception of one borderline sample that displayed low-quality RNA. This approach may therefore represent a rapid, accurate, and affordable procedure for detecting SARS-CoV-2.

The DETECTR (DNA Endonuclease-Targeted CRISPR Trans Reporter) method from Mammoth Biosciences involves purification of RNA extracted from patient specimens, amplification of extracted RNAs by loop-mediated amplification, and application of their Cas12-based technology. In this assay, guide RNAs (gRNAs) were designed to recognize portions of sequences corresponding to the SARS-CoV-2 genome, specifically the N2 and E regions ([603](#)). In the presence of SARS-CoV-2 genetic material, sequence recognition by the gRNAs results in double-stranded DNA cleavage by Cas12, as well as cleavage of a single-stranded DNA molecular beacon. The cleavage of this

molecular beacon acts as a colorimetric reporter that is subsequently read out in a lateral flow assay and indicates the presence of SARS-CoV-2 genetic material and therefore SARS-CoV-2 infection. The 40-minute assay is considered positive if there is detection of both the *E* and *N* genes or presumptive positive if there is detection of either of them. The assay had 95% positive predictive agreement and 100% negative predictive agreement with the US Centers for Disease Control and Prevention SARS-CoV-2 RT-qPCR assay. The estimated limit of detection was 10 copies per μl reaction, versus 1 copy per μl reaction for the CDC assay.

These results have been confirmed by other DETECTR approaches. Using RT-RPA for amplification, another group detected 10 copies of synthetic SARS-CoV-2 RNA per μl of input within 60 minutes of RNA sample preparation in a proof-of-principle evaluation (604). Through a similar approach, another group reported detection at 1 copy per μl (605). The DETECTR protocol was improved by combining RT-RPA and CRISPR-based detection in a one-pot reaction that incubates at a single temperature and by using dual CRISPR RNAs, which increases sensitivity. This new assay, known as All-In-One Dual CRISPR-Cas12a, detected 4.6 copies of SARS-CoV-2 RNA per μl of input in 40 minutes (606). Another single-tube, constant-temperature approach using Cas12b instead of Cas12a achieved a detection limit of 5 copies/ μl in 40-60 minutes (607).

It was also reported that electric field gradients can be used to control and accelerate CRISPR assays by co-focusing Cas12-gRNA, reporters, and target (608). The authors generated an appropriate electric field gradient using a selective ionic focusing technique known as isotachopheresis (ITP) implemented on a microfluidic chip. They also used ITP for automated purification of target RNA from raw nasopharyngeal swab samples. Combining this ITP purification with loop-mediated isothermal amplification, their ITP-enhanced assay achieved detection of SARS-CoV-2 RNA (from raw sample to result) in 30 minutes.

All these methods require upstream nucleic acid amplification prior to CRISPR-based detection. They rely on type V (Cas12-based) and type IV (Cas13-based) CRISPR systems. In contrast, type III CRISPR systems have the unique property of initiating a signaling cascade, which could boost the sensitivity of direct RNA detection. In type III CRISPR systems, guide CRISPR RNAs (crRNAs) are bound by several Cas proteins (609) and can target both DNA and RNA molecules (610, 611). A study tested this hypothesis using the type III-A crRNA-guided surveillance complex from *Thermus thermophilus* (612). The authors showed that activation of the Cas10 polymerase generates three products (cyclic nucleotides, protons, and pyrophosphates) that can all be used to detect SARS-CoV-2 RNA. Detection of viral RNA in patient samples still required an initial nucleic acid amplification step, but improvements may in the future remove that requirement.

This goal of amplification-free detection was later achieved for a Cas13a-based system (613). This approach combined multiple CRISPR RNAs to increase Cas13a activation, which is detected by a fluorescent reporter. Importantly, because the viral RNA is detected directly, the test yields a quantitative measurement rather than a binary result. The study also shows that fluorescence can be measured in a custom-made dark box with a mobile

phone camera and a low-cost laser illumination and collection optics. This approach is a truly portable assay for point-of-care diagnostics. The authors achieved detection of 100 copies/μl of pre-isolated RNA in 30 minutes, and correctly identified all SARS-CoV-2-positive patient RNA samples tested in 5 minutes (n = 20).

There is an increasing body of evidence that CRISPR-based assays offer a practical solution for rapid, low-barrier testing in areas that are at greater risk of infection, such as airports and local community hospitals. In the largest study to date, DETECTR was compared to RT-qPCR on 378 patient samples (614). The authors reported 95% reproducibility. Both techniques were equally sensitive in detecting SARS-CoV-2. Lateral flow strips showed 100% correlation to the high-throughput DETECTR assay. Importantly, DETECTR was 100% specific for SARS-CoV-2 and did not detect other human coronaviruses. A method based on a Cas9 ortholog from *Francisella novicida* known as FnCas9 achieved 100% sensitivity and 97% specificity in clinical samples, and the diagnostic kit is reported to have completed regulatory validation in India (615).

3.4.4 Immunoassays for the Detection of Antigens

Immunoassays can detect molecular indicators of SARS-CoV-2 infection, such as the proteins that act as antigens from the SARS-CoV-2 virus. They offer the advantage of generally being faster and requiring less specialized equipment than other molecular tests, especially those involving PCR. As a result, immunoassays hold particular interest for implementation at home and in situations where resources for PCR testing are limited. The trade-off is that these tests typically have a lower sensitivity, and sometimes a lower specificity, than other molecular tests. However, these tests tend to return a positive result five to 12 days after symptom onset, which may therefore correlate more closely with the timeframe during which viral replication occurs (616). Immunoassays for the detection of the SARS-CoV-2 antigen can include LFTs and ELISA, as discussed here, as well as CLIA and chromatographic immunoassays (617), as described in the serological testing section below.

3.4.4.1 Lateral Flow Tests

LFTs provide distinct value relative to PCR tests. They can return results within 30 minutes and can be performed without specialized equipment and at low cost. They also do not require training to operate and are cheap to produce. Thus, they can be distributed widely to affected populations making them an important public health measure to curb pandemic spread. LFTs rely on the detection of viral protein with an antibody. Often this is done with an antibody sandwich format, where one antibody conjugated to a dye binds at one site on the antigen, and an immobilized antibody on the strip binds at another site (558). This design allows the dye to accumulate to form a characteristic positive test line on the strip (558). Outside of COVID-19 diagnostics, the applications of LFTs are broad; they are routinely used for home pregnancy tests, disease detection, and even drugs of abuse detection in urine (618).

A recent review surveyed the performance of LFTs for detection of current SARS-CoV-2 infection ([619](#)). This review covered 24 studies that included more than 26,000 total LFTs. They reported significant heterogeneity in test sensitivities, with estimates ranging from 37.7% to 99.2%. The estimated specificities of these tests were more homogeneous, spanning 92.4% to 100.0%.

Despite having lower sensitivity than PCR tests, LFTs occupy an important niche in the management of SARS-CoV-2. Current infection detection by LFTs enables the scale and speed of testing that is beneficial to managing viral spread. LFTs were available freely to citizens in the United Kingdom until April 1, 2022 ([620](#)) and to citizens of the United States in early 2022 ([621](#)). These tests are particularly useful for ruling out SARS-CoV-2 infection in cases where the likelihood of infection is low (e.g., asymptomatic individuals) and positives (including false positives) can be validated with testing by alternate means ([622](#)).

3.4.4.2 Enzyme-Linked Immunosorbent Assay

ELISA is a very sensitive immunoassay that can be considered a gold standard for the detection of biological targets, including antibodies and antigenic proteins ([559](#)). It can be used to generate either quantitative or qualitative results that can be returned within a few hours ([623](#)). ELISA builds on the idea that antibodies and antigens bind together to form complexes ([559](#)) and utilizes an enzyme covalently linked to an antibody against the antigen to produce assay signal, usually a color change ([624](#)). The main advantage of ELISA is that it enables signal amplification through the enzyme's activity, which increases sensitivity. With sandwich ELISA, antibodies are immobilized on a surface such as a plate, and viral protein antigens in the sample bind and are retained ([625](#)). A second antibody is added that binds to another site on the antigen is then added, and that second antibody is covalently linked to an enzyme. A substrate for that enzyme is then added to produce signal, usually light or a color change. The exact strategy for tagging with a reporter enzyme varies among different types of ELISA ([559](#), [625](#)). For COVID-19 diagnostics, ELISAs have been designed to detect the antigenic Spike protein ([626](#)).

One of these assays uses two monoclonal antibodies specific to the nucleocapsid of SARS-CoV-2 to evaluate the relationship between the effect of (estimated) viral load on the ability of the assay to detect the SARS-CoV-2 antigen ([627](#)). This study analyzed 339 naso-oropharyngeal samples that were also analyzed with RT-qPCR as a gold standard. RT-qPCR identified 147 samples as positive and 192 as negative. The authors estimated the overall sensitivity and specificity to be 61.9% and 99.0%, respectively. Sensitivity increased with higher C_t . This study also assessed the performance of the ELISA test under different conditions in order to evaluate how robust it would be to the challenges of testing in real-world settings globally. Higher sensitivity was achieved for samples that were stored under ideal conditions (immediate placement in -80°C). Therefore, while immediate access to laboratory equipment is an advantage, it is not strictly necessary for ELISA to detect the antigen.

3.4.5 Limitations of Molecular Tests

Tests that identify SARS-CoV-2 using molecular technologies will identify only individuals with current infections and are not appropriate for identifying individuals who have recovered from a previous infection. Among molecular tests, different technologies have different sensitivities and specificities. In general, specificity is high, and even then, the public health repercussions of a false positive can generally be mitigated with follow-up testing. On the other hand, a test's sensitivity, which indicates the risk of a false-negative response, can pose significant challenge to large-scale testing. False negatives are a significant concern for several reasons. Importantly, clinical reports indicate that it is imperative to exercise caution when interpreting the results of molecular tests for SARS-CoV-2 because negative results do not necessarily mean a patient is virus-free ([628](#)). To reduce occurrence of false negatives, correct execution of the analysis is crucial ([629](#)). Additionally, PCR-based tests can remain positive for a much longer time than the virus is likely to be actively replicating ([616](#)), raising concerns about their informativeness after the acute phase of the disease. Hence, the CDC has advised individuals who suspect they have been re-infected with SARS-CoV-2 to avoid using diagnostic tests within 90 days of receiving a previous positive test ([630](#)).

Additionally, the emerging nature of the COVID-19 pandemic has introduced some challenges related to uncertainty surrounding interactions between SARS-CoV-2 and its human hosts. For example, viral shedding kinetics are still not well understood but are expected to introduce a significant effect of timing of sample collection on test results ([629](#)). Similarly, the type of specimen could also influence outcomes, as success in viral detection varies among clinical sample types ([570](#), [572](#), [629](#)). With CRISPR-based testing strategies, the gRNA can recognize off-target interspersed sequences in the viral genome ([631](#)), potentially resulting in false positives and a loss of specificity.

There are also significant practical and logistical concerns related to the widespread deployment of molecular tests. Much of the technology used for molecular tests is expensive, and while it might be available in major hospitals and/or diagnostic centers, it is often not available to smaller facilities ([632](#)). At times during the pandemic, the availability of supplies for testing, including swabs and testing media, has also been limited ([633](#)). Similarly, processing times can be long, and tests might take up to 4 days to return results ([632](#)), especially during times of high demand, such as spikes in case numbers ([634](#)). Countries have employed various and differing molecular testing strategies as a tool to reduce viral transmission, even among high-income countries ([635](#)). The rapid development of molecular tests has provided a valuable, albeit imperfect, tool to identify active SARS-CoV-2 infections.

3.5 Serological Tests to Identify Recovered Individuals

Although several molecular diagnostic tests to detect viral genetic material have high specificity and sensitivity, they provide information only about active infection, and therefore offer just a snapshot-in-time perspective on

the spread of a disease. Most importantly, they would not work on a patient who has fully recovered from the virus at the time of sample collection. In such contexts, serological tests are informative.

Serological tests use many of the same technologies as the immunoassays used to detect the presence of an antigen but are instead used to evaluate the presence of antibodies against SARS-CoV-2 in a serum sample. These tests are particularly useful for insight into population-level dynamics and can also offer a glimpse into the development of antibodies by individual patients during the course of a disease. Immunoassays can detect antibodies produced by the adaptive immune system in response to viral threat. Understanding the acquisition and retention of antibodies is important both to the diagnosis of prior (inactive) infections and to the development of vaccines. The two immunoglobulin classes that are most pertinent to these goals are immunoglobulin M (IgM), which are the first antibodies produced in response to an infection, and immunoglobulin G (IgG), which are the most abundant antibodies ([636](#), [637](#)). Serological tests detect these antibodies, offering a mechanism through which prior infection can be identified. However, the complexity of the human immune response means that there are many facets to such analyses.

In general, SARS-CoV-2 infection will induce the immune system to produce antibodies fairly quickly. Prior research is available about the development of antibodies to SARS-CoV-1 during the course of the associated disease, severe acute respiratory syndrome (SARS). IgM and IgG antibodies were detected in the second week following SARS-CoV-1 infection. IgM titers peaked by the first month post-infection, and then declined to undetectable levels after day 180. IgG titers peaked by day 60 and persisted in all donors through the two-year duration of study ([638](#)). Such tests can also illuminate the progression of viral disease, as IgM are the first antibodies produced by the body and indicate that the infection is active. Once the body has responded to the infection, IgG are produced and gradually replace IgM, indicating that the body has developed immunogenic memory ([639](#)). Therefore, it was hoped that the development of assays to detect the presence of IgM and IgG antibodies against SARS-CoV-2 would allow the identification of cases from early in the infection course (via IgM) and for months or years afterwards (via IgG). Several technologies have been used to develop serological tests for COVID-19, including ELISA, lateral flow immunoassay, chemiluminescence immunoassay, and neutralizing antibody assays ([640](#)).

3.5.1 ELISA

The application of ELISA to serological testing is complementary to its use in molecular diagnostics (see above). Instead of using an enzyme-labeled antibody as a probe that binds to the target antigen, the probe is an antigen and the target is an antibody. The enzyme used for detection and signal amplification is on a secondary antibody raised generally against human IgG or IgM. In March 2020, the Krammer lab proposed an ELISA test that detects IgG and IgM that react against the receptor-binding domain (RBD) of the spike proteins (S) of the virus ([641](#)). A subsequent ELISA test developed to detect SARS-CoV-2 IgG based on the RBD reported a specificity of over 99% and a sensitivity of up to 88.24%, which was observed in samples collected 21 to 27 days after the onset of infection (approximated with symptom onset or

positive PCR test) ([642](#)). Earlier in the disease course, sensitivity was lower: 53.33% between days 0 and 13 and 80.47% between days 14 and 20. This study reported that their laboratory ELISA outperformed two commercial kits that also used an ELISA design ([642](#)). Therefore, while analysis with ELISA requires laboratory support and equipment, these results do suggest that ELISA achieves relatively high sensitivity, especially in the weeks following infection. Efforts have been made to develop low-cost strategies for conducting these tests that will make them more accessible worldwide ([643](#)).

3.5.2 Chemiluminescence Immunoassay

Another early approach investigated for detection of antibodies against SARS-CoV-2 was CLIA. Like ELISA, CLIA is a type of enzyme immunoassay (EIA) ([644](#)). While the technique varies somewhat, in one approach, a bead is coated with the antigen and then washed with the sample ([645](#)). If the antibody is present in the sample, it will bind to the bead. Then the bead is exposed to a label, a luminescent molecule that will bind to the antigen/antibody complex and can therefore be used as an indicator ([645](#)). One CLIA approach to identify COVID-19 used a synthetic peptide derived from the amino acid sequence of the SARS-CoV-2 S protein ([646](#)). It was highly specific to SARS-CoV-2 and detected IgM in 57.2% and IgG in 71.4% of serum samples from 276 COVID-19 cases confirmed with RT-qPCR. IgG could be detected within two days of the onset of fever, but IgM could not be detected any earlier ([646](#)), which has been supported by other analyses as well ([647](#)). This pattern was consistent with observations in Middle East respiratory syndrome, which is also caused by an HCoV. In comparisons of different commercial immunoassays, accuracy of CLIA tests were often roughly comparable to other EIAs ([648](#)), although one CLIA did not perform as well as several other EIAs ([647](#), [649](#)). The sensitivity and specificities reported vary among CLIA tests and for the detection of IgM versus IgG, but sensitivities and specificities as high as 100% have been reported among various high-throughput tests ([649–651](#)). CLIA has previously been used to develop tests that can be used at point of care (e.g., ([644](#))) which may allow for this technique to become more widely accessible in the future.

3.5.3 Lateral Flow Immunoassay

The first serological test approved for emergency use in the United States was developed by Cellex ([652](#)). The Cellex qSARS-CoV-2 IgG/IgM Rapid Test is a chromatographic immunoassay, also known as a lateral flow immunoassay, designed to qualitatively detect IgM and IgG antibodies against SARS-CoV-2 in the plasma of patients suspected to have developed a SARS-CoV-2 infection ([652](#)). The Cellex test cassette contains a pad of SARS-CoV-2 antigens and a nitrocellulose strip with lines for each of IgG and IgM, as well as a control (goat IgG) ([652](#)). In a specimen that contains antibodies against the SARS-CoV-2 antigen, the antibodies will bind to the strip and be captured by the IgM and/or IgG line(s), resulting in a change of color ([652](#)). With this particular assay, results can be read within 15 to 20 minutes ([652](#)). Lateral flow immunoassays are often available at point of care but can have very low sensitivity ([649](#)).

3.5.4 Neutralizing Antibody Assays

Neutralizing antibody assays play a functional role in understanding immunity that distinguishes them from other serological tests. The tests described above are all binding antibody tests. On the other hand, rather than simply binding an antibody to facilitate detection, neutralizing antibody assays determine whether an antibody response is present that would prevent infection (653, 654). Therefore, these tests serve the purpose of evaluating the extent to which a sample donor has acquired immunity that will reduce susceptibility to SARS-CoV-2. As a result, neutralizing antibody assays have been used widely to characterize the duration of immunity following infection, to assess vaccine candidates, and to establish correlates of protection against infection and disease (655–657). These tests are typically performed in a laboratory (653), and in SARS-CoV-2, the results of neutralizing antibody assays are often correlated with the results of binding antibody tests (653).

The gold standard for assessing the presence of neutralizing antibodies is the plaque reduction neutralization test (PRNT), but this approach does not scale well (654). An early high-throughput neutralizing antibody assay designed against SARS-CoV-2 used a fluorescently labeled reporter virus that was incubated with different dilutions of patient serum (654). The cells used for incubation would turn green if antibodies were not present. Essentially, this assay evaluates whether the virus is able to infect the cell in the presence of the serum. The specificity of this assay was 100%, and the correlation between the results of this assay and of PRNT was 0.85 with the results suggesting that the sensitivity of the high-throughput approach was higher than that of PRNT (654). While this approach was performed on a plate and using cells, other methods have been developed using methods such as bead arrays (658).

3.5.5 Duration of Immune Indicators

While the adaptive immune system produces antibodies in response to SARS-CoV-2 viral challenge, these indicators of seroconversion are unlikely to remain in circulation permanently. Previously, a two-year longitudinal study following convalesced SARS patients with a mean age of 29 found that IgG antibodies were detectable in all 56 patients surveyed for at least 16 months and remained detectable in all but 4 patients (11.8%) through the full two-year study period (659). These results suggest that immunity to SARS-CoV-1 is sustained for at least a year. Circulating antibody titers to other coronaviruses have been reported to decline significantly after one year (660). Evidence to date suggests that sustained immunity to the SARS-CoV-2 virus remains for a shorter period of time but at least 6 to 8 months after infection (661–664). However, this does not mean that all serological evidence of infection dissipates, but rather that the immune response becomes insufficient to neutralize the virus.

In order to study the persistence of SARS-CoV-2 antibodies, one study assessed sustained immunity using 254 blood samples from 188 COVID-19 positive patients (662). The samples were collected at various time points between 6 and 240 days post-symptom onset; some patients were assessed longitudinally. Of the samples, 43 were collected at least 6 months after symptom onset. After one month, 98% of patients were seropositive for IgG to S. Moreover, S IgG titers were stable and heterogeneous among patients

over a period of 6 to 8 months post-symptom onset, with 90% of subjects seropositive at 6 months. Similarly, at 6 to 8 months 88% of patients were seropositive for RBD IgG, and 90% were seropositive for SARS-CoV-2 neutralizing antibodies. Another study examined 119 samples from 88 donors who had recovered from mild to severe cases of COVID-19 (664). A relatively stable level of IgG and plasma neutralizing antibodies was identified up to 6 months post diagnosis. Significantly lower but considerable levels of anti-SARS-CoV-2 IgG antibodies were still present in 80% of samples obtained 6 to 8 months post-symptom onset.

Titers of IgM and IgG antibodies against the RBD were found to decrease from 1.3 to 6.2 months post infection in a study of 87 individuals (665). However, the decline of IgA activity (15%) was less pronounced than that of IgM (53%) or IgG (32%). It was noted that higher levels of anti-RBD IgG and anti-N total antibodies were detected in individuals that reported persistent post-acute symptoms at both study visits. Moreover, plasma neutralizing activity decreased five-fold between 1.3 and 6.2 months in an assay of HIV-1 virus pseudotyped with SARS-CoV-2 S protein, and this neutralizing activity was directly correlated with IgG anti-RBD titers (665). These findings are in accordance with other studies that show that the majority of seroconverters have detectable, albeit decreasing, levels of neutralizing antibodies at least 3 to 6 months post infection (666–668).

Determining the potency of anti-RBD antibodies early in the course of an infection may be important moving forward, as their neutralizing potency may be prognostic for disease severity and survival (669). The duration of immunity might also vary with age (670) or ABO blood type (671). Autopsies of lymph nodes and spleens from severe acute COVID-19 patients showed a loss of T follicular helper cells and germinal centers that may explain some of the impaired development of antibody responses (672). Therefore, serological testing may be time-limited in its ability to detect prior infection.

Other immune indicators of prior infection have also been evaluated to see how they persist over time. SARS-CoV-2 memory CD8⁺ T cells were slightly decreased (50%) 6 months post-symptom onset. In this same subset of COVID-19 patients, 93% of subjects had detectable levels of SARS-CoV-2 memory CD4⁺ T cells, of which 42% had more than 1% SARS-CoV-2-specific CD4⁺ T cells. At 6 months, 92% of patients were positive for SARS-CoV-2 memory CD4⁺ T cells. Indeed, the abundance of S-specific memory CD4⁺ T cells over time was similar to that of SARS-CoV-2-specific CD4⁺ T cells overall (662). T cell immunity to SARS-CoV-2 at 6 to 8 months following symptom onset has also been confirmed by other studies (664, 673, 674). In another study, T cell reactivity to SARS-CoV-2 epitopes was also detected in some individuals never been exposed to SARS-CoV-2. This finding suggests the potential for cross-reactive T cell recognition between SARS-CoV-2 and pre-existing circulating HCoV that are responsible for the “common cold” (675), but further research is required. Therefore, whether T cells will provide a more stable measure through which to assess prior infection remains unknown. Notably, commercial entities have tried to develop tests specifically for T cells, some of which have been authorized by the United States Food and Drug Administration (676, 677) to identify people with adaptive T cell immune responses to SARS-CoV-2, either from a previous or ongoing infection.

3.5.6 Applications of Serological Tests

In addition to the limitations posed by the fact that antibodies are not permanent indicators of prior infection, serological immunoassays carry a number of limitations that influence their utility in different situations. Importantly, false positives can occur due to cross-reactivity with other antibodies according to the clinical condition of the patient (652). Due to the long incubation times and delayed immune responses of infected patients, serological immunoassays are insufficiently sensitive for a diagnosis in the early stages of an infection. Therefore, such tests must be used in combination with RNA detection tests if intended for diagnostic purposes (678). False positives are particularly harmful if they are erroneously interpreted to mean that a population is more likely to have acquired immunity to a disease (679). Similarly, while serological tests may be of interest to individuals who wish to confirm they were infected with SARS-CoV-2 in the past, their potential for false positives means that they are not currently recommended for this use. However, in the wake of vaccines becoming widely available, accurate serological tests that could be administered at point of care were investigated in the hope that they could help to prioritize vaccine recipients (680). Another concern with serological testing is the potential for viral evolution to reduce the sensitivity of assays, especially for neutralizing antibody assays. Chen et al. performed a systematic re-analysis of published data examining the neutralizing effect of serum from vaccinated or recovered individuals on four VOC (681). They found reduced neutralizing titers against these variants relative to the lineages used for reference. These findings suggest that such techniques will need to be modified over time as SARS-CoV-2 evolves.

These limitations make serological tests far less useful for diagnostics and for test-and-trace strategies; however, serological testing is valuable for public health monitoring at the population level. Serosurveys provide a high-level perspective of the prevalence of a disease and can provide insight into the susceptibility of a population as well as variation in severity, e.g., between geographic regions (679). From a public health perspective, they can also provide insight into the effectiveness of mitigation efforts and to gain insight into risk factors influencing susceptibility (682). EIA methods are high-throughput (683, 684), and, as with molecular tests, additional efforts have been made to scale up the throughput of serological tests (685). Therefore, serological tests can be useful to developing strategies for the management of viral spread.

Early in the course of the pandemic, it was also hoped that serological tests would provide information relevant to advancing economic recovery. Some infectious agents can be controlled through “herd immunity”, which is when a critical mass within the population acquires immunity through vaccination and/or infection, preventing an infectious agent from spreading widely. It was hoped that people who had recovered and developed antibodies might be able to return to work (686, 687). This strategy would have relied on recovered individuals acquiring long-term immunity, which has not been borne out (688). Additionally, it was hoped that identifying seroconverters and specifically those who had mounted a strong immune response would reveal strong candidates for convalescent plasma donation (641); however,

convalescent plasma has not been found to offer therapeutic benefit (reviewed in [\(3\)](#)). While these hopes have not been realized, serological tests have been useful for gaining a better understanding of the pandemic ([682](#)).

3.6 Possible Alternatives to Current Diagnostic Practices

One possible alternative or complement to molecular and serological testing would be diagnosing COVID-19 cases based on symptomatology. COVID-19 can present with symptoms similar to other types of pneumonia, and symptoms can vary widely among COVID-19 patients; therefore, clinical presentation is often insufficient as a sole diagnostic criterion. In addition, identifying and isolating mild or asymptomatic cases is critical to efforts to manage outbreaks. Even among mildly symptomatic patients, a predictive model based on clinical symptoms had a sensitivity of only 56% and a specificity of 91% ([689](#)). More problematic is that clinical symptom-based tests are only able to identify already symptomatic cases, not presymptomatic or asymptomatic cases. They may still be important for clinical practice and for reducing tests needed for patients deemed unlikely to have COVID-19.

In some cases, clinical signs may also provide information that can inform diagnosis. Using computed tomography of the chest in addition to RT-qPCR testing was found to provide a higher sensitivity than either measure alone ([690](#)). X-ray diagnostics have been reported to have high sensitivity but low specificity in some studies ([691](#)). Other studies have shown that specificity varies between radiologists ([692](#)), though the sensitivity reported here was lower than that published in the previous paper. While preliminary machine-learning results suggested that chest X-rays might provide high sensitivity and specificity and potentially facilitate the detection of asymptomatic and presymptomatic infections (e.g., ([693](#))), further investigation suggested that such approaches are prone to bias and are unlikely to be clinically useful ([694](#)). Given the above, the widespread use of X-ray tests on otherwise healthy adults is likely inadvisable.

Finally, in addition to genomic and serological surveillance, other types of monitoring have proven useful in managing the pandemic ([695](#)). One that has received significant attention is wastewater surveillance. This approach can use several of the technologies described for molecular testing, such as qPCR and dPCR, as well as *in vitro* culturing ([696](#)) and can provide insight into trends in the prevalence of SARS-CoV-2 regionally.

3.7 Strategies and Considerations for Testing

Deciding whom to test, when to test, and which test to use have proven challenging as the COVID-19 pandemic has unfolded. Early in the COVID-19 pandemic, testing was typically limited to individuals considered high risk for developing serious illness ([697](#)). This approach often limited testing to people with severe symptoms and people showing mild symptoms that had been in contact with a person who had tested positive. Individuals who were asymptomatic (i.e., potential spreaders) and individuals who were able to

recover at home were thus often unaware of their status. Therefore, this method of testing administration misses a high proportion of infections and does not allow for test-and-trace methods to be used. For instance, a study from Imperial College estimates that in Italy, the true number of infections was around 5.9 million in a total population of ~60 million, compared to the 70,000 detected as of March 28, 2020 (314). Another analysis, which examined New York state, indicated that as of May 2020, approximately 300,000 cases had been reported in a total population of approximately 20 million (698). This corresponded to ~1.5% of the population, but ~12% of individuals sampled statewide were estimated as positive through antibody tests (along with indications of spatial heterogeneity at higher resolution) (698). Technological advancements that facilitate widespread, rapid testing would therefore be valuable for accurately assessing the rate of infection and aid in controlling the virus' spread. Additionally, the trade off of accessibility, sensitivity, and time to results has raised some complex questions around which tests are best suited to certain situations. Immunoassays, including serological tests, have much higher limits of detection than PCR tests do (699).

Changes in public attitudes and the lifting of COVID-19 restrictions due to the multifactorial desire to stimulate economic activities has required a shift of testing paradigms in 2022, despite warnings from public health officials against a hard exit from public health restrictions (700, 701). An important strategy for testing moving forward is to determine when someone becomes infectious or is no longer infectious following a positive test for COVID-19. Generally, patient specimens tend to not contain culturable virus past day 5 of symptom onset (702, 703). However, due to their sensitivity to post-infectious viral RNA in specimens, PCR-based methods may mislead individuals to believe that they are still infectious several days after symptom onset (678). Furthermore, detection of viral RNA can occur days and weeks after an active infection due to the sensitivity of PCR-based methods (568, 704, 705).

In contrast, LFTs were thought to have poor sensitivity and their value for identifying infections and managing the pandemic was questioned (706, 707). However, LFTs do reliably detect SARS-CoV-2 proteins when there is a high viral load, which appears to correlate with a person's infectiousness (616, 708). Therefore, LFTs are an important diagnostic tool to determine infectiousness with fast turnaround times, ease of use, and accessibility by the general public (678, 709). One study has suggested that the test sensitivity of LFTs appears to be less important than accessibility to LFTs, frequent testing, and fast reporting times for reducing the impact of viral spread (710). While PCR-based methods are important for COVID-19 surveillance, their use is labor intensive and time consuming, and laboratories are often slow to report results, rendering such methods limited in their surveillance capacity (678).

These limitations are demonstrated by the estimated 10-fold under-reporting of cases in the United States in 2020 due to shortages in testing and slow rollout of testing and slow reporting of results (711). However, one strategy that may balance the strengths and weaknesses of both types of tests is to corroborate a positive LFT result using a PCR-based method. Indeed, in

December 2021 sufficient surveillance and reduction of COVID-19 spread using this joint LFT-PCR strategy was demonstrated in Liverpool, U.K., where there was an estimated 21% reduction of cases ([709](#), [712](#)).

3.8 What Lies Ahead

Diagnostic tools have played an important role during the COVID-19 pandemic. Different tests offer different advantages (Figure 2). Specifically, the results of SARS-CoV-2 diagnostic tests (typically qPCR or LFT-based tests) have been used to estimate the number of infections in the general population, thus informing public health strategies around the globe ([574](#)). During the surges caused by the different SARS-CoV-2 variants between 2020 and 2021, government-sponsored efforts to conduct mass testing and to provide free diagnostic tests to the population were a common occurrence in many parts of the world ([713–715](#)). However, recent reports indicate that such public health policies are starting to change during 2022. For example, it is known that the UK plans to dismantle its COVID-19 testing program and scale back its daily reporting requirements ([716](#), [717](#)). A similar approach can be seen in the US as well, where multiple state-run testing facilities are closing, despite some groups advocating to keep them open ([718](#), [719](#)). These ongoing changes in testing policy are likely to have a direct effect on how the pandemic is managed moving forward. SARS-CoV-2 diagnostic tests can be used effectively to slow the spread of the disease only when 1) they are used to share testing results in a timely manner so that they can reasonably be used to approximate the number of infections in the population and 2) those tests are easily accessible by the general public.

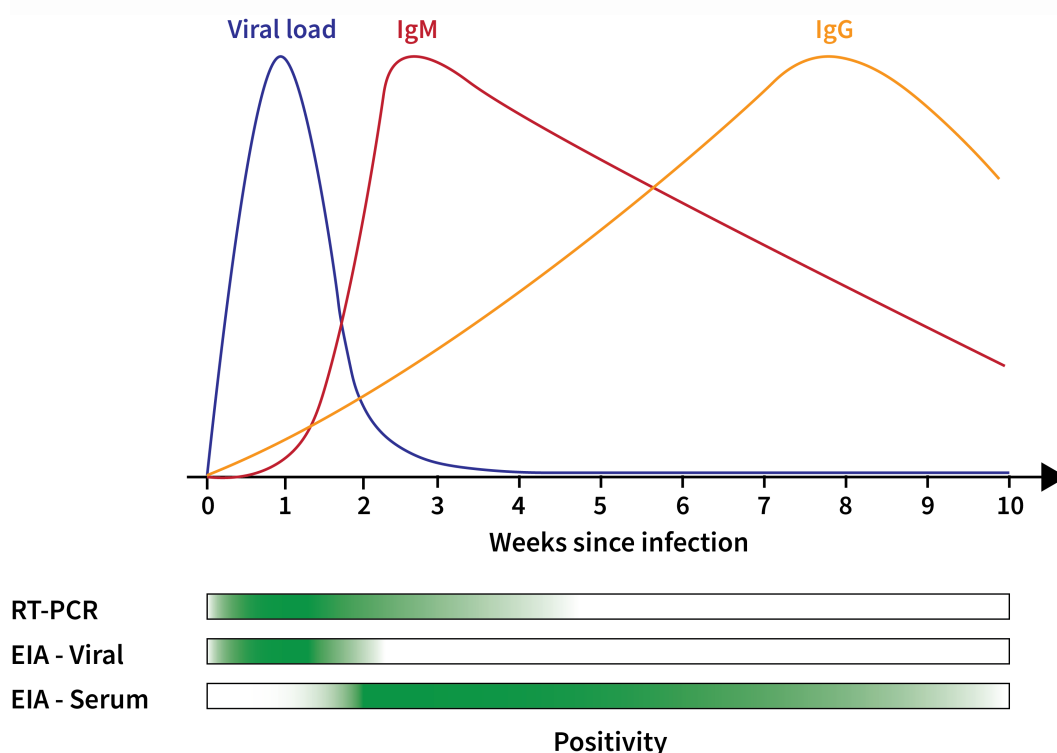


Figure 2: Summary of Diagnostic Technologies used in COVID-19 Testing. The immune response to SARS-CoV-2 means that different diagnostic approaches offer different views of COVID-19. Early in the infection course, viral load is high. This means that PCR-based testing and EIA testing for antigens are likely to return positives (as indicated by the green bars at the bottom). As viral load decreases, EIA antigen tests become negative, but PCR-based tests can still detect even very low viral loads. From a serological perspective, IgM peaks in the first few weeks following infection and then decreases, while IgG peaks much

later in the infection course. Therefore, serological tests are likely to return positives in first few months following the acute infection course. Additional detail is available above and in several analyses and reviews ([1](#), [678](#), [705](#), [720](#)).

Children are one segment of the population where the importance of the two aforementioned conditions can be exemplified. This group is particularly vulnerable as there are ongoing challenges with testing in schools, increased COVID-19 mortality rates, and COVID-19-associated orphanhood. In this regard, although there is evidence of the efficacy of routine diagnostic testing to reduce the probability of having infectious students ([721](#), [722](#)), as of March of 2022 there is an increasing number of schools that have stopped or plan to stop contact tracing efforts ([723](#), [724](#)), in line with an announcement made by the CDC where it no longer recommended contact tracing as a strategy to contain the virus ([725](#)). An estimated 197 children have died in the US from COVID-19 during the first three months of 2022 ([726](#)), compared to 735 deaths in the preceding 20 months of the pandemic ([727](#)), and millions of children have been orphaned as a consequence of parent or caregiver death due to COVID-19 ([728](#)). It is likely that reducing or eliminating testing capacity in schools will directly exacerbate these negative outcomes for the remainder of 2022.

The SARS-CoV-2 diagnostic tools presented in this paper are far less useful if they are difficult to obtain, or if their limited use results in biased data that would lead to ill-informed public health strategies. Under conditions of limited supply, different strategies for testing are needed ([729](#)). The pandemic is still an ongoing public health threat and it is worrying that active testing and tracing efforts are a low priority for public health authorities in many countries. If this trend continues, the lack of testing could result in increased morbidity and mortality and an overall failure to manage the pandemic.

4 Identification and Development of Therapeutics for COVID-19

4.1 Abstract

After emerging in China in late 2019, the novel coronavirus SARS-CoV-2 spread worldwide and as of mid-2021 remains a significant threat globally. Only a few coronaviruses are known to infect humans, and only two cause infections similar in severity to SARS-CoV-2: *Severe acute respiratory syndrome-related coronavirus*, a closely related species of SARS-CoV-2 that emerged in 2002, and *Middle East respiratory syndrome-related coronavirus*, which emerged in 2012. Unlike the current pandemic, previous epidemics were controlled rapidly through public health measures, but the body of research investigating severe acute respiratory syndrome and Middle East respiratory syndrome has proven valuable for identifying approaches to treating and preventing novel coronavirus disease 2019 (COVID-19). Building on this research, the medical and scientific communities have responded rapidly to the COVID-19 crisis to identify many candidate therapeutics. The approaches used to identify candidates fall into four main categories: adaptation of clinical approaches to diseases with related pathologies,

adaptation based on virological properties, adaptation based on host response, and data-driven identification of candidates based on physical properties or on pharmacological compendia. To date, a small number of therapeutics have already been authorized by regulatory agencies such as the Food and Drug Administration (FDA), while most remain under investigation. The scale of the COVID-19 crisis offers a rare opportunity to collect data on the effects of candidate therapeutics. This information provides insight not only into the management of coronavirus diseases, but also into the relative success of different approaches to identifying candidate therapeutics against an emerging disease.

4.2 Importance

The COVID-19 pandemic is a rapidly evolving crisis. With the worldwide scientific community shifting focus onto the SARS-CoV-2 virus and COVID-19, a large number of possible pharmaceutical approaches for treatment and prevention have been proposed. What was known about each of these potential interventions evolved rapidly throughout 2020 and 2021. This fast-paced area of research provides important insight into how the ongoing pandemic can be managed and also demonstrates the power of interdisciplinary collaboration to rapidly understand a virus and match its characteristics with existing or novel pharmaceuticals. As illustrated by the continued threat of viral epidemics during the current millennium, a rapid and strategic response to emerging viral threats can save lives. In this review, we explore how different modes of identifying candidate therapeutics have borne out during COVID-19.

4.3 Introduction

The novel coronavirus *Severe acute respiratory syndrome-related coronavirus 2* (SARS-CoV-2) emerged in late 2019 and quickly precipitated the worldwide spread of novel coronavirus disease 2019 (COVID-19). COVID-19 is associated with symptoms ranging from mild or even asymptomatic to severe, and up to 2% of patients diagnosed with COVID-19 die from COVID-19-related complications such as acute respiratory disease syndrome (ARDS) (1). As a result, public health efforts have been critical to mitigating the spread of the virus. However, as of mid-2021, COVID-19 remains a significant worldwide concern (Figure 3), with 2021 cases in some regions surging far above the numbers reported during the initial outbreak in early 2020. While a number of vaccines have been developed and approved in different countries starting in late 2020 (5), vaccination efforts have not proceeded at the same pace throughout the world and are not yet close to ending the pandemic.

Due to the continued threat of the virus and the severity of the disease, the identification and development of therapeutic interventions have emerged as significant international priorities. Prior developments during other recent outbreaks of emerging diseases, especially those caused by human coronaviruses (HCoV), have guided biomedical research into the behavior and treatment of this novel coronavirus infection. However, previous emerging HCoV-related disease threats were controlled much more quickly

than SARS-CoV-2 through public health efforts (Figure 3). The scale of the COVID-19 pandemic has made the repurposing and development of pharmaceuticals more urgent than in previous coronavirus epidemics.

4.3.1 Lessons from Prior HCoV Outbreaks

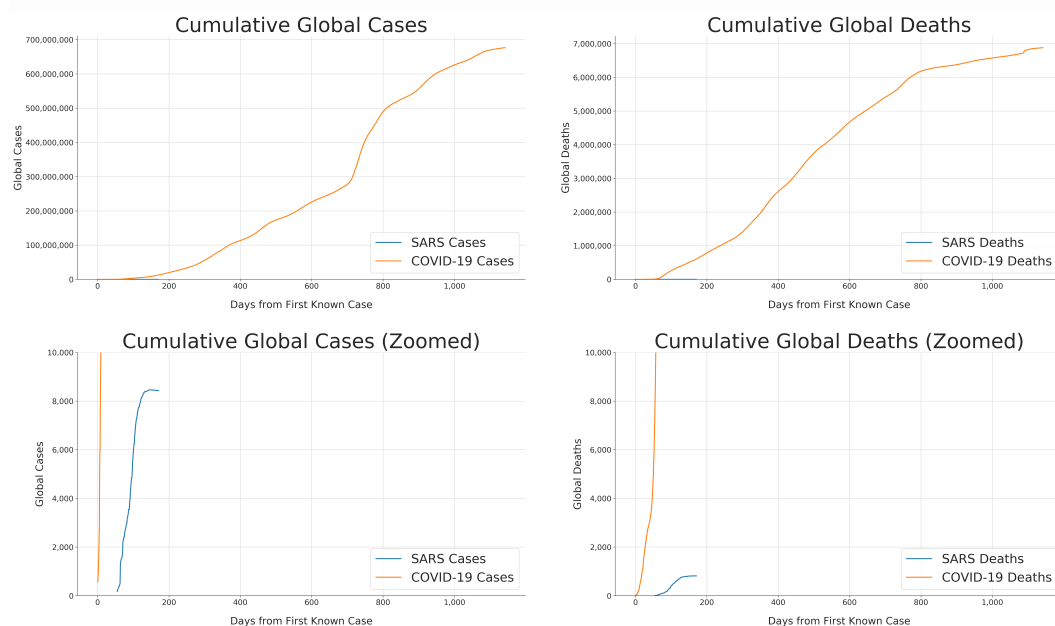


Figure 3: Cumulative global incidence of COVID-19 and SARS. As of March 9, 2023, 676,570,149 COVID-19 cases and 6,881,802 COVID-19 deaths had been reported worldwide since January 22, 2020. A total of 8,432 cases and 813 deaths were reported for SARS from March 17 to July 11, 2003. SARS-CoV-1 was officially contained on July 5, 2003, within 9 months of its appearance (730). In contrast, SARS-CoV-2 remains a significant global threat nearly two years after its emergence. COVID-19 data are from the COVID-19 Data Repository by the Center for Systems Science and Engineering at Johns Hopkins University (731, 732). SARS data are from the WHO (733) and were obtained from a dataset on GitHub (734). See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

At first, SARS-CoV-2's rapid shift from an unknown virus to a significant worldwide threat closely paralleled the emergence of *Severe acute respiratory syndrome-related coronavirus* (SARS-CoV-1), which was responsible for the 2002-03 SARS epidemic. The first documented case of COVID-19 was reported in Wuhan, China in November 2019, and the disease quickly spread worldwide in the early months of 2020. In comparison, the first case of SARS was reported in November 2002 in the Guangdong Province of China, and it spread within China and then into several countries across continents during the first half of 2003 (250, 340, 730). In fact, genome sequencing quickly revealed the virus causing COVID-19 to be a novel betacoronavirus closely related to SARS-CoV-1 (11).

While similarities between these two viruses are unsurprising given their close phylogenetic relationship, there are also some differences in how the viruses affect humans. SARS-CoV-1 infection is severe, with an estimated case fatality rate (CFR) for SARS of 9.5% (340), while estimates of the CFR associated with COVID-19 are much lower, at up to 2% (1). SARS-CoV-1 is highly contagious and spread primarily by droplet transmission, with a basic reproduction number (R_0) of 4 (i.e., each person infected was estimated to infect four other people) (340). There is still some controversy whether SARS-

CoV-2 is primarily spread by droplets or is primarily airborne ([258](#), [259](#), [262](#), [735](#)). Most estimates of its R_0 fall between 2.5 and 3 ([1](#)). Therefore, SARS is thought to be a deadlier and more transmissible disease than COVID-19.

With the 17-year difference between these two outbreaks, there were major differences in the tools available to efforts to organize international responses. At the time that SARS-CoV-1 emerged, no new HCoV had been identified in almost 40 years ([250](#)). The identity of the virus underlying the SARS disease remained unknown until April of 2003, when the SARS-CoV-1 virus was characterized through a worldwide scientific effort spearheaded by the World Health Organization (WHO) ([250](#)). In contrast, the SARS-CoV-2 genomic sequence was released on January 3, 2020 ([11](#)), only days after the international community became aware of the novel pneumonia-like illness now known as COVID-19. While SARS-CoV-1 belonged to a distinct lineage from the two other HCoVs known at the time of its discovery ([340](#)), SARS-CoV-2 is closely related to SARS-CoV-1 and is a more distant relative of another HCoV characterized in 2012, *Middle East respiratory syndrome-related coronavirus* ([19](#), [736](#)). Significant efforts had been dedicated towards understanding SARS-CoV-1 and MERS-CoV and how they interact with human hosts. Therefore, SARS-CoV-2 emerged under very different circumstances than SARS-CoV-1 in terms of scientific knowledge about HCoVs and the tools available to characterize them.

Despite the apparent advantages for responding to SARS-CoV-2 infections, COVID-19 has caused many orders of magnitude more deaths than SARS did (Figure [3](#)). The SARS outbreak was officially determined to be under control in July 2003, with the success credited to infection management practices such as mask wearing ([250](#)). *Middle East respiratory syndrome-related coronavirus* (MERS-CoV) is still circulating and remains a concern; although the fatality rate is very high at almost 35%, the disease is much less easily transmitted, as its R_0 has been estimated to be 1 ([340](#)). The low R_0 in combination with public health practices allowed for its spread to be contained ([340](#)). Neither of these trajectories are comparable to that of SARS-CoV-2, which remains a serious threat worldwide over a year and a half after the first cases of COVID-19 emerged (Figure [3](#)).

4.3.2 Potential Approaches to the Treatment of COVID-19

Therapeutic interventions can utilize two approaches: they can either mitigate the effects of an infection that harms an infected person, or they can hinder the spread of infection within a host by disrupting the viral life cycle. The goal of the former strategy is to reduce the severity and risks of an active infection, while for the latter, it is to inhibit the replication of a virus once an individual is infected, potentially freezing disease progression. Additionally, two major approaches can be used to identify interventions that might be relevant to managing an emerging disease or a novel virus: drug repurposing and drug development. Drug repurposing involves identifying an existing compound that may provide benefits in the context of interest ([737](#)). This strategy can focus on either approved or investigational drugs, for which there may be applicable preclinical or safety information ([737](#)). Drug development, on the other hand, provides an opportunity to identify or develop a compound specifically relevant to a particular need, but it is often

a lengthy and expensive process characterized by repeated failure (738). Drug repurposing therefore tends to be emphasized in a situation like the COVID-19 pandemic due to the potential for a more rapid response.

Even from the early months of the pandemic, studies began releasing results from analyses of approved and investigational drugs in the context of COVID-19. The rapid timescale of this response meant that, initially, most evidence came from observational studies, which compare groups of patients who did and did not receive a treatment to determine whether it may have had an effect. This type of study can be conducted rapidly but is subject to confounding. In contrast, randomized controlled trials (RCTs) are the gold-standard method for assessing the effects of an intervention. Here, patients are prospectively and randomly assigned to treatment or control conditions, allowing for much stronger interpretations to be drawn; however, data from these trials take much longer to collect. Both approaches have proven to be important sources of information in the development of a rapid response to the COVID-19 crisis, but as the pandemic draws on and more results become available from RCTs, more definitive answers are becoming available about proposed therapeutics. Interventional clinical trials are currently investigating or have investigated a large number of possible therapeutics and combinations of therapeutics for the treatment of COVID-19 (Figure 4).

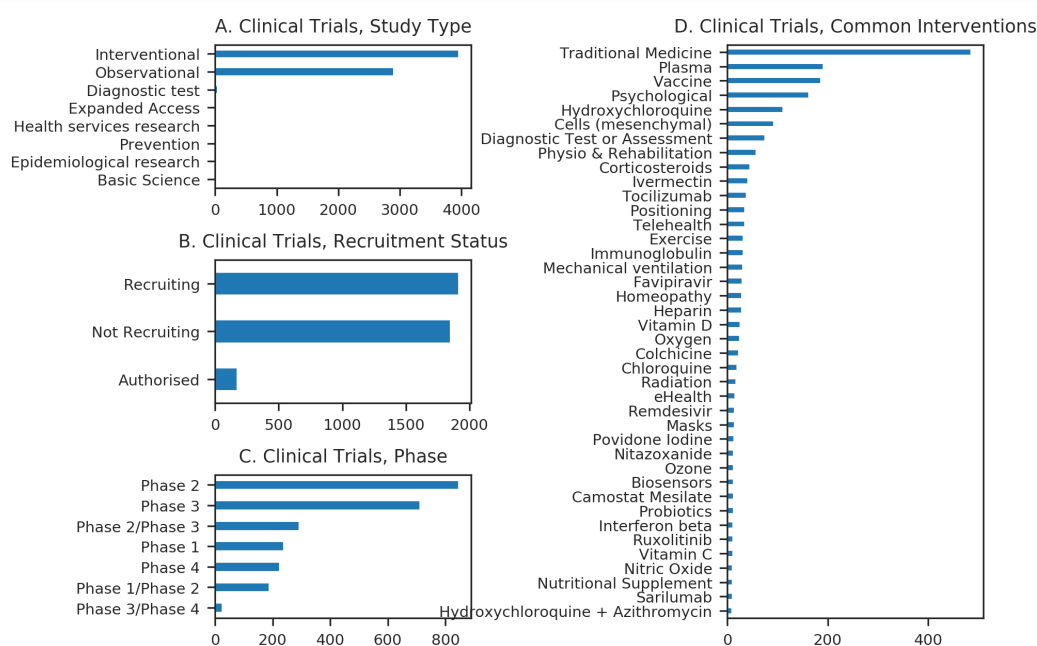


Figure 4: COVID-19 clinical trials. Trials data are from the University of Oxford Evidence-Based Medicine Data Lab's COVID-19 TrialsTracker (739). As of December 31, 2020, there were 6,987 COVID-19 clinical trials of which 3,962 were interventional. The study types include only types used in at least five trials. Only interventional trials are analyzed in the figures depicting status, phase, and intervention. Of the interventional trials, 98 trials had reported results as of December 31, 2020. Recruitment status and trial phase are shown only for interventional trials in which the status or phase is recorded. Common interventions refers to interventions used in at least ten trials. Combinations of interventions, such as hydroxychloroquine with azithromycin, are tallied separately from the individual interventions. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

The purpose of this review is to provide an evolving resource tracking the status of efforts to repurpose and develop drugs for the treatment of COVID-19. We highlight four strategies that provide different paradigms for the

identification of potential pharmaceutical treatments. The WHO guidelines (81) and a systematic review (740) are complementary living documents that summarize COVID-19 therapeutics.

4.4 Repurposing Drugs for Symptom Management

A variety of symptom profiles with a range of severity are associated with COVID-19 (1). In many cases, COVID-19 is not life threatening. A study of COVID-19 patients in a hospital in Berlin, Germany reported that the highest risk of death was associated with infection-related symptoms, such as sepsis, respiratory symptoms such as ARDS, and cardiovascular failure or pulmonary embolism (741). Similarly, an analysis in Wuhan, China reported that respiratory failure (associated with ARDS) and sepsis/multi-organ failure accounted for 69.5% and 28.0% of deaths, respectively, among 82 deceased patients (742). COVID-19 is characterized by two phases. The first is the acute response, where an adaptive immune response to the virus is established and in many cases can mitigate viral damage to organs (743). The second phase characterizes more severe cases of COVID-19. Here, patients experience a cytokine storm, whereby excessive production of cytokines floods into circulation, leading to systemic inflammation, immune dysregulation, and multiorgan dysfunction that can cause multiorgan failure and death if untreated (744). ARDS-associated respiratory failure can occur during this phase. Cytokine dysregulation was also identified in patients with SARS (745, 746).

In the early days of the COVID-19 pandemic, physicians sought to identify potential treatments that could benefit patients, and in some cases shared their experiences and advice with the medical community on social media sites such as Twitter (747). These on-the-ground treatment strategies could later be analyzed retrospectively in observational studies or investigated in an interventional paradigm through RCTs. Several notable cases involved the use of small-molecule drugs, which are synthesized compounds of low molecular weight, typically less than 1 kilodalton (kDa) (748). Small-molecule pharmaceutical agents have been a backbone of drug development since the discovery of penicillin in the early twentieth century (749). It and other antibiotics have long been among the best known applications of small molecules to therapeutics, but biotechnological developments such as the prediction of protein-protein interactions (PPIs) have facilitated advances in precise targeting of specific structures using small molecules (749). Small molecule drugs today encompass a wide range of therapeutics beyond antibiotics, including antivirals, protein inhibitors, and many broad-spectrum pharmaceuticals.

Many treatments considered for COVID-19 have relied on a broad-spectrum approach. These treatments do not specifically target a virus or particular host receptor, but rather induce broad shifts in host biology that are hypothesized to be potential inhibitors of the virus. This approach relies on the fact that when a virus enters a host, the host becomes the virus's environment. Therefore, the state of the host can also influence the virus's ability to replicate and spread. The administration and assessment of broad-spectrum small-molecule drugs on a rapid time course was feasible because

they are often either available in hospitals, or in some cases may also be prescribed to a large number of out-patients. One of the other advantages is that these well-established compounds, if found to be beneficial, are often widely available, in contrast to boutique experimental drugs.

In some cases, prior data was available from experiments examining the response of other HCoVs or HCoV infections to a candidate drug. In addition to non-pharmaceutical interventions such as encouraging non-intubated patients to adopt a prone position ([750](#)), knowledge about interactions between HCoVs and the human body, many of which emerged from SARS and MERS research over the past two decades, led to the suggestion that a number of common drugs might benefit COVID-19 patients. However, the short duration and low case numbers of prior outbreaks were less well-suited to the large-scale study of clinical applications than the COVID-19 pandemic is. As a result, COVID-19 has presented the first opportunity to robustly evaluate treatments that were common during prior HCoV outbreaks to determine their clinical efficacy. The first year of the COVID-19 pandemic demonstrated that there are several different trajectories that these clinically suggested, widely available candidates can follow when assessed against a widespread, novel viral threat.

One approach to identifying candidate small molecule drugs was to look at the approaches used to treat SARS and MERS. Treatment of SARS and MERS patients prioritized supportive care and symptom management ([340](#)). Among the clinical treatments for SARS and MERS that were explored, there was generally a lack of evidence indicating whether they were effective. Most of the supportive treatments for SARS were found inconclusive in meta-analysis ([751](#)), and a 2004 review reported that not enough evidence was available to make conclusions about most treatments ([752](#)). However, one strategy adopted from prior HCoV outbreaks is currently the best-known treatment for severe cases of COVID-19. Corticosteroids represent broad-spectrum treatments and are a well-known, widely available treatment for pneumonia ([753–758](#)) that have also been debated as a possible treatment for ARDS ([759–764](#)). Corticosteroids were also used and subsequently evaluated as possible supportive care for SARS and MERS. In general, studies and meta-analyses did not identify support for corticosteroids to prevent mortality in these HCoV infections ([765–767](#)); however, one found that the effects might be masked by variability in treatment protocols, such as dosage and timing ([752](#)). While the corticosteroids most often used to treat SARS were methylprednisolone and hydrocortisone, availability issues for these drugs at the time led to dexamethasone also being used in North America ([768](#)).

Dexamethasone (9 α -fluoro-16 α -methylprednisolone) is a synthetic corticosteroid that binds to glucocorticoid receptors ([769](#), [770](#)). It functions as an anti-inflammatory agent by binding to glucocorticoid receptors with higher affinity than endogenous cortisol ([771](#)). Dexamethasone and other steroids are widely available and affordable, and they are often used to treat community-acquired pneumonia ([772](#)) as well as chronic inflammatory conditions such as asthma, allergies, and rheumatoid arthritis ([773–775](#)). Immunosuppressive drugs such as steroids are typically contraindicated in the setting of infection ([776](#)), but because COVID-19 results in hyperinflammation that appears to contribute to mortality via lung damage, immunosuppression may be a helpful approach to treatment ([158](#)). A clinical

trial that began in 2012 recently reported that dexamethasone may improve outcomes for patients with ARDS (759), but a meta-analysis of a small amount of available data about dexamethasone as a treatment for SARS suggested that it may, in fact, be associated with patient harm (777). However, the findings in SARS may have been biased by the fact that all of the studies examined were observational and a large number of inconclusive studies were not included (778). The questions of whether and when to counter hyperinflammation with immunosuppression in the setting of COVID-19 (as in SARS (746)) was an area of intense debate, as the risks of inhibiting antiviral immunity needed to be weighed against the beneficial anti-inflammatory effects (779). As a result, guidelines early in the pandemic typically recommended avoiding treating COVID-19 patients with corticosteroids such as dexamethasone (777).

Despite this initial concern, dexamethasone was evaluated as a potential treatment for COVID-19 (Appendix 1). Dexamethasone treatment comprised one arm of the multi-site Randomized Evaluation of COVID-19 Therapy (RECOVERY) trial in the United Kingdom (780). This study found that the 28-day mortality rate was lower in patients receiving dexamethasone than in those receiving standard of care (SOC). However, this finding was driven by differences in mortality among patients who were receiving mechanical ventilation or supplementary oxygen at the start of the study. The report indicated that dexamethasone reduced 28-day mortality relative to SOC in patients who were ventilated (29.3% versus 41.4%) and among those who were receiving oxygen supplementation (23.3% versus 26.2%) at randomization, but not in patients who were breathing independently (17.8% versus 14.0%). These findings also suggested that dexamethasone may have reduced progression to mechanical ventilation, especially among patients who were receiving oxygen support at randomization. Other analyses have supported the importance of disease course in determining the efficacy of dexamethasone: additional results suggest greater potential for patients who have experienced symptoms for at least seven days and patients who were not breathing independently (781). A meta-analysis that evaluated the results of the RECOVERY trial alongside trials of other corticosteroids, such as hydrocortisone, similarly concluded that corticosteroids may be beneficial to patients with severe COVID-19 who are receiving oxygen supplementation (782). Thus, it seems likely that dexamethasone is useful for treating inflammation associated with immunopathy or cytokine release syndrome (CRS), which is a condition caused by detrimental overactivation of the immune system (1). In fact, corticosteroids such as dexamethasone are sometimes used to treat CRS (783). Guidelines were quickly updated to encourage the use of dexamethasone in severe cases (784), and this affordable and widely available treatment rapidly became a valuable tool against COVID-19 (785), with demand surging within days of the preprint's release (786).

4.5 Approaches Targeting the Virus

Therapeutics that directly target the virus itself hold the potential to prevent people infected with SARS-CoV-2 from developing potentially damaging symptoms (Figure 5). Such drugs typically fall into the broad category of antivirals. Antiviral therapies hinder the spread of a virus within the host, rather than destroying existing copies of the virus, and these drugs can vary

in their specificity to a narrow or broad range of viral targets. This process requires inhibiting the replication cycle of a virus by disrupting one of six fundamental steps (787). In the first of these steps, the virus attaches to and enters the host cell through endocytosis. Then the virus undergoes uncoating, which is classically defined as the release of viral contents into the host cell. Next, the viral genetic material enters the nucleus where it gets replicated during the biosynthesis stage. During the assembly stage, viral proteins are translated, allowing new viral particles to be assembled. In the final step new viruses are released into the extracellular environment. Although antivirals are designed to target a virus, they can also impact other processes in the host and may have unintended effects. Therefore, these therapeutics must be evaluated for both efficacy and safety. As the technology to respond to emerging viral threats has also evolved over the past two decades, a number of candidate treatments have been identified for prior viruses that may be relevant to the treatment of COVID-19.

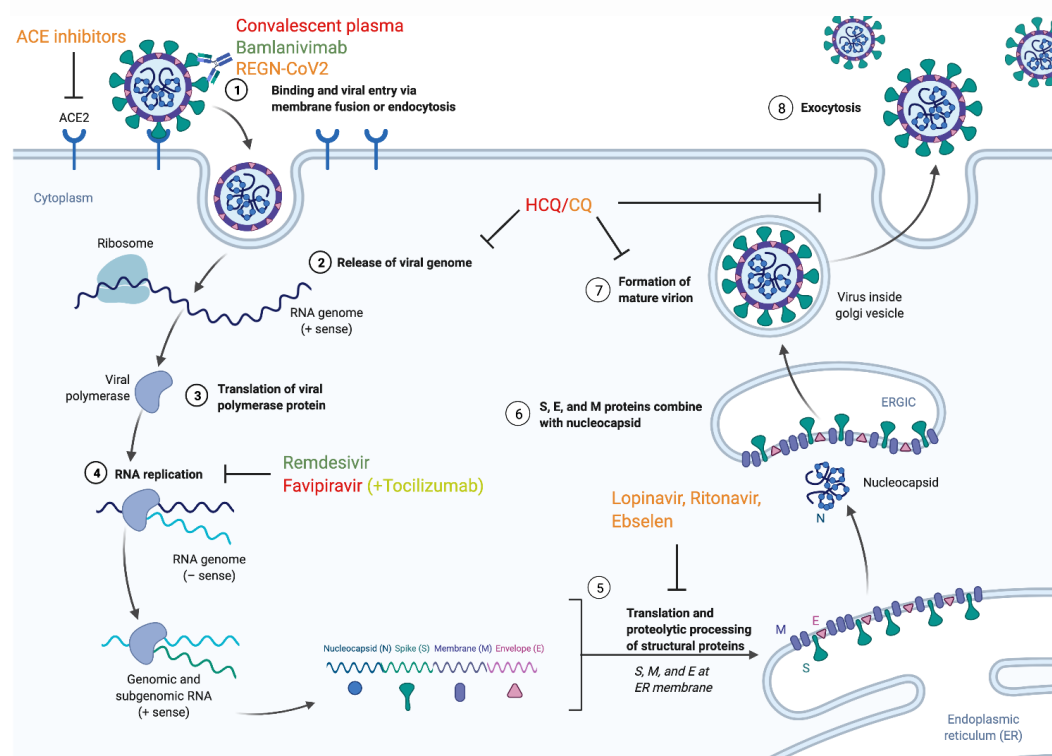


Figure 5: Mechanisms of Action for Potential Therapeutics Potential therapeutics currently being studied can target the SARS-CoV-2 virus or modify the host environment through many different mechanisms. Here, the relationships between the virus, host cells, and several therapeutics are visualized. Drug names are color-coded according to the grade assigned to them by the Center for Cytokine Storm Treatment & Laboratory's CORONA Project (788) (Green = A, Lime = B, Orange = C, and Red = D).

Many antiviral drugs are designed to inhibit the replication of viral genetic material during the biosynthesis step. Unlike DNA viruses, which can use the host enzymes to propagate themselves, RNA viruses like SARS-CoV-2 depend on their own polymerase, the RNA-dependent RNA polymerase (RdRP), for replication (789, 790). RdRP is therefore a potential target for antivirals against RNA viruses. Disruption of RdRP is the proposed mechanism underlying the treatment of SARS and MERS with ribavirin (791). Ribavirin is an antiviral drug effective against other viral infections that was often used in combination with corticosteroids and sometimes interferon (IFN) medications to treat SARS and MERS (250). However, analyses of its effects in retrospective and *in vitro* analyses of SARS and the SARS-CoV-1 virus,

respectively, have been inconclusive (250). While IFNs and ribavirin have shown promise in *in vitro* analyses of MERS, their clinical effectiveness remains unknown (250). The current COVID-19 pandemic has provided an opportunity to assess the clinical effects of these treatments. As one example, ribavirin was also used in the early days of COVID-19, but a retrospective cohort study comparing patients who did and did not receive ribavirin revealed no effect on the mortality rate (792).

Since nucleotides and nucleosides are the natural building blocks for RNA synthesis, an alternative approach has been to explore nucleoside and nucleotide analogs for their potential to inhibit viral replication. Analogs containing modifications to nucleotides or nucleosides can disrupt key processes including replication (793). A single incorporation does not influence RNA transcription; however, multiple events of incorporation lead to the arrest of RNA synthesis (794). One candidate antiviral considered for the treatment of COVID-19 is favipiravir (Avigan), also known as T-705, which was discovered by Toyama Chemical Co., Ltd. (795). It was previously found to be effective at blocking viral amplification in several influenza subtypes as well as other RNA viruses, such as *Flaviviridae* and *Picornaviridae*, through a reduction in plaque formation (796) and viral replication in Madin-Darby canine kidney cells (797). Favipiravir (6-fluoro-3-hydroxy-2-pyrazinecarboxamide) acts as a purine and purine nucleoside analogue that inhibits viral RNA polymerase in a dose-dependent manner across a range of RNA viruses, including influenza viruses (798–802). Biochemical experiments showed that favipiravir was recognized as a purine nucleoside analogue and incorporated into the viral RNA template. In 2014, the drug was approved in Japan for the treatment of influenza that was resistant to conventional treatments like neuraminidase inhibitors (803). Though initial analyses of favipiravir in observational studies of its effects on COVID-19 patients were promising, recent results of two small RCTs suggest that it is unlikely to affect COVID-19 outcomes (Appendix 1).

In contrast, another nucleoside analog, remdesivir, is one of the few treatments against COVID-19 that has received FDA approval. Remdesivir (GS-5734) is an intravenous antiviral that was proposed by Gilead Sciences as a possible treatment for Ebola virus disease. It is metabolized to GS-441524, an adenosine analog that inhibits a broad range of polymerases and then evades exonuclease repair, causing chain termination (804–806). Gilead received an emergency use authorization (EUA) for remdesivir from the FDA early in the pandemic (May 2020) and was later found to reduce mortality and recovery time in a double-blind, placebo-controlled, phase III clinical trial performed at 60 trial sites, 45 of which were in the United States (807–810). Subsequently, the WHO Solidarity trial, a large-scale, open-label trial enrolling 11,330 adult in-patients at 405 hospitals in 30 countries around the world, reported no effect of remdesivir on in-hospital mortality, duration of hospitalization, or progression to mechanical ventilation (811). Therefore, additional clinical trials of remdesivir in different patient pools and in combination with other therapies may be needed to refine its use in the clinic and determine the forces driving these differing results. Remdesivir offers proof of principle that SARS-CoV-2 can be targeted at the level of viral replication, since remdesivir targets the viral RNA polymerase at high potency. Identification of such candidates depends on knowledge about the virological properties of a novel threat. However, the success and relative

lack of success, respectively, of remdesivir and favipiravir underscore the fact that drugs with similar mechanisms will not always produce similar results in clinical trials.

4.6 Disrupting Host-Virus Interactions

4.6.1 Interrupting Viral Colonization of Cells

Some of the most widely publicized examples of efforts to repurpose drugs for COVID-19 are broad-spectrum, small-molecule drugs where the mechanism of action made it seem that the drug might disrupt interactions between SARS-CoV-2 and human host cells (Figure 5). However, the exact outcomes of such treatments are difficult to predict *a priori*, and there are several examples where early enthusiasm was not borne out in subsequent trials. One of the most famous examples of an analysis of whether a well-known medication could provide benefits to COVID-19 patients came from the assessment of chloroquine (CQ) and hydroxychloroquine (HCQ), which are used for the treatment and prophylaxis of malaria as well as the treatment of lupus erythematosus and rheumatoid arthritis in adults (812). These drugs are lysosomotropic agents, meaning they are weak bases that can pass through the plasma membrane. It was thought that they might provide benefits against SARS-CoV-2 by interfering with the digestion of antigens within the lysosome and inhibiting CD4 T-cell stimulation while promoting the stimulation of CD8 T-cells (813). These compounds also have anti-inflammatory properties (813) and can decrease the production of certain key cytokines involved in the immune response, including interleukin-6 (IL-6) and inhibit the stimulation of Toll-like receptors (TLR) and TLR signaling (813).

In vitro analyses reported that CQ inhibited cell entry of SARS-CoV-1 (814) and that both CQ and HCQ inhibited viral replication within cultured cells (815), leading to early hope that it might provide similar therapeutic or protective effects in patients. However, while the first publication on the clinical application of these compounds to the inpatient treatment of COVID-19 was very positive (816), it was quickly discredited (817). Over the following months, extensive evidence emerged demonstrating that CQ and HCQ offered no benefits for COVID-19 patients and, in fact, carried the risk of dangerous side effects (Appendix 1). The nail in the coffin came when findings from the large-scale RECOVERY trial were released on October 8, 2020. This study enrolled 11,197 hospitalized patients whose physicians believed it would not harm them to participate and used a randomized, open-label design to study the effects of HCQ compared to standard of care (SOC) at 176 hospitals in the United Kingdom (818). Rates of COVID-19-related mortality did not differ between the control and HCQ arms, but patients receiving HCQ were slightly more likely to die due to cardiac events. Patients who received HCQ also had a longer duration of hospitalization than patients receiving usual care and were more likely to progress to mechanical ventilation or death (as a combined outcome). As a result, enrollment in the HCQ arm of the RECOVERY trial was terminated early (819). The story of CQ/HCQ therefore illustrates how initial promising *in vitro* analyses can fail to translate to clinical usefulness.

A similar story has arisen with the broad-spectrum, small-molecule anthelmintic ivermectin, which is a synthetic analog of avermectin, a bioactive compound produced by a microorganism known as *Streptomyces avermectinus* and *Streptomyces avermitilis* ([820](#), [821](#)). Avermectin disrupts the ability of parasites to avoid the host immune response by blocking glutamate-gated chloride ion channels in the peripheral nervous system from closing, leading to hyperpolarization of neuronal membranes, disruption of neural transmission, and paralysis ([820](#), [822](#), [823](#)). Ivermectin has been used since the early 1980s to treat endo- and ecto-parasitic infections by helminths, insects, and arachnids in veterinary contexts ([820](#), [824](#)) and since the late 1980s to treat human parasitic infections as well ([820](#), [822](#)). More recent research has indicated that ivermectin might function as a broad-spectrum antiviral by disrupting the trafficking of viral proteins by both RNA and DNA viruses ([823](#), [825](#), [826](#)), although most of these studies have demonstrated this effect *in vitro* ([826](#)). The potential for antiviral effects on SARS-CoV-2 were investigated *in vitro*, and ivermectin was found to inhibit viral replication in a cell line derived from Vero cells (Vero-hSLAM) ([827](#)). However, inhibition of viral replication was achieved at concentrations that were much higher than that explored by existing dosage guidelines ([828](#), [829](#)), which are likely to be associated with significant side effects due to the increased potential that the compound could cross the mammalian blood-brain barrier ([830](#), [831](#)).

Retrospective studies and small RCTs began investigating the effects of standard doses of this low-cost, widely available drug. One retrospective study reported that ivermectin reduced all-cause mortality ([832](#)) while another reported no difference in clinical outcomes or viral clearance ([833](#)). Small RCTs enrolling less than 50 patients per arm have also reported a wide array of positive ([834–838](#)) and negative results ([839](#), [840](#)). A slightly larger RCT enrolling 115 patients in two arms reported inconclusive results ([841](#)). Hope for the potential of ivermectin peaked with the release of a preprint reporting results of a multicenter, double-blind RCT where a four-day course of ivermectin was associated with clinical improvement and earlier viral clearance in 400 symptomatic patients and 200 close contacts ([842](#)); however, concerns were raised about both the integrity of the data and the paper itself ([843](#), [844](#)), and this study was removed by the preprint server Research Square ([845](#)). A similarly sized RCT suggested no effect on the duration of symptoms among 400 patients split evenly across the intervention and control arms ([846](#)), and although meta-analyses have reported both null ([847](#), [848](#)) and beneficial ([849–856](#)) effects of ivermectin on COVID-19 outcomes, the certainty is likely to be low ([850](#)). These findings are potentially biased by a small number of low-quality studies, including the preprint that has been taken down ([857](#)), and the authors of one ([858](#)) have issued a notice ([849](#)) that they will revise their study with the withdrawn study removed. Thus, much like HCQ/CQ, enthusiasm for research that either has not or should not have passed peer review has led to large numbers of patients worldwide receiving treatments that might not have any effect or could even be harmful. Additionally, comments on the now-removed preprint include inquiries into how best to self-administer veterinary ivermectin as a prophylactic ([845](#)), and the FDA has posted information explaining why veterinary ivermectin should not be taken by humans concerned about COVID-19 ([859](#)). Ivermectin is now one of several candidate therapeutics being investigated in the large-scale TOGETHER ([860](#)) and PRINCIPLE ([861](#))

clinical trials. The TOGETHER trial, which previously demonstrated no effect of HCQ and lopinavir-ritonavir (862), released preliminary results in early August 2021 suggesting that ivermectin also has no effect on COVID-19 outcomes (863).

While CQ/HCQ and ivermectin are well-known medications that have long been prescribed in certain contexts, investigation of another well-established type of pharmaceutical was facilitated by the fact that it was already being taken by a large number of COVID-19 patients. Angiotensin-converting enzyme inhibitors (ACEIs) and angiotensin II receptor blockers (ARBs) are among today's most commonly prescribed medications, often being used to control blood pressure (864, 865). In the United States, for example, they are prescribed well over 100,000,000 times annually (866). Prior to the COVID-19 pandemic, the relationship between ACE2, ACEIs, and SARS had been considered as possible evidence that ACE2 could serve as a therapeutic target (867), and the connection had been explored through *in vitro* and molecular docking analysis (868) but ultimately was not pursued clinically (869). Data from some animal models suggest that several, but not all, ACEIs and several ARBs increase ACE2 expression in the cells of some organs (870), but clinical studies have not established whether plasma ACE2 expression is increased in humans treated with these medications (871). In this case, rather than introducing ARBs/ACEIs, a number of analyses have investigated whether discontinuing use affects COVID-19 outcomes. An initial observational study of the association of exposure to ACEIs or ARBs with outcomes in COVID-19 was retracted from the *New England Journal of Medicine* (872) due to concerns related to data availability (873). As RCTs have become available, they have demonstrated no effect of continuing versus discontinuing ARBs/ACEIs on patient outcomes (874, 875) (Appendix 1). Thus, once again, despite a potential mechanistic association with the pathology of SARS-CoV-2 infection, these medications were not found to influence the trajectory of COVID-19 illness.

For medications that are widely known and common, clinical research into their efficacy against a novel threat can be developed very quickly. This feasibility can present a double-edged sword. For example, HCQ and CQ were incorporated into SOC in many countries early in the pandemic and had to be discontinued once their potential to harm COVID-19 patients became apparent (876, 877). Dexamethasone remains the major success story from this category of repurposed drugs and is likely to have saved a large number of lives since summer 2020 (785).

4.6.2 Manipulating the Host Immune Response

Treatments based on understanding a virus and/or how a virus interacts with the human immune system can fall into two categories: they can interact with the innate immune response, which is likely to be a similar response across viruses, or they can be specifically designed to imitate the adaptive immune response to a particular virus. In the latter case, conservation of structure or behavior across viruses enables exploring whether drugs developed for one virus can treat another. During the COVID-19 pandemic, a number of candidate therapeutics have been explored in these categories, with varied success.

Knowledge gained from trying to understand SARS-CoV-1 and MERS-CoV from a fundamental biological perspective and characterize how they interact with the human immune system provides a theoretical basis for identifying candidate therapies. Biologics are a particularly important class of drugs for efforts to address HCoV through this paradigm. They are produced from components of living organisms or viruses, historically primarily from animal tissues, but have become increasingly feasible to produce as recombinant technologies have advanced ([878](#)).

There are many differences on the development side between biologics and synthesized pharmaceuticals, such as small molecule drugs. Typically, biologics are orders of magnitude larger than small molecule drugs and are catabolized by the body to their amino acid components ([879](#)). They are often heat sensitive, and their toxicity can vary, as it is not directly associated with the primary effects of the drug; in general, their physiochemical properties are much less understood compared to small molecules ([879](#)). Biologics include significant medical breakthroughs such as insulin for the management of diabetes and vaccines, as well monoclonal antibodies (mAbs) and interferons (IFNs), which can be used to target the host immune response after infection.

mAbs have revolutionized the way we treat human diseases and have become some of the best-selling drugs in the pharmaceutical market in recent years ([880](#)). There are currently 79 FDA approved mAbs on the market, including antibodies for viral infections (e.g. Ibalizumab for *Human immunodeficiency virus* and Palivizumab for *Respiratory syncytial virus*) ([880](#), [881](#)). Virus-specific neutralizing antibodies commonly target viral surface glycoproteins or host structures, thereby inhibiting viral entry through receptor binding interference ([882](#), [883](#)). This interference is predicted to reduce the viral load, mitigate disease, and reduce overall hospitalization. mAbs can be designed for a particular virus, and significant advances have been made in the speed at which new mAbs can be identified and produced. At the time of the SARS and MERS epidemics, interest in mAbs to reduce infection was never realized ([884](#), [885](#)), but this allowed for mAbs to quickly be considered among the top candidates against COVID-19.

4.6.2.1 Biologics and the Innate Immune Response

Deaths from COVID-19 often occur when inflammation becomes dysregulated following an immune response to the SARS-CoV-2 virus. Therefore, one potential approach to reducing COVID-19 mortality rates is to manage the inflammatory response in severely ill patients. One candidate therapeutic identified that uses this mechanism is tocilizumab (TCZ). TCZ is a mAb that was developed to manage chronic inflammation caused by the continuous synthesis of the cytokine IL-6 ([886](#)). IL-6 is a pro-inflammatory cytokine belonging to the interleukin family, which is comprised by immune system regulators that are primarily responsible for immune cell differentiation. Often used to treat chronic inflammatory conditions such as rheumatoid arthritis ([886](#)), TCZ has become a pharmaceutical of interest for the treatment of COVID-19 because of the role IL-6 plays in this disease. It has also been approved to treat CRS caused by CAR-T treatments ([887](#)). While the secretion of IL-6 can be associated with chronic conditions, IL-6 is a key player in the innate immune response and is secreted by macrophages in

response to the detection of pathogen-associated molecular patterns and damage-associated molecular patterns (886). An analysis of 191 in-patients at two Wuhan hospitals revealed that blood concentrations of IL-6 differed between patients who did and did not recover from COVID-19. Patients who ultimately died had higher IL-6 levels at admission than those who recovered (83). Additionally, IL-6 levels remained higher throughout the course of hospitalization in the patients who ultimately died (83).

Currently, TCZ is being administered either as a monotherapy or in combination with other treatments in 73 interventional COVID-19 clinical trials (Figure 4). A number of retrospective studies have been conducted in several countries (888–893). In general, these studies have reported a positive effect of TCZ on reducing mortality in COVID-19 patients, although due to their retrospective designs, significant limitations are present in all of them (Appendix 1). It was not until February 11, 2021 that a preprint describing preliminary results of the first RCT of TCZ was released as part of the RECOVERY trial (894). TCZ was found to reduce 28-day mortality from 33% in patients receiving SOC alone to 29% in those receiving TCZ. Combined analysis of the RECOVERY trial data with data from smaller RCTs suggested a 13% reduction in 28-day mortality (894). While this initial report did not include the full results expected from the RECOVERY trial, this large-scale, RCT provides strong evidence that TCZ may offer benefits for COVID-19 patients. The RECOVERY trial along with results from several other RCTs (895–899) were cited as support for the EUA issued for TCZ in June 2021 (900). However, the fact that TCZ suppresses the immune response means that it does carry risks for patients, especially a potential risk of secondary infection (Appendix 1).

TCZ is just one example of a candidate drug targeting the host immune response and specifically excessive inflammation. For example, interferons (IFNs) have also been investigated; these are a family of cytokines critical to activating the innate immune response against viral infections. Synairgen has been investigating a candidate drug, SNG001, which is an IFN- β -1a formulation to be delivered to the lungs via inhalation (901) that they reported reduced progression to ventilation in a double-blind, placebo-controlled, multi-center study of 101 patients with an average age in the late 50s (902, 903). However, these findings were not supported by the large-scale WHO Solidarity trial, which reported no significant effect of IFN- β -1a on patient survival during hospitalization (811), although differences in the designs of the two studies, and specifically the severity of illness among enrolled patients, may have influenced their divergent outcomes (Appendix 1). Other biologics influencing inflammation are also being explored (Appendix 1). It is also important that studies focused on inflammation as a possible therapeutic target consider the potential differences in baseline inflammation among patients from different backgrounds, which may be caused by differing life experiences (see (339)).

4.6.2.2 Biologics and the Adaptive Immune Response

While TCZ is an example of an mAb focused on managing the innate immune response, other treatments are more specific, targeting the adaptive immune response after an infection. In some cases, treatments can utilize biologics obtained directly from recovered individuals. From the very early days of the

COVID-19 pandemic, polyclonal antibodies from convalescent plasma were investigated as a potential treatment for COVID-19 ([904](#), [905](#)). Convalescent plasma was used in prior epidemics including SARS, Ebola Virus Disease, and even the 1918 Spanish Influenza ([904](#), [906](#)). Use of convalescent plasma transfusion (CPT) over more than a century has aimed to reduce symptoms and improve mortality in infected people ([906](#)), possibly by accelerating viral clearance ([904](#)). However, it seems unlikely that this classic treatment confers any benefit for COVID-19 patients. Several systematic reviews have investigated whether CPT reduced mortality in COVID-19 patients, and although findings from early in the pandemic (up to April 19, 2020) did support use of CPT ([906](#)), the tide has shifted as the body of available literature has grown ([907](#)). While titer levels were suggested as a possible determining factor in the success of CPT against COVID-19 ([908](#)), the large-scale RECOVERY trial evaluated the effect of administering high-titer plasma specifically and found no effect on mortality or hospital discharge over a 28-day period ([909](#)). These results thus suggest that, despite initial optimism and an EUA from the FDA, CPT is unlikely to be an effective therapeutic for COVID-19.

A different narrative is shaping up around the use of mAbs specifically targeting SARS-CoV-2. During the first SARS epidemic in 2002, neutralizing antibodies (nAbs) were found in SARS-CoV-1-infected patients ([910](#), [911](#)). Several studies following up on these findings identified various S-glycoprotein epitopes as the major targets of nAbs against SARS-CoV-1 ([912](#)). Coronaviruses use trimeric spike (S) glycoproteins on their surface to bind to the host cell, allowing for cell entry ([25](#), [33](#)). Each S glycoprotein protomer is comprised of an S1 domain, also called the receptor binding domain (RBD), and an S2 domain. The S1 domain binds to the host cell while the S2 domain facilitates the fusion between the viral envelope and host cell membranes ([912](#)). The genomic identity between the RBD of SARS-CoV-1 and SARS-CoV-2 is around 74% ([913](#)). Due to this high degree of similarity, preexisting antibodies against SARS-CoV-1 were initially considered candidates for neutralizing activity against SARS-CoV-2. While some antibodies developed against the SARS-CoV-1 spike protein showed cross-neutralization activity with SARS-CoV-2 ([914](#), [915](#)), others failed to bind to SARS-CoV-2 spike protein at relevant concentrations ([16](#)). Cross-neutralizing activities were dependent on whether the epitope recognized by the antibodies were conserved between SARS-CoV-1 and SARS-CoV-2 ([914](#)).

Technological advances in antibody drug design as well as in structural biology massively accelerated the discovery of novel antibody candidates and the mechanisms by which they interact with the target structure. Within just a year of the structure of the SARS-CoV-2 spike protein being published, an impressive pipeline of monoclonal antibodies targeting SARS-CoV-2 entered clinical trials, with hundreds more candidates in preclinical stages. The first human monoclonal neutralizing antibody specifically against the SARS-CoV-2 S glycoprotein was developed using hybridoma technology ([916](#)), where antibody-producing B-cells developed by mice are inserted into myeloma cells to produce a hybrid cell line (the hybridoma) that is grown in culture. The 47D11 antibody clone was able to cross-neutralize SARS-CoV-1 and SARS-CoV-2. This antibody (now ABVV-47D11) has recently entered clinical trials in collaboration with AbbVie. Additionally, an extensive monoclonal neutralizing antibody pipeline has been developed to combat the ongoing pandemic, with

over 50 different antibodies in clinical trials ([917](#)). Thus far, the monotherapy sotrovimab and two antibody cocktails (bamlanivimab/etesevimab and casirivimab/imdevimab) have been granted EUAs by the FDA.

One of the studied antibody cocktails consists of bamlanivimab and etesevimab. Bamlanivimab (Ly-CoV555) is a human mAb that was derived from convalescent plasma donated by a recovered COVID-19 patient, evaluated in research by the National Institute of Allergy and Infectious Diseases (NIAID), and subsequently developed by AbCellera and Eli Lilly. The neutralizing activity of bamlanivimab was initially demonstrated *in vivo* using a nonhuman primate model ([918](#)). Based on these positive preclinical data, Eli Lilly initiated the first human clinical trial for a monoclonal antibody against SARS-CoV-2. The phase 1 trial, which was conducted in hospitalized COVID-19 patients, was completed in August 2020 ([919](#)). Etesevimab (LY-CoV016 or JS-016) is also a monoclonal neutralizing antibody against the spike protein of SARS-CoV-2. It was initially developed by Junshi Biosciences and later licensed and developed through Eli Lilly. A phase 1 clinical trial to assess the safety of etesevimab was completed in October 2020 ([920](#)). Etesevimab was shown to bind a different epitope on the spike protein than bamlanivimab, suggesting that the two antibodies used as a combination therapy would further enhance their clinical use compared to a monotherapy ([921](#)). To assess the efficacy and safety of bamlanivimab alone or in combination with etesevimab for the treatment of COVID-19, a phase 2/3 trial (BLAZE-1) ([922](#)) was initiated. The interim analysis of the phase 2 portion suggested that bamlanivimab alone was able to accelerate the reduction in viral load ([923](#)). However, more recent data suggests that only the bamlanivimab/etesevimab combination therapy is able to reduce viral load in COVID-19 patients ([921](#)). Based on this data, the combination therapy received an EUA for COVID-19 from the FDA in February 2021 ([924](#)).

A second therapy is comprised of casirivimab and imdevimab (REGN-COV2). Casirivimab (REGN10933) and imdevimab (REGN10987) are two monoclonal antibodies against the SARS-CoV-2 spike protein. They were both developed by Regeneron in a parallel high-throughput screening (HTS) to identify neutralizing antibodies from either humanized mice or patient-derived convalescent plasma ([925](#)). In these efforts, multiple antibodies were characterized for their ability to bind and neutralize the SARS-CoV-2 spike protein. The investigators hypothesized that an antibody cocktail, rather than each individual antibody, could increase the therapeutic efficacy while minimizing the risk for virus escape. Therefore, the authors tested pairs of individual antibodies for their ability to simultaneously bind the RBD of the spike protein. Based on this data, casirivimab and imdevimab were identified as the lead antibody pair, resulting in the initiation of two clinical trials ([926](#), [927](#)). Data from this phase 1-3 trial published in the *New England Journal of Medicine* shows that the REGN-COV2 antibody cocktail reduced viral load, particularly in patients with high viral load or whose endogenous immune response had not yet been initiated ([928](#)). However, in patients who already initiated an immune response, exogenous addition of REGN-COV2 did not improve the endogenous immune response. Both doses were well tolerated with no serious events related to the antibody cocktail. Based on this data, the FDA granted an EUA for REGN-COV2 in patients with mild to moderate

COVID-19 who are at risk of developing severe disease ([929](#)). Ongoing efforts are trying to evaluate the efficacy of REGN-COV2 to improve clinical outcomes in hospitalized patients ([926](#)).

Sotrovimab is the most recent mAb to receive an EUA. It was identified in the memory B cells of a 2003 survivor of SARS ([930](#)) and was found to be cross-reactive with SARS-CoV-2 ([915](#)). This cross-reactivity is likely attributable to conservation within the epitope, with 17 out of 22 residues conserved between the two viruses, four conservatively substituted, and one semi-conservatively substituted ([915](#)). In fact, these residues are highly conserved among sarbecoviruses, a clade that includes SARS-CoV-1 and SARS-CoV-2 ([915](#)). This versatility has led to it being characterized as a “super-antibody” ([931](#)), a potent, broadly neutralizing antibody ([932](#)). Interim analysis of data from a clinical trial ([933](#)) reported high safety and efficacy of this mAb in 583 COVID-19 patients ([934](#)). Compared to placebo, sotrovimab was found to be 85% more effective in reducing progression to the primary endpoint, which was the proportion of patients who, within 29 days, were either hospitalized for more than 24 hours or died. Additionally, rates of adverse events were comparable, and in some cases lower, among patients receiving sotrovimab compared to patients receiving a placebo. Sotrovimab therefore represents a mAb therapeutic that is effective against SARS-CoV-2 and may also be effective against other sarbecoviruses.

Several potential limitations remain in the application of mAbs to the treatment of COVID-19. One of the biggest challenges is identifying antibodies that not only bind to their target, but also prove to be beneficial for disease management. Currently, use of mAbs is limited to people with mild to moderate disease that are not hospitalized, and it has yet to be determined whether they can be used as a successful treatment option for severe COVID-19 patients. While preventing people from developing severe illness provides significant benefits, patients with severe illness are at the greatest risk of death, and therefore therapeutics that provide benefits against severe illness are particularly desirable. It remains to be seen whether mAbs confer any benefits for patients in this category.

Another concern about therapeutics designed to amplify the response to a specific viral target is that they may need to be modified as the virus evolves. With the ongoing global spread of new SARS-CoV-2 variants, there is a growing concern that mutations in SARS-CoV-2 spike protein could escape antibody neutralization, thereby reducing the efficacy of monoclonal antibody therapeutics and vaccines. A comprehensive mutagenesis screen recently identified several amino acid substitutions in the SARS-CoV-2 spike protein that can prevent antibody neutralization ([935](#)). While some mutations result in resistance to only one antibody, others confer broad resistance to multiple mAbs as well as polyclonal human sera, suggesting that some amino acids are “hotspots” for antibody resistance. However, it was not investigated whether the resistance mutations identified result in a fitness advantage. Accordingly, an impact on neutralizing efficiency has been reported for the B.1.1.7 (Alpha) variant first identified in the UK and the B.1.351 (Beta) variant first identified in in South Africa ([936–938](#)). As of June 25, 2021, the CDC recommended a pause in the use of bamlanivimab and etesevimab due to decreased efficacy against the P.1 (Gamma) and B.1.351 (Beta) variants of SARS-CoV-2 ([939](#)). While the reported impact on antibody neutralization

needs to be confirmed *in vivo*, it suggests that some adjustments to therapeutic antibody treatments may be necessary to maintain the efficacy that was reported in previous clinical trials.

Several strategies have been employed to try to mitigate the risk of diminished antibody neutralization. Antibody cocktails such as those already holding an EUA may help overcome the risk for attenuation of the neutralizing activity of a single monoclonal antibody. These cocktails consist of antibodies that recognize different epitopes on the spike protein, decreasing the likelihood that a single amino acid change can cause resistance to all antibodies in the cocktail. However, neutralizing resistance can emerge even against an antibody cocktail if the individual antibodies target subdominant epitopes (937). Another strategy is to develop broadly neutralizing antibodies that target structures that are highly conserved, as these are less likely to mutate (940, 941) or to target epitopes that are insensitive to mutations (942). Sotrovimab, one such “super-antibody”, is thought to be somewhat robust to neutralization escape (943) and has been found to be effective against all variants assessed as of August 12, 2021 (944). Another antibody (ADG-2) targets a highly conserved epitope that overlaps the hACE2 binding site of all clade 1 sarbecoviruses (945). Prophylactic administration of ADG-2 in an immunocompetent mouse model of COVID-19 resulted in protection against viral replication in the lungs and respiratory burden. Since the epitope targeted by ADG-2 represents an Achilles’ heel for clade 1 sarbecoviruses, this antibody, like sotrovimab, might be a promising candidate against all circulating variants as well as emerging SARS-related coronaviruses. To date, it has fared well against the Alpha, Beta, Gamma, and Delta variants (944).

The development of mAbs against SARS-CoV-2 has made it clear that this technology is rapidly adaptable and offers great potential for the response to emerging viral threats. However, additional investigation may be needed to adapt mAb treatments to SARS-CoV-2 as it evolves and potentially to pursue designs that confer benefits for patients at the greatest risk of death. While polyclonal antibodies from convalescent plasma have been evaluated as a treatment for COVID-19, these studies have suggested fewer potential benefits against SARS-CoV-2 than mAbs; convalescent plasma therapy has been thoroughly reviewed elsewhere (904, 905). Thus, advances in biologics for COVID-19 illustrate that an understanding of how the host and virus interact can guide therapeutic approaches. The FDA authorization of two combination mAb therapies, in particular, underscores the potential for this strategy to allow for a rapid response to a novel pathogen. Additionally, while TCZ is not yet as established, this therapy suggests that the strategy of using biologics to counteract the cytokine storm response may provide therapies for the highest-risk patients.

4.7 High-Throughput Screening for Drug Repurposing

The drug development process is slow and costly, and developing compounds specifically targeted to an emerging viral threat is not a practical short-term solution. Screening existing drug compounds for alternative indications is a popular alternative (946–949). HTS has been a goal of

pharmaceutical development since at least the mid-1980s (950). Traditionally, phenotypic screens were used to test which compounds would induce a desired change in *in vitro* or *in vivo* models, focusing on empirical, function-oriented exploration naïve to molecular mechanism (951–953). In many cases, these screens utilize large libraries that encompass a diverse set of agents varying in many pharmacologically relevant properties (e.g., (954)). The compounds inducing a desired effect could then be followed up on. Around the turn of the millennium, advances in molecular biology allowed for HTS to shift towards screening for compounds interacting with a specific molecular target under the hypothesis that modulating that target would have a desired effect. These approaches both offer pros and cons, and today a popular view is that they are most effective in combination (951, 953, 955).

Today, some efforts to screen compounds for potential repurposing opportunities are experimental, but others use computational HTS approaches (946, 956). Computational drug repurposing screens can take advantage of big data in biology (737) and as a result are much more feasible today than during the height of the SARS and MERS outbreaks in the early 2000s and early 2010s, respectively. Advancements in robotics also facilitate the experimental component of HTS (948). For viral diseases, the goal of drug repurposing is typically to identify existing drugs that have an antiviral effect likely to impede the virus of interest. While both small molecules and biologics can be candidates for repurposing, the significantly lower price of many small molecule drugs means that they are typically more appealing candidates (957).

Depending on the study design, screens vary in how closely they are tied to a hypothesis. As with the candidate therapeutics described above, high-throughput experimental or computational screens can proceed based on a hypothesis. Just as remdesivir was selected as a candidate antiviral because it is a nucleoside analog (958), so too can high-throughput screens select libraries of compounds based on a molecular hypothesis. Likewise, when the library of drugs is selected without basis in a potential mechanism, a screen can be considered hypothesis free (958). Today, both types of analyses are common both experimentally and computationally. Both strategies have been applied to identifying candidate therapeutics against SARS-CoV-2.

4.7.1 Hypothesis-Driven Screening

Hypothesis-driven screens often select drugs likely to interact with specific viral or host targets or drugs with desired clinical effects, such as immunosuppressants. There are several properties that might identify a compound as a candidate for an emerging viral disease. Drugs that interact with a target that is shared between pathogens (i.e., a viral protease or a polymerase) or between a viral pathogen and another illness (i.e., a cancer drug with antiviral potential) are potential candidates, as are drugs that are thought to interact with additional molecular targets beyond those they were developed for (956). Such research can be driven by *in vitro* or *in silico* experimentation. Computational analyses depend on identifying compounds that modulate pre-selected proteins in the virus or host. As a result, they build on experimental research characterizing the molecular features of the virus, host, and candidate compounds (949).

One example of the application of this approach to COVID-19 research comes from work on protease inhibitors. Studies have shown that viral proteases play an important role in the life cycle of viruses, including coronaviruses, by modulating the cleavage of viral polyprotein precursors (959). Several FDA-approved drugs target proteases, such as lopinavir and ritonavir for HIV infection and simeprevir for hepatitis C virus infection. Serine protease inhibitors were previously suggested as possible treatments for SARS and MERS (960). One early study (33) suggested that camostat mesylate, a protease inhibitor, could block the entry of SARS-CoV-2 into lung cells *in vitro*. Two polyproteins encoded by the SARS-CoV-2 replicase gene, pp1a and pp1ab, are critical for viral replication and transcription (961). These polyproteins must undergo proteolytic processing, which is usually conducted by main protease (M^{Pro}), a 33.8-kDa SARS-CoV-2 protease that is therefore fundamental to viral replication and transcription. Therefore, it was hypothesized that compounds targeting M^{Pro} could be used to prevent or slow the replication of the SARS-CoV-2 virus.

Both computational and experimental approaches facilitated the identification of compounds that might inhibit SARS-CoV-2 M^{Pro} . In 2005, computer-aided design facilitated the development of a Michael acceptor inhibitor, now known as N3, to target M^{Pro} of SARS-like coronaviruses (962). N3 binds in the substrate binding pocket of M^{Pro} in several HCoV (962–965). The structure of N3-bound SARS-CoV-2 M^{Pro} has been solved, confirming the computational prediction that N3 would similarly bind in the substrate binding pocket of SARS-CoV-2 (961). N3 was tested *in vitro* on SARS-CoV-2-infected Vero cells, which belong to a line of cells established from the kidney epithelial cells of an African green monkey, and was found to inhibit SARS-CoV-2 (961). A library of approximately 10,000 compounds was screened in a fluorescence resonance energy transfer assay constructed using SARS-CoV-2 M^{Pro} expressed in *Escherichia coli* (961).

Six leads were identified in this hypothesis-driven screen. *In vitro* analysis revealed that ebselen had the strongest potency in reducing the viral load in SARS-CoV-2-infected Vero cells (961). Ebselen is an organoselenium compound with anti-inflammatory and antioxidant properties (966). Molecular dynamics analysis further demonstrated the potential for ebselen to bind to M^{Pro} and disrupt the protease's enzymatic functions (967). However, ebselen is likely to be a promiscuous binder, which could diminish its therapeutic potential (961, 968), and compounds with higher specificity may be needed to translate this mechanism effectively to clinical trials. In July 2020, phase II clinical trials commenced to assess the effects of SPI-1005, an investigational drug from Sound Pharmaceuticals that contains ebselen (969), on 60 adults presenting with each of moderate (970) and severe (971) COVID-19. Other M^{Pro} inhibitors are also being evaluated in clinical trials (972, 973, 973). Pending the results of clinical trials, N3 remains a computationally interesting compound based on both computational and experimental data, but whether these potential effects will translate to the clinic remains unknown.

4.7.2 Hypothesis-Free Screening

Hypothesis-free screens use a discovery-driven approach, where screens are not targeted to specific viral proteins, host proteins, or desired clinical modulation. Hypothesis-free drug screening began twenty years ago with the testing of libraries of drugs experimentally. Today, like many other areas of biology, *in silico* analyses have become increasingly popular and feasible through advances in biological big data ([958](#), [974](#)). Many efforts have collected data about interactions between drugs and SARS-CoV-2 and about the host genomic response to SARS-CoV-2 exposure, allowing for hypothesis-free computational screens that seek to identify new candidate therapeutics. Thus, they utilize a systems biology paradigm to extrapolate the effect of a drug against a virus based on the host interactions with both the virus and the drug ([949](#)).

Resources such as the COVID-19 Drug and Gene Set Library, which at the time of its publication contained 1,620 drugs sourced from 173 experimental and computational drug sets and 18,676 human genes sourced from 444 gene sets ([975](#)), facilitate such discovery-driven approaches. Analysis of these databases indicated that some drugs had been identified as candidates across multiple independent analyses, including high-profile candidates such as CQ/HCQ and remdesivir ([975](#)). Computational screening efforts can then mine databases and other resources to identify potential PPIs among the host, the virus, and established and/or experimental drugs ([976](#)). Subject matter expertise from human users may be integrated to varying extents depending on the platform (e.g., ([976](#), [977](#))). These resources have allowed studies to identify potential therapeutics for COVID-19 without an *a priori* reason for selecting them.

One example of a hypothesis-free screen for COVID-19 drugs comes from a PPI-network-based analysis that was published early in the pandemic ([193](#)). Here, researchers cloned the proteins expressed by SARS-CoV-2 *in vitro* and quantified 332 viral-host PPI using affinity purification mass spectrometry ([193](#)). They identified two SARS-CoV-2 proteins (Nsp6 and Orf9c) that interacted with host Sigma-1 and Sigma-2 receptors. Sigma receptors are located in the endoplasmic reticulum of many cell types, and type 1 and 2 Sigma receptors have overlapping but distinct affinities for a variety of ligands ([978](#)). Molecules interacting with the Sigma receptors were then analyzed and found to have an effect on viral infectivity *in vitro* ([193](#)). A follow-up study evaluated the effect of perturbing these 332 proteins in two cells lines, A549 and Caco-2, using knockdown and knockout methods, respectively, and found that the replication of SARS-CoV-2 in cells from both lines was dependent on the expression of *SIGMAR1*, which is the gene that encodes the Sigma-1 receptor ([979](#)). Following these results, drugs interacting with Sigma receptors were suggested as candidates for repurposing for COVID-19 (e.g., ([980](#))). Because many well-known and affordable drugs interact with the Sigma receptors ([193](#), [981](#)), they became a major focus of drug repurposing efforts. Some of the drugs suggested by the apparent success of Sigma receptor-targeting drugs were already being investigated at the time. HCQ, for example, forms ligands with both Sigma-1 and Sigma-2 receptors and was already being explored as a candidate therapeutic for COVID-19 ([193](#)). Thus, this computational approach yielded interest in drugs whose antiviral activity was supported by initial *in vitro* analyses.

Follow-up research, however, called into question whether the emphasis on drugs interacting with Sigma receptors might be based on a spurious association (982). This study built on the prior work by examining whether antiviral activity among compounds correlated with their affinity for the Sigma receptors and found that it did not. The study further demonstrated that cationic amphiphilicity was a shared property among many of the candidate drugs identified through both computational and phenotypic screens and that it was likely to be the source of many compounds' proposed antiviral activity (982). Cationic amphiphilicity is associated with the induction of phospholipidosis, which is when phospholipids accumulate in the lysosome (983). Phospholipidosis can disrupt viral replication by inhibiting lipid processing (984) (see discussion of HCQ in Appendix 1). However, phospholipidosis is known to translate poorly from *in vitro* models to *in vivo* models or clinical applications. Thus, this finding suggested that these screens were identifying compounds that shared a physiochemical property rather than a specific target (982). The authors further demonstrated that antiviral activity against SARS-CoV-2 *in vitro* was correlated with the induction of phospholipidosis for drugs both with and without cationic amphiphilicity (982). This finding supports the idea that the property of cationic amphiphilicity was being detected as a proxy for the shared effect of phospholipidosis (982). They demonstrated that phospholipidosis-inducing drugs were not effective at preventing viral propagation *in vivo* in a murine model of COVID-19 (982). Therefore, removing hits that induce phospholipidosis from computational and *in vitro* experimental repurposing screens (e.g., (985)) may help emphasize those that are more likely to provide clinical benefits. This work illustrates the importance of considering confounding variables in computational screens, a principle that has been incorporated into more traditional approaches to drug development (986).

One drug that acts on Sigma receptors does, however, remain a candidate for the treatment of COVID-19. Several psychotropic drugs target Sigma receptors in the central nervous system and thus attracted interest as potential COVID-19 therapeutics following the findings of two host-virus PPI studies (987). For several of these drugs, the *in vitro* antiviral activity (979) was not correlated with their affinity for the Sigma-1 receptor (982, 987) but was correlated with phospholipidosis (982). However, fluvoxamine, a selective serotonin reuptake inhibitor that is a particularly potent Sigma-1 receptor agonist (987), has shown promise as a preventative of severe COVID-19 in a preliminary analysis of data from the large-scale TOGETHER trial (863). As of August 6, 2021, this trial had collected data from over 1,400 patients in the fluvoxamine arm of their study, half of whom received a placebo (863). Only 74 patients in the fluvoxamine group had progressed to hospitalization for COVID-19 compared to 107 in the placebo group, corresponding to a relative risk of 0.69; additionally, the relative risk of mortality between the two groups was calculated at 0.71. These findings support the results of small clinical trials that have found fluvoxamine to reduce clinical deterioration relative to a placebo (988, 989). However, the ongoing therapeutic potential of fluvoxamine does not contradict the finding that hypothesis-free screening hits can be driven by confounding factors. The authors point out that its relevance would not just be antiviral as it has a potential immunomodulatory mechanism (988). It has been found to be protective against septic shock in an *in vivo* mouse model (990). It is possible that fluvoxamine also exerts an antiviral effect (991). Thus, Sigma-1 receptor

activity may contribute to fluvoxamine's potential effects in treating COVID-19, but is not the only mechanism by which this drug can interfere with disease progression.

4.7.3 Potential and Limitations of High-Throughput Analyses

Computational screening allows for a large number of compounds to be evaluated to identify those most likely to display a desired behavior or function. This approach can be guided by a hypothesis or can aim to discover underlying characteristics that produce new hypotheses about the relationship between a host, a virus, and candidate pharmaceuticals. The examples outlined above illustrate that HTS-based evaluations of drug repurposing can potentially provide valuable insights. Computational techniques were used to design compounds targeting M^{Pro} based on an understanding of how this protease aids viral replication, and M^{Pro} inhibitors remain promising candidates ([948](#)), although the clinical trial data is not yet available. Similarly, computational analysis correctly identified the Sigma-1 receptor as a protein of interest. Although the process of identifying which drugs might modulate the interaction led to an emphasis on candidates that ultimately have not been supported, fluvoxamine remains an appealing candidate. The difference between the preliminary evidence for fluvoxamine compared to other drugs that interact with Sigma receptors underscores a major critique of hypothesis-free HTS in particular: while these approaches allow for brute force comparison of a large number of compounds against a virus of interest, they lose the element of expertise that is associated with most successes in drug repurposing ([958](#)).

There are also practical limitations to these methods. One concern is that computational analyses inherently depend on the quality of the data being evaluated. The urgency of the COVID-19 pandemic led many research groups to pivot towards computational HTS research without familiarity with best practices in this area ([948](#)). As a result, there is an excessive amount of information available from computational studies ([992](#)), but not all of it is high-quality. Additionally, the literature used to identify and validate targets can be difficult to reproduce ([993](#)), which may pose challenges to target-based experimental screening and to *in silico* screens. Some efforts to repurpose antivirals have focused on host, rather than viral, proteins ([949](#)), which might be expected to translate poorly *in vivo* if the targeted proteins serve essential functions in the host. Concerns about the practicality of hypothesis-free screens to gain novel insights are underscored by the fact that very few or possibly no success stories have emerged from hypothesis-free screens over the past twenty years ([958](#)). These findings suggest that data-driven research can be an important component of the drug repurposing ecosystem, but that drug repurposing efforts that proceed without a hypothesis, an emphasis on biological mechanisms, or an understanding of confounding effects may not produce viable candidates.

4.8 Considerations in Balancing Different Approaches

The approaches described here offer a variety of advantages and limitations in responding to a novel viral threat and building on existing bodies of knowledge in different ways. Medicine, pharmacology, basic science (especially virology and immunology), and biological data science can all provide different insights and perspectives for addressing the challenging question of which existing drugs might provide benefits against an emerging viral threat. A symptom management-driven approach allows clinicians to apply experience with related diseases or related symptoms to organize a rapid response aimed at saving the lives of patients already infected with a new disease. Oftentimes, the pharmaceutical agents that are applied are small-molecule, broad-spectrum pharmaceuticals that are widely available and affordable to produce, and they may already be available for other purposes, allowing clinicians to administer them to patients quickly either with an EUA or off-label. In this vein, dexamethasone has emerged as the strongest treatment against severe COVID-19 (Table 1).

Alternatively, many efforts to repurpose drugs for COVID-19 have built on information gained through basic scientific research of HCoV. Understanding how related viruses function has allowed researchers to identify possible pharmacological strategies to disrupt pathogenesis (Figure 5). Some of the compounds identified through these methods include small-molecule antivirals, which can be boutique and experimental medications like remdesivir (Table 1). Other candidate drugs that intercept host-pathogen interactions include biologics, which imitate the function of endogenous host compounds. Most notably, several mAbs that have been developed (casirivimab, imdevimab, bamlanivimab and etesevimab) or repurposed (sotrovimab, tocilizumab) have now been granted EUAs (Table 1). Although not discussed here, several vaccine development programs have also met huge success using a range of strategies (5, 6).

Table 1: Summary table of candidate therapeutics examined in this manuscript. “Grade” is the rating given to each treatment by the Systematic Tracker of Off-label/Repurposed Medicines Grades (STORM) maintained by the Center for Cytokine Storm Treatment & Laboratory (CSTL) at the University of Pennsylvania (788). A grade of A indicates that a treatment is considered effective, B that all or most RCTs have shown positive results, C that RCT data are not yet available, and D that multiple RCTs have produced negative results. Treatments not in the STORM database are indicated as N/A. FDA status is also provided where available. The evidence available is based on the progression of the therapeutic through the pharmaceutical development pipeline, with RCTs as the most informative source of evidence. The effectiveness is summarized based on the current available evidence; large trials such as RECOVERY and Solidarity are weighted heavily in this summary. This table was last updated on August 20, 2021.

Treatment	Grade	Category	FDA Status	Evidence Available	Suggested Effectiveness
Dexamethasone	A	Small molecule, broad spectrum	Used off-label	RCT	Supported: RCT shows improved outcomes over SOC, especially in severe cases such as CRS

Treatment	Grade	Category	FDA Status	Evidence Available	Suggested Effectiveness
Remdesivir	A	Small molecule, antiviral, adenosine analog	Approved for COVID-19 (and EUA for combination with baricitinib)	RCT	Mixed: Conflicting evidence from large WHO-led Solidarity trial vs US-focused RCT and other studies
Tocilizumab	A	Biologic, monoclonal antibody	EUA	RCT	Mixed: It appears that TCZ may work well in combination with dexamethasone in severe cases, but not as monotherapy
Sotrovimab	N/A	Biologic, monoclonal antibody	EUA	RCT	Supported: Phase 2/3 clinical trial showed reduced hospitalization/death
Bamlanivimab and etesevimab	B & N/A	Biologic, monoclonal antibodies	EUA	RCT	Supported: Phase 2 clinical trial showed reduction in viral load, but FDA pause recommended because may be less effective against Delta variant
Casirivimab and imdevimab	N/A	Biologic, monoclonal antibodies	EUA	RCT	Supported: Reduced viral load at interim analysis
Fluvoxamine	B	Small-molecule, Sigma-1 receptor agonist	N/A	RCT	Supported: Support from two small RCTs and preliminary support from interim analysis of TOGETHER
SNG001	B	Biologic, interferon	None	RCT	Mixed: Support from initial RCT but no effect found in WHO's Solidarity trial
M ^{Pro} Protease Inhibitors	N/A	Small molecule, protease inhibitor	None	Computational prediction, <i>in vitro</i> studies	Unknown

Treatment	Grade	Category	FDA Status	Evidence Available	Suggested Effectiveness
ARBs & ACEIs	C	Small molecule, broad spectrum	None	Observational studies and some RCTs	Not supported: Observational study retracted, RCTs suggest no association
Favipiravir	D	Small molecule, antiviral, nucleoside analog	None	RCT	Not supported: RCTs do not show significant improvements for individuals taking this treatment, good safety profile
HCQ/CQ	D	Small molecule, broad spectrum	None	RCT	Not supported, possibly harmful: Non-blinded RCTs showed no improvement over SOC, safety profile may be problematic
Convalescent plasma transfusion	D	Biologic, polyclonal antibodies	EUA	RCT	Mixed: Supported in small trials but not in large-scale RECOVERY trial
Ivermectin	D	Small molecule, broad spectrum	None	RCT	Mixed: Mixed results from small RCTs, major supporting RCT now withdrawn, preliminary results of large RCT (TOGETHER) suggest no effect on emergency room visits or hospitalization for COVID-19

All of the small-molecule drugs evaluated and most of the biologics are repurposed, and thus hinge on a theoretical understanding of how the virus interacts with a human host and how pharmaceuticals can be used to modify those interactions rather than being designed specifically against SARS-CoV-2 or COVID-19. As a result, significant attention has been paid to computational approaches that automate the identification of potentially desirable interactions. However, work in COVID-19 has made it clear that relevant compounds can also be masked by confounds, and spurious associations can drive investment in candidate therapeutics that are unlikely to translate to the clinic. Such spurious hits are especially likely to impact hypothesis-free screens. However, hypothesis-free screens may still be able to contribute to the drug discovery or repurposing ecosystem, assuming the computational arm of HTS follows the same trends seen in its experimental arm. In 2011, a landmark study in drug discovery demonstrated that although more new drugs were discovered using target-based rather than phenotypic approaches, the majority of drugs with a novel molecular mechanism of action (MMOA) were identified in phenotypic screens ([994](#)). This pattern

applied only to first-in-class drugs, with most follower drugs produced by target-based screening (952). These findings suggest that target-based drug discovery is more successful when building on a known MMOA, and that modulating a target is most valuable when the target is part of a valuable MMOA (953). Building on this, many within the field suggested that mechanism-informed phenotypic investigations may be the most useful approach to drug discovery (951, 953, 955). As it stands, data-driven efforts to identify patterns in the results of computational screens allowed researchers to notice the shared property of cationic amphiphilicity among many of the hits from computational screening analyses (982). While easier said than done, efforts to fill in the black box underlying computational HTS and recognize patterns among the identified compounds aid in moving data-oriented drug repurposing efforts in this direction.

The unpredictable nature of success and failure in drug repurposing for COVID-19 thus highlights one of the tenets of phenotypic screening: there are a lot of “unknown unknowns”, and a promising mechanism at the level of an MMOA will not necessarily propagate up to the pathway, cellular, or organismal level (951). Despite the fact that apparently mechanistically relevant drugs may exist, identifying effective treatments for a new viral disease is extremely challenging. Targets of repurposed drugs are often non-specific, meaning that the MMOA can appear to be relevant to COVID-19 without a therapeutic or prophylactic effect being observed in clinical trials. The difference in the current status of remdesivir and favipiravir as treatments for COVID-19 (Table 1) underscores how difficult to predict whether a specific compound will produce a desired effect, even when the mechanisms are similar. Furthermore, the fact that many candidate COVID-19 therapeutics were ultimately identified because of their shared propensity to induce phospholipidosis underscores how challenging it can be to identify a mechanism *in silico* or *in vitro* that will translate to a successful treatment. While significant progress has been made thus far in the pandemic, the therapeutic landscape is likely to continue to evolve as more results become available from clinical trials and as efforts to develop novel therapeutics for COVID-19 progress.

4.9 Towards the Next HCoV Threat

Only very limited testing of candidate therapies was feasible during the SARS and MERS epidemics, and as a result, few treatments were available at the outset of the COVID-19 pandemic. Even corticosteroids, which were used to treat SARS patients, were a controversial therapeutic prior to the release of the results of the large RECOVERY trial. The scale and duration of the COVID-19 pandemic has made it possible to conduct large, rigorous RCTs such as RECOVERY, Solidarity, TOGETHER, and others. As results from these trials have continued to emerge, it has become clear that small clinical trials often produce spurious results. In the case of HCQ/CQ, the therapeutic had already attracted so much attention based on small, preliminary (and in some cases, methodologically concerning) studies that it took the results of multiple large studies before attention began to be redirected to more promising candidates (995). In fact, most COVID-19 clinical trials lack the statistical power to reliably test their hypotheses (996, 997). In the face of an urgent crisis like COVID-19, the desire to act quickly is understandable, but it is imperative that studies maintain strict standards of scientific rigor (948, 986),

especially given the potential dangers of politicization, as illustrated by HCQ/CQ ([998](#)). Potential innovations in clinical trial structure, such as adaptable clinical trials with master protocols ([999](#)) or the sharing of data among small clinical trials ([997](#)) may help to address future crises and to bolster the results from smaller studies, respectively.

In the long-term, new drugs specific for treatment of COVID-19 may also enter development. Development of novel drugs is likely to be guided by what is known about the pathogenesis and molecular structure of SARS-CoV-2. For example, understanding the various structural components of SARS-CoV-2 may allow for the development of small molecule inhibitors of those components. Crystal structures of the SARS-CoV-2 main protease have been resolved ([961](#), [1000](#)). Much work remains to be done to determine further crystal structures of other viral components, understand the relative utility of targeting different viral components, perform additional small molecule inhibitor screens, and determine the safety and efficacy of the potential inhibitors. While still nascent, work in this area is promising. Over the longer term, this approach and others may lead to the development of novel therapeutics specifically for COVID-19 and SARS-CoV-2. Such efforts are likely to prove valuable in managing future emergent HCoV, just as research from the SARS and MERS pandemic has provided a basis for the COVID-19 response.

5 Appendix: Identification and Development of Therapeutics for COVID-19

5.1 Dexamethasone

In order to understand how dexamethasone reduces inflammation, it is necessary to consider the stress response broadly. In response to stress, corticotropin-releasing hormone stimulates the release of neurotransmitters known as catecholamines, such as epinephrine, and steroid hormones known as glucocorticoids, such as cortisol ([1001](#), [1002](#)). While catecholamines are often associated with the fight-or-flight response, the specific role that glucocorticoids play is less clear, although they are thought to be important to restoring homeostasis ([1003](#)). Immune challenge is a stressor that is known to interact closely with the stress response. The immune system can therefore interact with the central nervous system; for example, macrophages can both respond to and produce catecholamines ([1001](#)). Additionally, the production of both catecholamines and glucocorticoids is associated with inhibition of proinflammatory cytokines such as IL-6, IL-12, and tumor necrosis factor- α (TNF- α) and the stimulation of anti-inflammatory cytokines such as IL-10, meaning that the stress response can regulate inflammatory immune activity ([1002](#)). Administration of dexamethasone has been found to correspond to dose-dependent inhibition of IL-12 production, but not to affect IL-10 ([1004](#)); the fact that this relationship could be disrupted by administration of a glucocorticoid-receptor antagonist suggests that it is regulated by the receptor itself ([1004](#)). Thus, the administration of dexamethasone for COVID-19 is likely to simulate the release of

glucocorticoids endogenously during stress, resulting in binding of the synthetic steroid to the glucocorticoid receptor and the associated inhibition of the production of proinflammatory cytokines. In this model, dexamethasone reduces inflammation by stimulating the biological mechanism that reduces inflammation following a threat such as immune challenge.

Initial support for dexamethasone as a treatment for COVID-19 came from the United Kingdom's RECOVERY trial ([780](#)), which assigned over 6,000 hospitalized COVID-19 patients to the standard of care (SOC) or treatment (dexamethasone) arms of the trial at a 2:1 ratio. At the time of randomization, some patients were ventilated (16%), others were on non-invasive oxygen (60%), and others were breathing independently (24%). Patients in the treatment arm were administered dexamethasone either orally or intravenously at 6 mg per day for up to 10 days. The primary end-point was the patient's status at 28-days post-randomization (mortality, discharge, or continued hospitalization), and secondary outcomes analyzed included the progression to invasive mechanical ventilation over the same period. The 28-day mortality rate was found to be lower in the treatment group than in the SOC group (21.6% vs 24.6%, $p < 0.001$). However, the effect was driven by improvements in patients receiving mechanical ventilation or supplementary oxygen. One possible confounder is that patients receiving mechanical ventilation tended to be younger than patients who were not receiving respiratory support (by 10 years on average) and to have had symptoms for a longer period. However, adjusting for age did not change the conclusions, although the duration of symptoms was found to be significantly associated with the effect of dexamethasone administration. Thus, this large, randomized, and multi-site, albeit not placebo-controlled, study suggests that administration of dexamethasone to patients who are unable to breathe independently may significantly improve survival outcomes. Additionally, dexamethasone is a widely available and affordable medication, raising the hope that it could be made available to COVID-19 patients globally.

It is not surprising that administration of an immunosuppressant would be most beneficial in severe cases where the immune system was dysregulated towards inflammation. However, it is also unsurprising that care must be taken in administering an immunosuppressant to patients fighting a viral infection. In particular, the concern has been raised that treatment with dexamethasone might increase patient susceptibility to concurrent (e.g., nosocomial) infections ([1005](#)). Additionally, the drug could potentially slow viral clearance and inhibit patients' ability to develop antibodies to SARS-CoV-2 ([777](#), [1005](#)), with the lack of data about viral clearance being put forward as a major limitation of the RECOVERY trial ([1006](#)). Furthermore, dexamethasone has been associated with side effects that include psychosis, glucocorticoid-induced diabetes, and avascular necrosis ([777](#)), and the RECOVERY trial did not report outcomes with enough detail to be able to determine whether they observed similar complications. The effects of dexamethasone have also been found to differ among populations, especially in high-income versus middle- or low-income countries ([1007](#)). However, since the RECOVERY trial's results were released, strategies have been proposed for administering dexamethasone alongside more targeted

treatments to minimize the likelihood of negative side effects ([1005](#)). Given the available evidence, dexamethasone is currently the most promising treatment for severe COVID-19.

5.2 Favipiravir

The effectiveness of favipiravir for treating patients with COVID-19 is currently under investigation. Evidence for the drug inhibiting viral RNA polymerase are based on time-of-drug addition studies that found that viral loads were reduced with the addition of favipiravir in early times post-infection ([798](#), [801](#), [802](#)). An open-label, nonrandomized, before-after controlled study for COVID-19 was recently conducted ([1008](#)). The study included 80 COVID-19 patients (35 treated with favipiravir, 45 control) from the isolation ward of the National Clinical Research Center for Infectious Diseases (The Third People's Hospital of Shenzhen), Shenzhen, China. The patients in the control group were treated with other antivirals, such as lopinavir and ritonavir. It should be noted that although the control patients received antivirals, two subsequent large-scale analyses, the WHO Solidarity trial and the Randomized Evaluation of COVID-19 Therapy (RECOVERY) trial, identified no effect of lopinavir or of a lopinavir-ritonavir combination, respectively, on the metrics of COVID-19-related mortality that each assessed ([811](#), [1009](#), [1010](#)). Treatment was applied on days 2-14; treatment stopped either when viral clearance was confirmed or at day 14. The efficacy of the treatment was measured by, first, the time until viral clearance using Kaplan-Meier survival curves, and, second, the improvement rate of chest computed tomography (CT) scans on day 14 after treatment. The study found that favipiravir increased the speed of recovery, measured as viral clearance from the patient by RT-PCR, with patients receiving favipiravir recovering in four days compared to 11 days for patients receiving antivirals such as lopinavir and ritonavir. Additionally, the lung CT scans of patients treated with favipiravir showed significantly higher improvement rates (91%) on day 14 compared to control patients (62%, $p = 0.004$). However, there were adverse side effects in 4 (11%) favipiravir-treated patients and 25 (56%) control patients. The adverse side effects included diarrhea, vomiting, nausea, rash, and liver and kidney injury. Despite the study reporting clinical improvement in favipiravir-treated patients, several study design issues are problematic and lower confidence in the overall conclusions. For example, the study was neither randomized nor blinded. Moreover, the selection of patients did not take into consideration important factors such as previous clinical conditions or sex, and there was no age categorization. Additionally, it should be noted that this study was temporarily retracted and then restored without an explanation ([1011](#)).

In late 2020 and early 2021, the first randomized controlled trials of favipiravir for the treatment of COVID-19 released results ([1012](#)–[1014](#)). One study ([1013](#)) was retracted in November 2021 due to concerns about the data. Of the two remaining, the first ([1012](#)) used a randomized, controlled, open-label design to compare two drugs, favipiravir and baloxavir marboxil, to SOC alone. Here, SOC included antivirals such as lopinavir/ritonavir and was administered to all patients. The primary endpoint analyzed was viral clearance at day 14. The sample size for this study was very small, with 29 total patients enrolled, and no significant effect of the treatments was found for the primary or any of the secondary outcomes analyzed, which included

mortality. The second trial examined 60 patients and reported a significant effect of favipiravir on viral clearance at four days (a secondary endpoint), but not at 10 days (the primary endpoint) ([1014](#)). This study, as well as a prior study of favipiravir ([1015](#)), also reported that the drug was generally well-tolerated. Thus, in combination, these small studies suggest that the effects of favipiravir as a treatment for COVID-19 cannot be determined based on the available evidence, but additionally, none raise major concerns about the safety profile of the drug.

5.3 Remdesivir

At the outset of the COVID-19 pandemic, remdesivir did not have any FDA-approved use. A clinical trial in the Democratic Republic of Congo found some evidence of effectiveness against ebola virus disease (EVD), but two antibody preparations were found to be more effective, and remdesivir was not pursued ([1016](#)). Remdesivir also inhibits polymerase and replication of the coronaviruses MERS-CoV and SARS-CoV-1 in cell culture assays with submicromolar IC50s ([1017](#)). It has also been found to inhibit SARS-CoV-2, showing synergy with CQ *in vitro* ([806](#)).

Remdesivir was first used on some COVID-19 patients under compassionate use guidelines ([1020](#)). All were in late stages of COVID-19 infection, and initial reports were inconclusive about the drug's efficacy. Gilead Sciences, the maker of remdesivir, led a recent publication that reported outcomes for compassionate use of the drug in 61 patients hospitalized with confirmed COVID-19. Here, 200 mg of remdesivir was administered intravenously on day 1, followed by a further 100 mg/day for 9 days ([810](#)). There were significant issues with the study design, or lack thereof. There was no randomized control group. The inclusion criteria were variable: some patients only required low doses of oxygen, while others required ventilation. The study included many sites, potentially with variable inclusion criteria and treatment protocols. The patients analyzed had mixed demographics. There was a short follow-up period of investigation. Eight patients were excluded from the analysis mainly due to missing post-baseline information; thus, their health was unaccounted for. Therefore, even though the study reported clinical improvement in 68% of the 53 patients ultimately evaluated, due to the significant issues with study design, it could not be determined whether treatment with remdesivir had an effect or whether these patients would have recovered regardless of treatment. Another study comparing 5- and 10-day treatment regimens reported similar results but was also limited because of the lack of a placebo control ([1021](#)). These studies did not alter the understanding of the efficacy of remdesivir in treating COVID-19, but the encouraging results provided motivation for placebo-controlled studies.

The double-blind placebo-controlled ACTT-1 trial ([807](#), [808](#)) recruited 1,062 patients and randomly assigned them to placebo treatment or treatment with remdesivir. Patients were stratified for randomization based on site and the severity of disease presentation at baseline ([807](#)). The treatment was 200 mg on day 1, followed by 100 mg on days 2 through 10. Data was analyzed from a total of 1,059 patients who completed the 29-day course of the trial, with 517 assigned to remdesivir and 508 to placebo ([807](#)). The two groups were well matched demographically and clinically at baseline. Those who

received remdesivir had a median recovery time of 10 days, as compared with 15 days in those who received placebo (rate ratio for recovery, 1.29; 95% confidence interval (CI), 1.12 to 1.49; $p < 0.001$). The Kaplan-Meier estimates of mortality by 14 days were 6.7% with remdesivir and 11.9% with placebo, with a hazard ratio (HR) for death of 0.55 and a 95% CI of 0.36 to 0.83, and at day 29, remdesivir corresponded to 11.4% and the placebo to 15.2% (HR: 0.73; 95% CI, 0.52 to 1.03). Serious adverse events were reported in 131 of the 532 patients who received remdesivir (24.6%) and in 163 of the 516 patients in the placebo group (31.6%). This study also reported an association between remdesivir administration and both clinical improvement and a lack of progression to more invasive respiratory intervention in patients receiving non-invasive and invasive ventilation at randomization (807). Largely on the results of this trial, the FDA reissued and expanded the EUA for remdesivir for the treatment of hospitalized COVID-19 patients ages twelve and older (1022). Additional clinical trials (806, 1023–1026) are currently underway to evaluate the use of remdesivir to treat COVID-19 patients at both early and late stages of infection and in combination with other drugs (Figure 4). As of October 22, 2020, remdesivir received FDA approval based on three clinical trials (1027).

However, results suggesting no effect of remdesivir on survival were reported by the WHO Solidarity trial (811). Patients were randomized in equal proportions into four experimental conditions and a control condition, corresponding to four candidate treatments for COVID-19 and SOC, respectively; no placebo was administered. The 2,750 patients in the remdesivir group were administered 200 mg intravenously on the first day and 100 mg on each subsequent day until day 10 and assessed for in-hospital death (primary endpoint), duration of hospitalization, and progression to mechanical ventilation. There were also 2,708 control patients who would have been eligible and able to receive remdesivir were they not assigned to the control group. A total of 604 patients among these two cohorts died during initial hospitalization, with 301 in the remdesivir group and 303 in the control group. The rate ratio of death between these two groups was therefore not significant (0.95, $p = 0.50$), suggesting that the administration of remdesivir did not affect survival. The two secondary analyses similarly did not find any effect of remdesivir. Additionally, the authors compared data from their study with data from three other studies of remdesivir (including (807)) stratified by supplemental oxygen status. A meta-analysis of the four studies yielded an overall rate ratio for death of 0.91 ($p = 0.20$). These results thus do not support the previous findings that remdesivir reduced median recovery time and mortality risk in COVID-19 patients.

In response to the results of the Solidarity trial, Gilead, which manufactures remdesivir, released a statement pointing to the fact that the Solidarity trial was not placebo-controlled or double-blind and at the time of release, the statement had not been peer reviewed (1028); these sentiments have been echoed elsewhere (1029). Other critiques of this study have noted that antivirals are not typically targeted at patients with severe illness, and therefore remdesivir could be more beneficial for patients with mild rather than severe cases (1010, 1030). However, the publication associated with the trial sponsored by Gilead did purport an effect of remdesivir on patients with severe disease, identifying an 11 versus 18 day recovery period (rate ratio for

recovery: 1.31, 95% CI 1.12 to 1.52) ([807](#)). Additionally, a smaller analysis of 598 patients, of whom two-thirds were randomized to receive remdesivir for either 5 or 10 days, reported a small effect of treatment with remdesivir for five days relative to standard of care in patients with moderate COVID-19 ([1031](#)). These results suggest that remdesivir could improve outcomes for patients with moderate COVID-19, but that additional information would be needed to understand the effects of different durations of treatment. Therefore, the Solidarity trial may point to limitations in the generalizability of other research on remdesivir, especially since the broad international nature of the Solidarity clinical trial, which included countries with a wide range of economic profiles and a variety of healthcare systems, provides a much-needed global perspective in a pandemic ([1010](#)). On the other hand, only 62% of patients in the Solidarity trial were randomized on the day of admission or one day afterwards ([811](#)), and concerns have been raised that differences in disease progression could influence the effectiveness of remdesivir ([1010](#)). Despite the findings of the Solidarity trial, remdesivir remains available for the treatment of COVID-19 in many places. Remdesivir has also been investigated in combination with other drugs, such as baricitinib, which is an inhibitor of Janus kinase 1 and 2 ([1032](#)); the FDA has issued an EUA for the combination of remdesivir and baricitinib in adult and pediatric patients ([1033](#)). Follow-up studies are needed and, in many cases, are underway to further investigate remdesivir-related outcomes.

Similarly, the extent to which the remdesivir dosing regimen could influence outcomes continues to be under consideration. A randomized, open-label trial compared the effect of remdesivir on 397 patients with severe COVID-19 over 5 versus 10 days ([809](#), [1021](#)), complementing the study that found that a 5-day course of remdesivir improved outcomes for patients with moderate COVID-19 but a 10-day course did not ([1031](#)). Patients in the two groups were administered 200 mg of remdesivir intravenously on the first day, followed by 100 mg on the subsequent four or nine days, respectively. The two groups differed significantly in their clinical status, with patients assigned to the 10-day group having more severe illness. This study also differed from most because it included not only adults, but also pediatric patients as young as 12 years old. It reported no significant differences across several outcomes for patients receiving a 5-day or 10-day course, when correcting for baseline clinical status. The data did suggest that the 10-day course might reduce mortality in the most severe patients at day 14, but the representation of this group in the study population was too low to justify any conclusions ([1021](#)). Thus, additional research is also required to determine whether the dosage and duration of remdesivir administration influences outcomes.

In summary, remdesivir is the first FDA approved anti-viral against SARS-CoV-2 as well as the first FDA approved COVID-19 treatment. Early investigations of this drug established proof of principle that drugs targeting the virus can benefit COVID-19 patients. Moreover, one of the most successful strategies for developing therapeutics for viral diseases is to target the viral replication machinery, which are typically virally encoded polymerases. Small molecule drugs targeting viral polymerases are the backbones of treatments for other viral diseases including human immunodeficiency virus (HIV) and herpes. Notably, the HIV and herpes polymerases are a reverse transcriptase and a DNA polymerase, respectively, whereas SARS-CoV-2 encodes an RdRP, so most of the commonly used polymerase inhibitors are not likely to be active

against SARS-CoV-2. In clinical use, polymerase inhibitors show short term benefits for HIV patients, but for long term benefits they must be part of combination regimens. They are typically combined with protease inhibitors, integrase inhibitors, and even other polymerase inhibitors. Remdesivir provides evidence that a related approach may be beneficial for the treatment of COVID-19.

5.4 Hydroxychloroquine and Chloroquine

CQ and hydroxychloroquine (HCQ) increase cellular pH by accumulating in their protonated form inside lysosomes ([813](#), [1034](#)). This shift in pH inhibits the breakdown of proteins and peptides by the lysosomes during the process of proteolysis ([813](#)). Interest in CQ and HCQ for treating COVID-19 was catalyzed by a mechanism observed in *in vitro* studies of both SARS-CoV-1 and SARS-CoV-2. In one study, CQ inhibited viral entry of SARS-CoV-1 into Vero E6 cells, a cell line that was derived from Vero cells in 1968, through the elevation of endosomal pH and the terminal glycosylation of ACE2 ([814](#)). Increased pH within the cell, as discussed above, inhibits proteolysis, and terminal glycosylation of ACE2 is thought to interfere with virus-receptor binding. An *in vitro* study of SARS-CoV-2 infection of Vero cells found both HCQ and CQ to be effective in inhibiting viral replication, with HCQ being more potent ([815](#)). Additionally, an early case study of three COVID-19 patients reported the presence of antiphospholipid antibodies in all three patients ([100](#)). Antiphospholipid antibodies are central to the diagnosis of the antiphospholipid syndrome, a disorder that HCQ has often been used to treat ([1035–1037](#)). Because the 90% effective concentration (EC₉₀) of CQ in Vero E6 cells (6.90 μM) can be achieved in and tolerated by rheumatoid arthritis (RA) patients, it was hypothesized that it might also be possible to achieve the effective concentration in COVID-19 patients ([1038](#)). Additionally, clinical trials have reported HCQ to be effective in treating HIV ([1039](#)) and chronic Hepatitis C ([1040](#)). Together, these studies triggered initial enthusiasm about the therapeutic potential for HCQ and CQ against COVID-19. HCQ/CQ has been proposed both as a treatment for COVID-19 and a prophylaxis against SARS-CoV-2 exposure, and trials often investigated these drugs in combination with azithromycin (AZ) and/or zinc supplementation. However, as more evidence has emerged, it has become clear that HCQ/CQ offer no benefits against SARS-CoV-2 or COVID-19.

5.4.1 Trials Assessing Therapeutic Administration of HCQ/CQ

The initial study evaluating HCQ as a treatment for COVID-19 patients was published on March 20, 2020 by Gautret et al. ([816](#)). This non-randomized, non-blinded, non-placebo clinical trial compared HCQ to SOC in 42 hospitalized patients in southern France. It reported that patients who received HCQ showed higher rates of virological clearance by nasopharyngeal swab on days 3-6 when compared to SOC. This study also treated six patients with both HCQ + AZ and found this combination therapy to be more effective than HCQ alone. However, the design and analyses used showed weaknesses that severely limit interpretability of results, including the small sample size and the lack of: randomization, blinding, placebo (no “placebo pill” given to SOC group), Intention-To-Treat analysis, correction for

sequential multiple comparisons, and trial pre-registration. Furthermore, the trial arms were entirely confounded by the hospital and there were false negative outcome measurements (see [\(1041\)](#)). Two of these weaknesses are due to inappropriate data analysis and can therefore be corrected *post hoc* by recalculating the p-values (lack of Intention-To-Treat analysis and multiple comparisons). However, all other weaknesses are fundamental design flaws and cannot be corrected for. Thus, the conclusions cannot be generalized outside of the study. The International Society of Antimicrobial Chemotherapy, the scientific organization that publishes the journal where the article appeared, subsequently announced that the article did not meet its expected standard for publications ([817](#)), although it has not been officially retracted.

Because of the preliminary data presented in this study, HCQ treatment was subsequently explored by other researchers. About one week later, a follow-up case study reported that 11 consecutive patients were treated with HCQ + AZ using the same dosing regimen ([1042](#)). One patient died, two were transferred to the intensive care unit (ICU), and one developed a prolonged QT interval, leading to discontinuation of HCQ + AZ administration. As in the Gautret et al. study, the outcome assessed was virological clearance at day 6 post-treatment, as measured from nasopharyngeal swabs. Of the ten living patients on day 6, eight remained positive for SARS-CoV-2 RNA. Like in the original study, interpretability was severely limited by the lack of a comparison group and the small sample size. However, these results stand in contrast to the claims by Gautret et al. that all six patients treated with HCQ + AZ tested negative for SARS-CoV-2 RNA by day 6 post-treatment. This case study illustrated the need for further investigation using robust study design to evaluate the efficacy of HCQ and/or CQ.

On April 10, 2020, a randomized, non-placebo trial of 62 COVID-19 patients at the Renmin Hospital of Wuhan University was released ([1043](#)). This study investigated whether HCQ decreased time to fever break or time to cough relief when compared to SOC ([1043](#)). This trial found HCQ decreased both average time to fever break and average time to cough relief, defined as mild or no cough. While this study improved on some of the methodological flaws in Gautret et al. by randomizing patients, it also had several flaws in trial design and data analysis that prevent generalization of the results. These weaknesses include the lack of placebo, lack of correction for multiple primary outcomes, inappropriate choice of outcomes, lack of sufficient detail to understand analysis, drastic disparities between pre-registration ([1044](#)) and published protocol (including differences in the inclusion and exclusion criteria, the number of experimental groups, the number of patients enrolled, and the outcome analyzed), and small sample size. The choice of outcomes may be inappropriate as both fevers and cough may break periodically without resolution of illness. Additionally, for these outcomes, the authors reported that 23 of 62 patients did not have a fever and 25 of 62 patients did not have a cough at the start of the study, but the authors failed to describe how these patients were included in a study assessing time to fever break and time to cough relief. It is important to note here that the authors claimed “neither the research performers nor the patients were aware of the treatment assignments.” This blinding seems impossible in a non-placebo trial because at the very least, providers would know whether they were administering a medication or not, and this knowledge could lead

to systematic differences in the administration of care. Correction for multiple primary outcomes can be adjusted *post hoc* by recalculating p-values, but all of the other issues were design and statistical weaknesses that cannot be corrected for. Additionally, disparities between the pre-registered and published protocols raise concerns about experimental design. The design limitations mean that the conclusions cannot be generalized outside of the study.

A second randomized trial, conducted by the Shanghai Public Health Clinical Center, analyzed whether HCQ increased rates of virological clearance at day 7 in respiratory pharyngeal swabs compared to SOC ([1045](#)). This trial was published in Chinese along with an abstract in English, and only the English abstract was read and interpreted for this review. The trial found comparable outcomes in virological clearance rate, time to virological clearance, and time to body temperature normalization between the treatment and control groups. The small sample size is one weakness, with only 30 patients enrolled and 15 in each arm. This problem suggests the study is underpowered to detect potentially useful differences and precludes interpretation of results. Additionally, because only the abstract could be read, other design and analysis issues could be present. Thus, though these studies added randomization to their assessment of HCQ, their conclusions should be interpreted very cautiously. These two studies assessed different outcomes and reached differing conclusions about the efficacy of HCQ for treating COVID-19; the designs of both studies, especially with respect to sample size, meant that no general conclusions can be made about the efficacy of the drug.

Several widely reported studies on HCQ also have issues with data integrity and/or provenance. A Letter to the Editor published in *BioScience Trends* on March 16, 2020 claimed that numerous clinical trials have shown that HCQ is superior to control treatment in inhibiting the exacerbation of COVID-19 pneumonia ([1046](#)). This letter has been cited by numerous primary literature, review articles, and media alike ([1047](#), [1048](#)). However, the letter referred to 15 pre-registration identifiers from the Chinese Clinical Trial Registry. When these identifiers are followed back to the registry, most trials claim they are not yet recruiting patients or are currently recruiting patients. For all of these 15 identifiers, no data uploads or links to publications could be located on the pre-registrations. At the very least, the lack of availability of the primary data means the claim that HCQ is efficacious against COVID-19 pneumonia cannot be verified. Similarly, a recent multinational registry analysis ([1049](#)) analyzed the efficacy of CQ and HCQ with and without a macrolide, which is a class of antibiotics that includes Azithromycin, for the treatment of COVID-19. The study observed 96,032 patients split into a control and four treatment conditions (CQ with and without a macrolide; HCQ with and without a macrolide). They concluded that treatment with CQ or HCQ was associated with increased risk of *de novo* ventricular arrhythmia during hospitalization. However, this study has since been retracted by *The Lancet* due to an inability to validate the data used ([1050](#)). These studies demonstrate that increased skepticism in evaluation of the HCQ/CQ and COVID-19 literature may be warranted, possibly because of the significant attention HCQ and CQ have received as possible treatments for COVID-19 and the politicization of these drugs.

Despite the fact that the study suggesting that CQ/HCQ increased risk of ventricular arrhythmia in COVID-19 patients has now been retracted, previous studies have identified risks associated with HCQ/CQ. A patient with systemic lupus erythematosus developed a prolonged QT interval that was likely exacerbated by use of HCQ in combination with renal failure ([1051](#)). A prolonged QT interval is associated with ventricular arrhythmia ([1052](#)). Furthermore, a separate study ([1053](#)) investigated the safety associated with the use of HCQ with and without macrolides between 2000 and 2020. The study involved 900,000 cases treated with HCQ and 300,000 cases treated with HCQ + AZ. The results indicated that short-term use of HCQ was not associated with additional risk, but that HCQ + AZ was associated with an enhanced risk of cardiovascular complications (such as a 15% increased risk of chest pain, calibrated HR = 1.15, 95% CI, 1.05 to 1.26) and a two-fold increased 30-day risk of cardiovascular mortality (calibrated HR = 2.19; 95% CI, 1.22 to 3.94). Therefore, whether studies utilize HCQ alone or HCQ in combination with a macrolide may be an important consideration in assessing risk. As results from initial investigations of these drug combinations have emerged, concerns about the efficacy and risks of treating COVID-19 with HCQ and CQ have led to the removal of CQ/HCQ from SOC practices in several countries ([1054](#), [1055](#)). As of May 25, 2020, WHO had suspended administration of HCQ as part of the worldwide Solidarity Trial ([1056](#)), and later the final results of this large-scale trial that compared 947 patients administered HCQ to 906 controls revealed no effect on the primary outcome, mortality during hospitalization (rate ratio: 1.19; $p = 0.23$)

Additional research has emerged largely identifying HCQ/CQ to be ineffective against COVID-19 while simultaneously revealing a number of significant side effects. A randomized, open-label, non-placebo trial of 150 COVID-19 patients was conducted in parallel at 16 government-designated COVID-19 centers in China to assess the safety and efficacy of HCQ ([1057](#)). The trial compared treatment with HCQ in conjunction with SOC to SOC alone in 150 infected patients who were assigned randomly to the two groups (75 per group). The primary endpoint of the study was the negative conversion rate of SARS-CoV-2 in 28 days, and the investigators found no difference in this parameter between the groups (estimated difference between SOC plus HCQ and SOC 4.1%; 95% CI, -10.3% to 18.5%). The secondary endpoints were an amelioration of the symptoms of the disease such as axillary temperature $\leq 36.6^{\circ}\text{C}$, SpO₂ >94% on room air, and disappearance of symptoms like shortness of breath, cough, and sore throat. The median time to symptom alleviation was similar across different conditions (19 days in HCQ + SOC versus 21 days in SOC, $p = 0.97$). Additionally, 30% of the patients receiving SOC+HCQ reported adverse outcomes compared to 8.8% of patients receiving only SOC, with the most common adverse outcome in the SOC+HCQ group being diarrhea (10% versus 0% in the SOC group, $p = 0.004$). However, there are several factors that limit the interpretability of this study. Most of the enrolled patients had mild-to-moderate symptoms (98%), and the average age was 46. SOC in this study included the use of antivirals (Lopinavir-Ritonavir, Arbidol, Oseltamivir, Virazole, Entecavir, Ganciclovir, and Interferon alfa), which the authors note could influence the results. Thus, they note that an ideal SOC would need to exclude the use of antivirals, but that ceasing antiviral treatment raised ethical concerns at the time that the study was conducted. In this trial, the samples used to test for the presence of the SARS-CoV-2 virus were collected from the upper respiratory tract, and

the authors indicated that the use of upper respiratory samples may have introduced false negatives (e.g., [\(71\)](#)). Another limitation of the study that the authors acknowledge was that the HCQ treatment began, on average, at a 16-day delay from the symptom onset. The fact that this study was open-label and lacked a placebo limits interpretation, and additional analysis is required to determine whether HCQ reduces inflammatory response. Therefore, despite some potential areas of investigation identified in *post hoc* analysis, this study cannot be interpreted as providing support for HCQ as a therapeutic against COVID-19. This study provided no support for HCQ against COVID-19, as there was no difference between the two groups in either negative seroconversion at 28 days or symptom alleviation, and in fact, more severe adverse outcomes were reported in the group receiving HCQ.

Additional evidence comes from a retrospective analysis ([1058](#)) that examined data from 368 COVID-19 patients across all United States Veteran Health Administration medical centers. The study retrospectively investigated the effect of the administration of HCQ (n=97), HCQ + AZ (n=113), and no HCQ (n=158) on 368 patients. The primary outcomes assessed were death and the need for mechanical ventilation. Standard supportive care was rendered to all patients. Due to the low representation of women (N=17) in the available data, the analysis included only men, and the median age was 65 years. The rate of death was 27.8% in the HCQ-only treatment group, 22.1% in the HCQ + AZ treatment group, and 14.1% in the no-HCQ group. These data indicated a statistically significant elevation in the risk of death for the HCQ-only group compared to the no-HCQ group (adjusted HR: 2.61, $p = 0.03$), but not for the HCQ + AZ group compared to the no-HCQ group (adjusted HR: 1.14; $p = 0.72$). Further, the risk of ventilation was similar across all three groups (adjusted HR: 1.43, $p = 0.48$ (HCQ) and 0.43, $p = 0.09$ (HCQ + AZ) compared to no HCQ). The study thus showed evidence of an association between increased mortality and HCQ in this cohort of COVID-19 patients but no change in rates of mechanical ventilation among the treatment conditions. The study had a few limitations: it was not randomized, and the baseline vital signs, laboratory tests, and prescription drug use were significantly different among the three groups. All of these factors could potentially influence treatment outcome. Furthermore, the authors acknowledge that the effect of the drugs might be different in females and pediatric subjects, since these subjects were not part of the study. The reported result that HCQ + AZ is safer than HCQ contradicts the findings of the previous large-scale analysis of twenty years of records that found HCQ + AZ to be more frequently associated with cardiac arrhythmia than HCQ alone ([1053](#)); whether this discrepancy is caused by the pathology of COVID-19, is influenced by age or sex, or is a statistical artifact is not presently known.

Finally, findings from the RECOVERY trial were released on October 8, 2020. This study used a randomized, open-label design to study the effects of HCQ compared to SOC in 11,197 patients at 176 hospitals in the United Kingdom ([818](#)). Patients were randomized into either the control group or one of the treatment arms, with twice as many patients enrolled in the control group as any treatment group. Of the patients eligible to receive HCQ, 1,561 were randomized into the HCQ arm, and 3,155 were randomized into the control arm. The demographics of the HCQ and control groups were similar in terms of average age (65 years), proportion female (approximately 38%), ethnic make-up (73% versus 76% white), and prevalence of pre-existing conditions

(56% versus 57% overall). In the HCQ arm of the study, patients received 800 mg at baseline and again after 6 hours, then 400 mg at 12 hours and every subsequent 12 hours. The primary outcome analyzed was all-cause mortality, and patient vital statistics were reported by physicians upon discharge or death, or else at 28 days following HCQ administration if they remained hospitalized. The secondary outcome assessed was the combined risk of progression to invasive mechanical ventilation or death within 28 days. By the advice of an external data monitoring committee, the HCQ arm of the study was reviewed early, leading to it being closed due a lack of support for HCQ as a treatment for COVID-19. COVID-19-related mortality was not affected by HCQ in the RECOVERY trial (rate ratio, 1.09; 95% CI, 0.97 to 1.23; $p = 0.15$), but cardiac events were increased in the HCQ arm (0.4 percentage points), as was the duration of hospitalization (rate ratio for discharge alive within 28 days: 0.90; 95% CI, 0.83 to 0.98) and likelihood of progression to mechanical ventilation or death (risk ratio 1.14; 95% CI, 1.03 to 1.27). This large-scale study thus builds upon studies in the United States and China to suggest that HCQ is not an effective treatment, and in fact may negatively impact COVID-19 patients due to its side effects. Therefore, though none of the studies have been blinded, examining them together makes it clear that the available evidence points to significant dangers associated with the administration of HCQ to hospitalized COVID-19 patients, without providing any support for its efficacy.

5.4.2 HCQ for the Treatment of Mild Cases

One additional possible therapeutic application of HCQ considered was the treatment of mild COVID-19 cases in otherwise healthy individuals. This possibility was assessed in a randomized, open-label, multi-center analysis conducted in Catalonia (Spain) ([1059](#)). This analysis enrolled adults 18 and older who had been experiencing mild symptoms of COVID-19 for fewer than five days. Participants were randomized into an HCQ arm (N=136) and a control arm (N=157), and those in the treatment arm were administered 800 mg of HCQ on the first day of treatment followed by 400 mg on each of the subsequent six days. The primary outcome assessed was viral clearance at days 3 and 7 following the onset of treatment, and secondary outcomes were clinical progression and time to complete resolution of symptoms. No significant differences between the two groups were found: the difference in viral load between the HCQ and control groups was 0.01 (95% CI, -0.28 to 0.29) at day 3 and -0.07 (95% CI -0.44 to 0.29) at day 7, the relative risk of hospitalization was 0.75 (95% CI, 0.32 to 1.77), and the difference in time to complete resolution of symptoms was -2 days ($p = 0.38$). This study thus suggests that HCQ does not improve recovery from COVID-19, even in otherwise healthy adult patients with mild symptoms.

5.4.3 Prophylactic Administration of HCQ

An initial study of the possible prophylactic application of HCQ utilized a randomized, double-blind, placebo-controlled design to analyze the administration of HCQ prophylactically ([1060](#)). Asymptomatic adults in the United States and Canada who had been exposed to SARS-CoV-2 within the past four days were enrolled in an online study to evaluate whether administration of HCQ over five days influenced the probability of developing COVID-19 symptoms over a 14-day period. Of the participants, 414 received

HCQ and 407 received a placebo. No significant difference in the rate of symptomatic illness was observed between the two groups (11.8% HCQ, 14.3% placebo, $p = 0.35$). The HCQ condition was associated with side effects, with 40.1% of patients reporting side effects compared to 16.8% in the control group ($p < 0.001$). However, likely due to the high enrollment of healthcare workers (66% of participants) and the well-known side effects associated with HCQ, a large number of participants were able to correctly identify whether they were receiving HCQ or a placebo (46.5% and 35.7%, respectively). Furthermore, due to a lack of availability of diagnostic testing, only 20 of the 107 cases were confirmed with a PCR-based test to be positive for SARS-CoV-2. The rest were categorized as “probable” or “possible” cases by a panel of four physicians who were blind to the treatment status. One possible confounder is that a patient presenting one or more symptoms, which included diarrhea, was defined as a “possible” case, but diarrhea is also a common side effect of HCQ. Additionally, four of the twenty PCR-confirmed cases did not develop symptoms until after the observation period had completed, suggesting that the 14-day trial period may not have been long enough or that some participants also encountered secondary exposure events. Finally, in addition to the young age of the participants in this study, which ranged from 32 to 51, there were possible impediments to generalization introduced by the selection process, as 2,237 patients who were eligible but had already developed symptoms by day 4 were enrolled in a separate study. It is therefore likely that asymptomatic cases were over-represented in this sample, which would not have been detected based on the diagnostic criteria used. Therefore, while this study does represent the first effort to conduct a randomized, double-blind, placebo-controlled investigation of HCQ’s effect on COVID-19 prevention after SARS-CoV-2 exposure in a large sample, the lack of PCR tests and several other design flaws significantly impede interpretation of the results. However, in line with the results from therapeutic studies, once again no evidence was found suggesting an effect of HCQ against COVID-19.

A second study ([1061](#)) examined the effect of administering HCQ to healthcare workers as a pre-exposure prophylactic. The primary outcome assessed was the conversion from SARS-CoV-2 negative to SARS-CoV-2 positive status over the 8 week study period. This study was also randomized, double-blind, and placebo-controlled, and it sought to address some of the limitations of the first prophylactic study. The goal was to enroll 200 healthcare workers, preferentially those working with COVID-19 patients, at two hospitals within the University of Pennsylvania hospital system in Philadelphia, PA. Participants were randomized 1:1 to receive either 600 mg of HCQ daily or a placebo, and their SARS-CoV-2 infection status and antibody status were assessed using RT-PCR and serological testing, respectively, at baseline, 4 weeks, and 8 weeks following the beginning of the treatment period. The statistical design of the study accounted for interim analyses at 50 and 100 participants in case efficacy or futility of HCQ for prophylaxis became clear earlier than completion of enrollment. The 139 individuals enrolled comprised a study population that was fairly young (average age 33) and made of largely of people who were white, women, and without pre-existing conditions. At the second interim analysis, more individuals in the treatment group than the control group had contracted COVID-19 (4 versus

3), causing the estimated z-score to fall below the pre-established threshold for futility. As a result, the trial was terminated early, offering additional evidence against the use of HCQ for prophylaxis.

5.4.4 Summary of HCQ/CQ Research Findings

Early *in vitro* evidence indicated that HCQ could be an effective therapeutic against SARS-CoV-2 and COVID-19, leading to significant media attention and public interest in its potential as both a therapeutic and prophylactic. Initially it was hypothesized that CQ/HCQ might be effective against SARS-CoV-2 in part because CQ and HCQ have both been found to inhibit the expression of CD154 in T-cells and to reduce TLR signaling that leads to the production of pro-inflammatory cytokines ([1062](#)). Clinical trials for COVID-19 have more often used HCQ rather than CQ because it offers the advantages of being cheaper and having fewer side effects than CQ. However, research has not found support for a positive effect of HCQ on COVID-19 patients. Multiple clinical studies have already been carried out to assess HCQ as a therapeutic agent for COVID-19, and many more are in progress. To date, none of these studies have used randomized, double-blind, placebo-controlled designs with a large sample size, which would be the gold standard. Despite the design limitations (which would be more likely to produce false positives than false negatives), initial optimism about HCQ has largely dissipated. The most methodologically rigorous analysis of HCQ as a prophylactic ([1060](#)) found no significant differences between the treatment and control groups, and the WHO's global Solidarity trial similarly reported no effect of HCQ on mortality ([811](#)). Thus, HCQ/CQ are not likely to be effective therapeutic or prophylactic agents against COVID-19. One case study identified drug-induced phospholipidosis as the cause of death for a COVID-19 patient treated with HCQ ([984](#)), suggesting that in some cases, the proposed mechanism of action may ultimately be harmful. Additionally, one study identified an increased risk of mortality in older men receiving HCQ, and administration of HCQ and HCQ + AZ did not decrease the use of mechanical ventilation in these patients ([1058](#)). HCQ use for COVID-19 could also lead to shortages for anti-malarial or anti-rheumatic use, where it has documented efficacy. Despite significant early attention, these drugs appear to be ineffective against COVID-19. Several countries have now removed CQ/HCQ from their SOC for COVID-19 due to the lack of evidence of efficacy and the frequency of adverse effects.

5.5 ACE Inhibitors and Angiotensin II Receptor Blockers

Several clinical trials testing the effects of ACEIs or ARBs on COVID-19 outcomes are ongoing ([1063–1069](#)). Clinical trials are needed because the findings of the various observational studies bearing on this topic cannot be interpreted as indicating a protective effect of the drug ([1070](#), [1071](#)). Two analyses ([1063](#), [1069](#)) have reported no effect of continuing or discontinuing ARBs and ACEIs on patients admitted to the hospital for COVID-19. The first, known as REPLACE COVID ([874](#)), was a randomized, open-label study that enrolled patients who were admitted to the hospital for COVID-19 and were taking an ACEI at the time of admission. They enrolled 152 patients at 20 hospitals across seven countries and randomized them into two arms,

continuation (n=75) and discontinuation (n=77). The primary outcome evaluated was a global rank score that integrated several dimensions of illness. The components of this global rank score, such as time to death and length of mechanical ventilation, were evaluated as secondary endpoints. This analysis reported no differences between the two groups in the primary or any of the secondary outcomes.

Similarly, a second study ([875](#)) used a randomized, open-label design to examine the effects of continuing versus discontinuing ARBs and ACEIs on patients hospitalized for mild to moderate COVID-19 at 29 hospitals in Brazil. This study enrolled 740 patients but had to exclude one trial site from all analyses due to the discovery of violations of Good Clinical Trial practice and data falsification. After this exclusion, 659 patients remained, with 334 randomized to discontinuation and 325 to continuation. In this study, the primary endpoint analyzed was the number of days that patients were alive and not hospitalized within 30 days of enrollment. The secondary outcomes included death (including in-hospital death separately), number of days hospitalized, and specific clinical outcomes such as heart failure or stroke. Once again, no significant differences were found between the two groups. Initial studies of randomized interventions therefore suggest that ACEIs and ARBs are unlikely to affect COVID-19 outcomes. These results are also consistent with findings from observational studies (summarized in [874](#)). Additional information about ACE2, observational studies of ACEIs and ARBs in COVID-19, and clinical trials on this topic have been summarized ([1072](#)). Therefore, despite the promising potential mechanism, initial results have not provided support for ACEIs and ARBs as therapies for COVID-19.

5.6 Tocilizumab

Human IL-6 is a 26-kDa glycoprotein that consists of 184 amino acids and contains two potential N-glycosylation sites and four cysteine residues. It binds to a type I cytokine receptor (IL-6R α or glycoprotein 80) that exists in both membrane-bound (IL-6R α) and soluble (sIL-6R α) forms ([1073](#)). It is not the binding of IL-6 to the receptor that initiates pro- and/or anti-inflammatory signaling, but rather the binding of the complex to another subunit, known as IL-6R β or glycoprotein 130 (gp130) ([1073](#), [1074](#)). Unlike membrane-bound IL-6R α , which is only found on hepatocytes and some types of leukocytes, gp130 is found on most cells ([1075](#)). When IL-6 binds to sIL-6R α , the complex can then bind to a gp130 protein on any cell ([1075](#)). The binding of IL-6 to IL-6R α is termed classical signaling, while its binding to sIL-6R α is termed trans-signaling ([1075–1077](#)). These two signaling processes are thought to play different roles in health and illness. For example, trans-signaling may play a role in the proliferation of mucosal T-helper TH2 cells associated with asthma, while an earlier step in this proliferation process may be regulated by classical signaling ([1075](#)). Similarly, IL-6 is known to play a role in Crohn's Disease via trans-, but not classical, signaling ([1075](#)). Both classical and trans-signaling can occur through three independent pathways: the Janus-activated kinase-STAT3 pathway, the Ras/Mitogen-Activated Protein Kinases pathway and the Phosphoinositol-3 Kinase/Akt pathway ([1073](#)). These signaling pathways are involved in a variety of different functions, including cell type differentiation, immunoglobulin synthesis, and cellular survival signaling pathways, respectively ([1073](#)). The ultimate result of the IL-6 cascade is to direct transcriptional activity of various promoters of pro-

inflammatory cytokines, such as IL-1, TFN, and even IL-6 itself, through the activity of NF- κ B (1073). IL-6 synthesis is tightly regulated both transcriptionally and post-transcriptionally, and it has been shown that viral proteins can enhance transcription of the IL-6 gene by strengthening the DNA-binding activity between several transcription factors and IL-6 gene-cis-regulatory elements (1078). Therefore, drugs inhibiting the binding of IL-6 to IL-6R α or sIL-6R α are of interest for combating the hyperactive inflammatory response characteristic of cytokine release syndrome (CRS) and cytokine storm syndrome (CSS). TCZ is a humanized monoclonal antibody that binds both to the insoluble and soluble receptor of IL-6, providing de facto inhibition of the IL-6 immune cascade. Interest in TCZ as a possible treatment for COVID-19 was piqued by early evidence indicating that COVID-19 deaths may be induced by the hyperactive immune response, often referred to as CRS or CSS (83), as IL-6 plays a key role in this response (150). The observation of elevated IL-6 in patients who died relative to those who recovered (83) could reflect an over-production of proinflammatory interleukins, suggesting that TCZ could potentially palliate some of the most severe symptoms of COVID-19 associated with increased cytokine production.

This early interest in TCZ as a possible treatment for COVID-19 was bolstered by a very small retrospective study in China that examined 20 patients with severe symptoms in early February 2020 and reported rapid improvement in symptoms following treatment with TCZ (893). Subsequently, a number of retrospective studies have been conducted in several countries. Many studies use a retrospective, observational design, where they compare outcomes for COVID-19 patients who received TCZ to those who did not over a set period of time. For example, one of the largest retrospective, observational analyses released to date (888), consisting of 1,351 patients admitted to several care centers in Italy, compared the rates at which patients who received TCZ died or progressed to invasive medical ventilation over a 14-day period compared to patients receiving only SOC. Under this definition, SOC could include other drugs such as HCQ, azithromycin, lopinavir-ritonavir or darunavir-cobicistat, or heparin. While this study was not randomized, a subset of patients who were eligible to receive TCZ were unable to obtain it due to shortages; however, these groups were not directly compared in the analysis. After adjusting for variables such as age, sex, and SOFA (sequential organ failure assessment) score, they found that patients treated with TCZ were less likely to progress to invasive medical ventilation and/or death (adjusted HR = 0.61, CI 0.40-0.92, p = 0.020); analysis of death and ventilation separately suggests that this effect may have been driven by differences in the death rate (20% of control versus 7% of TCZ-treated patients). The study reported particular benefits for patients whose PaO₂/FiO₂ ratio, also known as the Horowitz Index for Lung Function, fell below a 150 mm Hg threshold. They found no differences between groups administered subcutaneous versus intravenous TCZ.

Another retrospective observational analysis of interest examined the charts of patients at a hospital in Connecticut, USA where 64% of all 239 COVID-19 patients in the study period were administered TCZ based on assignment by a standardized algorithm (889). They found that TCZ administration was associated with more similar rates of survivorship in patients with severe versus nonsevere COVID-19 at intake, defined based on the amount of

supplemental oxygen needed. They therefore proposed that their algorithm was able to identify patients presenting with or likely to develop CRS as good candidates for TCZ. This study also reported higher survivorship in Black and Hispanic patients compared to white patients when adjusted for age. The major limitation with interpretation for these studies is that there may be clinical characteristics that influenced medical practitioners decisions to administer TCZ to some patients and not others. One interesting example therefore comes from an analysis of patients at a single hospital in Brescia, Italy, where TCZ was not available for a period of time ([890](#)). This study compared COVID-19 patients admitted to the hospital before and after March 13, 2020, when the hospital received TCZ. Therefore, patients who would have been eligible for TCZ prior to this arbitrary date did not receive it as treatment, making this retrospective analysis something of a natural experiment. Despite this design, demographic factors did not appear to be consistent between the two groups, and the average age of the control group was older than the TCZ group. The control group also had a higher percentage of males and a higher incidence of comorbidities such as diabetes and heart disease. All the same, the multivariate HR, which adjusted for these clinical and demographic factors, found a significant difference between survival in the two groups (HR=0.035, CI=0.004-0.347, $p = 0.004$). The study reported improvement of survival outcomes after the addition of TCZ to the SOC regime, with 11 of 23 patients (47.8%) admitted prior to March 13th dying compared to 2 of 62 (3.2%) admitted afterwards (HR=0.035; 95% CI, 0.004 to 0.347; $p = 0.004$). They also reported a reduced progression to mechanical ventilation in the TCZ group. However, this study also holds a significant limitation: the time delay between the two groups means that knowledge about how to treat the disease likely improved over this timeframe as well. All the same, the results of these observational retrospective studies provide support for TCZ as a pharmaceutical of interest for follow-up in clinical trials.

Other retrospective analyses have utilized a case-control design to match pairs of patients with similar baseline characteristics, only one of whom received TCZ for COVID-19. In one such study, TCZ was significantly associated with a reduced risk of progression to intensive care unit (ICU) admission or death ([891](#)). This study examined only 20 patients treated with TCZ (all but one of the patients treated with TCZ in the hospital during the study period) and compared them to 25 patients receiving SOC. For the combined primary endpoint of death and/or ICU admission, only 25% of patients receiving TCZ progressed to an endpoint compared to 72% in the SOC group ($p = 0.002$, presumably based on a chi-square test based on the information provided in the text). When the two endpoints were examined separately, progression to invasive medical ventilation remained significant (32% SOC compared to 0% TCZ, $p = 0.006$) but not for mortality (48% SOC compared to 25% TCZ, $p = 0.066$). In contrast, a study that compared 96 patients treated with TCZ to 97 patients treated with SOC only in New York City found that differences in mortality did not differ between the two groups, but that this difference did become significant when intubated patients were excluded from the analysis ([892](#)). Taken together, these findings suggest that future clinical trials of TCZ may want to include intubation as an endpoint. However, these studies should be approached with caution, not only because of the small number of patients enrolled and the retrospective design, but also because they performed a large number of

statistical tests and did not account for multiple hypothesis testing. In general, caution must be exercised when interpreting subgroup analyses after a primary combined endpoint analysis. These last findings highlight the need to search for a balance between impairing a harmful immune response, such as the one generated during CRS/CSS, and preventing the worsening of the clinical picture of the patients by potential new viral infections. Early meta-analyses and systematic reviews have investigated the available data about TCZ for COVID-19. One meta-analysis ([1079](#)) evaluated 19 studies published or released as preprints prior to July 1, 2020 and found that the overall trends were supportive of the frequent conclusion that TCZ does improve survivorship, with a significant HR of 0.41 ($p < 0.001$). This trend improved when they excluded studies that administered a steroid alongside TCZ, with a significant HR of 0.04 ($p < 0.001$). They also found some evidence for reduced invasive ventilation or ICU admission, but only when excluding all studies except a small number whose estimates were adjusted for the possible bias introduced by the challenges of stringency during the enrollment process. A systematic analysis of sixteen case-control studies of TCZ estimated an odds ratio of mortality of 0.453 (95% CI 0.376–0.547, $p < 0.001$), suggesting possible benefits associated with TCZ treatment ([1080](#)). Although these estimates are similar, it is important to note that they are drawing from the same literature and are therefore likely to be affected by the same potential biases in publication. A different systematic review of studies investigating TCZ treatment for COVID-19 analyzed 31 studies that had been published or released as pre-prints and reported that none carried a low risk of bias ([1081](#)). Therefore, the present evidence is not likely to be sufficient for conclusions about the efficacy of TCZ.

On February 11, 2021, a preprint describing the first randomized control trial of TCZ was released as part of the RECOVERY trial ([894](#)). Of the 21,550 patients enrolled in the RECOVERY trial at the time, 4,116 adults hospitalized with COVID-19 across the 131 sites in the United Kingdom were assigned to the arm of the trial evaluating the effect of TCZ. Among them, 2,022 were randomized to receive TCZ and 2,094 were randomized to SOC, with 79% of patients in each group available for analysis at the time that the initial report was released. The primary outcome measured was 28-day mortality, and TCZ was found to reduce 28-day mortality from 33% of patients receiving SOC alone to 29% of those receiving TCZ, corresponding to a rate ratio of 0.86 (95% CI 0.77-0.96; $p = 0.007$). TCZ was also significantly associated with the probability of hospital discharge within 28 days for living patients, which was 47% in the SOC group and 54% in the TCZ group (rate ratio 1.22, 95% CI 1.12-1.34, $p < 0.0001$). A potential statistical interaction between TCZ and corticosteroids was observed, with the combination providing greater mortality benefits than TCZ alone, but the authors note that caution is advisable in light of the number of statistical tests conducted. Combining the RECOVERY trial data with data from seven smaller randomized control trials indicates that TCZ is associated with a 13% reduction in 28-day mortality (rate ratio 0.87, 95% CI 0.79-0.96, $p = 0.005$) ([894](#)).

There are possible risks associated with the administration of TCZ for COVID-19. TCZ has been used for over a decade to treat RA ([1082](#)), and a recent study found the drug to be safe for pregnant and breastfeeding women ([1083](#)). However, TCZ may increase the risk of developing infections ([1082](#)), and RA patients with chronic hepatitis B infections had a high risk of hepatitis

B virus reactivation when TCZ was administered in combination with other RA drugs ([1084](#)). As a result, TCZ is contraindicated in patients with active infections such as tuberculosis ([1085](#)). Previous studies have investigated, with varying results, a possible increased risk of infection in RA patients administered TCZ ([1086](#), [1087](#)), although another study reported that the incidence rate of infections was higher in clinical practice RA patients treated with TCZ than in the rates reported by clinical trials ([1088](#)). In the investigation of 544 Italian COVID-19 patients, the group treated with TCZ was found to be more likely to develop secondary infections, with 24% compared to 4% in the control group ($p < 0.0001$) ([888](#)). Reactivation of hepatitis B and herpes simplex virus 1 was also reported in a small number of patients in this study, all of whom were receiving TCZ. A July 2020 case report described negative outcomes of two COVID-19 patients after receiving TCZ, including one death; however, both patients were intubated and had entered septic shock prior to receiving TCZ ([1089](#)), likely indicating a severe level of cytokine production. Additionally, D-dimer and sIL2R levels were reported by one study to increase in patients treated with TCZ, which raised concerns because of the potential association between elevated D-dimer levels and thrombosis and between sIL2R and diseases where T-cell regulation is compromised ([889](#)). An increased risk of bacterial infection was also identified in a systematic review of the literature, based on the unadjusted estimates reported ([1079](#)). In the RECOVERY trial, however, only three out of 2,022 participants in the group receiving TCZ developed adverse reactions determined to be associated with the intervention, and no excess deaths were reported ([894](#)). TCZ administration to COVID-19 patients is not without risks and may introduce additional risk of developing secondary infections; however, while caution may be prudent when treating patients who have latent viral infections, the results of the RECOVERY trial indicate that adverse reactions to TCZ are very rare among COVID-19 patients broadly.

In summary, approximately 33% of hospitalized COVID-19 patients develop ARDS ([1090](#)), which is caused by an excessive early response of the immune system which can be a component of CRS/CSS ([889](#), [1085](#)). This overwhelming inflammation is triggered by IL-6. TCZ is an inhibitor of IL-6 and therefore may neutralize the inflammatory pathway that leads to the cytokine storm. The mechanism suggests TCZ could be beneficial for the treatment of COVID-19 patients experiencing excessive immune activity, and the RECOVERY trial reported a reduction in 28-day mortality. Interest in TCZ as a treatment for COVID-19 was also supported by two meta-analyses ([1079](#), [1091](#)), but a third meta-analysis found that all of the available literature at that time carried a risk of bias ([1081](#)). Additionally, different studies used different dosages, number of doses, and methods of administration. Ongoing research may be needed to optimize administration of TCZ ([1092](#)), although similar results were reported by one study for intravenous and subcutaneous administration ([888](#)). Clinical trials that are in progress are likely to provide additional insight into the effectiveness of this drug for the treatment of COVID-19 along with how it should be administered.

5.7 Interferons

IFNs are a family of cytokines critical to activating the innate immune response against viral infections. Interferons are classified into three categories based on their receptor specificity: types I, II and III (150). Specifically, IFNs I (IFN- α and β) and II (IFN- γ) induce the expression of antiviral proteins (1093). Among these IFNs, IFN- β has already been found to strongly inhibit the replication of other coronaviruses, such as SARS-CoV-1, in cell culture, while IFN- α and γ were shown to be less effective in this context (1093). There is evidence that patients with higher susceptibility to ARDS indeed show deficiency in IFN- β . For instance, infection with other coronaviruses impairs IFN- β expression and synthesis, allowing the virus to escape the innate immune response (1094). On March 18 2020, Synairgen plc received approval to start a phase II trial for SNG001, an IFN- β -1a formulation to be delivered to the lungs via inhalation (901). SNG001, which contains recombinant interferon beta-1a, was previously shown to be effective in reducing viral load in an *in vivo* model of swine flu and *in vitro* models of other coronavirus infections (1095). In July 2020, a press release from Synairgen stated that SNG001 reduced progression to ventilation in a double-blind, placebo-controlled, multi-center study of 101 patients with an average age in the late 50s (902). These results were subsequently published in November 2020 (903). The study reports that the participants were assigned at a ratio of 1:1 to receive either SNG001 or a placebo that lacked the active compound, by inhalation for up to 14 days. The primary outcome they assessed was the change in patients' score on the WHO Ordinal Scale for Clinical Improvement (OSCI) at trial day 15 or 16. SNG001 was associated with an odds ratio of improvement on the OSCI scale of 2.32 (95% CI 1.07 – 5.04, $p = 0.033$) in the intention-to-treat analysis and 2.80 (95% CI 1.21 – 6.52, $p = 0.017$) in the per-protocol analysis, corresponding to significant improvement in the SNG001 group on the OSCI at day 15/16. Some of the secondary endpoints analyzed also showed differences: at day 28, the OR for clinical improvement on the OSCI was 3.15 (95% CI 1.39 – 7.14, $p = 0.006$), and the odds of recovery at day 15/16 and at day 28 were also significant between the two groups. Thus, this study suggested that IFN- β 1 administered via SNG001 may improve clinical outcomes.

In contrast, the WHO Solidarity trial reported no significant effect of IFN- β -1a on patient survival during hospitalization (811). Here, the primary outcome analyzed was in-hospital mortality, and the rate ratio for the two groups was 1.16 (95% CI, 0.96 to 1.39; $p = 0.11$) administering IFN- β -1a to 2050 patients and comparing their response to 2,050 controls. However, there are a few reasons that the different findings of the two trials might not speak to the underlying efficacy of this treatment strategy. One important consideration is the stage of COVID-19 infection analyzed in each study. The Synairgen trial enrolled only patients who were not receiving invasive ventilation, corresponding to a less severe stage of disease than many patients enrolled in the SOLIDARITY trial, as well as a lower overall rate of mortality (1096). Additionally, the methods of administration differed between the two trials, with the SOLIDARITY trial administering IFN- β -1a subcutaneously (1096). The differences in findings between the studies suggests that the method of administration might be relevant to outcomes, with nebulized IFN- β -1a more directly targeting receptors in the lungs. A trial that analyzed the effect of subcutaneously administered IFN- β -1a on patients with ARDS between 2015 and 2017 had also reported no effect on 28-day mortality (1097), while a smaller study analyzing the effect of subcutaneous IFN administration did

find a significant improvement in 28-day mortality for COVID-19 ([1098](#)). At present, several ongoing clinical trials are investigating the potential effects of IFN- β -1a, including in combination with therapeutics such as remdesivir ([1099](#)) and administered via inhalation ([901](#)). Thus, as additional information becomes available, a more detailed understanding of whether and under which circumstances IFN- β -1a is beneficial to COVID-19 patients should develop.

5.8 Potential Avenues of Interest for Therapeutic Development

Given what is currently known about these therapeutics for COVID-19, a number of related therapies beyond those explored above may also prove to be of interest. For example, the demonstrated benefit of dexamethasone and the ongoing potential of tocilizumab for treatment of COVID-19 suggests that other anti-inflammatory agents might also hold value for the treatment of COVID-19. Current evidence supporting the treatment of severe COVID-19 with dexamethasone suggests that the need to curtail the cytokine storm inflammatory response transcends the risks of immunosuppression, and other anti-inflammatory agents may therefore benefit patients in this phase of the disease. While dexamethasone is considered widely available and generally affordable, the high costs of biologics such as tocilizumab therapy may present obstacles to wide-scale distribution of this drug if it proves of value. At the doses used for RA patients, the cost for tocilizumab ranges from \$179.20 to \$896 per dose for the IV form and \$355 for the pre-filled syringe ([1100](#)). Several other anti-inflammatory agents used for the treatment of autoimmune diseases may also be able to counter the effects of the cytokine storm induced by the virus, and some of these, such as cyclosporine, are likely to be more cost-effective and readily available than biologics ([1101](#)). While tocilizumab targets IL-6, several other inflammatory markers could be potential targets, including TNF- α . Inhibition of TNF- α by a compound such as Etanercept was previously suggested for treatment of SARS-CoV-1 ([1102](#)) and may be relevant for SARS-CoV-2 as well. Another anti-IL-6 antibody, sarilumab, is also being investigated ([1103](#), [1104](#)). Baricitinib and other small molecule inhibitors of the Janus-activated kinase pathway also curtail the inflammatory response and have been suggested as potential options for SARS-CoV-2 infections ([1105](#)). Baricitinib, in particular, may be able to reduce the ability of SARS-CoV-2 to infect lung cells ([1106](#)). Clinical trials studying baricitinib in COVID-19 have already begun in the US and in Italy ([1107](#), [1108](#)). Identification and targeting of further inflammatory markers that are relevant in SARS-CoV-2 infection may be of value for curtailing the inflammatory response and lung damage.

In addition to immunosuppressive treatments, which are most beneficial late in disease progression, much research is focused on identifying therapeutics for early-stage patients. For example, although studies of HCQ have not supported the early theory-driven interest in this antiviral treatment, alternative compounds with related mechanisms may still have potential. Hydroxyferroquine derivatives of HCQ have been described as a class of bioorganometallic compounds that exert antiviral effects with some

selectivity for SARS-CoV-1 *in vitro* ([1109](#)). Future work could explore whether such compounds exert antiviral effects against SARS-CoV-2 and whether they would be safer for use in COVID-19.

Another potential approach is the development of antivirals, which could be broad-spectrum, specific to coronaviruses, or targeted to SARS-CoV-2. Development of new antivirals is complicated by the fact that none have yet been approved for human coronaviruses. Intriguing new options are emerging, however. Beta-D-N4-hydroxycytidine is an orally bioavailable ribonucleotide analog showing broad-spectrum activity against RNA viruses, which may inhibit SARS-CoV-2 replication *in vitro* and *in vivo* in mouse models of HCoVs ([1110](#)). A range of other antivirals are also in development. Development of antivirals will be further facilitated as research reveals more information about the interaction of SARS-CoV-2 with the host cell and host cell genome, mechanisms of viral replication, mechanisms of viral assembly, and mechanisms of viral release to other cells; this can allow researchers to target specific stages and structures of the viral life cycle. Finally, antibodies against viruses, also known as antiviral monoclonal antibodies, could be an alternative as well and are described in detail in an above section. The goal of antiviral antibodies is to neutralize viruses through either cell-killing activity or blocking of viral replication ([1111](#)). They may also engage the host immune response, encouraging the immune system to hone in on the virus. Given the cytokine storm that results from immune system activation in response to the virus, which has been implicated in worsening of the disease, a neutralizing antibody (nAb) may be preferable. Upcoming work may explore the specificity of nAbs for their target, mechanisms by which the nAbs impede the virus, and improvements to antibody structure that may enhance the ability of the antibody to block viral activity.

Some research is also investigating potential therapeutics and prophylactics that would interact with components of the innate immune response. For example, TLRs are pattern recognition receptors that recognize pathogen- and damage-associated molecular patterns and contribute to innate immune recognition and, more generally, promotion of both the innate and adaptive immune responses ([147](#)). In mouse models, poly(I:C) and CpG, which are agonists of Toll-like receptors TLR3 and TLR9, respectively, showed protective effects when administered prior to SARS-CoV-1 infection ([1112](#)). Therefore, TLR agonists hold some potential for broad-spectrum prophylaxis.

6 Application of Traditional Vaccine Development Strategies to SARS-CoV-2

6.1 Abstract

Over the past 150 years, vaccines have revolutionized the relationship between people and disease. During the COVID-19 pandemic, technologies such as mRNA vaccines have received attention due to their novelty and

successes. However, more traditional vaccine development platforms have also yielded important tools in the worldwide fight against the SARS-CoV-2 virus.

A variety of approaches have been used to develop COVID-19 vaccines that are now authorized for use in countries around the world. In this review, we highlight strategies that focus on the viral capsid and outwards, rather than on the nucleic acids inside. These approaches fall into two broad categories: whole-virus vaccines and subunit vaccines. Whole-virus vaccines use the virus itself, either in an inactivated or attenuated state. Subunit vaccines contain instead an isolated, immunogenic component of the virus. Here, we highlight vaccine candidates that apply these approaches against SARS-CoV-2 in different ways. In a companion manuscript, we review the more recent and novel development of nucleic-acid based vaccine technologies.

We further consider the role that these COVID-19 vaccine development programs have played in prophylaxis at the global scale. Well-established vaccine technologies have proved especially important to making vaccines accessible in low- and middle-income countries. Vaccine development programs that use established platforms have been undertaken in a much wider range of countries than those using nucleic-acid-based technologies, which have been led by wealthy Western countries. Therefore, these vaccine platforms, though less novel from a biotechnological standpoint, have proven to be extremely important to the management of SARS-CoV-2.

6.2 Importance

As of March 9, 2023, there have been over 676,570,149 SARS-CoV-2 cases, and the virus has taken the lives of at least 6,881,802 people globally ([732](#)). The development, production, and distribution of vaccines is imperative to saving lives, preventing illness, and reducing the economic and social burdens caused by the COVID-19 pandemic. Vaccines that use cutting-edge biotechnology have played an important role in mitigating the effects of SARS-CoV-2. However, more traditional methods of vaccine development that were refined throughout the twentieth century have been especially critical to increasing vaccine access worldwide. Effective deployment is necessary to reducing the susceptibility of the world's population, which is especially important in light of emerging variants. In this review, we discuss the safety, immunogenicity, and distribution of vaccines developed using established technologies. In a separate review, we describe the vaccines developed using nucleic acid-based vaccine platforms. From the current literature, it is clear that the well-established vaccine technologies are also highly effective against SARS-CoV-2 and are being used to address the challenges of COVID-19 globally, including in low- and middle-income countries. This worldwide approach is critical for reducing the devastating impact of SARS-CoV-2.

6.3 Introduction

The development of vaccines is widely considered one of the most important medical advances in human history. Over the past 150 years, several approaches to vaccination have been developed and refined ([1113](#)). The COVID-19 pandemic has produced unusual circumstances compared to past

health crises, leading to differences in vaccine development strategies. One way in which the COVID-19 pandemic differs from prior global health crises is that the SARS-CoV-2 viral genome was sequenced, assembled, and released very early in the course of the pandemic (January 2020). This genomic information has informed the biomedical response to this novel pathogen across several dimensions ([1](#), [3](#)). All the same, vaccines have been developed since long before the concept of a virus or a viral genome was known, and as early as September 2020, there were over 180 vaccine candidates against SARS-CoV-2 in development, many of which employed more traditional vaccine technologies ([1114](#)). However, public attention in the United States and elsewhere has largely focused on vaccine development platforms that use new technologies, especially mRNA vaccines. We review vaccine technologies used for SARS-CoV-2 in two parts: here, the application of established vaccine development platforms to SARS-CoV-2, and separately, novel nucleic acid-based approaches ([6](#)).

Understanding vaccine development programs that are using well-established technologies is important for a global perspective on COVID-19. As of May 3, 2023, 50 SARS-CoV-2 vaccines have been approved for use in at least one country ([1115](#)). A resource tracking the distribution of 28 vaccines indicates that, as of May 31, 2023, 13.0 billion doses have been administered across 223 countries ([1116](#)). Many of these vaccines use platforms that do not require information about the viral genome, with 20 developed using subunit and 11 using whole-virus approaches ([1115](#)). The types of vaccines available varies widely throughout the world, as the process of developing and deploying a vaccine is complex and often requires coordination between government, industry, academia, and philanthropic entities ([1117](#)).

Another difference between prior global health crises and the COVID-19 pandemic is the way that vaccines are evaluated. A vaccine's success is often discussed in terms of vaccine efficacy (VE), which describes the protection conferred during clinical trials ([1118](#)). The real-world protection offered by a vaccine is referred to as its effectiveness ([1118](#)). Additionally, protection can mean different things in different contexts. In general, the goal of a vaccine is to prevent disease, especially severe disease, rather than infection itself. As a proxy for VE, vaccine developers often test their candidates for serum neutralizing activity, which has been proposed as a biomarker for adaptive immunity in other respiratory illnesses ([1119](#)). The duration and intensity of the COVID-19 pandemic has made it possible to test multiple vaccines in phase III trials, where the effect of the vaccines on a cohort's likelihood of contracting SARS-CoV-2 can be evaluated, whereas this has not always been feasible for other infectious diseases. In some cases (e.g., SARS), the pathogen has been controlled before vaccine candidates were available, while in others (e.g., MERS), the scale of the epidemic has been smaller. Vaccine development is traditionally a slow process, but the urgency of the COVID-19 pandemic created an atypical vaccine development ecosystem where fast development and production was prioritized. Estimates of VE have been released for many vaccine candidates across a number of technology types based on phase III trial data.

However, efficacy is not a static value, and both trial efficacy and real-world effectiveness can vary across location and over time. Shifts in effectiveness in particular have been an especially heightened topic of concern since late

2021 given the potential for variants of SARS-CoV-2 to influence VE. Due to viral evolution, vaccine developers are in an arms race with a pathogen that benefits from mutations that reduce its susceptibility to adaptive immunity. The evolution of several variants of concern (VOC) presents significant challenges for vaccines developed based on the index strain identified in Wuhan in late 2019. We discuss these variants in depth elsewhere ([341](#)). To date, the most significant VOC identified are Alpha (2020), Beta (2020), Gamma (2020), Delta (2021), and Omicron (2021), with various subvariants of Omicron being the most recently identified (2022). The relative timing of studies relative to dominant VOC in the region where participants are recruited is important context for a complete picture of efficacy. Therefore, the efficacy and/or effectiveness of vaccines in the context of these variants is discussed where information is available.

Beyond the variability introduced by time and geography, efficacy within a trial and effectiveness in the real-world setting can also differ due to cohort differences. Patients participating in a clinical trial are likely to receive more medical oversight, resulting in better follow-up, adherence, and patient engagement ([1120](#)). Additionally, the criteria for participant inclusion in a trial often bias trials towards selection of younger, healthier individuals ([1121](#)). The ability of an RCT to accurately assess safety can be biased by the fact that a clinical trial might not reveal rare adverse events (AEs) that might become apparent on a larger scale ([1121](#)). Therefore, while clinical trials are the gold standard for evaluating vaccines for COVID-19, the results of these trials must be considered in a broader context when real-world data is available.

While the relationship between a vaccine and a pathogen is not static, the data clearly demonstrates that a variety of efficacious vaccines have been developed against SARS-CoV-2. Here we discuss a selection of programs that use well-established vaccine biotechnologies. These programs have been undertaken worldwide, in complement to the more cutting-edge approaches developed and distributed in the United States (U.S.), the European Union (E.U.), the United Kingdom (U.K.), and Russia ([6](#)). In this review, we discuss vaccine development using two well-established technologies, whole-virus vaccination and subunit vaccination, and the role these technologies have played in the global response to the COVID-19 pandemic.

6.4 Whole-Virus Vaccines

Whole-virus vaccines have the longest history among vaccine development approaches. Variolation, which is widely considered the first vaccination strategy in human history, is one example ([1122](#), [1123](#)). Famously, variolation was employed against smallpox when healthy individuals were exposed to pus from an individual infected with what was believed to be either cowpox or horsepox ([1122–1125](#)). This approach worked by inducing a mild case of a disease. Therefore, while whole-virus vaccines confer adaptive immunity, they also raise safety concerns ([1124](#), [1126](#), [1127](#)). As of 2005, most vaccines still used whole-virus platforms ([1128](#)), and these technologies remain valuable tools in vaccine development today ([1113](#)). Whole virus vaccine candidates have been developed for COVID-19 using both live attenuated viruses and inactivated whole viruses.

6.4.1 Live-Attenuated Virus Vaccines

Live-attenuated virus vaccines (LAV), also known as replication-competent vaccines, use a weakened, living version of a disease-causing virus or a version of a virus that is modified to induce an immune response ([1114](#)). Whether variolation is the first example of a LAV being used to induce immunity is debated ([1113](#), [1126](#)). The first deliberate (albeit pathogen-naïve) attempt to develop an attenuated viral vaccine dates back to Louis Pasteur's efforts in 1885 to inoculate a child against rabies ([1129](#)). The next intentional LAVs were developed against the yellow fever virus in 1935 and influenza in 1936 ([1130](#)).

Early efforts in LAV development relied on either the identification of a related virus that was less virulent in humans (e.g., cowpox/horsepox or rotavirus vaccines) or the culturing of a virus *in vitro* ([1113](#), [1124](#)). Today, a virus can be attenuated by passaging it in a foreign host until, due to selection pressure, the virus loses its efficacy in the original host. Alternatively, selective gene deletion or codon deoptimization can be utilized to attenuate the virus ([1114](#)), or foreign antigens can be integrated into an attenuated viral vector ([1131](#)). LAVs tend to be restricted to viral replication in the tissues around the location of inoculation ([1130](#)), and some can be administered intranasally ([1114](#)).

Today, LAVs are used globally to prevent diseases caused by viruses such as measles, mumps, rubella, polio, influenza, varicella zoster, and the yellow fever virus ([1132](#)). There were attempts to develop LAVs against both SARS-CoV-1 and MERS-CoV ([1133](#)), but no vaccines were approved. It is generally recognized that LAVs induce an immune response similar to natural infection, and they are favored because they induce long-lasting and robust immunity that can protect from disease. This strong protective effect is induced in part by the immune response to the range of viral antigens available from LAV, which tend to be more immunogenic than those from non-replicating vaccines ([1126](#), [1133](#), [1134](#)).

6.4.2 LAV Vaccines and COVID-19

To date, LAVs have not been widely deployed against SARS-CoV-2 and COVID-19. All the same, there are several COVID-19 LAV candidates in the early (preclinical/phase I) stages of investigation. These candidates utilize different approaches. Interestingly, several candidates (Meissa Vaccines' MV-014-212 and Codagenix's COVI-VAC, specifically) are administered intranasally, potentially improving accessibility by eliminating the need for sterile needles and reducing manufacturing costs, targeting conferring mucosal immunity, and reducing some issues related to vaccine hesitancy ([1135](#), [1136](#)). Additionally, although no phase III trial data is available for LAV vaccine candidates, some manufacturers have proactively sought to respond to the emergence of VOC. Therefore, the original approach to vaccination may still prove extremely advantageous in the high-tech landscape of COVID-19 vaccine development.

6.4.2.1 YF-S0

One candidate in the preclinical stage is YF-S0, a single-dose LAV developed at Belgium's Katholieke Universiteit Leuven that uses live-attenuated yellow fever 17D (YF17D) as a vector for a noncleavable prefusion conformation of the spike antigen of SARS-CoV-2 ([1131](#)). YF-S0 induced a robust immune response in three animal models and prevented SARS-CoV-2 infection in macaques and hamsters ([1131](#)). Additionally, the protective effect of YF-S0 against VOC has been investigated in hamsters ([1137](#)). Even for a small number of hamsters that developed breakthrough infections after exposure to the index strain or the Alpha variant, viral loads were very low ([1137](#)). However, much higher rates of breakthrough infection and higher viral loads were observed when the hamsters were exposed to the Beta variant ([1137](#)). Reduced seroconversion and nAb titers were also observed against the Beta and Gamma variants ([1137](#)). As a result, a modified version of YF-S0, called YF-S0*, was developed to include a modified spike protein intended to increase immunogenicity by including the full spectrum of amino acids found in the Gamma VOC as well as stabilizing the S protein's conformation ([1137](#)). The updated vaccine was again tested in Syrian golden hamsters ([1137](#)). No breakthrough infections were observed following vaccination with YF-S0* and exposure to the index strain and the Alpha, Beta, Gamma, and Delta variants ([1137](#)). YF-S0* also reduced the infectious viral load in the lungs of several VOCs (Alpha, Beta, Gamma, and Delta) relative to a sham comparison ([1137](#)), and the likelihood of the Delta variant spreading to unvaccinated co-housed hamsters was significantly reduced by YF-S0* ([1137](#)). The updated vaccine was also associated with the increased production of nAbs against the Omicron variant compared to YF-S0 ([1137](#)).

6.4.2.2 COVI-VAC

Other programs are developing codon deoptimized LAV candidates ([1138–1140](#)). This approach follows the synthetic attenuated virus engineering (SAVE) strategy to select codon substitutions that are suboptimal for the virus ([1140, 1141](#)). New York-based Codagenix and the Serum Institute of India reported a successful preclinical investigation ([1140](#)) of an intranasally administered deoptimized SARS-CoV-2 LAV known as COVI-VAC, and COVI-VAC entered phase I human trials and dosed its first participants in January 2021 ([1139, 1142](#)). This vaccine is optimized through the removal of the furin cleavage site (see ([1](#)) for a discussion of this site's importance) and deoptimization of 283 codons ([1143](#)). The results of the COVI-VAC phase I human trials are expected soon ([1142](#)).

Other results suggest both potential benefits and risks to the COVI-VAC vaccine candidate. Preclinical results suggest that the vaccine candidate may confer some protection against VOC even though it was designed based on the index strain: a poster reported that Syrian golden hamsters who received COVI-VAC were significantly less likely to lose weight following viral challenge with the Beta VOC ([1143](#)). On the other hand, some concerns have arisen about the possibility of spillover from LAV vaccines. A December 2022 study analyzed SARS-CoV-2 samples isolated from COVID-19 patients in India and identified two extremely similar sequences collected on June 30, 2020 that showed a high level of recombination relative to the dominant strains at the time ([1144](#)). Comparing these samples to a database of SARS-CoV-2 sequences revealed they were most similar to the sequence used for COVI-VAC ([1144](#)). Based on phylogenetic reconstruction, the authors argued that

these SARS-CoV-2 isolates were most likely to have spilled over from COVI-VAC trials ([1144](#)). If this was a case of spillover, the effect seems to have been limited, as these sequences were just two among over 1,600 analyzed. However, these concerns may be one consideration in the development of LAV vaccines for COVID-19.

6.4.2.3 Meissa Vaccines MV-014-212

Another company, Meissa Vaccines in Kansas, U.S., which also develops vaccines for respiratory syncytial virus (RSV), has developed an intranasal live-attenuated chimeric vaccine MV-014-212 ([1145](#)). Chimeric vaccines integrate genomic content from multiple viruses to create a more stable LAV ([1146](#)). To develop a SARS-CoV-2 vaccine candidate, Meissa Vaccines built on their prior work developing RSV vaccines ([1145](#)). A live attenuated recombinant strain of RSV previously investigated as a vaccine candidate was modified to replace two surface glycoproteins with a chimeric protein containing components of the SARS-CoV-2 Spike protein as well as the RSV fusion (F) protein ([1145](#)). Preclinical results describing the intranasal administration of MV-014-212 to African green monkeys and mice indicated that the vaccine candidate produced neutralizing antibodies (nAb) as well as a cellular immune response to SARS-CoV-2 challenge, including the Alpha, Beta, and Delta VOC ([1145](#)). Enrollment for phase I human trials began in March 2021 and recruitment is ongoing ([1139](#), [1147](#)).

6.4.2.4 Bacillus Calmette-Guerin Vaccines

Finally, Bacillus Calmette-Guerin (BCG) vaccines that use LAVs are being investigated for the prophylaxis of COVID-19 (see online Appendix ([1148](#))). The purpose of the BCG vaccine is to prevent tuberculosis, but non-specific effects against other respiratory illnesses have suggested a possible benefit against COVID-19 ([1149](#)). However, a multicenter trial that randomly assigned participants 60 years and older to vaccination with BCG (n = 1,008) or placebo (n = 1,006) found that BCG vaccination had no effect on the incidence of SARS-CoV-2 or other respiratory infections over the course of 12 months ([1150](#)). Despite these findings, BCG vaccination was associated with a stronger cytokine (specifically, IL-6) response following *ex vivo* stimulation of peripheral blood mononuclear cells in patients with no known history of COVID-19 ([1150](#)). Additionally, SARS-CoV-2-positive individuals who had received the BCG vaccine one year prior showed increased immunoglobulin (Ig) responses to the SARS-CoV-2 spike protein and receptor binding domain (RBD) relative to individuals who had received a placebo vaccine ([1150](#)). Currently, investigations of BCG vaccines against COVID-19 are being sponsored by institutes in Australia in collaboration with the Bill and Melinda Gates Foundation ([1151](#)) and by Texas A&M University in collaboration with numerous other U.S. institutions ([1152](#)).

6.4.2.5 Summary of LAV Vaccine Development

LAV vaccines for COVID-19 have not advanced as far in development as vaccines developed using other technologies. As of December 2022, COVI-VAC was the only LAV vaccine candidate in phase III clinical trials ([1153](#)). As a result, safety data is not yet available for human studies of these vaccines. In

general, though, safety concerns previously associated with LAV have been largely mitigated in the modern manufacturing process. Manufacturers use safe and reliable methods to produce large quantities of vaccines once they have undergone rigorous preclinical studies and clinical trials to evaluate their safety and efficacy. However, one remaining safety concern may be contributing to the relatively slow emergence of LAV candidates against COVID-19: they still may present risk to individuals who are immunocompromised ([1154](#)), which is an even greater concern when dealing with a novel virus and disease. Additional data are needed to ascertain how this technology performs in the case of SARS-CoV-2 and whether rare cases of spillover have indeed occurred. Additionally, modifications to the design of individual vaccine candidates may make this protection more robust as SARS-CoV-2 evolves, as the limited data about LAV performance against VOC suggests. Despite the long and trusted history of LAV development, this vaccine strategy has not been favored against COVID-19, as other technologies have shown greater expediency and safety compared to the time-consuming nature of developing LAVs for a novel virus.

6.5 Inactivated Whole-Virus Vaccines

Inactivated whole-virus (IWV) vaccines are another well-established vaccine platform. This platform uses full virus particles generally produced via cell culture that have been rendered non-infectious by chemical (i.e., formaldehyde or β -propiolactone ([1155](#))) or physical (i.e., heat or ultraviolet radiation) means. In general, these vaccines mimic the key properties of the virus that stimulate a robust immune response, but the risk of adverse reactions is reduced because the virus is inactivated and thus unable to replicate. Though these viral particles are inactivated, they retain the capacity to prime the immune system. The size of the viral particle makes it ideal for uptake by antigen-presenting cells, which leads to the stimulation of helper T-cells ([1156](#)). Additionally, the array of epitopes on the surface of the virus increases antibody binding efficiency ([1156](#)). The native conformation of the surface proteins, which is also important for eliciting an immune response, is preserved using these techniques ([1157](#)). Membrane proteins, which support B-cell responses to surface proteins, are also induced by this method ([1158](#)).

IWV vaccines have been a valuable tool in efforts to control many viruses. Some targets of IWV vaccines have included influenza viruses, poliovirus, and hepatitis A virus. Inactivated vaccines can generally be generated relatively quickly once the pathogenic virus has been isolated and can be passaged in cell culture ([1133](#), [1159](#)). During COVID-19, though the World Health Organization (WHO) has been slower to approve IWV vaccine candidates than those developed with nucleic acid-based technologies, IWV vaccine development was also fast. In China, the first emergency use authorization (EUA) was granted to an IWV vaccine in July 2020, with full approval following that December ([1160](#), [1161](#)). The fact that these vaccines have not received as much public attention (at least in Western media) as nucleic acid vaccines for SARS-CoV-2 may be due at least in part to the novelty of nucleic acid vaccine technologies ([1162](#)), which are more modular and immunogenic ([6](#)).

Past applications to human coronaviruses (HCoV) have focused predominantly on SARS-CoV-1. Preclinical studies have demonstrated that IWV SARS-CoV-1 vaccine candidates elicited immune responses *in vivo*. These vaccines generated nAb titers at concentrations similar to those evoked by recombinant protein vaccines (1157, 1163). Studies in ferrets and non-human primates demonstrated that IWV vaccines can offer protection against infection due to nAb and SARS-CoV-1-specific T cell responses (1164). However, several attempts to develop IWV vaccines against both SARS-CoV-1 and MERS-CoV were hindered by incidences of vaccine-associated disease enhancement (VADE) in preclinical studies (1165). In one example of a study in macaques, an inactivated SARS-CoV-1 vaccine induced even more severe lung damage than the virus due to an enhanced immune reaction (1166). Independent studies in mice also demonstrated evidence of lung immunopathology due to VADE in response to MERS-CoV IWV vaccination (1167, 1168). The exact mechanisms responsible for VADE remain elusive due to the specificity of the virus-host interactions involved, but VADE is the subject of investigation in preclinical SARS-CoV-2 vaccine studies to ensure the safety of any potential vaccines that may reach phase I trials and beyond (1165).

6.5.1 Application to COVID-19

Table 2: Inactivated whole-virus vaccines approved in at least one country (1169) as of May 3, 2023 (1115).

Vaccine	Company
Covaxin	Bharat Biotech
KoviVac	Chumakov Center
Turkovac	Health Institutes of Turkey
FAKHRAVAC (MIVAC)	Organization of Defensive Innovation and Research
QazVac	Research Institute for Biological Safety Problems (RIBSP)
KCONVAC	Shenzhen Kangtai Biological Products Co
COVIran Barekat	Shifa Pharmed Industrial Co
Covilo	Sinopharm (Beijing)
Inactivated (Vero Cells)	Sinopharm (Wuhan)
CoronaVac	Sinovac
VLA2001	Valneva

Several whole-virus vaccines have been developed against COVID-19 and are available in countries around the world (Table 2). As of May 31, 2023, 10 vaccines developed with IWV technology are being distributed in 120 countries (Figure 6). Evidence about the value of these vaccines to combat SARS-CoV-2 is available not only from clinical trials, but also from their roll-out following approval. Here, a major consideration has been that vaccines often lose efficacy as mutations accumulate in the epitopes of the circulating virus; IWV vaccines may be particularly affected in such cases (1127). This loss of specificity over time is likely to be influenced by the evolution of the virus,

and specifically by the rate of evolution in the region of the genome that codes for the antigenic spike protein. Here we review three vaccine development programs and their successes in a real-world setting.

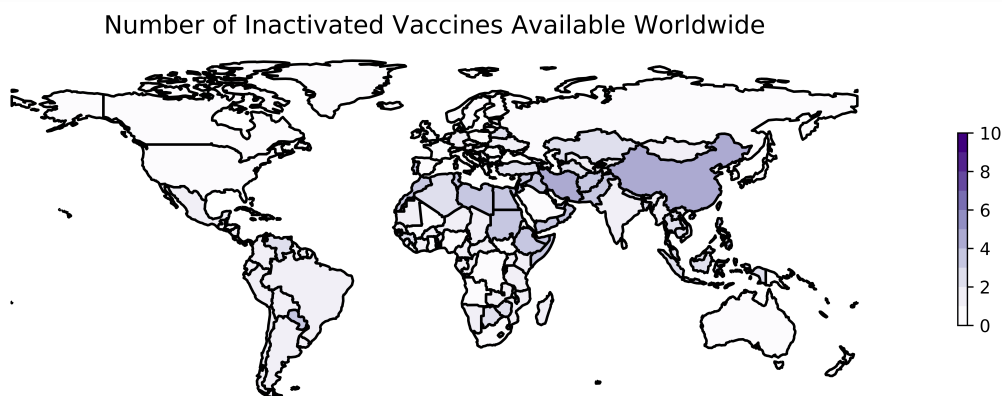


Figure 6: Worldwide availability of vaccines developed using inactivated whole viruses. This figure reflects the number of vaccines based on whole inactivated virus technology that were available in each country as of May 31, 2023. These data are retrieved from Our World in Data ([532](#)) and plotted using geopandas ([1170](#)). The color scale is based on the number of vaccines of this type included in the OWID dataset as a whole, not the maximum observed in a single country. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

6.5.1.1 Sinovac's CoronaVac

One IWV vaccine, CoronaVac, was developed by Beijing-based biopharmaceutical company Sinovac. The developers inactivated a SARS-CoV-2 strain collected in China with β -propiolactone and propagated it using Vero cells ([1133](#)). The vaccine is coupled with an aluminum adjuvant to increase immunogenicity ([1133](#)). Administration follows a prime-boost regimen using a 0.5 ml dose containing 3 μ g of inactivated SARS-CoV-2 virus per dose ([1171](#)). In phase I and II clinical trials, CoronaVac elicited a strong immunogenic response in animal models and the development of nAbs in human participants ([1172–1174](#)). The phase I/II clinical trials were conducted in adults 18–59 years old ([1174](#)) and adults over 60 years old ([1172](#)) in China. Safety analysis of the CoronaVac vaccine during the phase II trial revealed that most adverse reactions were either mild (grade 1) or moderate (grade 2) in severity ([1174](#)). In adults aged 18 to 59 years receiving a variety of dosage schedules, site injection pain was consistently the most common symptom reported ([1174](#)). In older adults, the most common local and systemic reactions were pain at the injection site (9%) and fever (3%), respectively ([1172](#)).

As of December 2022, a total of 17 CoronaVac trials had been registered in a variety of countries including the Philippines and Hong Kong ([1175](#)). Two of the earliest phase III trials to produce results examined a two-dose regimen of CoronaVac following a 14-day prime boost regimen ([1176](#), [1177](#)). These trials were conducted in Turkey ([1177](#)) and Chile ([1176](#)) and enrolled participants over an identical period from September 2020 and January 2021. The Chilean trial, which reported interim results regarding safety and immunogenicity, identified specific IgG nAbs against the S1 RBD and a robust IFN- γ secreting T cell response was induced via immunization with CoronaVac ([1176](#)). In the Turkish trial, VE was estimated to be 83.5% against

symptomatic COVID-19 ([1177](#)). In the safety and immunogenicity study, minimal AEs were reported ([1177](#)), and 18.9% of participants in the vaccine arm of the Turkish trial reported AEs compared to 16.9% of participants in the placebo group ([1177](#)). However, 2% (n=7) of Turkish participants aged 18 to 59 reported severe AEs ([1172](#)), causing the trial to be halted for investigation ([1178](#)). The investigation determined that these events were unrelated to the vaccine ([1172](#), [1178](#)).

An additional phase III trial was conducted in Brazil between July and December 2020 following a randomized, multicenter, endpoint driven, double-blind, placebo-controlled design and enrolling nearly 10,000 healthcare workers ([1179](#), [1180](#)). The preprint reporting the results of this study ([1180](#)) reports an efficacy of 50.7% against symptomatic COVID-19 and 100% against moderate to severe cases. A large percentage of participants, 77.1% in the vaccine group and 66.4% in the placebo group, reported AEs, including two deaths, but all of the serious AEs were determined not to be related to the vaccine ([1180](#)). CoronaVac also appears to be suitable for use in immunocompromised patients such as those with autoimmune rheumatic diseases according to phase IV trials ([1181](#)), and the vaccine was also well tolerated and induced humoral responses in phase I trials in children aged 3 to 17 years, which will now be examined in phase II and III clinical trials ([1182](#)).

Estimates of CoronaVac's VE have varied across trials. The 50.7% VE reported from the Brazilian trial was contested by Turkish officials, as at the time the efficacy in the Turkish trial appeared to be 91.25% ([1183](#), [1184](#)). Ultimately, after multiple announcements, the efficacy debate was settled at over 50% ([1183](#), [1184](#)). Subsequently, the VE for the Turkish trial was finalized at 83.5% ([1177](#)), and a prospective national cohort study in Chile reported an adjusted estimated effectiveness of 66% for the prevention of COVID-19 with an estimated 90% and 87% prevention of hospitalization and death, respectively ([1185](#)).

Based on these results, CoronaVac was approved in China, and has now been distributed in 66 countries across Africa, Asia, Europe, North America, and South America, including Brazil, Cambodia, Chile, Colombia, Laos, Malaysia, Mexico, Turkey, Ukraine, and Uruguay ([1116](#), [1186](#)). As of August 2021, Sinovac had reportedly produced over a billion doses of CoronaVac ([1186](#)). Outside of trials, rare cases of VADE have been reported in association with the CoronaVac vaccine ([1187](#)). In one case study, two male patients both presented with COVID-19 pneumonia following vaccination with CoronaVac ([1187](#)). This study identified the timeline of disease presentation, vaccination, and known COVID-19 exposure in the two patients and suggests that the inflammatory response induced by the vaccine could have caused an asymptomatic case of COVID-19 to present with symptoms ([1187](#)). However, no causal relationship between CoronaVac and COVID-19 symptom onset was evaluated, and the reports are extremely rare.

The effectiveness has also been questioned based on real-world data, such as when concerns were raised about the vaccine's effectiveness following reports that over 350 doctors became ill with COVID-19 in Indonesia despite being immunized with CoronaVac ([1188](#)). One possible explanation for such outbreaks was the evolution of the virus. Sera from individuals vaccinated

with CoronaVac was found to show reduced neutralizing activity against the Alpha, Beta, and Delta VOC relative to the index strain ([1189](#)). Similarly, a second study of 25 patients in Hong Kong in late 2021 evaluated serum neutralizing activity against the index strain and the Beta VOC, Delta VOC, and two Omicron isolates ([1190](#)). They reported that all individuals were seropositive for nAbs against the index strain, 68% against the Delta variant, and 0% against the Beta VOC and Omicron isolates ([1190](#)). The Beta variant appears to be more resistant to nAbs in sera from individuals immunized with CoronaVac than the Alpha variant or wildtype virus, indicating that emerging variants may be of concern ([1191](#)). Finally, a fourth study examined sera from 180 Thai healthcare workers vaccinated with CoronaVac and reported that neutralizing activity was significantly reduced against the Alpha, Delta, and Beta variants relative to the index strain ([1192](#)). Together, these results suggest that viral evolution is likely to pose a significant challenge to immunity acquired from the CoronaVac vaccine.

Therefore, studies have also evaluated whether booster doses would provide additional protection to individuals vaccinated with CoronaVac. This strategy is supported by the fact that the antibody response elicited by CoronaVac has been found to wane following the second dose, though it was still detected six months out ([1193](#)). A phase I/II clinical trial of CoronaVac in an elderly cohort (adults 60 years and older) in China determined that by 6 to 8 months following the second dose, nAb titers were detected below the seropositive cutoff ([1194](#)). Data from two phase II trials indicated that nAb response had declined 6 months after the second dose of the primary series, but a booster dose of CoronaVac administered 8 months after the second dose markedly increased geometric mean titers of SARS-CoV-2 nAbs ([1195](#)). However, the reduction of nAbs was ameliorated by a booster dose administered 8 months after the second CoronaVac dose ([1195](#)). Furthermore, Chinese ([1196](#)) and Chilean ([1197](#)) researchers have opted to investigate options to administer different vaccines (e.g., an mRNA vaccine dose) as a booster dose to individuals who have already received two doses of the IIV vaccine CoronaVac. Another study determined that using a viral-vectored vaccine (CanSino's Convidecia) or an mRNA vaccine (Pfizer/BioNTech's BNT162b2) instead of CoronaVac in a prime-boost vaccination regimen could induce a more robust immune response ([1198](#), [1199](#)). The WHO now suggests that a booster dose, either homologous or heterologous, can be considered 4 to 6 months after the primary series, especially for high-risk groups ([1200](#)).

6.5.1.2 Sinopharm's Covilo

Two additional IIV vaccine candidates were developed following a similar approach by the state-owned China National Pharmaceutical Group Co., Ltd., more commonly known as Sinopharm CNBG. One, known as BBIBP-CorV or Covilo, was developed in Beijing using the HB02 variant of SARS-CoV-2. The other was developed at Sinopharm CNBG's Wuhan Institute using the WIV04 variant of SARS-CoV-2 ([1201](#)). The viruses were purified, propagated using Vero cells, isolated, and inactivated using β -propiolactone ([1201](#), [1202](#)). Both vaccines are adjuvanted with aluminum hydroxide ([1201](#), [1202](#)). Here, we focus on Covilo.

Preclinical studies indicated that Covilo induced sufficient nAb titers in mice, and a prime-boost immunization scheme of 2 µg/dose was sufficient to protect rhesus macaques from disease ([1202](#)). In phase II trials, the Covilo vaccine appeared well-tolerated, with 23% of participants in the vaccine condition (482 total participants, 3:1, vaccine:placebo) reporting at least one adverse reaction characterized as mild to moderate ([1203](#)). No evidence of VADE was detected using this vaccine in phase II data ([1204](#)). In phase III trials conducted between July and December 2020, Covilo achieved an efficacy of 72.8% and was well tolerated ([1205](#)). However, questions were raised about efficacy when Sinopharm affiliates in the UAE in early December 2020 claimed the vaccine had 86% efficacy, which was later at odds with a Sinopharm Beijing affiliate that stated that Covilo had a 79.34% efficacy later that same month ([1206](#)).

Studies have also investigated expected differences in real-world effectiveness of Covilo given the continuing evolution of SARS-CoV-2. The antibody response elicited by Covilo was found to wane, but still to be detectable, by six months following the second dose ([1193](#)). One study showed that the Alpha variant exhibited very little resistance to neutralization by sera of those immunized with Covilo, but the Beta variant was more resistant to neutralization by almost a factor of 3 ([1191](#)). Another study examined sera from 282 participants and used a surrogate neutralizing assay, a test that generally correlates with nAbs, to determine that Covilo appears to induce nAbs against the binding of the RBD of wild type SARS-CoV-2 and the Alpha, Beta, and Delta variants to ACE2 ([1207](#)). Notably, a preprint reported that antisera (i.e., the antibody-containing component of the sera) from 12 people immunized with Covilo exhibited nAb capacity against the Beta variant (B.1.351), wild type SARS-CoV-2 (NB02), and one of the original variants of SARS-CoV-2 (D614G) ([1208](#)). As with many other vaccines, booster doses are being evaluated to mitigate some of the issues arising from viral evolution. A study of healthcare workers in China has since indicated that a booster shot of Covilo elevates B cell and T cell responses and increases nAb titers ([1209](#)). In May 2021, the UAE announced it would consider booster shots for all citizens who had been immunized with Covilo, which was shortly followed by a similar announcement in Bahrain, and by August 29, 2021, the UAE mandated booster shots for all residents who had received Covilo ([1186](#)).

6.5.1.3 Bharat Biotech's Covaxin

Another IWW vaccine candidate was developed by Bharat Biotech International Ltd., which is the biggest producer of vaccines globally, in collaboration with the Indian Council of Medical Research (ICMR) National Institute of Virology (NIV). This candidate is known as Covaxin or BBV152. Preclinical studies of Covaxin in hamsters ([1210](#)) and macaques ([1211](#)) indicated that the vaccine induced protective responses deemed sufficient to move forward to human trials. Phase I (July 2020) and phase I/II (September to October 2020) studies indicated that Covaxin adjuvanted with alum and a Toll-like receptor 7/8 (TLR7/8) agonist was safe and immunogenic ([1212](#), [1213](#)). These two studies demonstrated that the vaccine induced significant humoral and cell-mediated responses, as assessed by measuring binding ([1212](#)) and neutralizing ([1212](#), [1213](#)) antibodies, cytokines ([1212](#), [1213](#)), CD3⁺, CD4⁺, and CD8⁺ T-cells ([1212](#)), with some formulations also eliciting Th1-

skewed memory T-cell responses ([1213](#)). Only mild to moderate side-effects were reported upon immunization ([1212](#), [1213](#)), and in phase II trials, the Covilo vaccine appeared well-tolerated ([1203](#)).

In India, the Covaxin vaccine received emergency authorization on January 3, 2021, but the phase III data was not released until March 3, 2021, and even then it was communicated via press release ([1214](#)). This press release reported 80.6% efficacy in 25,800 participants ([1214](#), [1215](#)), spurring Zimbabwe to follow suit and authorize the use of Covaxin ([1216](#)). A detailed preprint describing the double-blind, randomized, controlled phase III trial that enrolled between November 2020 and January 2021 became available in July 2021 ([1217](#)), and the results collected as of May 17, 2021 were published in December 2021 ([1218](#)). Based on a final enrollment of 25,798 people (~1:1 vaccine:placebo), overall VE against symptomatic COVID-19 was estimated at 77.8% and against severe disease and asymptomatic infection was reported as 93.4% and 63.6%, respectively ([1218](#)). The vaccine was also reported to be well tolerated, with fewer severe events occurring in the Covaxin group (0.3%) than in the placebo group (0.5%) ([1218](#)). One case of a serious AE potentially related to the vaccine, immune thrombocytopenic purpura, was reported, although this patient was seropositive for SARS-CoV-2 at the baseline observation point ([1218](#)). As of June 1, 2023, Covaxin was approved for emergency use in 31 countries across Africa, Asia, Europe, and South America, including Guyana, India, Iran, Zimbabwe, Nepal, Mauritius, Mexico, Nepal, Paraguay, and the Philippines ([1219](#)).

Like with all vaccines, the continued evolution of SARS-CoV-2 poses a challenge to the effectiveness of Covaxin. In this case, the phase III clinical trial did evaluate the efficacy of Covaxin in response to variants circulating in mid-to-late 2020 ([1218](#)). In agreement with previous studies demonstrating sera from individuals vaccinated with Covaxin efficiently neutralized the Alpha variant (B.1.1.7) and the Delta variant (B.1.617.2) ([1220–1222](#)), the phase III trial reported a 65.2% efficacy against the Delta variant (B.1.617.2) ([1218](#)). Another study reported that sera from individuals immunized with Covaxin produced effective nAbs against the Delta variant and the so-called Delta plus variant (AY.1) ([1223](#)). Indeed, sera obtained from Covaxin boosted individuals (n=13) ([1224](#)) and those who were vaccinated with Covaxin but recovered from a breakthrough infection (n=31) also neutralized the Omicron variant ([1225](#)). Therefore, the data suggest that the vaccine does continue to confer protection to VOC.

The authorization of Covaxin has also offered opportunities to monitor how well the clinical trial results translate into a real-world setting. Additionally, an effort to monitor AEs and COVID-19 cases following vaccine roll-out reported that most side effects were mild and that cases were rare, even though this data would seem to have been collected during the severe wave of COVID-19 brought on by the Delta VOC in India in early 2021; at the same time, the sample sizes were extremely small ([1226](#)). Similarly, larger studies of adults (June to September 2021) ([1227](#)) and adolescents (beginning in January 2022) ([1228](#)) who received the vaccine outside of a trial setting reported that safety was similar across age groups, with no severe AEs reported in adults and with no serious AEs reported in adolescents, although 0.9% (6 individuals) reported severe AEs. However, a much lower effectiveness (22-29%) was estimated in a real-world setting during an analysis of cases in healthcare

workers from April to May 2021 ([1229](#)). All the same, monitoring of hospitalized COVID-19 patients between April and June 2021 indicated that the vaccines were highly effective against preventing severe illness ([1230](#)).

It is not yet clear what level of protection Covaxin offers beyond 6 to 8 months post the second vaccine; consequently, the potential requirement of a booster immunization is being explored ([1231](#)). Furthermore, Bharat Biotech is considering other vaccine regimens such as providing one initial immunization with Covaxin followed by two immunizations with its intranasal vaccine (BBV154) ([1232](#)). U.S.-based Ocugen Inc., a co-development partner of Bharat Biotech, is leading the application for an Emergency Use Authorization (EUA) for Covaxin intended for the U.S. market. It has been reported that Bharat Biotech will soon release its phase II and III pediatric trial results ([1233](#)).

However, the WHO approval of the Covaxin has been delayed ([1234](#)), and in April 2022, the WHO suspended procurement of Covaxin due to concerns about deviation from good manufacturing practice in their production facilities ([1235](#), [1236](#)). All the same, no safety issues had been reported in association with this vaccine, and the suspension was unlikely to affect distribution given that Bharat Biotech had not been supplying doses through this mechanism ([1237](#)). Clinical trials had recommenced in the United States as of May 2022 ([1237](#)).

6.5.2 Summary of IWV Vaccine Development

In the past, problems that arose during the manufacturing of IWV vaccines could present safety issues, but oversight of the manufacturing process has helped to improve IWV vaccine safety ([1238](#)). Nevertheless, the departure from norms necessitated by the COVID-19 crisis raised concerns about whether oversight would occur at pre-pandemic standards ([1238](#)). In general, the IWV COVID-19 vaccines have reported very few issues with safety. Additionally, safety audits have proactively identified concerns, as demonstrated with the WHO's suspension of Covaxin.

More concern has arisen around the issue of effectiveness due to the reduced neutralizing activity of IWV vaccines against VOC relative to the index strain. In several cases, estimates of VE have varied widely across different trials of a single vaccine. Such issues are likely to be exacerbated by spatiotemporal differences in viral evolution, though in the case of the very high estimate generated by the Turkish trial of CoronaVac ([1177](#)), the design of the study may have inflated the VE estimate ([1239](#)). Regardless, the authors of the original trial argued that all of the trials suggest a very high efficacy against severe disease ([1240](#)), as is the case for all of the IWV vaccines discussed here. In addition to issues related to the evolution of SARS-CoV-2, it is important to consider the duration of immunity over time. With IWV vaccines, heterologous vaccine boosters are being considered in many cases. Today, the WHO has developed recommendations for booster immunization for several whole-virus vaccines. In some cases (Valneva-VLA2001 ([1241](#)), Covaxin ([1242](#)), Covilo ([1243](#)), Sinopharm-WIBP Inactivated (Vero Cell) ([1244](#))), boosters are recommended only for high-risk and/or high-priority groups (e.g., the immunocompromised and medical professionals, respectively), while for Sinovac's CoronaVac ([1200](#)), they are recommended

more broadly. Studies are also investigating the effects of booster doses in other vaccines ([1245–1247](#)), though some are being investigated or deployed primarily as heterologous boosters in populations vaccinated with a different primary series ([1246](#)).

As new vaccines are approved by the WHO, more time elapses since many received the primary series, and new variants emerge, booster recommendations are likely to increase. Therefore, IWV vaccines have played an important role in vaccine access during the initial phase of vaccination against COVID-19, but many IWV vaccinees may receive booster doses developed with emergent vaccine technologies like DNA and mRNA. In head-to-head comparisons, these types of vaccines were typically found to outperform IWV vaccines (e.g., ([1190](#), [1192](#), [1250](#))). At the same time, IWV vaccines are among the easiest to store and transport due to requiring refrigeration only at 2 to 8°C and remaining stable for years at a time ([1205](#)). Therefore, these vaccines are likely to continue to play an important role in vaccine equity and accessibility.

6.6 Subunit Vaccines

Efforts to overcome the limitations of live-virus vaccines led to the development of approaches first to inactivate viruses (circa 1900), leading to IWV vaccines, and then to purifying proteins from viruses cultured in eggs (circa 1920) ([1113](#), [1251](#)). The purification of proteins then set the stage for the development of subunit vaccines based on the principle that the immune system can be stimulated by introducing one or more proteins or peptides isolated from the virus. Today, such approaches often use antigens isolated from the surface of the viral particle that are key targets of the immune system (protein subunit vaccines). Advances in biological engineering have also facilitated the development of approaches like viral-like particle (VLP) vaccines using nanotechnology ([1252](#)). VLPs share the conformation of a virus's capsid, thereby acting as an antigen, but lack the replication machinery ([1253](#)).

Unlike whole-virus vaccines, which introduce the whole virus, subunit vaccines stimulate the immune system by introducing one or more proteins or peptides of the virus that have been isolated. The main advantage of this platform is that subunit vaccines are considered very safe, as the antigen alone cannot cause an infection ([1254](#)). Both protein subunit and VLP vaccines thus mimic the principle of whole virus vaccines but lack the genetic material required for replication, removing the risk of infection ([1255](#)). Protein subunit vaccines can stimulate antibodies and CD4⁺ T-cell responses ([1253](#), [1256](#)).

The subunit approach is also favored for its consistency in production. The components can be designed for a highly targeted immune response to a specific pathogen using synthetic immunogenic particles, allowing the vaccine to be engineered to avoid allergen and reactogenic sequences ([1257](#)). One limitation is that, in the case of protein subunit vaccines, adjuvants are usually required to boost the immune response ([1258](#)) (see online Appendix ([1148](#))). Adjuvants, which are compounds that elicit an immunogenic effect,

include alum (aluminum hydroxide), squalene- or saponin-based adjuvants, and Freund's incomplete/complete adjuvants, although the latter is avoided in human and veterinary medicine due to high toxicity ([1257](#), [1259](#), [1260](#)).

Protein subunit vaccine development efforts for both SARS-CoV-1 and MERS-CoV explored a variety of immunogens as potential targets. The search for a potential SARS-CoV-1 vaccine included the development of vaccines based on the full-length or trimeric S protein ([1261–1263](#)), those focused on the RBD protein only ([1264–1267](#)) or non-RBD S protein fragments ([1262](#), [1268](#)), and those targeting the N and M proteins ([1269–1271](#)). These efforts have been thoroughly reviewed elsewhere ([1272](#)). There have been examples of successful preclinical research including candidate RBD219N-1, a 218-amino-acid residue of the SARS-CoV-1 RBD that, when adjuvanted to aluminum hydroxide, was capable of eliciting a high antibody response of both nAbs and RBD-specific monoclonal antibodies in both pseudovirus and live virus infections of immunized mice ([1273](#)).

Similarly to the SARS-CoV-1 vaccine candidates, the MERS-CoV protein subunit vaccine candidates generally target the RBD ([1264](#), [1272](#), [1274–1277](#)), with some targeting the full length S protein ([1278](#)), non-RBD protein fragments such as the SP3 peptide ([1279](#)), and the recombinant N-terminal domain (rNTD) ([1280](#)). Other strategies investigating the potential use of the full length S DNA have also been investigated in mice and rhesus macaques, which elicited immune responses ([1281](#)), but these responses were not as effective as the combination of S DNA and the S1 subunit protein together ([1281](#), [1282](#)). No protein subunit vaccine for MERS-CoV has progressed beyond preclinical research to date. VLPs have been investigated for development of vaccines against MERS and SARS ([1283](#), [1284](#)) including testing in animal models ([1285](#), [1286](#)), but once again, only preclinical data against HCoV has been collected ([1287](#)). However, protein subunit vaccines do play a role in public health and have contributed to vaccination against hepatitis B ([1288](#)) and pertussis ([1289](#), [1290](#)) since the 1980s and human papillomavirus since 2006 ([1291](#)). They are likely to continue to contribute to public health for the foreseeable future due to ongoing research in vaccines against influenza, SARS-CoV-2, Epstein-Barr virus, dengue virus, and human papillomavirus among others ([1292–1294](#)).

6.6.1 Application to COVID-19

Table 3: Subunit vaccines approved for use in at least one country ([1169](#)) as of May 3, 2023 ([1115](#)).

Vaccine	Company	Platform
Zifivax	Anhui Zhifei Longcom	protein subunit
Noora vaccine	Bagheiat-allah University of Medical Sciences	protein subunit
Corbevax	Biological E Limited	protein subunit
Abdala	Center for Genetic Engineering and Biotechnology (CIGB)	protein subunit

Vaccine	Company	Platform
Soberana 02	Instituto Finlay de Vacunas Cuba	protein subunit
Soberana Plus	Instituto Finlay de Vacunas Cuba	protein subunit
V-01	Livzon Mabpharm Inc	protein subunit
Covifenz	Medicago	VLP
MVC-COV1901	Medigen	protein subunit
Recombinant SARS-CoV-2 Vaccine (CHO Cell)	National Vaccine and Serum Institute	protein subunit
Nuvaxovid	Novavax	protein subunit
IndoVac	PT Bio Farma	protein subunit
Razi Cov Pars	Razi Vaccine and Serum Research Institute	protein subunit
VidPrevtyn Beta	Sanofi/GSK	protein subunit
COVOVAX (Novavax formulation)	Serum Institute of India	protein subunit
SKYCovione	SK Bioscience Co Ltd	protein subunit
TAK-019 (Novavax formulation)	Takeda	protein subunit
SpikoGen	Vaxine/CinnaGen Co.	protein subunit
Aurora-CoV	Vector State Research Center of Virology and Biotechnology	protein subunit
EpiVacCorona	Vector State Research Center of Virology and Biotechnology	protein subunit

The development of subunit vaccines against SARS-CoV-2 is a remarkable achievement given the short period of time since the emergence of SARS-CoV-2 in late 2019, particularly considering these types of vaccines have not played a major role in previous pandemics compared to LAV and IWV vaccines. More than 20 protein subunit vaccines from companies such as Sanofi/GlaxoSmithKline, Nanogen, and the Serum Institute of India have entered clinical trials for COVID-19 since the beginning of the pandemic ([1293](#)), 20 have been approved, and at least 9 are being administered worldwide ([1115](#), [1116](#)) (Table [3](#)). As of May 31, 2023, protein subunit vaccines are being distributed in at least 42 countries (Figure [7](#)).

Number of Protein Subunit Vaccines Available Worldwide

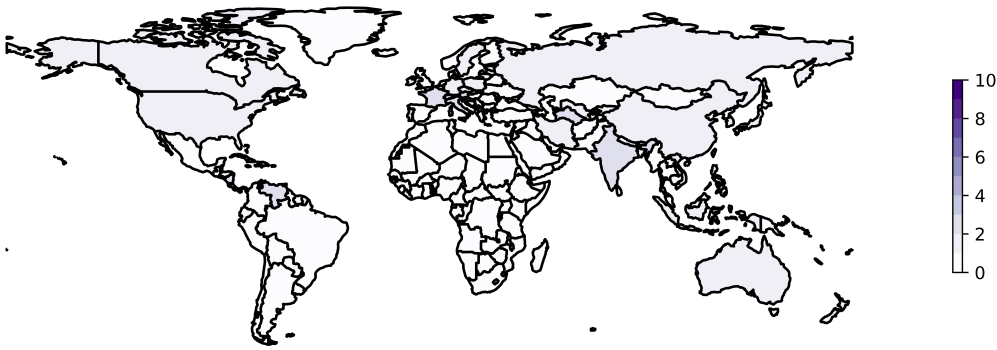


Figure 7: Worldwide availability of vaccines developed using protein subunit. This figure reflects the number of vaccines based on protein subunit technology that were available in each country as of May 31, 2023. These data are retrieved from Our World in Data ([532](#), [1116](#)) and plotted using geopandas ([1170](#)). The color scale is based on the number of vaccines of this type included in the OWID dataset as a whole, not the maximum observed in a single country. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

VLP vaccines have not progressed as rapidly. Programs seeking to develop VLP vaccines have used either the full-length S protein or the RBD of the S protein specifically as an antigen, although some use several different SARS-CoV-2 proteins ([1254](#)). As of May 31, 2023, only one VLP was available in one country (Canada) ([1116](#)).

Number of VLP Vaccines Available Worldwide



Figure 8: Worldwide availability of vaccines developed with VLPs. This figure reflects the number of vaccines based on VLP technology that were available in each country as of May 31, 2023. These data are retrieved from Our World in Data ([532](#)) and plotted using geopandas ([1170](#)). The color scale is based on the number of vaccines of this type included in the OWID dataset as a whole, not the maximum observed in a single country. See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily.

6.6.2 Novavax's Nuvaxovid

NVX-CoV2373, also known as Nuvaxovid or Covovax ([1295](#)), is a protein subunit vaccine for SARS-CoV-2 produced by U.S. company Novavax and partners. Nuvaxovid is a protein nanoparticle vaccine constructed from a mutated prefusion SARS-CoV-2 spike protein in combination with a specialized saponin-based adjuvant to elicit an immune response against SARS-CoV-2 ([1296](#)). The spike protein is recombinantly expressed in Sf9 insect cells ([1297](#)), which have previously been used for several other FDA-approved protein therapeutics ([1298](#)), and contains mutations in the furin cleavage site

(682-RRAR-685 to 682-QQAQ-685) along with two proline substitutions (K986P and V987P) that stabilize the protein ([1299](#)), including improving thermostability ([1297](#)).

In preclinical mouse models, Nuvaxovid elicited high anti-spike IgG titers 21 to 28 days post-vaccination that could neutralize the SARS-CoV-2 virus and protect the animals against viral challenge, with particularly strong effects when administered with the proprietary adjuvant Matrix-M™ ([1297](#)). In a phase I/II trial, a two-dose regimen of Nuvaxovid was found to induce anti-spike IgG levels and nAb titers exceeding those observed in convalescent plasma donated by symptomatic patients ([1296](#)). In line with the preclinical studies, the use of Matrix-M adjuvant further increased anti-spike immunoglobulin levels and induced a Th1 response.

In a phase III randomized, observer-blinded, placebo-controlled clinical trial in the U.K., 14,039 participants received two 5-µg doses of Nuvaxovid or placebo administered 21 days apart in a 1:1 ratio from late September to late November 2020 ([1300](#)). The primary endpoint of the trial was the occurrence or absence of PCR-confirmed, symptomatic mild, moderate or severe COVID-19 from 7 days after the second dose onward ([1300](#)). The VE was reported to be 89.7%, with a total of 10 patients developing COVID-19 in the vaccine group versus 96 in the placebo group ([1300](#)). No hospitalizations or deaths were reported in the vaccine group ([1300](#)). An additional phase III randomized, observer-blinded, placebo-controlled trial was conducted in the U.S. and Mexico, enrolling 29,949 participants and administering at least 1 vaccine in a 2:1 ratio from late December 2020 to late February 2021 ([1301](#)). This trial ([1301](#)) used the same primary endpoints as the initial phase III trial conducted in the U.K. ([1300](#)). A vaccine efficacy of 90.4% was reported based on 77 cases total, 63 of which occurred in the placebo group ([1301](#)). All moderate to severe cases of COVID-19 occurred in the placebo group ([1301](#)). Hospitalization and death were not evaluated as individual secondary endpoints, but were instead included in the definition of severe COVID-19; all-cause mortality was comparable between the placebo and treatment conditions ([1301](#)).

In both trials, the vaccine was found to be well-tolerated ([1300](#), [1302](#)). Analysis of 2,310 participants in the U.K. trial revealed that solicited AEs were much more common in the vaccine group than the placebo group across both doses, but the rate of unsolicited events was, while still higher in the vaccine group, much more similar ([1300](#)). A small number of severe AEs were reported by vaccine recipients, including one case of myocarditis; however, the myocarditis was determined to be viral myocarditis ([1300](#)). Common AEs were generally considered mild, with low incidences of headache, muscle pain, and fatigue ([1301](#)). In the trial conducted in the U.S. and Mexico, once again, the most common symptoms included headache, fatigue, and pain, as well as malaise ([1301](#)). Here, severe AEs were balanced across the vaccine and placebo groups ([1301](#)). Thus, both trials suggested that the Nuvaxovid vaccine is safe and effective against COVID-19.

However, Novavax experienced significant challenges in preparing Nuvaxovid for distribution. Prior to the pandemic's onset, Novavax had sold their manufacturing facilities and reduced their staff dramatically ([1303](#)). As a result, once they began producing Nuvaxovid, they struggled to establish a

stable relationship with contractors who could produce the vaccine (1304), especially given the challenge of producing vaccines at scale (1305). Additionally, Novavax was not able to meet the purity standards laid out by the FDA (1306). Eventually, the manufacturing issues were resolved (1307), and Nuvaxovid has since been authorized by the WHO (1308) and by political entities, including the United Kingdom (1309), the E.U. (1310), and the U.S. (1311). These delays obstructed some of the goals of the vaccine development program, which was undertaken with significant investment from the U.S. government through Operation Warp Speed (1306). Novavax was supposed to provide over a billion doses of Nuvaxovid to countries around the world through the COVID-19 Vaccines Global Access (COVAX) Facility (1306). However, following the delays, Gavi (which oversees COVAX) terminated the agreement, leading to ongoing legal disagreements between the nonprofit and Novavax as of late 2022 (1307, 1312).

As with other vaccines, the question of how well Nuvaxovid continues to provide protection as SARS-CoV-2 evolves has been raised. *Post hoc* analysis in the phase III trial indicated a VE of 86.3% against the Alpha variant (identified based on the presence of the 69–70del polymorphism) and 96.4% against viral specimens lacking the 69-70del polymorphism (1300). In the second phase III trial (1302), whole-genome sequencing was obtained from 61 of the 77 observed cases, and 79% of infections were identified as a VOC or variant of interest (VOI) known at the time of the study. Vaccine efficacy against cases caused by VOC, among which the Alpha variant was predominant (88.6%), was reported to be 92.6% (1302). In late 2020, an analysis of efficacy in South African adults revealed an overall efficacy of 60.1% and a slightly lower efficacy of 50.1% against the Beta variant (B.1.351) in particular (1313).

The company has also initiated the development of new constructs to select candidates that can be used as a booster against new strains and plans to initiate clinical trials for these new constructs in the second quarter of 2021. An analysis of a booster dose of Nuvaxovid administered six months after the primary series revealed a significant increase in neutralizing activity against VOC including Delta and Omicron (1314). This trial was conducted at 18 sites across the United States and Australia (1315). Novavax has also initiated booster trials in the U.K. (1186). Boosters may be especially important given that Omicron and related variants, in particular, may be associated with significantly reduced efficacy of Nuvaxovid (1316).

Given the apparent need for boosters, interest has also emerged in whether booster doses of Nuvaxovid can be safely administered along with annual flu vaccines. In a subgroup of approximately 400 patients enrolled from the U.K. phase III trial who received either Nuvaxovid or a placebo at a ratio of 1:1, a concomitant dose of adjuvanted seasonal influenza vaccines (either a trivalent vaccine or a quadrivalent vaccine) was administered (1317). This study demonstrated that the vaccines could be safely administered together (1317). While no change to the immune response was noted for the influenza vaccine, a notable reduction of the antibody response elicited by Nuvaxovid was reported, but efficacy was still high at 87.5% (1317). Novavax has since started phase I/II trials to investigate the administration of its own influenza vaccine, NanoFlu, concomitantly with Nuvaxovid (1318). The combination appeared to be safe and effective in preclinical studies (1319).

6.6.3 The Cuban Center for Genetic Engineering and Biotechnology's Abdala Vaccine

Another notable protein subunit vaccine development program came out of Cuba. Concerned about their ability to access vaccines, especially given the U.S.'s embargo ([1320](#)), health officials in this developing country made the decision in March 2020 to undertake vaccine development on their own ([1321](#)). Today, three Cuban protein subunit vaccines have been approved for use: Abdala, which was developed at the Cuban Genetic Engineering and Biotechnology Center and SOBERANA 02 and SOBERANA Plus, which were developed at Cuba's Finlay Vaccine Institute (Instituto Finlay de Vacunas Cuba) ([1321](#)). Here, we focus on the development of the Abdala vaccine, but SOBERANA 01/02/Plus vaccine development program has also achieved great success and reported VEs of over 90% in the three-dose regimen ([1322](#), [1323](#)).

Abdala, also known as CIGB-66, was developed using yeast as a low-cost alternative to mammalian cell expression systems (e.g., human embryonic kidney cells) to cultivate the recombinant proteins that form the basis of this protein subunit vaccine ([1324](#)). A sequence corresponding to the RBD of the Spike protein in the index strain of SARS-CoV-2 was codon optimized for expression in yeast, and the RBD proteins were then purified and used to inoculate mice, rats, and African green monkeys ([1324](#)). In addition to the proteins, the vaccine candidate included an adjuvant, aluminum hydroxide gel ([1324](#)). Comparing the immunogenicity of the yeast-cultivated proteins to those cultivated in human embryonic kidney cells revealed no significant difference in the immune response ([1324](#)).

Based on promising results in laboratory animal testing, Abdala moved to phase I/II trials in human subjects ages 19 to 80, recruiting participants between December 2020 and February 2021 ([1325](#)). The three-dose vaccine elicited no serious AEs across either phase I or II, and the vaccine was found to produce a strong immune response ([1325](#)). In March 2021, phase III trials began ([1326](#)), and by June, officials were reporting the VE to be 92.28% ([1327](#), [1328](#)). This high efficacy estimate, along with the short timeline of data collection, initially elicited skepticism, especially given that the data were not made public ([1329](#)). However, the trials were designed to enroll a large number of participants and were carried out during a wave of infections due to the arrival of variants carrying the D614G mutation in Cuba, which would be expected to allow an expedited timeline for interim analysis ([1329](#)). Based on the reported results, Abdala gained emergency use authorization in Cuba in July 2021 ([1330](#)), and by December 2021, cases in Cuba had dropped dramatically ([1331](#)). The results of the phase III trial were posted to *medRxiv* in September 2022, describing the results of a randomized, placebo-controlled, multicenter, double-blind investigation of the Abdala vaccine candidate in 48,000 participants between March 22 and April 3, 2021 ([1332](#)). The final results evaluated 42 symptomatic cases of COVID-19 among participants in the placebo condition compared to only 11 cases among participants who received the vaccine, yielding the reported VE of 92.28% ([1332](#)). In terms of secondary endpoints, the VE was 91.96% against mild/moderate COVID-19, 94.46% against severe COVID-19, and 100% against critical illness and death ([1332](#)). The vaccine was also found to be very safe,

with the overall incidence of AEs only 2.5% in vaccine recipients compared to 1.9% in the placebo recipients (1332). Therefore, the phase III trial suggests that this vaccine is highly effective and safe.

Evidence from the deployment of the vaccine also suggests it is highly effective. A retrospective cohort study conducted between May and August 2021 evaluated public health data from over a million people in the city of Havana and found that the real-world effectiveness of the vaccine met or exceeded estimates of VE during the trial, with 98.2% effectiveness against severe disease and 98.7% effectiveness against death observed in fully vaccinated subjects (1333). Notably, Cuba has vaccinated a high percentage of its population, with 87% of the population vaccinated by January 2022 and 90.3% by the end of December 2022 (1334, 1335). Therefore, one consideration in interpreting retrospective cohort studies is that the vaccination rate in Cuba is so high that the two cohorts might not be directly comparable. All the same, the fact that the efficacy and effectiveness of the Abdala vaccine have both been estimated to be over 90% against severe illness suggests that this vaccine is highly effective for mitigating the risk of COVID-19. As of December 2022, the vaccine had been authorized for distribution in five additional countries, including Mexico and Vietnam, although its evaluation for WHO approval was ongoing (1336, 1337).

However, limited data is available about the Abdala's vaccine's robustness to evolutionary changes in SARS-CoV-2. An *in silico* analysis identified several potential changes in the epitopes of the Omicron VOC relative to the sequence used in the development of Abdala (1338). Instead, Cuban health officials have prioritized boosters. A representative of the Cuban state business group reportedly stated that immunity remains high at six months after the primary course but that some people may be prone to infection (1339), suggesting waning immunity. The Cuban government authorized boosters in January 2022 in an effort to mitigate the effects of the Omicron variant (1339–1341). Additional support for the efficacy of Abdala and other Cuban vaccines comes from the fact that Cuba's COVID-19 death rate has virtually flatlined since fall 2021, with less than 250 deaths reported during the entire year of 2022 in a population of 11.3 million (1342). Therefore, in addition to developing a vaccine with an estimated VE paralleling that of vaccines developed using cutting-edge nucleic acid technologies (6), Cuba's vaccine roll out has also been much more successful than in nearly all similarly sized countries. This remarkable vaccine program underscores the continued importance of established, cost-effective vaccine development strategies (1340) that make it possible for countries that have not traditionally been a leader in biotechnological innovation but have developed a solid vaccine production sector (1343) to develop and produce vaccines that will serve their own population's needs. Additionally, Cuba's vaccines are uniquely accessible to many countries around the world (1340).

6.6.4 Medicago's Covifenz

The leading example of a VLP approach applied to COVID-19 comes from Covifenz, developed by Canadian company Medicago (1344). This vaccine was developed using plant-based VLP technology (1345) that the company had been investigating in order to develop a high-throughput quadrivalent VLP platform to provide protection against influenza (1346). The approach utilizes

Nicotiana benthamiana, an Australian relative of the tobacco plant, as an upstream bioreactor ([1346](#), [1347](#)). Specifically, the *S* gene from SARS-CoV-2 in its prefusion conformation is inserted into a bacterial vector (*Agrobacterium tumefaciens*) that then infects the plant cells ([1346](#), [1347](#)). Expression of the *S* glycoprotein causes the production of VLPs composed of *S* trimers anchored in a lipid envelope that accumulate between the plasma membrane and the cell wall of the plant cell ([1347](#)). Because these VLPs do not contain the SARS-CoV-2 genome, they offer similar advantages to whole-virus vaccines while mitigating the risks ([1346](#), [1347](#)).

In the phase I study, 180 Canadian adults ages 18 to 55 years old were administered Covifenz as two doses, 21 days apart, with three different dosages evaluated ([1347](#)). This study reported that when the VLPs were administered with AS03, an oil-in-water emulsion containing α -tocopherol and squalene ([1348](#)), as an adjuvant, the vaccine elicited an nAb response that was significantly (approximately 10 times) higher than that in convalescent sera ([1347](#)). The phase III trial examined 24,141 adults assigned to the treatment and control conditions at a 1:1 ratio between March and September of 2021 ([1349](#)).

Covifenz was reported to be 71% effective in preventing COVID-19 in the per-protocol analysis ([1349](#)). Efficacy was only slightly lower in the intention-to-treat group at 69%, with the VE for the prevention of moderate-to-severe disease in this group estimated at 78.8% ([1349](#)). Over 24,000 participants were included in the safety analysis, which reported that 92.3% of vaccine recipients reported local AEs compared to 45.5% of placebo recipients, with rates for systemic AEs at 87.3% and 65.0%, respectively ([1349](#)). The adverse effects reported were generally mild to moderate, with the most common adverse effects being injection site pain, headache, myalgia, fatigue, and general discomfort ([1349](#)). Only three patients (two in the vaccine group) reported grade 4 events, all after the second dose ([1349](#)). The vaccine was approved for use in adults ages 18 to 65 by Health Canada in February 2022 ([1350](#)).

Plant-based expression systems such as the one used in Covifenz are relatively new ([1347](#)) but are likely to offer unparalleled feasibility at scale given the speed and low-cost associated with the platform ([1351](#)). Additionally, the Covifenz vaccine offers the advantage of being stored at 2 to 8°C. However, the worldwide footprint of Covifenz, and of VLP-based technologies against SARS-CoV-2 broadly, remains small, with only 1 VLP vaccine approved for distribution in 1 countries (Figure [8](#)). Approval and administration of Covifenz in countries outside of Canada has been limited by concerns at the WHO about ties between Medicago and the tobacco industry ([1344](#), [1352](#)). While other species of plants have been explored as the upstream bioreactors for plant-derived VLPs, the specific species of tobacco used increased yield dramatically ([1353](#)). In December 2022, tobacco industry investors in Medicago divested, opening new possibilities for the distribution of the vaccine ([1354](#)).

As a result of this limited roll-out and given that the phase III results were published only in May 2022, little is known about the real-world performance of Covifenz. However, it should be noted that the Covifenz trials were conducted in 2021, at a time during which the B.1.617.2 (Delta) and P.1

(Gamma) variants were predominant ([1349](#)). Genomic analysis of 122 out of 176 cases (165 in the per-protocol population) revealed that none of the COVID-19 cases reported were caused by the original Wuhan strain ([1349](#)). Instead, 45.9% of cases were identified as the Delta variant, 43.4% as Gamma, 4.9% as Alpha, and 5.8% as VOIs ([1349](#)). Therefore, Covifenz and Nuvaxovid, despite both being designed based on the index strain, were tested under circumstances where different VOC were dominant, and differences in the Spike proteins of different VOC relative to the index strain could affect vaccine efficacy. As of late 2022, Covifenz has not been authorized as a booster in Canada ([1355](#)), and no studies on booster doses had been released by Medicago ([1356](#)).

6.6.5 Subunit Vaccine Summary

Subunit vaccine technology is one of the best-represented platforms among COVID-19 vaccine candidates. Development programs are underway in many countries around the world, including low- and middle-income countries ([1293](#)). To date, data about the effect of viral evolution on the effectiveness of subunit vaccines has been limited. Because these vaccines were developed using the Spike protein from the index strain ([1297](#), [1349](#)), a potential concern has been that these vaccines could lose effectiveness against SARS-CoV-2 containing mutations in the Spike protein. Comparison of studies across vaccines suggests that some VOC, such as Alpha, may have minimal impact on vaccine efficacy/effectiveness ([1357](#)). Additionally, to the extent that data is available such as from the vaccine rollout in Cuba, it suggests that real-world effectiveness remains strong against severe illness and death.

Subunit platforms offer some unique advantages. Cuba's successful vaccine development programs highlights the fact that protein subunit vaccines can be developed using low-cost technologies. Additionally, they are more feasible to store and transport ([1358](#)). Hoping to build on Cuba's success and the continued lack of vaccine access in many countries, several Latin American nations have begun developing protein subunit vaccines ([1359](#)).

The efficacy and effectiveness of these vaccines is also very high, especially for Nuvaxovid, Abdala, and SOBERANA 01/02/Plus, where estimates exceeded 90%. Unfortunately, there seem to be limited studies directly comparing the immunogenicity of subunit vaccines to nucleic acid vaccines, and comparing efficacies across trials is subject to bias ([1360](#)). All the same, the evidence suggests that some protein subunit vaccines are able to provide extremely strong protection. Coupled with the reduced barriers to development and transportation relative to most nucleic acid vaccines, it is clear that subunit technologies are important to vaccine access.

6.7 Global Vaccine Status and Distribution

The unprecedented deployment of COVID-19 vaccines in under a year from the identification of SARS-CoV-2 led to a new challenge: the formation of rapid global vaccine production and distribution plans. The development of vaccines is costly and complicated, but vaccine distribution can be just as challenging. Logistical considerations such as transport, storage, equipment (e.g., syringes), the workforce to administer the vaccines, and a continual

supply from the manufacturers to meet global demands all must be accounted for and vary globally due to economic, geographic, and sociopolitical reasons ([1361–1363](#)). As of May 25, 2023, at least 13.0 billion vaccine doses had been administered in at least 223 countries worldwide using 28 different vaccines ([532](#)).> The daily global vaccination rate at this time was 8.0 per million.

However, the distribution of these doses is not uniform around the globe. Latin America leads world vaccination rates with at least 82% of individuals in this region receiving one vaccine dose followed by the U.S. and Canada (81%), Asia-Pacific (81%), Europe (70%), the Middle East (58%), and Africa with only 33% as of November 2022 ([1364](#)). It is estimated that only ~25% of individuals in low- and middle-income countries have received one vaccine dose ([1116](#), [1365](#)). Vaccine production and distribution varies from region to region and seems to depend on the availability of the vaccines and potentially a country's resources and wealth ([1366](#)).

One effort to reduce these disparities is COVAX, a multilateral initiative as part of the Access to COVID-19 Tools (ACT) Accelerator coordinated by the WHO, Gavi, the Vaccine Alliance, and the Coalition for Epidemic Preparedness Innovations (CEPI), the latter two of which are supported by the Bill and Melinda Gates Foundation. Their intention is to accelerate the development of COVID-19 vaccines, diagnostics, and therapeutics and to ensure the equitable distribution of vaccines to low- and middle-income countries ([1367](#), [1368](#)). COVAX invested in several vaccine programs to ensure they would have access to successful vaccine candidates ([1369](#)). However, the initiative has been less successful than was initially hoped due to less participation from high-income countries than was required for COVAX to meet its goals ([1370](#)).

Additionally, the vaccine technologies available differ widely around the globe. As we review elsewhere ([6](#)), wealthier nations have invested heavily in mRNA and DNA vaccines. In contrast, as we describe above, many countries outside of Europe and North America have developed highly effective vaccines using more traditional approaches. There is a clear relationship between a country's gross domestic product (GDP) and its access to these cutting-edge types of vaccines (Figure [9](#)). Whole-virus and subunit vaccine development programs are responsible for a much higher percentage of the vaccinated populous in lower-income countries. Therefore, vaccine development programs that utilized established vaccine technologies have played a critical role in providing protection against SARS-CoV at the global level.

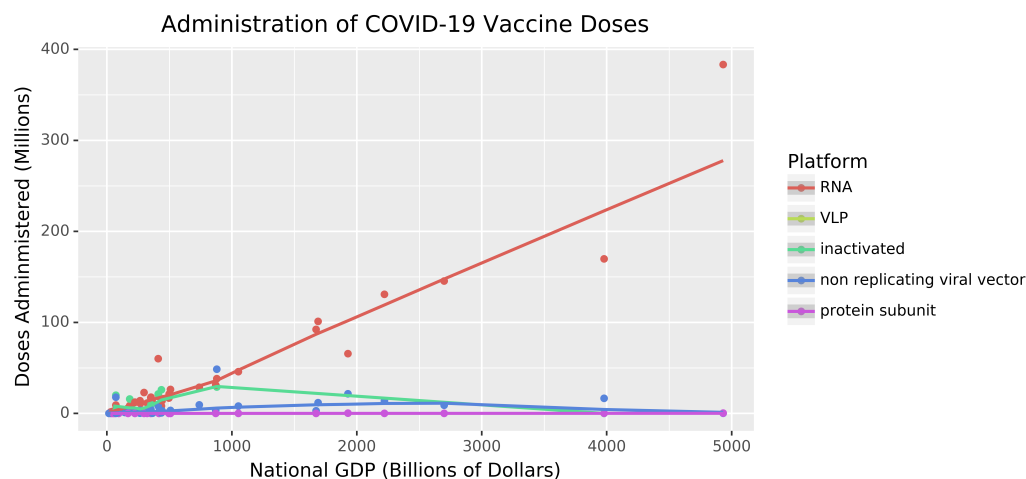


Figure 9: Vaccine Distribution across Platform Types as a Function of GDP. The total number of doses of the original formulation of each vaccine that were distributed within each country as of May 31, 2023, by platform type, is shown as a function of GDP. These data are retrieved from Our World in Data ([532](#), [1116](#)) and plotted using the Python package plotnine ([1371](#)). Lines show a general trend in the data and are drawn using `geom_smooth` ([1372](#)). The list of countries included in the dataset is available from OWID ([1373](#)). See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily. Axes are not scaled per capita because both variables are modulated by population size.

When vaccines first became available, the wealthy nations of North America and Europe secured most of the limited COVID-19 vaccine stocks ([1374](#)). Throughout 2021, low- and middle-income countries faced steep competition with high-income countries for vaccines, and the rates of vaccination reflected this unequal distribution ([1375](#)). While the wealthiest countries in these regions could compete with each other for vaccines independent of programs such as COVAX ([1375](#)), other countries in these regions have faced challenges in acquiring vaccines developed by the world's wealthiest nations. Fortunately, while mRNA and DNA vaccine development programs are not widespread, vaccine development using whole-virus and subunit technologies has been undertaken worldwide. China and India, in particular, have developed several vaccines that are now widely available in these densely populated countries (see online Appendix ([1148](#))).

Still, many nations, especially in Africa, are reliant on the COVAX Facility, who have promised 600 million doses to the continent ([1376](#)). The COVAX plan seeks to ensure that all participating countries would be allocated vaccines in proportion to their population sizes. Once each country has received vaccine doses to account for 20% of their population, the country's risk profile will determine its place in subsequent phases of vaccine distribution. However, several limitations of this framework exist, including that the COVAX scheme seems to go against the WHO's own ethical principles of human well-being, equal respect, and global equity and that other frameworks might have been more suitable ([1377](#)). Furthermore, COVAX is supposed to allow poorer countries access to affordable vaccines, but the vaccines are driven by publicly traded companies that are required to make a profit ([1366](#)).

In any case, COVAX provides access to COVID-19 vaccines that may otherwise have been difficult for some countries to obtain. COVAX aimed to distribute 2 billion vaccine doses globally by the end of 2021 ([1378](#)). According to Gavi, as of January 2022, COVAX had distributed over 1 billion vaccines to 144

participants of the program ([1379](#)), short of its target but still a major global achievement. It is envisaged that COVAX may also receive additional donations of doses from Western nations who purchased surplus vaccines in the race to vaccinate their populations, which will be a welcome boost to the vaccination programs of low- and middle-income countries ([1380](#)).

In general, deciding on the prioritization and allocation of the COVID-19 vaccines is also a challenging task due to ethical and operational considerations. Various frameworks, models, and methods have been proposed to tackle these issues with many countries, regions, or U.S. states devising their own distribution and administration plans ([1381–1385](#)). The majority of the distribution plans prioritized offering vaccines to key workers such as health care workers and those who are clinically vulnerable, such as the elderly, the immunocompromised, and individuals with comorbidities, before targeting the rest of the population, who are less likely to experience severe outcomes from COVID-19 ([1386](#)). The availability of vaccines developed in a variety of countries using a variety of platforms is likely to work in favor of worldwide vaccine access. The initiative by Texas Children's Hospital and Baylor College of Medicine to develop Corbevax, a patent-free COVID-19 protein subunit vaccine, is an important step towards vaccine equity because the manufacturing specifications can be shared globally. Corbevax can be produced at low cost using existing technology and is now licensed to Biological E. Limited (BioE), an Indian company specializing in low-cost vaccine production ([1387](#)). The vaccine has been approved for distribution in India and Botswana ([1388](#)).

Logistical challenges and geographical barriers also dictate the availability of certain vaccines. Many countries have had poor availability of ultra-low temperature freezers, leading to challenges of distribution for vaccines such as mRNA vaccines that require storage at very low temperatures ([1389–1391](#)). Furthermore, ancillary supplies such as vaccine containers, diluents for frozen or lyophilized vaccines, disinfecting wipes, bandages, needles, syringes, sharps and biological waste disposal containers are also required, which may not be readily available in geographically isolated locations and can be bulky and expensive to ship ([1389](#)). While some of these challenges in vaccine rollout in low- and middle-income countries are being addressed through COVAX ([1392](#)), many issues persist worldwide ([1393–1395](#)). COVAX also failed to distribute its promised two billion vaccine doses on time due to multiple complications ([1396](#)).

Another major challenge to global vaccine distribution is vaccine hesitancy, which the WHO has designated as a leading global health threat ([1397](#)). Polling in the U.S. in January 2021 suggested that 20% of individuals were reluctant to receive a vaccine at that time, with a further 31% expressing some hesitancy to a lesser extent ([1398, 1399](#)). A survey of 8,243 long-term healthcare workers in November 2020 (Indiana, USA) reported that only 69% of respondents would ever consider receiving an FDA-approved vaccine due to their perceived risk of side effects (70%), health concerns (34%), efficacy (20%), and religious beliefs (12%) ([1400](#)). Notably, almost a third of parents surveyed in the United States in March 2021 expressed concerns about vaccinating their children against COVID-19 ([1401](#)). Indeed, vaccine hesitancy has been reported as a significant barrier to vaccine distribution in countries in North and South America, Europe, Asia, and Africa ([1402–1406](#)). Various

factors have been associated with increased vaccine hesitancy including access to compelling misinformation via social media ([1407](#), [1408](#)), religious and conservative political beliefs ([1409–1412](#)), and safety and efficacy concerns ([1401](#)), to highlight a few. Many of the concerns regarding safety and efficacy have focused on the novel mRNA technologies due to the perceived speed of their development and expedited clinical trial process ([1413](#)); however, general vaccine hesitancy relating to traditional vaccine platforms existed long before the pandemic and the distribution of the novel mRNA vaccines ([1414](#), [1415](#)). While in the United States, it was hoped that Novavax's Nuvaxovid would appeal to the vaccine hesitant ([1416](#), [1417](#)), but this protein subunit vaccine has not led to the uptake hoped ([1418](#), [1419](#)).

Overall, the vaccine landscape remains heterogeneous even as the pandemic nears its third year, with certain vaccines much more accessible in high-income countries than in low- and middle-income countries. The vaccines described in this manuscript, which were developed using well-established technologies, have played a crucial role in improving the feasibility and accessibility of vaccination programs worldwide. While the novel technologies have received the bulk of public attention in countries like the U.S., these more traditional vaccine platforms also provide safe and highly effective protection against SARS-CoV-2. Although companies developing cutting-edge technologies, namely Moderna and Pfizer/BioNTech, reported very high efficacies greater than 90% in their clinical trials ([6](#)), the efficacies identified in whole-virus and subunit trials have also been very high. While the clinical trial efficacy estimates for IIV and subunit have been lower, some of these trials have also reported efficacies over 80% (e.g., Novavax's Nuvaxovid with 89.7% ([1300](#)) or Sinovac's CoronaVac with 83.5% ([1177](#))). Variation among studies investigating the efficacy of these vaccines, especially CoronaVac, clearly indicate that clinical trials of the same vaccine might not identify the same efficacy, depending on conditions such as the specific variants circulating in a clinical trial population during the trial period. Additionally, there are many cohort- and population-level characteristics that can introduce bias within and between clinical trials ([1360](#), [1420](#)), and the extent to which these different factors are present may influence trial outcomes. While head-to-head comparisons of VE across different studies may therefore not be appropriate, the results make it clear that effective vaccines have been developed with a wide variety of technologies. The vaccines discussed here, which took advantage of well-established approaches, have proven to be especially valuable in pursuing vaccine equity.

6.8 Conclusions

Much attention has focused on the most novel vaccine technologies that have been deployed against SARS-CoV-2, but the established vaccine platforms discussed here have all made a significant impact on human health during the twentieth century and in some cases even earlier. The COVID-19 pandemic has demonstrated new potential in these established technologies. In the early 2000s, these technologies were explored for managing SARS-CoV-1 ([1421](#), [1422](#)), but the epidemic was controlled before those efforts came to fruition ([1423](#)). Similarly, these technologies were explored for MERS-CoV, but outbreaks were sporadic and difficult to predict, making vaccine testing and the development of a vaccination strategy difficult ([1424](#)). However, in the COVID-19 pandemic, most of these

technologies have been used to accelerate vaccine development programs worldwide. Therefore, they are also offering the opportunity to respond quickly to an emergent pathogen.

While these tried-and-true technologies do not always produce vaccines with the highly desirable VE reported in mRNA clinical trials (which exceeded 90%), the efficacies are still very high, and these vaccines are extremely effective at preventing severe illness and death. Furthermore, some vaccine development programs using established technologies, especially protein subunit vaccines, have seen remarkably high VE and vaccine effectiveness. Some protein subunit vaccine phase III trials generated VE estimates of over 90%, comparable to those in the mRNA vaccine trials. Additionally, in some cases, such as Cuba's highly successful vaccine development program, these vaccines have been developed by and for low- and middle-income nations. As a result, the greater accessibility and stability of these vaccines makes them extremely valuable for the global effort to mitigate the loss of life from SARS-CoV-2. The outcomes of the response to COVID-19 suggests that these established vaccine technologies may continue to play an important role in tackling future viral threats.

7 Appendix: Additional Information about Established Vaccine Platforms for COVID-19

7.1 Sinovac's CoronaVac

The CoronaVac vaccine was developed by Sinovac, a Beijing-based biopharmaceutical company. The vaccine uses an inactivate whole virus with the addition of an aluminum adjuvant ([1425](#)). Pre-clinical trials were performed using BALB/c mice and rhesus macaques ([1173](#)). The SARS-CoV-2 strains used in this trial isolated from 11 hospitalized patients (5 from China, 3 from Italy, 1 from the United Kingdom (U.K.), 1 from Spain, 1 from Switzerland). A phylogenetic analysis demonstrated that the strains were representative of the variants circulating at the time. One of the strains from China, CN2, was used as the inactivated and purified virus while the other 10 strains were used to challenge. CN2 was grown in Vero cells. The immunogenicity of the vaccine candidate was evaluated with an ELISA assay. Ten mice were injected with the vaccine on day 0 and day 7 with varying doses (0, 1.5, 3, or 6 µg), and 10 mice were treated with physiological saline as the control. IgG developed in the serum of all vaccinated mice.

Using the same setup, immunogenicity was also assessed in macaques. Four macaques were assigned to each of four groups: treatment with 3 µg at day 0, 7, and 14, treatment with a high dose of 6 µg at day 0, 7, and 14, administration of a placebo vaccine, and administration of only the adjuvant. All vaccinated macaques induced IgGs and neutralizing antibodies. After challenge with SARS-CoV-2 strain CN1, vaccinated macaques were protected compared to control macaques (placebo or adjuvant only) based on histology and viral loads collected from different regions of the lung.

A single center, randomized, double-blind, placebo-controlled phase I/II trial was conducted in April 2020 in adults 18-59 years old. Patients in this study were recruited from the community in Suining County of Jiangsu province, China. For the phase I trial, 144 (of 185 screened) participants were enrolled, with 72 enrolled in the 14-day interval cohort (i.e., treated on day 0 and day 14) and 72 in the 28-day interval cohort. This group of 72 participants was split into 2 blocks for a low-dose (3 µg) and high-dose (6 µg) vaccine. Within each block, participants were randomly assigned vaccination with CoronaVac or placebo (aluminum diluent without the virus) at a 2:1 ratio. Both the vaccine and placebo were prepared in a Good Manufacturing Practice-accredited facility of Sinovac Life Sciences (Beijing, China).

The phase II trial followed the same organization of participants, this time using 300 enrolled participants in the 14-day and another 300 enrolled in the 28-day groups. One change of note was that the vaccine was produced using a highly automated bioreactor (ReadyToProcess WAVE 25, GE, Umeå, Sweden) to increase vaccine production capacity. This change resulted in a higher intact spike protein content. The authors of this study were not aware of this antigen-level difference between the vaccine batches for the phase I/II when the ethical approval for the trials occurred.

To assess adverse responses, participants were asked to record any events up to 7 days post-treatment. The reported adverse events were graded according to the China National Medical Products Administration guidelines. In the phase I trial, the overall incidence of adverse reactions was 29-38% of patients in the 0 to 14 day group and 13-17% in the 0 to 28 day vaccination group. The most common symptom was pain at the injection site, which was reported by 17-21% of patients in the 0 to 14 day cohort and 13% in the 0 to 28 day cohort. Most adverse reactions were mild (grade 1), with patients recovering within 48 hours. A single case of acute hypersensitivity with manifestation of urticaria 48 hours following the first dose of study drug was reported in the 6 µg group. Both the 14-day and 28-day cohorts had a strong neutralizing antibody (Ab) response. The neutralizing Ab response was measured using a micro cytopathogenic effect assay, which assesses the minimum dilution of neutralizing Ab to be 50% protective against structural changes in host cells in response to viral infection ([1426](#)). Additionally IgG antibody titers against the receptor binding domain were also measured using ELISA.

Another phase I/II study was performed with older patients (over 60 years of age) ([1172](#)). The study conducted a single-center, randomized, double-blind, placebo-controlled trial. The phase I trial looked at dose escalation using 3 dosages: 1.5, 3, and 6 µg. The mean age of participants was 65.8 years (standard deviation = 4.8). Of 95 screened participants, 72 were enrolled. These 72 participants were split into low (3 µg) and high (6 µg) dose groups. Within each group, 24 participants received the treatment and 12 the placebo. A neutralizing antibody response against live SARS-CoV-2 was detected compared to baseline using the same micro cytopathogenic effect assay. This response was similar across the two dose concentrations. Additionally, they did not observe a difference in response between age groups (60–64 years, 65–69 years, and ≥70 years).

In phase II the mean age was 66.6 years (standard deviation = 4.7). 499 participants were screened and 350 were enrolled. 300 were evenly split into 1.5, 3, and 6 µg dose groups, and the remaining 50 were assigned to the placebo group. Again, they found a neutralizing antibody response in phase II. There wasn't a significant difference between the response to 3 µg versus 6 µg, but the response in both these conditions was higher than in the 1.5 µg condition.

Participants were required to record adverse reaction events within the first 7 days after each dose. The safety results were combined across phase I and II. All adverse reactions were either mild (grade 1) or moderate (grade 2) in severity. The most common symptom was pain at the injection site (9%) and fever (3%). 2% (7 participants) reported severe adverse events (4 from the 1.5 µg group, 1 from the 3 µg group, 2 from the 6 µg group), though these were found to be unrelated to the vaccine.

Overall, the results from the pre-clinical and phase I/II clinical trials were promising, especially the fact that the immune response was consistent in older adults (> 60 years).

7.2 Sinopharm's Clinical Trials of Two Vaccine Candidates

Sinopharm Wuhan Institute developed their SARS-CoV-2 inactivated vaccine using the WIV04 strain isolated from a patient at the Jinyintan Hospital in Wuhan, China ([1201](#)). This vaccine is administered intramuscularly using 5 µg of virus per dose. Preclinical data providing supporting evidence for the use of this vaccine is not available publicly. Despite the lack of publicly available preclinical results, Sinopharm Wuhan Institute initiated phase I/II trials, which reported on varying dosing and prime-boost regimens.

A combined phase I/II RCT of Sinopharm's BBIBP-CorV, also known as Covilo, followed ([1203](#)). In phase I, 192 participants were randomized with varying doses of 2 µg, 4 µg, or 8 µg/dose or a placebo, and they received the same as a second dose 28 days later. Approximately 29% of participants reported at least 1 adverse event, most commonly fever, and neutralizing antibody titers were reported for all doses. In the phase II trial, 482 participants were enrolled (3:1, vaccine:placebo). Participants in the vaccine condition received either a single 8 µg dose or a double immunization of a 4 µg/dose that was administered 14, 21, or 28 days post the prime dose. Participants in the placebo condition received the placebo on one of the same four schedules. The vaccine appeared well-tolerated, with 23% reporting at least one adverse reaction characterized as mild to moderate. It was reported that all participants had a humoral immune response to the vaccines by day 42 but that the double immunization dosing regimen of 4 µg/dose achieved higher neutralizing antibody titers than a single dose of 8 µg and that the highest response was seen in the double-immunization regimen when at least 21 days separated the two doses ([1203](#)). Similar findings were reported in another phase I/II trial published by the same authors ([1205](#)). For the other vaccine, nAbs were detected in all groups 14 days after the final dose in the phase I part of the trial ([1204](#)), with similar findings reported in interim phase II data ([1204](#)).

7.3 Novavax's Nuvaxovid (NVX-CoV2373)

Novavax's Nuvaxovid is a particularly appealing candidate because the improved stability caused by the proline substitutions is particularly critical to facilitating global distribution, particularly to regions where local refrigerator/freezer capacities are limited. Importantly, these amino acid substitutions did not affect the ability of the spike protein to bind the hACE2 receptor (the target receptor of SARS-CoV-2 spike protein). The Novavax-CoV2373 vaccine candidate uses a proprietary, saponin-based Matrix-MTM adjuvant that contains two different 40nm-sized particles formed by formulating purified saponin with cholesterol and phospholipids ([1427](#)). In preclinical models, the use of the Matrix-M adjuvant potentiated the cellular and humoral immune responses to influenza vaccines ([1427–1430](#)). Importantly, Matrix-M adjuvant-containing vaccines have shown acceptable safety profiles in human clinical trials ([1431](#)).

Novavax-CoV2373 induced a multifunctional CD4/CD8 T-cell responses and generate high frequencies of follicular helper T-cells and B-cell germinal centers after vaccination. These findings were subsequently evaluated in a baboon primate model, in which Novavax-CoV2373 also elicited high antibody titers against the SARS-CoV-2 spike protein, as well as an antigen specific T-cell response. Based on this data Novavax initiated a phase I/II clinical trial to evaluate the safety and immunogenicity of Novavax-CoV2373 with Matrix-M ([1296](#), [1315](#)).

The phase I/II trial was a randomized, placebo-controlled study with 131 healthy adult participants in 5 treatment arms ([1296](#)). Participants that received the recombinant SARS-CoV-2 vaccine with or without the Matrix-M adjuvant got two injections, 21 days apart. Primary outcomes that were assessed include reactogenicity, lab-values (serum chemistry and hematology), and anti-spike IgG levels. Secondary outcomes measured included virus neutralization, T-cell responses, and unsolicited adverse events. The authors reported that no serious treatment-related adverse events occurred in any of the treatment arms. Reactogenicity was mostly absent and of short duration. The two-dose vaccine regimen induced anti-spike IgG levels and neutralizing antibody-titers exceeding those in the convalescent plasma of symptomatic patients. The outcomes of this trial suggest that Novavax-CoV2373 has an acceptable safety profile and is able to induce a strong immune response with high neutralizing antibody titers.

The phase II component of the trial was designed to identify the dose regimen for the next clinical trial stage ([1432](#)). Both younger (18-59 years) and older patients (60-84 years) were randomly assigned to receive either 5 µg or 25 µg Novavax-CoV2373 or a sodium-chloride placebo in two doses, 21 days apart. In line with the phase I data, reactogenicity remained mild to moderate, with no more than 1% of participants in any group reporting grade 3 AEs, and of short duration. Both dose levels were able to induce high anti-spike IgG titers as well as neutralizing antibody responses after the second dose. Based on both safety and efficacy data, the 5 µg dosing regimen was selected as the optimal dose regimen for the phase III trial.

Novavax announced an efficacy of 89.3% based on their phase III trial in the U.K. and South Africa ([1300](#), [1433](#), [1434](#)). This trial included over 15,000 participants in the U.K. and 4,000 participants in South Africa. The primary endpoint of the trial was the occurrence or absence of PCR-confirmed, symptomatic mild, moderate or severe COVID-19 from 7 days after the second dose onward. In the first interim analysis (U.K.), 56 cases of COVID-19 were observed in the placebo group compared to 6 cases in the treatment group. Importantly, the vaccine candidate also shows significant clinical efficacy against the prevalent U.K. and South African variants.

7.4 Protein Subunit Vaccine Development Programs Prior to SARS-CoV-2

Earlier studies examined the immunogenicity of a SARS-CoV-1 RBD fused with IgG1 Fc. This recombinant fusion protein could induce a robust long-lasting neutralizing antibody and cellular immune response that protected mice from SARS-CoV-1 ([1133](#), [1264](#), [1267](#)). While there have been other potential protein subunit vaccines for SARS-CoV-1 investigated *in vivo* ([1133](#), [1272](#)), none of these candidates have successfully completed clinical trials, more than likely due to the fact that the SARS-CoV-1 epidemic mostly ended by May 2004, and there was thus less of a demand or funding for SARS-CoV-1 vaccine research.

Similar vaccine candidates have emerged that target the RBD found in the S1 subunit of the trimeric MERS-CoV S protein, which binds to dipeptidyl-peptidase 4 (DPP4 also known as hCD26), the entry point through which MERS-CoV infects cells ([1435–1437](#)). After initially determining that an RBD subunit candidate (S377-588-Fc) could elicit neutralizing antibodies ([1438](#)), a study in mice determined that the administration of three sequential doses of RBD-Fc vaccine coupled with MF59, a squalene immunogenic adjuvant, induced humoral and systemic immunity in mice ([1439](#)). Mice that had been transduced with Ad5-hCD26 and subsequently challenged with MERS-CoV five days later did not show evidence of viral infection in the lungs versus control mice at ten days post vaccination ([1439](#)). Other variations of this vaccine approach include a stable S trimer vaccine whereby proline-substituted variants of S2 can maintain a stable prefusion conformation of the S2 domain ([1133](#)). This approach leads to broad and potent neutralizing antibodies ([1133](#)).

7.5 Complementary Approaches to Vaccine Development

A complementary approach to other vaccine development programs that is being investigated explores the potential for vaccines that are not made from the SARS-CoV-2 virus to confer what has been termed trained immunity. In a recent review ([1440](#)), trained immunity was defined as forms of memory that are temporary (e.g., months or years) and reversible. It is induced by exposure to whole-microorganism vaccines or other microbial stimuli that generates heterologous protective effects. Trained immunity can be displayed by innate immune cells or innate immune features of other cells, and it is characterized by alterations to immune responsiveness to future

immune challenges due to epigenetic and metabolic mechanisms. These alterations can take the form of either an increased or decreased response to immune challenge by a pathogen. Trained immunity elicited by non-SARS-CoV-2 whole-microorganism vaccines could potentially improve SARS-CoV-2 susceptibility or severity ([1441](#)).

One type of stimulus which research indicates can induce trained immunity is bacillus Calmette-Guerin (BCG) vaccination. BCG is an attenuated form of bacteria *Mycobacterium bovis*. The vaccine is most commonly administered for the prevention of tuberculosis in humans. Clinical trials in non-SARS-CoV-2-infected adults have been designed to assess whether BCG vaccination could have prophylactic effects against SARS-CoV-2 by reducing susceptibility, preventing infection, or reducing disease severity. A number of trials are now evaluating the effects of the BCG vaccine or the related vaccine VPM1002 ([1151](#), [1152](#), [1441–1453](#)).

The ongoing trials are using a number of different approaches. Some trials enroll healthcare workers, other trials hospitalized elderly adults without immunosuppression who get vaccinated with placebo or BCG at hospital discharge, and yet another set of trials older adults (>50 years) under chronic care for conditions like hypertension and diabetes. One set of trials, for example, uses time until first infection as the primary study endpoint; more generally, outcomes measured in some of these trials are related to incidence of disease and disease severity or symptoms. Some analyses have suggested a possible correlation at the country level between the frequency of BCG vaccination (or BCG vaccination policies) and the severity of COVID-19 ([1441](#)). Currently it is unclear whether this correlation has any connection to trained immunity. Many possible confounding factors are also likely to vary among countries, such as age distribution, detection efficiency, stochastic epidemic dynamic effects, differences in healthcare capacity over time in relation to epidemic dynamics, and these have not been adequately accounted for in current analyses. It is unclear whether there is an effect of the timing of BCG vaccination, both during an individual's life cycle and relative to the COVID-19 pandemic. Additionally, given that severe SARS-CoV-2 may be associated with a dysregulated immune response, it is unclear what alterations to the immune response would be most likely to be protective versus pathogenic (e.g., ([149](#), [1441](#), [1454](#), [1455](#))). The article ([1441](#)) proposes that trained immunity might lead to an earlier and stronger response, which could in turn reduce viremia and the risk of later, detrimental immunopathology. While trained immunity is an interesting possible avenue to complement vaccine development efforts through the use of an existing vaccine, additional research is required to assess whether the BCG vaccine is likely to confer trained immunity in the case of SARS-CoV-2.

7.6 India and China's Roles in Vaccine Innovation and Development

The nations of China and India have played a major role as COVID-19 vaccination developers and providers. Considering India produced approximately 60% of the world's vaccines prior to the pandemic, it is no surprise that the nation has developed and is developing several COVID-19 vaccine candidates. In addition to Covaxin, the Bio E subunit vaccine

CORBEVAX is being produced by Biological E in collaboration with U.S.-based Dynavax and the Baylor College of Medicine ([1456](#)). These two home-grown vaccines are now approved for adults and children as young as five (CORBEVAX) and six (Covaxin) ([1457](#)).

Other vaccines licensed by India were developed elsewhere but produced in India. For example, Novavax (developed in the United States) has signed an agreement with the Serum Institute of India allowing them to produce up to 2 billion doses a year ([1458](#)). Similarly, many people within India have been vaccinated with the AstraZeneca-University of Oxford vaccine, known as Covishield in India, which is also produced by the Serum Institute of India ([1456](#)). India is also developing vaccines using cutting-edge nucleic-acid-based platforms.. These include ZyCov-D, a DNA vaccine produced by Zydus Cadila, HGC019 and India's first mRNA vaccine, produced by Genova and HDT Biotech Corporation (of the U.S.) ([1456](#)). Additionally, in February 2021, Bharat Biotech received approval from Indian officials to commence a phase I study of an intranasal chimpanzee-adenovirus (ChAd) vectored SARS-CoV-2-S vaccine called BBV154 ([1459](#)).

In China, the Sinopharm-Beijing Institute vaccine, the Sinopharm-Wuhan Institute of Biological Products vaccine, the Sinovac Biotech (CoronaVac) vaccine, and CanSino Biologics vaccine are the main vaccines being distributed. Sinovac and Sinopharm aimed to produce 2 billion doses by the end of 2021, and they have distributed vaccines as aid to the Philippines and Pakistan ([1460](#)). In contrast, the Sinopharm-Wuhan vaccine, which has been approved for use in China since February 25, 2021, has been distributed almost exclusively within China, with limited supplies distributed to the United Arab Emirates ([1186](#)). On the same date, the CanSino vaccine was approved for use in China and has been granted emergency use in several other countries ([1186](#)).

However, the vaccine approval and distribution processes in China have come under increased scrutiny from other nations. China was criticized for administering vaccines to thousands of government officials and state-owned businesses in September 2020, prior to the completion of phase III clinical trials ([1461](#)). The behavior of Chinese officials has also come into question due to misinformation campaigns questioning the safety of Western vaccine candidates such as Moderna and Pfizer-BioNTech in a way that is intended to highlight the benefits of their own vaccine candidates ([1460](#)). China in particular took aim at mRNA technologies, but Chinese companies have since developed their own mRNA vaccines targeting the omicron variant, one of which is due to begin trials soon in the UAE ([1462](#)). Furthermore, delays in vaccine distribution have also caused issues, particularly in Turkey where 10 million doses of Sinovac were due to arrive by December 2020, but instead only 3 million were delivered in early January ([1460](#)). Similar delays and shortages of doses promised were reported by officials in the Philippines, Egypt, Morocco, and the United Arab Emirates ([1463](#), [1464](#)). All the same, Sinovac's vaccine has since been approved for use in countries around the world ([1186](#)).

8 The Coming of Age of Nucleic Acid Vaccines during COVID-19

8.1 Abstract

In the 21st century, several emergent viruses have posed a global threat. Each pathogen has emphasized the value of rapid and scalable vaccine development programs. The ongoing SARS-CoV-2 pandemic has made the importance of such efforts especially clear.

New biotechnological advances in vaccinology allow for recent advances that provide only the nucleic acid building blocks of an antigen, eliminating many safety concerns. During the COVID-19 pandemic, these DNA and RNA vaccines have facilitated the development and deployment of vaccines at an unprecedented pace. This success was attributable at least in part to broader shifts in scientific research relative to prior epidemics; the genome of SARS-CoV-2 was available as early as January 2020, facilitating global efforts in the development of DNA and RNA vaccines within two weeks of the international community becoming aware of the new viral threat. Additionally, these technologies that were previously only theoretical are not only safe but also highly efficacious.

Although historically a slow process, the rapid development of vaccines during the COVID-19 crisis reveals a major shift in vaccine technologies. Here, we provide historical context for the emergence of these paradigm-shifting vaccines. We describe several DNA and RNA vaccines and in terms of their efficacy, safety, and approval status. We also discuss patterns in worldwide distribution. The advances made since early 2020 provide an exceptional illustration of how rapidly vaccine development technology has advanced in the last two decades in particular and suggest a new era in vaccines against emerging pathogens.

8.2 Importance

The SARS-CoV-2 pandemic has caused untold damage globally, presenting unusual demands on but also unique opportunities for vaccine development. The development, production, and distribution of vaccines is imperative to saving lives, preventing severe illness, and reducing the economic and social burdens caused by the COVID-19 pandemic. Although vaccine technologies that provide the DNA or RNA sequence of an antigen had never previously been approved for use in humans, they have played a major role in the management of SARS-CoV-2. In this review we discuss the history of these vaccines and how they have been applied to SARS-CoV-2. Additionally, given that the evolution of new SARS-CoV-2 variants continues to present a significant challenge in 2022, these vaccines remain an important and evolving tool in the biomedical response to the pandemic.

8.3 Introduction

The SARS-CoV-2 virus emerged at the end of 2019 and soon spread around the world. In response, the Coalition for Epidemic Preparedness Innovations quickly began coordinating global health agencies and pharmaceutical companies to develop vaccines, as vaccination is one of the primary approaches available to combat the effects of a virus. Vaccines can bolster the immune response to a virus at both the individual and population levels, thereby reducing fatalities and severe illness and potentially driving a lower rate of infection even for a highly infectious virus like SARS-CoV-2. However, vaccines have historically required a lengthy development process due to both the experimental and regulatory demands.

As we review in a companion manuscript ([5](#)), vaccine technologies prior to the COVID-19 pandemic were largely based on triggering an immune response by introducing a virus or one of its components. Such vaccines are designed to induce an adaptive immune response without causing the associated viral illness. Each time a virus emerges that poses a significant global threat, as has happened several times over the past 20 years, the value of a rapid vaccine response is underscored. With progressive biotechnological developments, this objective has become increasingly tangible.

In the current century, significant advances in vaccine development have largely been built on genomics, as is somewhat unsurprising given the impact of the Genomic Revolution across all biology. This shift towards nucleic acid-based technologies opens a new frontier in vaccinology, where just the sequence encoding an antigen can be introduced to induce an immune response. While other platforms can carry some risks related to introducing all or part of a virus ([5](#)), nucleic acid-based platforms eliminate these risks entirely. Additionally, vaccine technologies that could be adjusted for novel viral threats are appealing because this modular approach would mean they could enter trials quickly in response to a new pathogen of concern.

8.4 Honing a 21st Century Response to Emergent Viral Threats

Recently, vaccine technologies have been developed and refined in response to several epidemics that did not reach the level of destruction caused by COVID-19. Emergent viral threats of the 21st century include severe acute respiratory syndrome (SARS), the H1N1 influenza strain known as swine flu, Middle East respiratory syndrome (MERS), Ebola virus disease, COVID-19, and, most recently, monkeypox, all of which have underscored the importance of a rapid global response to a new infectious virus. Because the vaccine development process has historically been slow, the use of vaccines to control most of these epidemics was limited.

One of the more successful recent vaccine development programs was for H1N1 influenza. This program benefited from the strong existing infrastructure for influenza vaccines along with the fact that regulatory agencies had determined that vaccines produced using egg- and cell-based platforms could be licensed under the regulations used for a strain change ([1423](#)). Although a monovalent H1N1 vaccine was not available before the pandemic peaked in the United States of America (U.S.A.) and Europe, it

became available soon afterward as a stand-alone vaccine that was eventually incorporated into commercially available seasonal influenza vaccines ([1423](#)). Critiques of the production and distribution of the H1N1 vaccine have stressed the need for alternative development-and-manufacturing platforms that can be readily adapted to new pathogens.

Efforts to develop such approaches had been undertaken prior to the COVID-19 pandemic. DNA vaccine development efforts began for SARS-CoV-1 but did not proceed past animal testing ([1422](#)). Likewise, the development of viral-vectored Ebola virus vaccines was undertaken, but the pace of vaccine development was behind the spread of the virus from early on ([1465](#)). Although a candidate Ebola vaccine V920 showed promise in preclinical and clinical testing, it did not receive breakthrough therapy designation until the summer of 2016, by which time the Ebola outbreak was winding down ([1466](#)). Therefore, the COVID-19 pandemic has been the first case where vaccines have been available early enough to significantly influence outcomes at the global scale.

The pandemic caused by SARS-CoV-2 has highlighted a confluence of circumstances that positioned vaccine development as a key player in efforts to control the virus and mitigate its damage. This virus did not follow the same trajectory as other emergent viruses of recent note, such as SARS-CoV-1, MERS-CoV, and Ebola virus, none of which presented a global threat for such a sustained duration (see visualization in [3](#)). Spread of the SARS-CoV-2 virus has remained out of control in many parts of the world into 2022, especially with the emergence of novel variants exhibiting increased rates of transmission ([1](#)). While, for a variety of reasons, SARS-CoV-2 was not controlled as rapidly as the viruses underlying prior 21st century epidemics, vaccine development technology had also progressed based on these and other prior viral threats to the point that a rapid international vaccine development response was possible.

8.5 Development of COVID-19 Vaccines using DNA and RNA Platforms

Vaccine development programs for COVID-19 emerged very quickly. The first administration of a dose of a COVID-19 vaccine to a human trial participant occurred on March 16, 2020 ([1467](#), [1468](#)), marking an extremely rapid response to the emergence of SARS-CoV-2. Within a few weeks of this first trial launching, at least 78 vaccine development programs were active ([1468](#)), and by September 2020, there were over 180 vaccine candidates against SARS-CoV-2 in development ([1114](#)). As of May 3, 2023, 50 SARS-CoV-2 vaccines have been approved world wide and 28 are being administered throughout the world, with 13.0 billion doses administered across 223 countries. The first critical step towards developing a vaccine against SARS-CoV-2 was characterizing the viral target, which happened extremely early in the COVID-19 outbreak with the sequencing and dissemination of the viral genome in early January 2020 ([555](#)) (Figure [10](#)). This genomic information allowed for an early identification of the sequence of the Spike (S) protein (Figure [10](#)), which is the antigen and induces an immune response ([1469](#), [1470](#)).

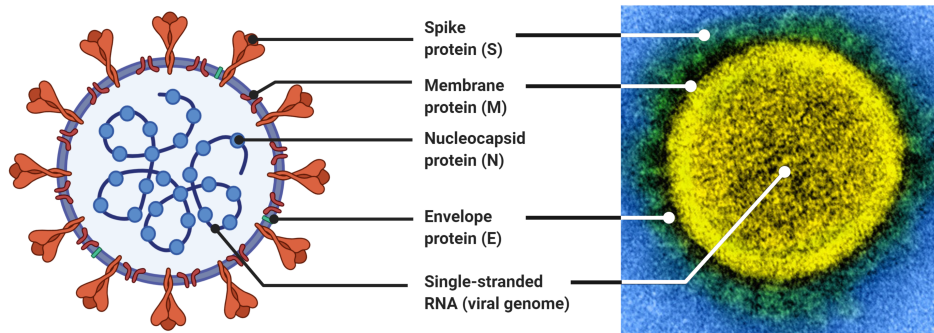


Figure 10: Structure of the SARS-CoV-2 virus. The development of vaccines depends on the immune system recognizing the virus. Here, the structure of SARS-CoV-2 is represented both in the abstract and against a visualization of the virion. The abstracted visualization was made using BioRender ([1471](#)) using the template “Human Coronavirus Structure” by BioRender (August 2020) ([1472](#)). The microscopy was conducted by the National Institute of Allergy and Infectious Diseases ([1473](#)).

During the development process, one measure used to assess whether a vaccine candidate is likely to provide protection is serum neutralizing activity ([1119](#)). This assay evaluates the presence of antibodies that can neutralize, or prevent infection by, the virus in question. Often, titration is used to determine the extent of neutralization activity. However, unlike in efforts to develop vaccines for prior viral threats, the duration of the COVID-19 pandemic has made it possible to also test vaccines in phase III trials where the effect of the vaccines on a cohort’s likelihood of contracting SARS-CoV-2 was evaluated.

8.6 Theory and Implementation of Nucleic Acid Vaccines

Biomedical research in the 21st century has been significantly influenced by the genomic revolution. While traditional methods of vaccine development, such as inactivated whole viruses are still used today ([5](#)), vaccine development is no exception. The shift towards omics-based approaches to vaccine development began to take hold with the meningococcal type B vaccine, which was developed using reverse vaccinology in the early 2010s ([1474](#), [1475](#)). Under this approach, the genome of a pathogen is screened to identify potential vaccine targets ([1475](#)), and pathogens of interest are then expressed *in vitro* and tested in animal models to determine their immunogenicity ([1475](#)). In this way, the genomic revolution catalyzed a fundamental shift in the development of vaccines. Such technologies could revolutionize the role of vaccines given their potential to address one of the major limitations of vaccines today and facilitate the design of therapeutic, rather than just prophylactic, vaccines ([1476](#)).

Nucleic-acid based approaches share an underlying principle: a vector that delivers the information needed to produce an antigen. When the host cells manufacture the antigen, it can then trigger an immune response. The fact that no part of the virus is introduced aside from the genetic code of the antigen means that these vaccines carry no risk of infection. Such approaches build on subunit vaccination strategies, where a component of a virus (e.g., an antigenic protein) is delivered by the vaccine. Platforms based on genomic sequencing began to be explored beginning in the 1980s as

genetic research became increasingly feasible. Advances in genetic engineering allowed for gene sequences of specific viral antigens to be grown *in vitro* (1113). Studies also demonstrated that model organisms could be induced to construct antigens that would trigger an immune response (1128, 1477, 1478). These two developments sparked interest in whether it could be possible to identify any or all of the antigens encoded by a virus's genome and train the immune response to recognize them.

The delivery and presentation of antigens is fundamental to inducing immunity against a virus. Vaccines that deliver nucleic acids allow the introduction of foreign substances to the body to induce both humoral and cellular immune responses (1479). Delivering a nucleic acid sequence to host cells allows the host to synthesize an antigen without exposure to a viral threat (1479). Host-synthesized antigens can activate both humoral and cellular immunity (1479), as they can be presented in complex with major histocompatibility complex (MHC) I and II, which can activate either T- or B-cells (1479). In contrast, prior approaches activated only MHC II (1478). Because these vaccines encode specific proteins, providing many of the benefits of a protein subunit vaccine, they do not carry any risk of DNA being live, replicating, or spreading, and their manufacturing process lends itself to scalability (1479). Here, opportunities can be framed in terms of the central dogma of genetics: instead of directly providing the proteins from the infectious agents, vaccines developers are exploring the potential for the delivery of DNA or RNA to induce the cell to produce proteins from the virus that in turn induce a host immune response.

8.7 Cross-Platform Considerations in Vaccine Development

Certain design decisions are relevant to vaccine development across multiple platforms. One applies to the platforms that deliver the antigen, which in the case of SARS-CoV-2 vaccines is the S protein. The prefusion conformation of the S protein, which is the structure before the virus fuses to the host cell membrane, is metastable (1480), and the release of energy during membrane fusion drives this process forward following destabilization (12, 1481). Due to the significant conformational changes that occur during membrane fusion (25, 1482, 1483), S protein immunogens that are stabilized in the prefusion conformation are of particular interest, especially because a prefusion stabilized *Middle East respiratory syndrome-related coronavirus* (MERS-CoV) S antigen was found to elicit an improved antibody response (1278). Moreover, the prefusion conformation offers an opportunity to target S2, a region of the S protein that accumulates mutations at a slower rate (23, 24, 1278) (see also (1)). Vaccine developers can stabilize the prefusion conformer by selecting versions of the S protein containing mutations that lock the position (1484). The immune response to the Spike protein when it is stabilized in this conformation is improved over other S structures (1485). Thus, vaccines that use this prefusion stabilized conformation are expected to not only offer improved immunogenicity, but also be more resilient to the accumulation of mutations in SARS-CoV-2.

Another cross-platform consideration is the use of adjuvants. Adjuvants include a variety of molecules or larger microbial-related products that affect the immune system broadly or an immune response of interest. They can either be comprised of or contain immunostimulants or immunomodulators. Adjuvants are sometimes included within vaccines in order to enhance the immune response. Different adjuvants can regulate different types of immune responses, so the type or combination of adjuvants used in a vaccine will depend on both the type of vaccine and concern related to efficacy and safety. A variety of possible mechanisms for adjuvants have been investigated ([1486–1488](#)).

Due to viral evolution, vaccine developers are in an arms race with a pathogen that benefits from mutations that reduce its susceptibility to adaptive immunity. The evolution of several variants of concern (VOC) presents significant challenges for vaccines developed based on the index strain identified in Wuhan in late 2019. We discuss these variants in depth elsewhere in the COVID-19 Review Consortium project ([1148](#)). To date, the most significant variants of concern identified are Alpha (2020), Beta (2020), Gamma (2020), Delta (2021), Omicron (2021), and related Omicron subvariants (2022). The effectiveness or efficacy (i.e., trial or real-world prevention, respectively) of vaccines in the context of these variants is discussed where information is available.

8.8 DNA Vaccine Platforms

DNA vaccine technologies have developed slowly over the past thirty years. These vaccines introduce a vector containing a DNA sequence that encodes antigen(s) selected to induce a specific immune response ([1478](#)). Early attempts revealed issues with low immunogenicity ([1128](#), [1478](#), [1489](#)). Additionally, initial skepticism about the approach suggested that DNA vaccines might bind to the host genome or induce autoimmune disease ([1479](#), [1490](#)), but pre-clinical and clinical studies have consistently disproved this hypothesis and indicated DNA vaccines to be safe ([1489](#)). Another concern, antibiotic resistance introduced during the plasmid selection process, did remain a concern during this initial phase of development ([1479](#)), but this issue was resolved through strategic vector design ([1491](#), [1492](#)). However, for many years, the immunogenicity of DNA vaccines failed to reach expectations ([1479](#)). Several developments during the 2010s led to greater efficacy of DNA vaccines ([1479](#)). However, no DNA vaccines had been approved for use in humans prior to the COVID-19 pandemic ([1489](#), [1493](#)). As of May 3, 2023, 10 vaccines have been approved worldwide (Table [4](#)). These vaccines fall into two categories, vaccines that are vectored with a plasmid and those that are vectored with another virus.

Table 4: DNA vaccines approved in at least one country ([1169](#)) as of May 3, 2023.

Vaccine	Company	Platform
iNCOVACC	Bharat Biotech	non replicating viral vector
Convitecia	CanSino	non replicating viral vector

Vaccine	Company	Platform
Convidecia Air	CanSino	non replicating viral vector
Gam-COVID-Vac	Gamaleya	non replicating viral vector
Sputnik Light	Gamaleya	non replicating viral vector
Sputnik V	Gamaleya	non replicating viral vector
Jcovden	Janssen (Johnson & Johnson)	non replicating viral vector
Vaxzevria	Oxford/AstraZeneca	non replicating viral vector
Covishield (Oxford/ AstraZeneca formulation)	Serum Institute of India	non replicating viral vector
ZyCoV-D	Zydus Cadila	plasmid vectored

8.8.1 Plasmid-Vectored DNA Vaccines

Many DNA vaccines use a plasmid vector-based approach, where the sequence encoding the antigen(s) against which an immune response is sought are cultivated in a plasmid and delivered directly to an appropriate tissue ([1494](#)). Plasmids can also be designed to act as adjuvants by targeting essential regulators of pathways such as the inflammasome or simply just specific cytokines ([1490](#), [1495](#)). The DNA itself may also stimulate the innate immune response ([1478](#), [1492](#)). Once the plasmid brings the DNA sequence to an antigen-presenting cell (APC), the host machinery can be used to construct antigen(s) from the transported genetic material, and the body can then synthesize antibodies in response ([1479](#)). The vectors are edited to remove extra sequences ([1492](#)). These types of manufacturing advances have improved the safety and throughput of this platform ([1492](#)).

8.8.1.1 Prior Applications

In the 1990s and 2000s, DNA vaccines delivered via plasmids sparked significant scientific interest, leading to a large number of preclinical trials ([1479](#)). Early preclinical trials primarily focused on long-standing disease threats, including viral diseases such as rabies and parasitic diseases such as malaria, and promising results led to phase I testing of the application of this technology to human immunodeficiency virus (HIV), influenza, malaria, and other diseases of concern during this period ([1479](#)). Although they were well-tolerated, these early attempts to develop vaccines were generally not very successful in inducing immunity to the target pathogen, with either limited T-cell or limited neutralizing antibody responses observed ([1479](#)).

Early plasmid-vectored DNA vaccine trials targeted HIV and subsequently diseases of worldwide importance such as malaria and hepatitis B ([1496](#)). The concern with these early development projects was immunogenicity, not safety ([1496](#)). Around the turn of the millennium, a hepatitis B vaccine development program demonstrated that these vaccines can induce both

antibody and cellular immune response ([1497](#)). Prior to COVID-19, however, plasmid-vectored DNA vaccines had been approved for commercial use only in veterinary populations ([1498–1500](#)). Between 2005 and 2006, several DNA vaccines were developed for non-human animal populations, including against viruses including a rhabdovirus in fish ([1501](#)), porcine reproductive and respiratory syndrome virus ([1502](#)), and West Nile virus in horses ([1503](#)). Within the past five years, additional plasmid-vectored vaccines for immunization against viruses were developed against a herpesvirus (in mice) ([1504](#)) and an alphavirus (in fish) ([1505](#)).

8.8.1.2 Applications to COVID-19

Several plasmid-vectored DNA vaccines have been developed against COVID-19 (Table [4](#)). In fact, the ZyCoV-D vaccines developed by India's Zydus Cadila is the first plasmid-vectored DNA vaccine to receive approval or to be used in human medicine ([1506–1508](#)). Another plasmid-vectored DNA vaccine, INO-4800 ([1509](#)), was developed by Inovio Pharmaceuticals Technology that uses electroporation as an adjuvant. Electroporation was developed as a solution to the issue of limited immunogenicity by increasing the permeability of cell membranes by delivering electrical pulses ([1510](#)). It has been shown that electroporation can enhance vaccine efficacy by up to 100-fold, as measured by increases in antigen-specific antibody titers ([1511](#)). The temporary formation of pores through electroporation facilitates the successful transportation of macromolecules into cells, allowing cells to robustly take up INO-4800 for the production of an antibody response. For INO-4800, a plasmid-vectored vaccine is delivered through intradermal injection which is then followed by electroporation with a device known as CELLECTRA® ([1512](#)). The safety of the CELLECTRA® device has been studied for over seven years, and these studies support the further development of electroporation as a safe vaccine delivery method ([1510](#)).

These vaccines therefore represent implementations of a new platform technology. In particular, they offer the advantage of a temperature-stable vaccine, facilitating worldwide administration ([1513](#)). Although an exciting development in DNA vaccines, the cost of vaccine manufacturing and electroporation may make scaling the use of this technology for prophylactic use for the general public difficult.

8.8.1.3 Trial Safety and Immunogenicity

The INO-4800 trials began with a phase I trial evaluating two different doses administered as a two-dose series ([1512](#)). This trial found the vaccine to be safe, with only six adverse events (AEs) reported by 39 participants, all grade 1, and effective, with all but three participants of 38 developing serum IgG binding titers to the SARS-CoV-2 S protein ([1512](#)). In the phase II trial of 401 adults at high risk of exposure to SARS-CoV-2 similarly supported the safety and efficacy of INO-4800. Only one treatment-related AE was observed and the vaccine was found to be associated with a significant increase in neutralizing activity ([1513](#)). Results of phase III trials are not yet available ([1514–1517](#)).

Trials of ZyCoV-D have progressed further. This vaccine uses a plasmid to deliver the expression-competent Spike protein and IgE signal peptides to the vaccinee ([1518](#)). During the phase I trial, vaccination with a needle versus a needle-free injection system was evaluated, and the vaccine can now be administered without a needle ([1506](#), [1507](#)). A phase III trial enrolling over 27,000 patients found no difference in AEs between the placebo and treatment groups and estimated the efficacy of ZyCoV-D to be 66.6% ([1519](#)). It was authorized for people ages 12 and older ([1508](#)). The highly portable design offers advantages over traditional vaccines ([1518](#)), especially as the emergence of variants continues to challenge the effectiveness of vaccines. As of August 2022, ZyCoV-D has only been approved in India ([1520](#)) and is not tracked by Our World in Data ([1116](#)).

8.8.1.4 Real-World Safety and Effectiveness

In terms of the ability of plasmid-vectored vaccines to neutralize VOC, varying information is available. The situation for ZyCoV-D is somewhat different, as their phase III trial occurred during the Delta wave in India ([1519](#)). At present, no major press releases have addressed the vaccine's ability to neutralize Omicron and related VOC, but reporting suggests that the manufacturers were optimistic about the vaccine in light of the Omicron variant as of late 2021 ([1521](#)).

As for INO-4800, studies have examined whether the induced immune response can neutralize existing VOC. They assessed neutralization of several VOC relative to the index strain and found no difference in neutralization between the index strain and the Gamma VOC (P.1) ([1522](#)). However, neutralization of the Alpha and Beta VOC was significantly lower (approximately two and seven times, respectively) ([1522](#)). These findings are in line with the shifts in effectiveness reported for other vaccines ([5](#)). In addition to loss of neutralizing activity due to viral evolution, studies have also evaluated the decline in neutralizing antibodies (nAbs) induced by INO-4800 over time. Levels of nAbs remained statistically significant relative to the pre-vaccination baseline for six months ([1523](#)). Administration of a booster dose induced a significant increase of titers relative to their pre-booster levels ([1523](#)). Given the timing of this trial (enrollment between April and July 2020), it is unlikely that participants were exposed to VOC associated with decreased efficacy.

In light of the emergence of VOC against which many vaccines show lower effectiveness, Inovio Pharmaceuticals began to develop a new vaccine with the goal of improving robustness against known and future VOC ([1524](#)). Known as INO-4802, this vaccine was designed to express a pan-Spike immunogen ([1525](#)). Booster studies in rodents ([1526](#)) and non-human primates ([1525](#)) suggest that it may be more effective than INO-4800 in providing immunity to VOC such as Delta and Omicron when administered as part of a heterologous boost regimen, although boosting with INO-4800 was also very effective in increasing immunity in rhesus macaques ([1525](#)). Therefore, boosting is likely to be an important strategy for this vaccine, especially as the virus continues to evolve.

8.8.2 Viral-Vectored DNA Vaccines

Plasmids are not the only vector that can be used to deliver sequences associated with viral antigens. Genetic material from the target virus can also be delivered using a second virus as a vector. Viral vectors have emerged as a safe and efficient method to furnish the nucleotide sequences of an antigen to the immune system ([1527](#)). The genetic content of the vector virus is often altered to prevent it from replicating, but replication-competent viruses can also be used under certain circumstances ([1528](#)). Once the plasmid or viral vector brings the DNA sequence to an APC, the host machinery can be used to construct antigen(s) from the transported genetic material, and the host can then synthesize antibodies in response ([1479](#)).

One of the early viral vectors explored was adenovirus, with serotype 5 (Ad5) being particularly effective ([1479](#)). This technology rose in popularity during the 2000s due to its being more immunogenic in humans and non-human primates than plasmid-vectored DNA vaccines ([1479](#)). In the 2000s, interest also arose in utilizing simian adenoviruses as vectors because of the reduced risk that human vaccine recipients would have prior exposure resulting in adaptive immunity ([1479](#), [1529](#)), and chimpanzee adenoviruses were explored as a potential vector in the development of a vaccine against MERS-CoV ([1530](#)).

Today, various viral-vector platforms including poxviruses ([1531](#), [1532](#)), adenoviruses ([1533](#)), and vesicular stomatitis viruses ([1534](#), [1535](#)) are being developed. Viral-vector vaccines are able to induce both an antibody and cellular response; however, the response is limited due to the immunogenicity of the viral vector used ([1533](#), [1536](#)). An important consideration in identifying potential vectors is the immune response to the vector. Both the innate and adaptive immune responses can potentially respond to the vector, limiting the ability of the vaccine to transfer information to the immune system ([1537](#)). Different vectors are associated with different levels of reactogenicity; for example, adenoviruses elicit a much stronger innate immune response than replication deficient adeno-associated viruses derived from parvoviruses ([1537](#)). Additionally, using a virus circulating widely in human populations as a vector presents additional challenges because vaccine recipients may already have developed an immune response to the vector ([1538](#)). Furthermore, repeated exposure to adenoviruses via viral-vectored DNA vaccines may increase reactivity to these vectors over time, presenting a challenge that will need to be considered in long-term development of these vaccines ([1539](#), [1540](#)).

8.8.2.1 Prior Applications

There are several viral vector vaccines that are available for veterinary use ([1479](#), [1541](#)), but prior to the COVID-19 pandemic, only one viral vector vaccine was approved by the United States' Food and Drug Administration (FDA) for use in humans. This vaccine is vectored with a recombinant vesicular stomatitis virus and targeted against the Ebola virus ([1542](#)). Additionally, several phase I and phase II clinical trials for other vaccines are ongoing ([1527](#)), and the technology is currently being explored for its potential against numerous infectious diseases including malaria ([1543](#), [1544](#)), Ebola ([1545–1547](#)), and HIV ([1548](#), [1549](#)).

The threat of MERS and SARS initiated interest in the application of viral vector vaccines to human coronaviruses ([1530](#)), but efforts to apply this technology to these pathogens had not yet led to a successful vaccine candidate. In the mid-to-late 2000s, adenoviral vectored vaccines against SARS were found to induce SARS-CoV-specific IgA in the lungs of mice ([1550](#)) but were later found to offer incomplete protection in ferret models ([1551](#)). Gamaleya National Center of Epidemiology and Microbiology in Moscow sought to use an adenovirus platform for the development of vaccines for MERS-CoV and Ebola virus, although neither of the previous vaccines were internationally licensed ([1552](#)).

In 2017, results were published from an initial investigation of two vaccine candidates against MERS-CoV containing the MERS-CoV *S* gene vectored with chimpanzee adenovirus, Oxford University #1 (ChAdOx1), a replication-deficient chimpanzee adenovirus ([1553](#)). This study reported that a candidate containing the complete *S* protein sequence induced a stronger neutralizing antibody response in mice than candidates vectored with modified vaccinia virus Ankara.

The candidate was pursued in additional research, and in the summer of 2020 results of two studies were published. The first reported that a single dose of ChAdOx1 MERS induced an immune response and inhibited viral replication in macaques ([1554](#)). The second reported promising results from a phase I trial that administered the vaccine to adults and measured safety, tolerability, and immune response ([1555](#)).

8.8.2.2 Application to COVID-19

Number of Non Replicating Viral Vector Vaccines Available Worldwide

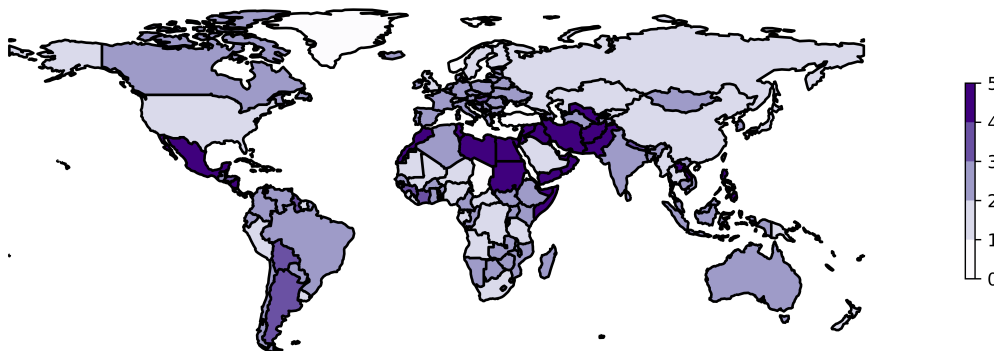


Figure 11: Worldwide availability of vaccines developed using non-replicating viral vectors. This figure reflects the number of vaccines using non-replicating viral vectors that were available in each country as of May 31, 2023. These data are retrieved from Our World in Data ([1116](#)) and plotted using geopandas ([1170](#)). See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily. Note that this figure draws from a different data source than Table [4](#) and does not necessarily include data for every vaccine developed within this category.

While not all of the above results were available at the time that vaccine development programs against SARS-CoV-2 began, at least three viral vector vaccines have also been developed against SARS-CoV-2 (Figure [11](#)). First, a collaboration between AstraZeneca and researchers at the University of Oxford successfully applied a viral vector approach to the development of a vaccine against SARS-CoV-2 using the replication-deficient ChAdOx1 vector

modified to encode the S protein of SARS-CoV-2 ([1556](#)). In a phase I trial, the immunogenic potential of vaccine candidate ChAdOx1 nCoV-19 was demonstrated through the immune challenge of two animal models, mice and rhesus macaques ([1556](#)). In a phase I/II trial, patients receiving the ChAdOx1 nCoV-19 vaccine developed antibodies to the SARS-CoV-2 Spike protein that peaked by day 28, with these levels remaining stable until a second observation at day 56 ([1557](#)).

Second, a viral vector approach was applied by Russia's Gamaleya Research Institute of Epidemiology and Microbiology to develop Sputnik V, a replication-deficient recombinant adenovirus (rAd) vaccine that combines two adenovirus vectors, rAd26-S and rAd5-S, that express the full-length SARS-CoV-2 Spike glycoprotein. These vectors are intramuscularly administered individually using two separate vaccines in a prime-boost regimen. The rAd26-S is administered first, followed by rAd5-S 21 days later. Both vaccines deliver 10^{11} viral particles per dose. This approach is designed to overcome any potential pre-existing immunity to adenovirus in the population ([1558](#)), as some individuals may possess immunity to Ad5 ([1559](#)). Sputnik V is the only recombinant adenovirus vaccine to utilize two vectors.

Third, Janssen Pharmaceuticals, Inc., a subsidiary of Johnson & Johnson, developed a viral vector vaccine in collaboration with and funded by the United States' "Operation Warp Speed" ([1560](#), [1561](#)). The vaccine candidate JNJ-78436735, formerly known as Ad26.COV2-S, is a monovalent vaccine that is composed of a replication-deficient adenovirus serotype 26 (Ad26) vector expressing the stabilized prefusion S protein of SARS-CoV-2 ([1485](#), [1562](#)). Unlike the other two viral vector vaccines available to date, JNJ-78436735 requires only a single dose, a characteristic that was expected to aid in global deployment ([1563](#)). JNJ-78436735 was selected from among a number of initial candidate designs ([1485](#)) and tested *in vivo* in Syrian golden hamsters and Rhesus macaques to assess safety and immunogenicity ([1485](#), [1563](#)–[1565](#)). The JNJ-78436735 candidate was selected for its favorable immunogenicity profile and ease of manufacturability ([1485](#), [1563](#)–[1565](#)) and was found to confer protection against SARS-CoV-2 in macaques even after six months ([1566](#)). The one- versus two-dose regimen was then tested in volunteers through a phase I/IIa trial ([1562](#), [1567](#)). A major difference between this vaccine and the other two in this category is that the S protein immunogen is stabilized in its prefusion conformation, while in the Sputnik V and AstraZeneca vaccines it is not.

As of May 31, 2023, data describing the distribution of 5 viral-vectored vaccines in 203 countries are available (Figure [11](#)). ChAdOx1 nCoV-19 was first approved for emergency use on December 30, 2020 in the U.K. ([1568](#)). Sputnik V was available soon after, and early as January 2021, Sputnik V had been administered to 1.5 million Russians ([1569](#)) and began distributing doses to other countries within Europe such as Belarus, Bosnia-Herzegovina, Hungary, San Marino, Serbia, and Slovakia ([1570](#)–[1572](#)).

8.8.2.3 Trial Estimates of Safety and Efficacy

The first DNA viral-vectored vaccine for which efficacy estimates became available was AstraZeneca's ChAdOx1 nCoV-19. In December 2020, preliminary results of the phase III trial were released detailing randomized

control trials conducted in the United Kingdom (U.K.), Brazil, and South Africa between April and November 2020 ([1469](#)). These trials compared ChAdOx1 nCoV-19 to a control, but the design of each study varied; pooling data across studies indicated an overall efficacy of 70.4%. For Sputnik V, the phase III trial indicated an overall vaccine efficacy of 91.6% for symptomatic COVID-19 ([1573](#)). As for Janssen, the vaccine was well-tolerated, and across all regions studied, it was found to be 66.9% effective after 28 days for the prevention of moderate to severe COVID-19 and to be 81.7% effective for the prevention of laboratory-confirmed severe COVID-19 ([1574](#)). There were no COVID-19-associated deaths in the vaccine group. However, the emergence of the Beta variant in the South African trial population was associated with a slightly reduced efficacy (64% two weeks after receipt), and all of the COVID-19-associated deaths in the trial occurred in the South African placebo cohort ([1574](#)). In February 2021, the FDA issued an EUA for the Janssen vaccine based on interim results from the phase III trial ([1575](#), [1576](#)).

Two of the three vaccines have faced a number of criticisms surrounding the implementation of their clinical trials. In the race to develop vaccines against SARS-CoV-2, President Vladimir Putin of Russia announced the approval of the Sputnik V vaccines on August 11, 2020 in the absence of clinical evidence ([1577](#)). A press release on November 11, 2020 indicated positive results from an interim analysis of the phase III Sputnik V trials, which reported 92% efficacy in 16,000 volunteers ([1578](#)). However, this release came only two days after both Pfizer and BioNTech reported that their vaccines had an efficacy over 90%, which led to significant skepticism of the Russian findings for a myriad of reasons including the lack of a published protocol and the “reckless” approval of the vaccine in Russia months prior to the publication of the interim results of the phase III trial ([1578](#), [1579](#)). Consequently, many international scientific agencies and public health bodies expressed concern that due diligence to the clinical trial process was subverted for the sake of expediency, leading many to question the safety and efficacy of Sputnik V ([1577](#), [1580](#), [1581](#)). Despite regulatory, safety, and efficacy concerns, pre-orders for 1 billion doses of the Sputnik V were reported within days of the vaccine’s approval in Russia ([1577](#)). Almost a month later, the phase I/II trial data was published ([1582](#)). It wasn’t until February 2021, six months after its approval in Russia, that interim results of the phase III trial were released ([1573](#)). This publication reported a VE of 91% and a low rate of serious AEs, although there were several serious AEs that were determined not to be associated with the vaccine by an independent data monitoring committee about which little other information was released ([1583](#)).

AstraZeneca’s clinical trial also faced criticism. The trial was paused in September 2020 following a severe adverse event in one participant ([1584](#)). It was restarted soon after ([1585](#)), but it seems that the recent pause was not mentioned to the FDA during a call the morning before the story broke ([1586](#)). Additionally, individual sites within the trial employed somewhat different designs but were combined for analysis. For example, in South Africa, the trial was double-blinded, whereas in the U.K. and Brazil it was single-blinded, and one of the two trials carried out in the U.K. evaluated two dosing regimens (low dose or standard dose, both followed by standard dose). Some of the trials used a meningococcal conjugate vaccine (MenACWY) as a control, while others used saline. Data was pooled across countries for analysis, a design decision that was approved by regulators but raised some

questions when higher efficacy was reported in a subgroup of patients who received a low-dose followed by a standard dose. This group came about because some participants in the U.K. were erroneously primed with a much lower dose, which turned out to have higher efficacy than the intended dose (1587). Combining the data then led to confusion surrounding the VE, as VE varied widely among conditions (e.g., 62% VE in the standard dose group vs 90% in the group that received a low prime dose (1469)). Subsequent research, however, suggests that reducing the prime dose may, in fact, elicit a superior immune response in the long-term despite a lower initial response (1588). Therefore, this error may serendipitously improve efficacy of vaccine-vectored vaccines broadly.

8.8.2.4 Real-World Safety and Efficacy

Following the trials, additional concerns have been raised about some of these vaccines. Within a few days to a few weeks following their first dose of the AstraZeneca vaccine, three women developed extensive venous sinus thrombosis (1589). In March 2021, administration of the vaccine was paused in several European countries while a possible link to thrombotic events was investigated (1590), as these adverse events had not been observed in clinical trials, but the European Medicine Agency (EMA) soon determined that 25 events were not related to the vaccine (1591). The following month, the United States paused administration of the Janssen vaccine for ten days due to 15 similar AEs (1592, 1593), but the EMA, U.S. Centers for Disease Control, and the FDA's Advisory Committee on Immunization Practices again identified the events as being very rare and the benefits of the vaccine as likely to outweigh its risks (1594–1597). In Denmark and Norway, population-based estimates suggested AstraZeneca's vaccine increased incidence of venous thromboembolic events by 11 cases over baseline per 100,000 doses (1598). Estimates of the incidence in other western countries have also been low (1599). In the US, thromboembolic events following the Janssen vaccine have also been very rare (1595). Subsequently, a potential mechanism was identified: the adenovirus vector binding to platelet factor 4 (1600, 1601). Because this adverse event is so rare, the risk is likely still outweighed by the risks associated with contracting COVID-19 (1602), which is also associated with thrombotic events) (1593, 1603). Similarly, concerns about Guillain-Barré syndrome arose in connection to the Janssen vaccine, but these events have similarly been determined to be very rare and the benefits to outweigh the risks (1597).

Given that vaccines from multiple platforms are now widely available, people at increased risk of a specific severe AE may have options to pursue vaccination with a platform that does not carry such risks. For example, a woman in the U.S. with a history of thromboembolic concerns might feel more comfortable with an mRNA vaccine (described below), where such AEs have not been identified in association with COVID-19 vaccination. However, within the U.S.A., no clear framework has been established for advising patients on whether a specific vaccine may be preferable for their individual concerns now that vaccines based on three different technologies are widely available (see (5) for information about Novavax, which is a protein subunit vaccine).

8.9 mRNA Vaccines

Building on DNA vaccine technology, RNA vaccines are an even more recent advancement for vaccine development. Interest in messenger RNA (mRNA) vaccines emerged around 1990 following *in vitro* and animal model studies that demonstrated that mRNA could be transferred into cells ([1604](#), [1605](#)). mRNA contains the minimum information needed to create a protein ([1605](#)). RNA vaccines are therefore nucleic-acid based modalities that code for viral antigens against which the human body elicits a humoral and cellular immune response.

The strategy behind mRNA vaccines operates one level above the DNA: instead of directly furnishing the gene sequence associated with an antigen to the host, it provides the mRNA transcribed from the DNA sequence. The mRNA is transcribed *in vitro* and delivered to cells via lipid nanoparticles (LNP) ([1606](#)). It is recognized by ribosomes *in vivo* and then translated and modified into functional proteins ([1127](#)). The resulting intracellular viral proteins are displayed on surface MHC proteins, provoking a strong CD8⁺ T cell response as well as a CD4⁺ T cell and B cell-associated antibody responses ([1127](#)). mRNA is naturally not very stable and can degrade quickly in the extracellular environment or the cytoplasm. The LNP covering protects the mRNA from enzymatic degradation outside of the cell ([1607](#)). Codon optimization to prevent secondary structure formation and modifications of the poly-A tail as well as the 5' untranslated region to promote ribosomal complex binding can increase mRNA expression in cells. Furthermore, purifying out double-stranded RNA and immature RNA with fast performance liquid chromatography and high performance liquid chromatography technology will improve translation of the mRNA in the cell ([1127](#), [1608](#)).

There are three types of RNA vaccines: non-replicating, *in vivo* self-replicating, and *in vitro* dendritic cell non-replicating ([1609](#)). Non-replicating mRNA vaccines consist of a simple open reading frame for the viral antigen flanked by the 5' UTR and 3' poly-A tail. *In vivo* self-replicating vaccines encode a modified viral genome derived from single-stranded, positive sense RNA alphaviruses ([1127](#), [1608](#)). The RNA genome encodes the viral antigen along with proteins of the genome replication machinery, including an RNA polymerase. Structural proteins required for viral assembly are not included in the engineered genome ([1127](#)). Self-replicating vaccines produce more viral antigens over a longer period of time, thereby evoking a more robust immune response ([1609](#)). Finally, *in vitro* dendritic cell non-replicating RNA vaccines limit transfection to dendritic cells. Dendritic cells are potent antigen-presenting immune cells that easily take up mRNA and present fragments of the translated peptide on their MHC proteins, which can then interact with T cell receptors. Ultimately, primed T follicular helper cells can stimulate germinal center B cells that also present the viral antigen to produce antibodies against the virus ([1610](#)). These cells are isolated from the patient, then grown and transfected *ex vivo* ([1611](#)). They can then be reintroduced to the patient ([1611](#)).

In addition to the benefits of nucleic acid vaccines broadly, mRNA confers specific advantages compared to DNA vaccines and other platforms ([1612](#)). Some of these advantages fall within the domain of safety. Unlike DNA vaccines, mRNA technologies are naturally degradable and non-integrating, and they do not need to cross the nuclear membrane in addition to the

plasma membrane for their effects to be seen (1127). Additionally, the half life can be regulated by the contents of the 5' and 3' untranslated regions (1613). In comparison to vaccines that use live attenuated viruses, mRNA vaccines are non-infectious and can be synthetically produced in an egg-free, cell-free environment, thereby reducing the risk of a detrimental immune response in the host (1614). Furthermore, mRNA vaccines are easily, affordably, and rapidly scalable, despite the fact that it took time to reach the scale needed to manufacture vaccines at a scale sufficient for the global population (1612).

8.9.0.1 Prior Applications

Although mRNA vaccines have been developed for therapeutic and prophylactic purposes, none have previously been licensed or commercialized. Challenges were caused by the instability of mRNA molecules, the design requirements of an efficient delivery system, and the potential for mRNA to elicit either a very strong immune response or to stimulate the immune system in secondary ways (1476, 1615). As of the 2010s, mRNA was still considered a promising technology for future advances in vaccine development (1605), but prior to 2020, no mRNA vaccines had been approved for use in humans, despite significant advances in the development of this technology (1611). This approach showed promise in animal models and preliminary clinical trials for several indications, including rabies, coronavirus, influenza, and cytomegalovirus (1616). Preclinical data previously identified effective antibody generation against full-length purified influenza hemagglutinin stalk-encoding mRNA in mice, rabbits, and ferrets (1617). Similar immunological responses for mRNA vaccines were observed in humans in phase I and II clinical trials operated by the pharmaceutical-development companies Curevac and Moderna for rabies, flu, and zika (1608). Positively charged bilayer LNPs carrying the mRNA attract negatively charged cell membranes, endocytose into the cytoplasm (1607), and facilitate endosomal escape. LNPs can be coated with modalities recognized and engulfed by specific cell types, and LNPs that are 150 nm or less effectively enter into lymphatic vessels (1607, 1618). Therefore, while these technologies elegantly capitalize on decades of research in vaccine development as well as the tools of the genomic revolution, it was largely unknown prior to the SARS-CoV-2 pandemic whether this potential could be realized in a real-world vaccination effort.

8.9.0.2 Application to COVID-19

Table 5: mRNA vaccines approved in at least one country (1169) as of May 3, 2023. As a note, this table includes licensing of existing mRNA technology, i.e., TAK-919 is used to describe Takeda's manufacturing of Moderna's formulation.

Vaccine	Company
GEMCOVAC-19	Gennova Biopharmaceuticals Limited
Spikevax	Moderna
Spikevax Bivalent Original/Omicron BA.1	Moderna
Spikevax Bivalent Original/Omicron BA.4/BA.5	Moderna
Comirnaty	Pfizer/BioNTech

Vaccine	Company
Comirnaty Bivalent Original/Omicron BA.1	Pfizer/BioNTech
Comirnaty Bivalent Original/Omicron BA.4/BA.5	Pfizer/BioNTech
TAK-919 (Moderna formulation)	Takeda
AWcorna	Walvax

Given the potential for mRNA technology to be quickly adapted for a new pathogen, it was favored as a potential vaccine against COVID-19, and fortunately, the prior work in mRNA vaccine development paid off, with 9 mRNA vaccines available in at least one country as of May 3, 2023 (Table 5). In the vaccines developed under this approach, the mRNA coding for a stabilized prefusion Spike protein, which is immunogenic (1619), is furnished to the immune system in order to train its response.

Two vaccine candidates in this category emerged with promising phase III results at the end of 2020. Both require two doses approximately one month apart. The first was Pfizer/BioNTech's BNT162b2, which contains the full prefusion stabilized, membrane-anchored SARS-CoV-2 Spike protein in a vaccine formulation based on modified mRNA (modRNA) technology (1620, 1621). The second mRNA vaccine, mRNA-1273 developed by ModernaTX, is comprised by a conventional LNP-encapsulated RNA encoding a full-length prefusion stabilized S protein for SARS-CoV-2 (1622). The vaccine candidates developed against SARS-CoV-2 using mRNA vectors utilize similar principles and technologies, although there are slight differences in implementation among candidates such as the formulation of the platform and the specific components of the Spike protein encapsulated (e.g., the full Spike protein vs. the RBD alone) (1623). As of May 31, 2023, 2 mRNA vaccines are available in 169 countries (Figure 12).

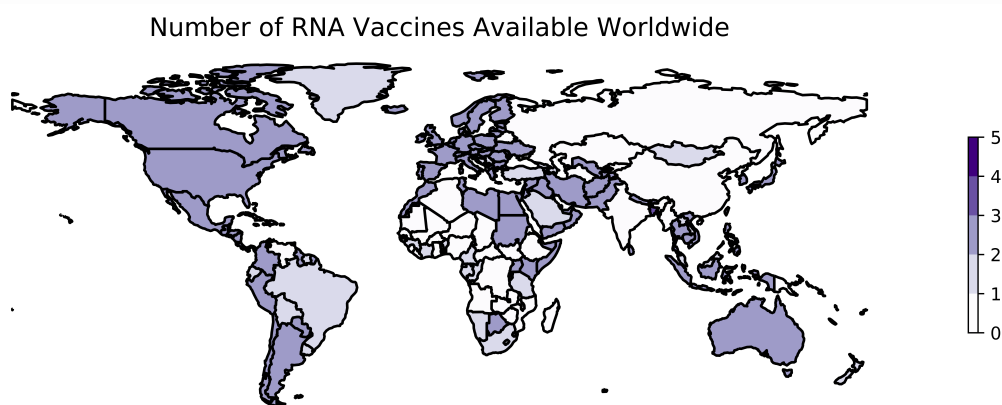


Figure 12: Worldwide availability of vaccines developed using mRNA. This figure reflects the number of vaccines based on mRNA technology that were available in each country as of May 31, 2023. These data are retrieved from Our World in Data (1116) and plotted using geopandas (1170). See <https://greenelab.github.io/covid19-review/> for the most recent version of this figure, which is updated daily. Note that this figure draws from a different data source than Table 5 and does not necessarily include data for every vaccine developed within this category.

The rapid and simultaneous development of these vaccines was met with some controversy related to intellectual property (IP). First, the National Institutes of Health (NIH) and Moderna became involved in a patent dispute,

after researchers at the NIH argued they were unfairly excluded from some patents filed based on their IP after they generated the stabilized modRNA sequence used in the vaccine ([1624](#)). Ultimately, in late 2021, Moderna backed down on the patent application ([1625](#)). However, in August 2022, the company filed their own suit against Pfizer/BioNTech over IP related to the modRNA used in the latter's COVID-19 vaccine ([1625](#), [1626](#)). The outcome of this suit remains to be seen.

8.9.0.3 Trial Safety and Immunogenicity

The VEs revealed by the Pfizer/BioNTech and Moderna clinical trials exceeded expectations. In a phase II/III multinational trial, the Pfizer/BioNTech's BNT162b2 vaccine was associated with a 95% efficacy against laboratory-confirmed COVID-19 and with mild-to-moderate local and systemic effects but a low risk of serious AEs when the prime-boost doses were administered 21 days apart ([516](#)). The ModernaTX mRNA-1273 vaccine was the second mRNA vaccine to release phase III results, despite being the first mRNA vaccine to enter phase I clinical trials and publish interim results of their phase III trial a few months later. Their study reported a 94.5% vaccine efficacy in preventing symptomatic COVID-19 in adults who received the vaccine at 99 sites around the United States ([1627](#)). Similar to BNT162b2, the mRNA-1273 vaccine was associated with mild-to-moderate AEs but with a low risk of serious AEs ([1627](#)). In late 2020, both vaccines received approval from the FDA under an emergency use authorization ([1628](#), [1629](#)), and these vaccines have been widely distributed, primarily in North America and the European Union ([1186](#)). As the first mRNA vaccines to make it to market, these two highly efficacious vaccines demonstrate the power of this emerging technology, which has previously attracted scientific interest because of its potential to be used to treat non-infectious as well as infectious diseases.

8.9.0.4 Real-World Safety and Effectiveness

As vaccines were rolled out, one study sought to monitor their effectiveness in a real-world setting. Between December 2020 and April 2021, this prospective cohort study obtained weekly nasal swabs from 3,975 individuals at high risk of SARS-CoV-2 exposure (health care workers, frontline workers, etc.) within the United States ([332](#)). Among these participants, 3,179 (80%) had received at least one dose of an mRNA vaccine, and of those, 2,686 (84%) were fully vaccinated, corresponding to 68% of trial participants overall. For each vaccinated participant (defined here as having received at least dose 1 more than 7 days ago) whose sample tested positive for SARS-CoV-2, they categorized the viral lineage(s) present in the sample as well as in samples from 3-4 unvaccinated individuals matched by site and testing date. Overall efficacy of mRNA vaccines was estimated at 91% with full vaccination, similar to the reports from the clinical trials. The occurrence of fevers was also lower in individuals who were partially or fully vaccinated, and the duration of symptoms was approximately 6 days shorter. Among the five cases in fully vaccinated and 11 cases in partially vaccinated participants, the rate of infection by VOC was much higher than in the unvaccinated population (30% versus 10%), suggesting that the vaccine was less effective against the VOC than the index strain.

The WHO continues to monitor the emergence of variants and their impact on vaccine efficacy ([529](#)). In general, mRNA vaccines remain highly effective against severe illness and death, but the effectiveness against infection generally has declined. A study monitoring infections in a Minnesota cohort from January to July 2021 estimated that the effectiveness of the Moderna vaccine fell to 86% and Pfizer to 76%, although protection against hospitalization remained at 91% and 85%, respectively ([1630](#)). In July of that year, as the Delta variant became dominant in the U.S.A., these estimates all fell, to an effectiveness of 76% for Moderna and 42% for Pfizer and effectiveness against hospitalization of 81% and 75%, respectively ([1630](#)).

With the emergence of the Omicron VOC, vaccine effectiveness has likely further declined. A study in a diverse cohort in Southern California, U.S.A. found the effectiveness of the Moderna vaccine in participants who had received only the primary course to be 44% ([1631](#)). A study in South Africa compared case and hospitalization records from a 4-week period where Omicron was dominant to a 2-month period where Delta was dominant and found that the effectiveness against hospitalization during the Omicron wave was approximately 70% compared to 93% during the Delta wave ([1632](#)). Similarly, a large study in England of 2.5 million individuals suggested that not only the variants circulating, but also the time since vaccination, played a large role in vaccine effectiveness ([1633](#)). Shortly after the BNT162b2 primary course, effectiveness against the Omicron VOC was as high as 65.5%, but this declined to below 10% by six months after the second dose. For mRNA-1273, the decline was from 75.1% to 14.9%. Therefore, it is unsurprising that in spite of vaccination programs, infection rates and hospitalization rates climbed in early 2022 in many Western countries including the United States ([1634](#), [1635](#)), especially given that many places simultaneously began to loosen public health restrictions designed to reduce viral spread.

On the side of safety, the only major concern that has been raised is a possible link between mRNA vaccination and myocarditis, especially in young men ([1597](#)). This concern began with case reports of a small number of cases of myocarditis following vaccination in several countries ([1636](#), [1637](#)). Following these reports, the Israeli Ministry of Health began surveillance to monitor the occurrence of myocarditis ([1638](#)). They identified 283 cases, almost exactly half of which occurred following vaccination with Pfizer's BNT162b2. Close analysis of these cases determined that the vaccine did have a significant effect on the incidence of myocarditis; however, the rate of myocarditis remained low overall ([1638](#)). The identification of young men as a population at particular risk of this AE was supported, and the risk was found to be greater after the second dose than the first. Both this study and a study evaluating data collected from US population-based surveillance identified an increased risk with additional doses ([1639](#)). However, most findings suggest that this AE does not have long-term negative effects; a 2021 meta-analysis identified 69 cases, all of which resulted in full recovery ([1640](#)). Although these events are very rare, as with the possible thromboembolic AEs associated with viral-vectored DNA vaccines, these findings suggest that it may be prudent to offer a framework for decision making for patients particularly concerned about specific AEs in settings where multiple vaccines are available.

8.10 Booster Doses

Due to waning effectiveness of vaccines over time, especially in light of viral evolution, boosters have emerged as an important strategy in retaining the benefits of vaccination over time. Booster shots are now recommended in many places, and boosters that account for multiple variants and strains of SARS-CoV-2 are now available in some places ([531](#)). For example, in the U.S.A., the FDA recently recommended bivalent booster doses designed to account for the Omicron VOC ([1641–1643](#)). In this case, bivalent refers to the fact that doses deliver both the original formulation and an updated vaccine designed for the Omicron subvariants circulating in summer 2022. The fact that the FDA did not require additional clinical trials from manufacturers for Omicron subvariants BA.4 and BA.5 specifically suggests that the rapid authorization of strain changes in response to emerging VOC may be increasingly attainable ([1644](#)). Results suggest that this fourth dose offered at least a short-term increase in VE against Omicron subvariants and also provided additional protection against hospitalization ([1645](#)).

Homologous booster doses have been investigated for most vaccines. For example, over 14,000 adults were administered a booster (second) dose of the Janssen Ad26.COV2.S vaccine ([1646](#)). The booster dose was highly efficacious, with severe COVID-19 and hospitalization prevented almost completely in the vaccinated group. A booster dose was also found to improve immune response for Sputnik V vaccinees ([1647](#)). For the AstraZeneca vaccine, a different approach was taken. In the interest of distributing first doses as widely as possible, in some places the time between the first and second doses was extended. One study assessed the immunogenicity and reactogenicity associated with delaying the second dose in the prime-boost series until up to 45 weeks after the first, reporting that an extended inter-dose period was associated with increased antibody titers 28 days after the second dose ([1648](#)). This analysis also revealed that a third dose provided an additional boost in neutralizing activity ([1648](#)).

Third and fourth doses have been introduced for at least some populations in many places in response to the Omicron variant. An early study in Israeli healthcare workers showed that the additional immunization was safe and immunogenic with antibody titers restored to peak-third dose titers. No severe illness was reported in the cohort studied (274 versus 426 age-matched controls), and vaccine efficacy against infection was reported at 30% for BNT162b2 and 11% for mRNA-1273 ([1649](#)). Other studies reported that a third dose of BNT162b2 raised vaccine effectiveness to 67.2% for approximately the first month but that the effectiveness dropped to 45.7% ([1633](#)). Reduced and even low efficacy against infection does not undermine the value of vaccination, considering the vaccines are intended to prevent severe disease, hospitalization, and death rather than infection generally. However, these findings do suggest that boosters will likely be needed as the virus continues to evolve.

Many trials have also investigated heterologous boosting approaches. In particular, the mRNA vaccines are a popular choice for booster doses regardless of primary series. In general, such approaches have been found to confer favorable immunogenicity relative to homologous boosters (e.g., [1650–1656](#)) and many other studies). Due to remaining concerns about rare thromboembolic events, vaccinees who received AstraZeneca for their primary course are advised in some countries to seek a heterologous booster

(1657), although such guidances are not supported by the evidence, which indicates that the first dose of AstraZeneca is most likely to be linked to these rare events (1658). In general, heterologous boosting with mRNA vaccines elicits a strong immune response. For patients who received BNT162b2 as a heterologous booster following a ChAdOx1 primary series, the vaccine effectiveness was estimated to be 62.4% initially, dropping to 39.6% after 10 weeks (1633). For a heterologous mRNA-1273 booster, the effectiveness was estimated to be slightly higher (70.1% and 60.9% following ChAdOx1 and 73.9% to 64.4% following BNT162b2) (1633). Therefore, subsequent booster doses may remain an ongoing component of strategies to combat SARS-CoV-2.

Although the vaccines developed based on the index strain remain highly effective at preventing severe illness and death, they serve much less utility at preventing illness broadly than they did early in the pandemic. Therefore, many manufacturers are exploring potential reformulations based on VOC that have emerged since the beginning of the pandemic. In June 2022, Moderna released data describing the effect of their bivalent mRNA booster, mRNA-1273.214, designed to protect against the Omicron variant (1659). A 50 µg dose of mRNA-1273.214 was administered to 437 participants. One month later, the neutralizing geometric mean titer ratio was assessed against several variants of SARS-CoV-2, including Omicron. The immune response was higher against all variants assessed, including Omicron, than for boosting with the original formulation (mRNA-1273). Another formulation, mRNA-1273.211, developed based on the Beta variant, has been associated with durable protection as long as six months after dosing. The associated publications suggest that this novel formulation offers significant protection against Omicron and other VOC (1660, 1661). In August 2022, Pfizer also announced successful development of a new formulation effective against Omicron (1662).

Modularity has been proposed as one of the advantages to developing DNA and mRNA vaccines. This design would allow for faster adaptation to viral evolution. However, in the arms race against SARS-CoV-2, the vaccines are still lagging behind the virus. This disadvantage may change as regulators become more familiar with these vaccines and as a critical mass of data is accumulated. Given the apparent need for boosters, interest has also emerged in whether updated formulations of SARS-CoV-2 vaccines can be administered along with annual flu vaccines to improve immunity to novel variants.

8.11 Conclusions

COVID-19 has seen the coming-of-age of vaccine technologies that have been in development since the late 20th century but had never before been authorized for use. Vaccines that employ DNA and RNA eliminate all concerns about potential infection due to the vaccine components. The vaccines described above demonstrate the potential for these technologies to facilitate a quick response to an emerging pathogen. Additionally, their efficacy in trials far exceeded expectations, especially in the case of RNA vaccines. These technologies hold significant potential to drive improvements in human health over the coming years.

Traditional vaccine technologies were built on the principle of using either a weakened version of the virus or a fragment of the virus. COVID-19 has highlighted the fact that in recent years, the field has undergone a paradigm shift towards reverse vaccinology. Reverse vaccinology emphasizes a discovery-driven approach to vaccine development based on knowledge of the viral genome ([1663](#)). This strategy was explored during development of a DNA vaccine against the Zika virus ([1664](#)). Though the disease was controlled before the vaccine became available ([1423](#)), the response demonstrated the potential for modular technologies to facilitate a response to emerging viral threats ([1664](#)). The potential for such vaccines to benefit the field of oncology has encouraged vaccine developers to invest in next-generation approaches, which has spurred the diversification of vaccine development programs ([1479](#), [1665](#)). As a result, during the COVID-19 pandemic, these modular technologies have taken center stage in controlling a viral threat for the first time.

The safety and efficacy of vaccines that use these new technologies has exceeded expectations. While there were rare reports of severe AEs such as myocarditis (mRNA platforms) and thromboembolic events (viral-vectored DNA platforms), widespread availability of both types of vaccines would allow individuals to choose (particularly relevant in this case because myocarditis has primarily been reported in men and thromboembolic events primarily in women). Estimates of efficacy have varied widely, but in all cases are high. Estimates of the efficacy of DNA vaccine platforms have typically fallen either in the range of approximately 67% (ZyCoV-D and Janssen) or 90% (Sputnik V). AstraZeneca's trial produced estimates in both ranges, with the standard dosage producing an efficacy of 62% and the lower prime dose producing a VE of 90%. The mRNA vaccine trials were somewhat higher, with VE estimated at approximately 95% for both the Moderna and Pfizer/BioNTech clinical trials. However, in all cases, the efficacy against severe illness and death were very high. Therefore, all of these vaccines are useful tools for combating COVID-19.

Furthermore, the fact that vaccine efficacy is not a static value has become particularly salient, as real-world effectiveness has changed with location and over time. COVID-19 vaccines have been challenged by the emergence of VOC. These VOC generally carry genetic mutations that code for an altered Spike protein (i.e., the antigen), so the antibodies resulting from immunization with vaccines developed from the index strain neutralize them less effectively ([1666](#), [1667](#)). Despite some reports of varying and reduced effectiveness or efficacy of the mRNA vaccines against the Alpha (B.1.1.7), Beta (B.1.351), and Delta (B.1.617.2) variants versus the original SARS-CoV-2 strain or the D614G variant ([1668–1670](#)), the greatest concern to date has been the Omicron variant (B.1.1.529), which was first identified in November 2021 ([1667](#), [1671](#)). As of March 2022, the Omicron variant accounted for 95% of all infections sequenced in the United States ([122](#)) and was linked to an increased risk of SARS-CoV-2 reinfection ([1666](#)) and further infection of those who have been vaccinated with the mRNA vaccines ([1672](#)).

One of the downsides of this leap in vaccine technologies, however, is that they have largely been developed by wealthy countries, including countries in the European Union, the United States, the U.K., and Russia. As a result, they are also largely available to residents of wealthy countries, primarily in

Europe and North America. Although the VE of DNA vaccines tends to be lower than that of mRNA vaccines ([1673](#)), they still provide excellent protection against severe illness and are much easier to distribute due to less complex demands for storage. Efforts such as COVAX that aim to expand access to vaccines developed by wealthy countries have not been as successful as hoped ([1674](#)). Fortunately, vaccine development programs using more established technologies have been undertaken in many middle-income countries, and those vaccines have been more accessible globally ([5](#)). Additionally, efforts to develop new formulations of DNA vaccines in lower- and middle-income countries are increasingly being undertaken ([1675](#)).

The modular nature of nucleic acid-based vaccine platforms has opened a new frontier in responding to emerging viral illnesses. The RNA vaccines received an EUA in only a few months more than it took to identify the pathogen causing SARS in 2002. Given the variety of options available for preventing severe illness and death, it is possible that certain vaccines may be preferable for certain demographics (e.g., young women might choose an mRNA vaccine to entirely mitigate the very low risk of blood clots ([1597](#))). However, this option is likely only available to people in high-income countries. In lower-income countries, access to vaccines broadly is a more critical issue. Different vaccines may confer advantages in different countries, and vaccine development in a variety of cultural contexts is therefore important ([1676](#)). Without widespread access to vaccines on the global scale, SARS-CoV-2 will continue evolving, presenting a threat to all nations.

9 Appendix: Additional Information about Novel Vaccine Platforms for COVID-19

9.1 Plasmid-Vectored DNA Vaccines

9.1.1 INO-4800

The phase I trial for INO-4800 began enrolling participants in April 2020 in Philadelphia, PA at the Perelman School of Medicine and at the Center for Pharmaceutical Research in Kansas City, MO. This trial examined two different doses administered in a two-dose regimen ([1512](#)). Among the 39 participants, only six AEs were reported and all were grade 1 ([1512](#)). Efficacy was evaluated based on blood samples collected pre- and post-vaccination, and all but three participants of 38 included in the analysis were found to have serum IgG binding titers to the spike protein after vaccination ([1512](#)).

Results from the phase II trial were released as a preprint in May 2021 and reported findings based on administering INO-4800 to 401 adult volunteers at high risk of exposure to SARS-CoV-2 ([1513](#)). The phase II results supported that the vaccine was safe, with 1,446 treatment-related AEs observed across 281 participants, all but one of which were grade 1 or grade 2. The single grade 3 event was joint stiffness ([1513](#)). The rates of AEs in the placebo group were not reported. To assess the immunogenicity of INO-4800, pre- and post-

vaccination blood samples were collected and evaluated for a humoral immune response to the spike protein, and the treatment group was identified to show significantly greater neutralizing activity than the placebo group ([1513](#)).

The phase II/III trials are ongoing in several countries, including the United States, Mexico, India, and Colombia ([1514–1517](#)). Therefore, vaccine efficacy data from a large study population is not yet available.

9.2 Viral-Vectored DNA Vaccines

9.2.1 ChAdOx1 nCoV-19 (AstraZeneca)

Prior analyses of viral vector vaccines against human coronaviruses (HCoVs) had indicated that this approach showed potential for inducing an immune response, but little information was available about the effect on real-world immunity. In the first phase of development, a candidate ChAdOx1 nCoV-19 was evaluated through the immune challenge of two animal models, mice and rhesus macaques ([1556](#)). Animals in the treatment condition were observed to develop neutralizing antibodies specific to SARS-CoV-2 (both macaques and mice) and to show reduced clinical scores when exposed to SARS-CoV-2 (macaques) ([1556](#)).

Next, a phase I/II trial was undertaken using a single-blind, randomized controlled design ([1557](#)). ChAdOx1 nCoV-19 and a control, the meningococcal conjugate vaccine MenACWY, were administered intramuscularly to adults ages 18 to 55 at five sites within the United Kingdom (U.K.) at a 1:1 ratio (n=543 and n=534, respectively). All but ten participants received a single dose; this small group received a booster 28 days after their first dose of ChAdOx1 nCoV-19. Commonly reported local adverse reactions included mild-to-moderate pain and tenderness at the injection site over the course of seven days, while the most common systemic adverse reactions were fatigue and headache; some patients reported severe adverse systemic effects. The study also reported that many common reactions could be reduced through the administration of paracetamol (acetaminophen), and paracetamol was not found to reduce immunogenicity. Patients receiving the ChAdOx1 nCoV-19 vaccine developed antibodies to the SARS-CoV-2 spike protein that peaked by day 28, with these levels remaining stable until a second observation at day 56 except in the ten patients who received a booster dose at day 28, in whom they increased by day 56. Analysis of serum indicated that participants developed antibodies to both S and the RBD, and that 100% of them achieved neutralizing titers by day 28. By day 35, the neutralization titers of vaccinated patients was comparable to that observed with plasma from convalescents. This initial study therefore suggested that the vaccine was likely to confer protection against SARS-CoV-2, although analysis of its efficacy in preventing COVID-19 was not reported.

The primary outcome assessed was symptomatic, laboratory-confirmed COVID-19. There were 131 cases observed among the 11,636 participants eligible for the primary efficacy analysis, corresponding to an overall efficacy of 70.4% (30 out of 5,807 in the vaccine arm and 101 out of 5,829 in the control arm); the 95.8% CI was reported as 54.8 to 80.6. However, a higher

efficacy was reported in the subgroup of patients who received a low-dose followed by a standard dose (90.0%, 95% CI 67.4 to 97.0). A total of ten cases of severe COVID-19 resulting in hospitalization were observed among trial participants, and all of these occurred in patients in the control arm of the study. In line with the previously reported safety profiling for this vaccine, serious adverse events were reported to be comparable across the two arms of the study, with only three events identified as potentially associated with the vaccine itself.

Additional data about the efficacy of this vaccine became available in a preprint released on March 2, 2021 ([1677](#)). This report provided data describing the efficacy of ChAdOx1 nCoV-19, along with Pfizer/BioNTech's BNT162b2, in the U.K. between December 8, 2020 and February 19, 2021 and specifically sought to evaluate the efficacy of the vaccine in the presence of a potentially more contagious variant of concern, B.1.1.7. All participants in this study were age 70 or older and the efficacy was estimated to increase from 60% at 28 days after vaccination to 73% at 35 days after vaccination, although the standard error also increased over this time. Therefore, preliminary results suggest that in a number of samples, this vaccine confers a high level of protection against SARS-CoV-2.

9.2.2 Sputnik-V (Gam-COVID-Vac and Gam-COVID-Vac-Lyo)

The vaccine Gam-COVID-Vac, nicknamed Sputnik V in reference to the space race and "V for vaccine", was developed by the Gamaleya National Center of Epidemiology and Microbiology in Moscow. The development of Sputnik V was financed by the Russian Direct Investment Fund (RDIF) ([1577](#), [1678](#)). The Sputnik V vaccines are available in both a lyophilized (Gam-COVID-Vac-Lyo) and frozen form (Gam-COVID-Vac), which are stored at 2-8°C and -18°C respectively ([1582](#)). The lyophilized vaccine is convenient for distribution and storage, particularly to remote or disadvantaged areas ([1679](#)).

In the phase I/II trial study conducted between late June and early August 2020, 76 participants (18-60 years old) were split into two groups of 38 participants, which were non-randomized in two hospitals in Russia. In phase I, 9 patients received rAd26 and 9 patients received rAd5-S to assess safety over 28 days. In phase II, at least 5 days after the completion of phase I, 20 patients received a prime-boost vaccination of rAd26-S on day 0 and rAd5-S on day 2, which was administered intramuscularly. The phase I/II trial reported that both vaccines were deemed safe and well tolerated. The most common adverse events reported were mild, such as pain at the injection site (58%), hypothermia (50%), headaches (42%), fatigue (28%), and joint and muscle pain (24%). Seroconversion was observed in all participants three weeks post the second vaccination (day 42), and all participants produced antibodies to the SARS-CoV-2 glycoprotein. RBD-specific IgG levels were high in both the frozen and lyophilized versions of the vaccine (14,703 and 11,143 respectively), indicating a sufficient immune response to both. Three weeks post the second vaccination, the virus-neutralizing geometric mean antibody titers were 49.25 and 45.95 from the frozen and lyophilized vaccines, respectively. At 28 days, median cell proliferation of 1.3% CD4⁺ and 1.1% CD8⁺ were reported for the lyophilized vaccine and 2.5% CD4⁺ and 1.3% CD8⁺ for the vaccine stored frozen. These results indicated that both forms

of Sputnik V appeared to be safe and induced a humoral and cellular response in human subjects ([1582](#)), which may be robust enough to persist and not wane rapidly ([1558](#)).

In February 2021, the interim results of the phase III randomized, double-blind, placebo-controlled trial were published in *The Lancet* ([1573](#)). The participants were randomly assigned to receive either a 0.5 mL/dose of vaccine or placebo, which was comprised of the vaccine buffer composition, that was delivered intramuscularly using the same prime-boost regimen as in the phase I/II trials. From September 7 to November 24, 19,866 participants completed the trial. Of the 14,964 participants who received the vaccine, 16 (0.1%) were confirmed to have COVID-19, whereas 62 of the 4,902 participants (1.3%) in the placebo group were confirmed to have COVID-19. Of these participants, no moderate or severe cases of COVID-19 were reported in the vaccine group, juxtaposed with 20 in the placebo group. However, only symptomatic individuals were confirmed for SARS-CoV-2 infection in this trial. Therefore, asymptomatic infections were not detected, thus potentially inflating the efficacy estimate. Overall, a vaccine efficacy of 91.6% (95% CI 85.6-95.2) was reported, where an efficacy of 91.8% was reported for those over 60 years old and 92.7% for those who were 51-60 years old. Indeed, 14 days after the first dose, 87.6% efficacy was achieved and the immunity required to prevent disease occurred within 18 days of vaccination.

Based on these results, scientists are investigating the potential for a single dose regimen of the rAd26-S sputnik V vaccine ([1680](#)). By the end of the trial, 7,485 participants reported adverse events, of which 94% were grade I. Of the 68 participants who experienced serious adverse events during the trial, 45 from the vaccine group and 23 from the placebo groups, none were reported to be associated with the vaccination. Likewise, 4 deaths occurred during the trial period that were not related to the vaccine ([1573](#)). The interim findings of the phase III trial indicate that the Sputnik V vaccine regimen appears to be safe with 91.6% efficacy. Gamaleya had intended to reach a total of 40,000 participants for the completion of their phase III trial. However, the trial has stopped enrolling participants and the numbers have been cut to 31,000 as many individuals in the placebo group dropped out of the study to obtain the vaccine ([1681](#)). Other trials involving Sputnik V are currently underway in Belarus, India, the United Arab Emirates, and Venezuela ([1682](#)).

Preliminary results of a trial of Argentinian healthcare workers in Buenos Aires who were vaccinated with the Sputnik V rAd26-R vector-based vaccine seems to support the short term safety of the first vaccination ([1683](#)). Of the 707 vaccinated healthcare workers, 71.3% of the 96.6% of respondents reported at least one adverse event attributed to the vaccine. Of these individuals, 68% experienced joint and muscle pain, 54% had injection site pain, 11% reported redness and swelling, 40% had a fever, and 5% reported diarrhea. Only 5% of the vaccinated participants experienced serious adverse events that required medical attention, of which one was monitored as an inpatient.

Additionally, an independent assessment of Sputnik V in a phase II clinical trial in India found the vaccine to be effective, but the data is not yet publicly available ([1684](#)). On December 21, 2020, Gamaleya, AstraZeneca, R-Pharm, and the Russian Direct Investment Fund agreed to assess the safety and immunogenicity of the combined use of components of the AstraZeneca and University of Oxford AZD1222 (ChAdOx1) vaccine and the rAd26-S component of the Sputnik V vaccine in clinical trials ([1685](#)). This agreement hopes to establish scientific and business relations between the entities with an aim to co-develop a vaccine providing long-term immunization. The trial, which will begin enrollment soon, will include 100 participants in a phase II open-label study and is hoped to be complete within 6 months. Participants will first receive an intramuscular dose of AZD1222 on day 1, followed by a dose of rAd26 on day 29 and be monitored from day 1 for 180 days in total. The primary outcomes measured will include incidence of serious adverse events post first dose until the end of the study. Secondary outcome measures will include incidence of local and systemic adverse events 7 days post each dose, a time course of antibody responses for the Spike protein and the presence of anti-SARS-CoV-2 neutralizing antibodies ([1686](#)).

Overall, there is hesitancy surrounding the management of the Sputnik V vaccine approval process and concerns over whether the efficacy data may be inflated due to a lack of asymptomatic testing within the trial. However, the interim results of the phase III study were promising and further trials are underway, which will likely shed light on the overall efficacy and safety of the Sputnik V vaccine regimen. There may be some advantage to the Sputnik V approach including the favorable storage conditions afforded by choice between a frozen and lyophilized vaccine. Furthermore, the producers of Gam-COVID-Vac state that they can produce the vaccine at a cost of less than \$10 per dose or less than \$20 per patient ([1687](#)).

9.3 Janssen's JNJ-78436735

The Johnson and Johnson (J&J) vaccine developed by Janssen Pharmaceuticals, Inc., a subsidiary of J&J, was conducted in collaboration with and funded by "Operation Warp Speed" ([1485](#), [1560–1562](#)). The vaccine was developed using Janssen's AdVac[®] and PER.C6 platforms that were previously utilized to develop the European Commission-approved Ebola vaccine (Ad26 ZEBOV and MVN-BN-Filo) and their Zika, respiratory syncytial virus, and human immunodeficiency virus investigational vaccine candidates ([1688](#)).

The development of a single-dose vaccine was desired by J&J from the outset, with global deployment being a key priority ([1563](#)). Using their AdVac[®] technology, the vaccine can remain stable for up to two years between -15 and -25°C and at least three months at 2 to 8°C ([1688](#)). This allows the vaccine to be distributed easily without the requirement for very low temperature storage, unlike many of the other COVID-19 vaccine candidates. J&J screened numerous potential vaccine candidates *in vitro* and in animal models using varying different designs of the S protein, heterologous signal peptides, and prefusion-stabilizing substitutions ([1485](#)). A select few candidates were further investigated as a single dose regimen in Syrian golden hamsters, a single dose regimen in rhesus macaques, and a single- and two-dose regimen in both adult and aged rhesus macaques ([1485](#), [1563](#)–

[1565](#)). A SARS-CoV-2 challenge study in rhesus macaques showed that vaccine doses as low as 2×10^9 viral particles/mL was sufficient to induce strong protection in bronchoalveolar lavage but that doses higher than 1.125×10^{10} were required to close achieve close to complete protection in nasal swabs ([1689](#)). Indeed, six months post-immunization, levels of S-binding and neutralizing antibodies in rhesus macaques indicated that the JNJ-78436735 vaccine conferred durable protection against SARS-CoV-2 ([1566](#)).

Following selection of the JNJ-78436735 vaccine, J&J began phase I/IIa trials. The interim phase I/IIa data was placed on the *medRxiv* preprint server on September 25th, 2020 ([1690](#)) and was later published in the *New England Journal of Medicine* on January 13th, 2021 ([1562](#)). The phase I/IIa multicenter, randomized, placebo-controlled trial enrolled 402 healthy participants between 18-55 years old and a further 403 healthy older participants ≥ 65 years old ([1562](#)). Patients were administered either a placebo, a low dose (5×10^{10} viral particles per mL), or a high dose (1×10^{11} viral particles per mL) intramuscularly as part of either a single- or two-dose regimen. All patients received injections 56 days apart, but participants in the single-dose condition received the placebo at the second appointment. Those who received only one dose of either vaccine received a placebo dose at their second vaccination visit. The primary endpoints of both the trial were safety and reactogenicity of each dose. Fatigue, headache, myalgia, and pain at the injection site were the most frequent solicited adverse events reported by participants. Although less common, particularly for those in the elderly cohort and those on the low dose regimen, the most frequent systemic adverse effect was fever. Overall, immunization was well tolerated, particularly at the lower dose concentration. In terms of reactogenicity, over 90% of those who received either the low or high dose demonstrated seroconversion in a neutralization assay using wild-type SARS-CoV-2, 29 days after immunization ([1562](#)). Neutralizing geometric mean ratio of antibody titers (GMT) between 224-354 were detected regardless of age. By day 57, 100% of the 18-55 year old participants had neutralizing GMT (288-488), which remained stable until day 71. In the ≥ 65 years old cohort, the incidence of seroconversion for the low- and high-dose was 96% and 88% respectively by day 29.

GMTs for the low and high doses were slightly lower for participants ≥ 65 years old (196 and 127 respectively), potentially indicating slightly lower immunogenicity. Seroconversion of the S antibodies was detected in 99% of individuals between 18-55 years old for the low and high doses (GMTs 528 and 695 respectively), with similar findings reported for the ≥ 65 years old. Indeed, both dose concentrations also induced robust Th1 cytokine-producing S-specific CD4⁺ T cells and CD8⁺ T cell responses in both age groups. The findings of the phase I/IIa study supported further investigation of a single immunization using the low dose vaccine. Therefore, 25 patients were enrolled for a second randomized double-blind, placebo-controlled phase I clinical trial currently being conducted in Boston, Massachusetts for 2 years ([1691](#)). Participants received either a single dose followed by a placebo, or a double dose of either a low dose (5×10^{10} viral particles/mL) or a high dose (1×10^{11} viral particles/mL) vaccine administered intramuscularly on day 1 or day 57. Placebo-only recipients received a placebo dose on day 1 and 57. Interim analyses conducted on day 71 indicated that binding and neutralizing antibodies developed 8 days after administration in 90% and

25% of vaccine recipients, respectively. Binding and neutralizing antibodies were detected in 100% of vaccine recipients by day 57 after a single dose immunization. Spike-specific antibodies were highly prevalent (GMT 2432 to 5729) as were neutralizing antibodies (GMT 242 to 449) in the vaccinated groups. Indeed, CD4⁺ and CD8⁺ T-cell responses were also induced, which may provide additional protection, particularly if antibodies wane or poorly respond to infection ([1692](#)).

On September 23rd, 2020, J&J launched its phase III trial ENSEMBLE and released the study protocol to the public ([1688](#), [1693](#)). The trial intended to enroll 60,000 volunteers to assess the safety and efficacy of the single vaccine dose versus placebo with primary endpoints of 14 and 28 days post-immunization ([1688](#)). The trial was conducted in Argentina, Brazil, Chile, Colombia, Mexico, Peru, South Africa, and the U.S. The trial was paused briefly in October 2020 to investigate a “serious medical event”, but resumed shortly after ([1694](#)).

An interim analysis was reported via press release on January 29th, 2021 ([1575](#), [1576](#)). The interim data included 43,783 participants who accrued 468 symptomatic cases of COVID-19. It was reported that the JNJ-78436735 vaccine was 66% effective across all regions studied for the prevention of moderate to severe COVID-19 28 days post-vaccination in those aged 18 years and older. Notably, JNJ-78436735 was 85% effective for the prevention of laboratory-confirmed severe COVID-19 and 100% protection against COVID-19-related hospitalization and death 28 days post-vaccination across all study sites. Efficacy of the vaccine against severe COVID-19 increased over time, and there were no cases of COVID-19 reported in immunized participants after day 49. The trial also determined that the vaccine candidate has a favorable safety profile as determined by an independent Data and Safety Monitoring Board. The vaccine was well tolerated, consistent with previous vaccines produced using the AdVac[®] platform. Fever occurred in 9% of vaccine recipients, with grade 3 fever occurring in only 0.2% of recipients. Serious adverse events were reportedly higher in the placebo group than the vaccine group, and no anaphylaxis was reported ([1576](#)).

At the time the phase III trial was being conducted, several concerning variants, including B.1.1.7 ([482](#)) and B.1.351 ([231](#)), were spreading across the globe. In particular, B.1.351 was first identified in South Africa, which was one of the JNJ-78436735 vaccine trial sites. Therefore, the J&J investigators also analyzed the efficacy of the JNJ-78436735 vaccine associated with their various trial sites to determine any potential risk of reduced efficacy as a result of the novel variants. It was determined that JNJ-78436735 was 72% effective in the U.S., 66% effective in Latin America, and 57% effective in South Africa 28 days post-vaccination. These findings underpin the importance of monitoring for the emergence of novel SARS-CoV-2 variants and determining their effects on vaccine efficacy.

Looking forward, Janssen are also running a phase III randomized, double-blind, placebo-controlled clinical trial, Ensemble 2, which aims to assess the efficacy, safety, and immunogenicity of a two-dose regimen of JNJ-78436735 administered 57 days apart. This trial will enroll 30,000 participants ≥ 18 years old from Belgium, Colombia, France, Germany, Philippines, South

Africa, Spain, U.K., and the U.S. ([1695](#)). This trial will also include participants with and without comorbidities associated with an increased risk of COVID-19.

9.4 RNA Vaccines

RNA vaccines are nucleic-acid based modalities that code for viral antigens against which the human body elicits a humoral and cellular immune response. The resulting intracellular viral proteins are displayed on surface MHC proteins, provoking a strong CD8+ T cell response as well as a CD4+ T cell and B cell-associated antibody responses ([1127](#)). Given the potential for this technology to be quickly adapted for a new pathogen, it has held significant interest for the treatment of COVID-19. The results of the interim analyses of two mRNA vaccine candidates became available at the end of 2020 and provided strong support for this emerging approach to vaccination. Below we describe in detail the results available as of February 2021 for two such candidates, mRNA-1273 produced by ModernaTX and BNT162b2 produced by Pfizer, Inc. and BioNTech. As of August 2022, the U.S. FDA has issues approvals or emergency use authorizations of versions of these vaccines for adults and for children 6 months and older ([1696](#)).

9.4.1 ModernaTX mRNA Vaccine

ModernaTX's mRNA-1273 vaccine was the first COVID-19 vaccine to enter a phase I clinical trial in the United States. An initial report described the results of enrolling forty-five participants who were administered intramuscular injections of mRNA-1273 in their deltoid muscle on day 1 and day 29, with the goal of following patients for the next twelve months ([1119](#)). Healthy males and non-pregnant females aged 18-55 years were recruited for this study and divided into three groups receiving 25, 100, or 250 µg of mRNA-1273. IgG ELISA assays on patient serology samples were used to examine the immunogenicity of the vaccine ([1622](#)). Binding antibodies were observed at two weeks after the first dose at all concentrations. At the time point one week after the second dose was administered on day 29, the pseudotyped lentivirus reporter single-round-of-infection neutralization assay, which was used to assess neutralizing activity, reached a median level similar to the median observed in convalescent plasma samples. Participants reported mild and moderate systemic adverse events after the day 1 injection, and one severe local event was observed in each of the two highest dose levels. The second injection led to severe systemic adverse events for three of the participants at the highest dose levels, with one participant in the group being evaluated at an urgent care center on the day after the second dose. The reported localized adverse events from the second dose were similar to those from the first.

Several months later, a press release from ModernaTX described the results of the first interim analysis of the vaccine ([1697](#)). On November 16, 2020, a report was released describing the initial results from phase III testing, corresponding to the first 95 cases of COVID-19 in the 30,000 enrolled participants ([1697](#)), with additional data released to the FDA on December 17, 2020 ([1698](#)). These results were subsequently published in a peer-reviewed journal (*The New England Journal of Medicine*) on December 30,

2020 ([1627](#)). The first group of 30,420 study participants was randomized to receive the vaccine or a placebo at a ratio of 1:1 ([1627](#)). Administration occurred at 99 sites within the United States in two sessions, spaced 28 days apart ([1627](#), [1699](#)). Patients reporting COVID-19 symptoms upon follow-up were tested for SARS-CoV-2 using a nasopharyngeal swab that was evaluated with RT-PCR ([1699](#)). The initial preliminary analysis reported the results of the cases observed up until a cut-off date of November 11, 2020. Of these first 95 cases reported, 90 occurred in participants receiving the placebo compared to 5 cases in the group receiving the vaccine ([1697](#)). These results suggested the vaccine is 94.5% effective in preventing COVID-19. Additionally, eleven severe cases of COVID-19 were observed, and all eleven occurred in participants receiving the placebo. The publication reported the results through an extended cut-off date of November 21, 2020, corresponding to 196 cases ([1627](#)). Of these, 11 occurred in the vaccine group and 185 in the placebo group, corresponding to an efficacy of 94.1%. Once again, all of the severe cases of COVID-19 observed (n=30) occurred in the placebo group, including one death. Thus, as more cases are reported, the efficacy of the vaccine has remained above 90%, and no cases of severe COVID-19 have yet been reported in participants receiving the vaccine.

These findings suggest the possibility that the vaccine might bolster immune defenses even for subjects who do still develop a SARS-CoV-2 infection. The study was designed with an explicit goal of including individuals at high risk for COVID-19, including older adults, people with underlying health conditions, and people of color ([1700](#)). The phase III trial population was comprised by approximately 25.3% adults over age 65 in the initial report and 24.8% in the publication ([1699](#)). Among the cases reported by both interim analyses, 16-17% occurred in older adults ([1627](#), [1697](#)). Additionally, approximately 10% of participants identified a Black or African-American background and 20% identified Hispanic or Latino ethnicity ([1627](#), [1699](#)). Among the first 95 cases, 12.6% occurred in participants identifying a Hispanic or Latino background and 4% in participants reporting a Black or African-American background ([1697](#)); in the publication, they indicated only that 41 of the cases reported in the placebo group and 1 case in the treatment group occurred in “communities of color”, corresponding to 21.4% of all cases ([1627](#)). While the sample size in both analyses is small relative to the study population of over 30,000, these results suggest that the vaccine is likely to be effective in people from a variety of backgrounds.

In-depth safety data was released by ModernaTX as part of their application for an EUA from the FDA and summarized in the associated publication ([1627](#), [1699](#)). Because the detail provided in the report is greater than that provided in the publication, here we emphasize the results observed at the time of the first analysis. Overall, a large percentage of participants reported adverse effects when solicited, and these reports were higher in the vaccine group than in the placebo group (94.5% versus 59.5%, respectively, at the time of the initial analysis) ([1699](#)). Some of these events met the criteria for grade 3 (local or systemic) or grade 4 (systemic only) toxicity ([1699](#)), but most were grade 1 or grade 2 and lasted 2-3 days ([1627](#)). The most common local adverse reaction was pain at the injection site, reported by 83.7% of participants receiving the first dose of the vaccine and 88.4% upon receiving the second dose, compared to 19.8% and 17.0%, respectively, of patients in the placebo condition ([1699](#)). Fewer than 5% of vaccine recipients reported

grade 3 pain at either administration. Other frequent local reactions included erythema, swelling, and lymphadenopathy (1699). For systemic adverse reactions, fatigue was the most common (1699). Among participants receiving either dose of the vaccine, 68.5% reported fatigue compared to 36.1% participants receiving the placebo (1699). The level of fatigue experienced was usually fairly mild, with only 9.6% and 1.3% of participants in the vaccine and placebo conditions, respectively, reporting grade 3 fatigue (1699), which corresponds to significant interference with daily activity (1701). Based on the results of the report, an EUA was issued on December 18, 2020 to allow distribution of this vaccine in the United States (1629), and it was shortly followed by an Interim Order authorizing distribution of the vaccine in Canada (1702) and a conditional marketing authorization by the European Medicines Agency to facilitate distribution in the European Union (1703).

9.4.2 Pfizer/BioNTech BNT162b2

ModernaTX was, in fact, the second company to release news of a successful interim analysis of an mRNA vaccine and receive an EUA. The first report came from Pfizer and BioNTech's mRNA vaccine BNT162b2 on November 9, 2020 (1704), and a preliminary report was published in the *New England Journal of Medicine* one month later (516). This vaccine candidate should not be confused with a similar candidate from Pfizer/BioNTech, BNT162b1, that delivered only the RBD of the spike protein (1705, 1706), which was not advanced to a phase III trial because of the improved reactogenicity/immunogenicity profile of BNT162b2 (517).

During the phase III trial of BNT162b2, 43,538 participants were enrolled 1:1 in the placebo and the vaccine candidate and received two 30-µg doses 21 days apart (516). Of these enrolled participants, 21,720 received BNT162b2 and 21,728 received a placebo (516). Recruitment occurred at 135 sites across six countries: Argentina, Brazil, Germany, South Africa, Turkey, and the United States. An initial press release described the first 94 cases, which were consistent with 90% efficacy of the vaccine at 7 days following the second dose (1704). The release of the full trial information covered a longer period and analyzed the first 170 cases occurring at least 7 days after the second dose, 8 of which occurred in patients who had received BNT162b2. The press release characterized the study population as diverse, reporting that 42% of the participants worldwide came from non-white backgrounds, including 10% Black and 26% Hispanic or Latino (1707). Within the United States, 10% and 13% of participants, respectively, identified themselves as having Black or Hispanic/Latino backgrounds (1707). Additionally, 41% of participants worldwide were 56 years of age or older (1707), and they reported that the efficacy of the vaccine in adults over 65 was 94% (1708). The primary efficacy analysis of the phase III study was concluded on November 18, 2020 (1708), and the final results indicted 94.6% efficacy of the vaccine (516).

The safety profile of the vaccine was also assessed (516). A subset of patients were followed for reactogenicity using electronic diaries, with the data collected from these 8,183 participants comprising the solicited safety events analyzed. Much like those who received the ModernaTX vaccine candidate, a large proportion of participants reported experiencing site injection pain within 7 days of vaccination. While percentages are broken down by age

group in the publication, these proportions correspond to approximately 78% and 73% of all participants after the first and second doses, respectively, overall. Only a small percentage of these events (less than 1%) were rated as serious, with the rest being mild or moderate, and none reached grade 4. Some participants also reported redness or swelling, and the publication indicates that in most cases, such events resolved within 1 to 2 days. Participants also experienced systemic effects, including fever (in most cases lower than 38.9°C and more common after dose 2), fatigue (25-50% of participants depending on age group and dose), headache (25-50% of participants depending on age group and dose), chills, and muscle or joint pain; more rarely, patients could experience gastrointestinal effects such as vomiting or diarrhea. As with the local events, these events were almost always grade 1 or 2. While some events were reported by the placebo groups, these events were much rarer than in the treatment group even though compliance was similar. Based on the efficacy and safety information released, the vaccine was approved in early December by the United Kingdom's Medicines and Healthcare Products Regulatory Agency with administration outside of a clinical trial beginning on December 8, 2020 ([1709](#), [1710](#)). As of December 11, 2020, the United States FDA approved this vaccine under an emergency use authorization ([1628](#)), and in August 2021, it received full approval for ages 16 and older ([1711](#)).

9.4.3 Neutralizing of VOC

Prior to studies examining the effectiveness of vaccines in real-world settings (summarized in [6](#)), several studies reported reduced efficacy of the mRNA vaccines based on the measurement of antibody titers. Plasma from individuals double-dosed with Pfizer/BioNTech's BNT162b2 vaccine had up to a 16-fold reduction in neutralizing capacity against the Omicron variant ([1712](#)) and a reduced efficacy (70%) ([1632](#)). Estimates for the mRNA vaccines range from a 2-fold to over a 20-fold drop in neutralisation titers ([1713](#)), hence the push for third and fourth doses of mRNA vaccines in many Western countries. A third mRNA vaccine dose does increase antibody titers, but these levels also wane with time ([1714](#)). Notably, immunocompromised individuals such as cancer patients seem to elicit a sufficient protective immune response against the Omicron variant when they have been boosted with a third dose of either mRNA vaccine, albeit a blunted response ([1715](#)). While antibody titers do correlate with protection ([1716–1720](#)), they are not the only mechanisms of immune protection. For example, T cell and non-neutralizing antibody responses may be unaffected or less affected by the new VOC, and they warrant further investigation.

9.5 Global Vaccine Status and Distribution

In North America, the majority of vaccines distributed until March 2021 have been produced by Pfizer-BioNTech and Moderna. In Canada, the vaccine approval process is conducted by Health Canada, which uses a fast-tracked process whereby vaccine producers can submit data as it becomes available to allow for rapid review. An approval may be granted following reviews of the available phase III clinical data. This is followed by a period of pharmacovigilance in the population using their post-market surveillance system, which will monitor the long-term safety and efficacy of any vaccines

([1721](#), [1722](#)). Health Canada has authorized the use of the Pfizer (December 9th, 2020), Moderna (December 23rd, 2020), Oxford-AstraZeneca (February 26th, 2021), and the Janssen (March 5th, 2021) vaccines, and the Novavax Inc vaccine is also under consideration ([1723](#)). While Canada initially projected that by the end of September 2021 a vaccine would be available for all Canadian adults, they now predict that it may be possible earlier as more vaccines have been approved and become available ([1724](#)).

In the U.S., vaccines are required to have demonstrated safety and efficacy in phase III trials before manufacturers apply for an emergency use authorization (EUA) from the FDA. If an EUA is granted, an additional evaluation of the safety and efficacy of the vaccines is conducted by the CDC's Advisory Committee on Immunization Practices (ACIP) who also provide guidance on vaccine prioritization. On December 1st, 2020, ACIP provided an interim phase 1a recommendation that healthcare workers and long-term care facility residents should be the first to be offered any vaccine approved ([1725](#)). This was shortly followed by an EUA on December 11th, 2020 for the use of the Pfizer-BioNTech COVID vaccine ([1726](#)), which was distributed and administered to the first healthcare workers on December 14th, 2020 ([1727](#)). Shortly thereafter, an EUA for the Moderna vaccine was issued on December 18th, 2020 ([1728](#)). On December 20th, 2020, ACIP updated their initial recommendations to suggest that vaccinations should be offered to people aged 75 years old and older and to non-healthcare frontline workers in phase 1b ([1729](#)). On the same date, it was recommended that phase 1c should include people aged 65-74 years old, individuals between the ages of 16-74 years old at high-risk due to health conditions, and essential workers ineligible in phase 1b ([1729](#)). On the following day, December 21st, 2020, the first Moderna vaccines used outside of clinical trials were administered to American healthcare workers, which was the same day that President-elect Biden and Dr. Biden received their first doses of the Pfizer-BioNTech vaccine live on television to instill confidence in the approval and vaccination process ([1730](#)).

On February 27th, 2020, the FDA issued an EUA for the Janssen COVID-19 Vaccine ([1731](#)). This was followed by an update on recommendations by ACIP for the use of the Janssen COVID-19 vaccine for those over 18 years old ([1732](#)). The Janssen vaccine was first distributed to healthcare facilities on March 1st, 2021. On March 12, 2021, the WHO added the Janssen vaccine to the list of safe and effective emergency tools for COVID-19 ([1733](#)). While the CDC's ACIP can provide recommendations, it is up to the public health authorities of each state, territory, and tribe to interpret the guidance and determine who will be vaccinated first ([1734](#)). Prior to distribution of the Janssen vaccine, over 103 million doses of the Moderna and Pfizer-BioNTech vaccines were delivered across the U.S., with almost 79 million doses administered. Of the total population, 15.6% have received at least one dose and 7.9% have received a second dose of either the Moderna (~38.3 million) or the Pfizer-BioNTech (~40.2 million) vaccines by February 28th, 2021 ([1735](#)). President Biden's administration has predicted that by the end of May 2021 there may be enough vaccine supply available for all adults in the U.S. ([1736](#), [1737](#)). However, vaccine production, approval, and distribution was not straightforward in the U.S., as information was initially sparse and the rollout of vaccines was complicated by poor planning and leadership due to political

activities prior to the change of administration in January 2021 ([1738](#)). These political complications highlight the importance of the transparent vaccine approval process conducted by the FDA ([1461](#)).

Outside the U.S., the Moderna and Pfizer-BioNTech vaccines have been administered in 29 and 69 other countries, respectively, mainly in Europe and North America ([1364](#)). The Janssen vaccine has so far only been administered in South Africa and the U.S. ([1364](#), [1739](#)), but it has also been approved in Bahrain, the European Union (E.U.), Iceland, Liechtenstein, and Norway ([1186](#)). On March 11th, 2021, Johnson & Johnson received approval from the European Medicines Agency (EMA) for conditional marketing authorization of their vaccine ([1740](#)). Notably, on March 2nd, 2021, rivals Johnson & Johnson and Merck announced that they entered an agreement to increase production of the Janssen vaccine to meet global demand ([1741](#)).

The U.K. was the first country to approve use of the Pfizer-BioNTech vaccine on December 2nd, 2020 ([1742](#)), and it was later approved by EMA on December 21st, 2020 ([1743](#)). The U.K. was also the first to administer the Pfizer-BioNTech vaccine, making it the first COVID-19 vaccine supported by phase III data to be administered outside of clinical trials on December 8th, 2020. The Oxford-AstraZeneca vaccine, was approved by the Medicines and Healthcare Products Regulatory Agency (MHRA) in the U.K. and by EMA in the E.U. on December 30th (2020) ([1744](#)) and January 29th (2021) ([1602](#)), respectively. The Oxford-AstraZeneca vaccine was first administered in the UK on January 4th, 2021 ([1745](#)), and it is now being used in 53 countries in total, including Brazil, India, Pakistan, Mexico, and spanning most of Europe ([1364](#)). The Moderna vaccine was authorized for use in the E.U. by EMA on January 6th, 2021 ([1746](#)) and in the U.K. by MHRA on January 8th, 2021 ([1747](#)). As of March 5th, 2021, 22 million people in the U.K. had received at least one vaccine dose ([1365](#)).

While the Pfizer-BioNTech vaccine was the first to be distributed following phase III clinical trials, the first COVID-19 vaccine to be widely administered to people prior to the completion of phase III clinical trials was Sputnik V. Sputnik V was administered to as many as 1.5 million Russians by early January ([1569](#)) due to the establishment of mass vaccination clinics in December 2020, prior to which only approximately 100,000 Russians had already been vaccinated ([1748](#), [1749](#)). Doses of Sputnik V have also been distributed to other parts of Europe ([1570](#)–[1572](#)). Hungary was the first E.U. member country to approve and distribute Sputnik V outside of Russia ([1750](#)), despite the EMA stating that they had neither approved nor received a request for approval of Sputnik V ([1751](#)). Hungary is also in talks with China to procure the Sinopharm vaccines, which have been approved by Hungarian health authorities but also have not received approval by EMA in the E.U. ([1750](#)). In Latin America, production facilities in both Brazil and Argentina will allow for increased production capacity of Sputnik V and doses have been distributed to Mexico, Argentina, Bolivia, Nicaragua, Paraguay, and Venezuela ([1752](#)). Guinea was the first African nation to administer Sputnik V in December 2020, and the Central African Republic, Zimbabwe, and the Ivory Coast have all registered their interest in purchasing doses of the vaccine ([1752](#)). In the Middle East, Iran has received its first doses of Sputnik V and the United Arab Emirates is conducting phase III trials ([1752](#)). In Asia, while China's vaccine candidates are favored, the Philippines, Nepal, and

Uzbekistan have sought Sputnik V doses ([1753](#)). In total, the RDIF claims to have received orders totalling 1.2 billion doses by over 50 countries worldwide ([1753](#)) and at least 18 countries are currently administering Sputnik V around the globe ([1364](#)). Sputnik V has been an attractive vaccine for many countries due to its relatively low price, high efficacy, and its favorable storage conditions. For some countries, Russia and China have also been more palatable politically than vaccine suppliers in the West ([1752](#), [1754](#)). For others, the delays in the distribution of the other, more-favored candidates has been a motivating factor for pursuing the Sputnik V and Chinese alternatives ([1571](#), [1754](#)). Additionally, Germany has stated that if Sputnik V were approved by EMA, it would be considered by the E.U. ([1755](#)). Russia is developing other vaccine candidates and has approved a third vaccine, CoviVac, which is an inactivated vaccine produced by the Chumakov Centre in Moscow, despite the fact the clinical trials have yet to begin ([1756](#)).

10 Dietary Supplements and Nutraceuticals Under Investigation for COVID-19 Prevention and Treatment

10.1 Abstract

Coronavirus disease 2019 (COVID-19) has caused global disruption and a significant loss of life. Existing treatments that can be repurposed as prophylactic and therapeutic agents could reduce the pandemic's devastation. Emerging evidence of potential applications in other therapeutic contexts has led to the investigation of dietary supplements and nutraceuticals for COVID-19. Such products include vitamin C, vitamin D, omega 3 polyunsaturated fatty acids, probiotics, and zinc, all of which are currently under clinical investigation. In this review, we critically appraise the evidence surrounding dietary supplements and nutraceuticals for the prophylaxis and treatment of COVID-19. Overall, further study is required before evidence-based recommendations can be formulated, but nutritional status plays a significant role in patient outcomes, and these products could help alleviate deficiencies. For example, evidence indicates that vitamin D deficiency may be associated with greater incidence of infection and severity of COVID-19, suggesting that vitamin D supplementation may hold prophylactic or therapeutic value. A growing number of scientific organizations are now considering recommending vitamin D supplementation to those at high risk of COVID-19. Because research in vitamin D and other nutraceuticals and supplements is preliminary, here we evaluate the extent to which these nutraceutical and dietary supplements hold potential in the COVID-19 crisis.

10.2 Importance

Sales of dietary supplements and nutraceuticals have increased during the pandemic due to their perceived “immune-boosting” effects. However, little is known about the efficacy of these dietary supplements and nutraceuticals against the novel coronavirus (SARS-CoV-2) or the disease it causes, COVID-19. This review provides a critical overview of the potential prophylactic and therapeutic value of various dietary supplements and nutraceuticals from the evidence available to date. These include vitamin C, vitamin D, and zinc, which are often perceived by the public as treating respiratory infections or supporting immune health. Consumers need to be aware of misinformation and false promises surrounding some supplements, which may be subject to limited regulation by authorities. However, considerably more research is required to determine whether dietary supplements and nutraceuticals exhibit prophylactic and therapeutic value against SARS-CoV-2 infection and COVID-19. This review provides perspective on which nutraceuticals and supplements are involved in biological processes that are relevant to recovery from or prevention of COVID-19.

10.3 Introduction

The year 2020 saw scientists and the medical community scrambling to repurpose or discover novel host-directed therapies against the coronavirus disease 2019 (COVID-19) pandemic caused by the spread of the novel *Severe acute respiratory syndrome-related coronavirus 2* (SARS-CoV-2). This rapid effort led to the identification of some promising pharmaceutical therapies for hospitalized patients, such as remdesivir and dexamethasone. Furthermore, most societies have adopted non-pharmacological preventative measures such as utilizing public health strategies that reduce the transmission of SARS-CoV-2. However, during this time, many individuals sought additional protections via the consumption of various dietary supplements and nutraceuticals that they believed to confer beneficial effects. While a patient’s nutritional status does seem to play a role in COVID-19 susceptibility and outcomes ([1757–1761](#)), the beginning of the pandemic saw sales of vitamins and other supplements soar despite a lack of any evidence supporting their use against COVID-19. In the United States, for example, dietary supplement and nutraceutical sales have shown modest annual growth in recent years (approximately 5%, or a \$345 million increase in 2019), but during the six-week period preceding April 5, 2020, they increased by 44% (\$435 million) relative to the same period in 2019 ([1762](#)). While growth subsequently leveled off, sales continued to boom, with a further 16% (\$151 million) increase during the six weeks preceding May 17, 2020 relative to 2019 ([1762](#)). In France, New Zealand, India, and China, similar trends in sales were reported ([1763–1766](#)). The increase in sales was driven by a consumer perception that dietary supplements and nutraceuticals would protect consumers from infection and/or mitigate the impact of infection due to the various “immune-boosting” claims of these products ([1767](#), [1768](#)).

Due to the significant interest from the general public in dietary additives, whether and to what extent nutraceuticals or dietary supplements can provide any prophylactic or therapeutic benefit remains a topic of interest for the scientific community. Nutraceuticals and dietary supplements are related but distinct non-pharmaceutical products. Nutraceuticals are classified as supplements with health benefits beyond their basic nutritional value ([1769](#),

[1770](#)). The key difference between a dietary supplement and a nutraceutical is that nutraceuticals should not only supplement the diet, but also aid in the prophylaxis and/or treatment of a disorder or disease ([1771](#)). However, dietary supplements and nutraceuticals, unlike pharmaceuticals, are not subject to the same regulatory protocols that protect consumers of medicines. Indeed, nutraceuticals do not entirely fall under the responsibility of the Food and Drug Administration (FDA), but they are monitored as dietary supplements according to the Dietary Supplement, Health and Education Act 1994 (DSHEA) ([1772](#)) and the Food and Drug Administration Modernization Act 1997 (FDAMA) ([1773](#)). Due to increases in sales of dietary supplements and nutraceuticals, in 1996 the FDA established the Office of Dietary Supplement Programs (ODSP) to increase surveillance. Novel products or nutraceuticals must now submit a new dietary ingredient notification to the ODSP for review. There are significant concerns that these legislations do not adequately protect the consumer as they ascribe responsibility to the manufacturers to ensure the safety of the product before manufacturing or marketing ([1774](#)). Manufacturers are not required to register or even seek approval from the FDA to produce or sell food supplements or nutraceuticals. Health or nutrient content claims for labeling purposes are approved based on an authoritative statement from the Academy of Sciences or relevant federal authorities once the FDA has been notified and on the basis that the information is known to be true and not deceptive ([1774](#)). Therefore, there is often a gap between perceptions by the American public about a nutraceutical or dietary supplement and the actual clinical evidence surrounding its effects.

Despite differences in regulations, similar challenges exist outside of the United States. In Europe, where the safety of supplements is monitored by the European Union (EU) under Directive 2002/46/EC ([1775](#)). However, nutraceuticals are not directly mentioned. Consequently, nutraceuticals can be generally described as either a medicinal product under Directive 2004/27/EC ([1776](#)) or as a 'foodstuff' under Directive 2002/46/EC of the European council. In order to synchronize the various existing legislations, Regulation EC 1924/2006 on nutrition and health claims was put into effect to assure customers of safety and efficacy of products and to deliver understandable information to consumers. However, specific legislation for nutraceuticals is still elusive. Health claims are permitted on a product label only following compliance and authorization according to the European Food Safety Authority (EFSA) guidelines on nutrition and health claims ([1777](#)). EFSA does not currently distinguish between food supplements and nutraceuticals for health claim applications of new products, as claim authorization is dependent on the availability of clinical data in order to substantiate efficacy ([1778](#)). These guidelines seem to provide more protection to consumers than the FDA regulations but potentially at the cost of innovation in the sector ([1779](#)). The situation becomes even more complicated when comparing regulations at a global level, as countries such as China and India have existing regulatory frameworks for traditional medicines and phytomedicines not commonly consumed in Western society ([1780](#)). Currently, there is debate among scientists and regulatory authorities surrounding the development of a widespread regulatory framework to deal with the challenges of safety and health claim substantiation for nutraceuticals ([1774](#), [1778](#)), as these products do not necessarily follow the same rigorous clinical trial frameworks used to approve the use of pharmaceuticals. Such

regulatory disparities have been highlighted by the pandemic, as many individuals and companies have attempted to profit from the vulnerabilities of others by overstating claims in relation to the treatment of COVID-19 using supplements and nutraceuticals. The FDA has written several letters to prevent companies marketing or selling products based on false hyperbolic promises about preventing SARS-CoV-2 infection or treating COVID-19 ([1781–1783](#)). These letters came in response to efforts to market nutraceutical prophylactics against COVID-19, some of which charged the consumer as much as \$23,000 ([1784](#)). There have even been some incidents highlighted in the media because of their potentially life threatening consequences; for example, the use of oleandrin was touted as a potential “cure” by individuals close to the former President of the United States despite its high toxicity ([1785](#)). Thus, heterogeneous and at times relaxed regulatory standards have permitted high-profile cases of the sale of nutraceuticals and dietary supplements that are purported to provide protection against COVID-19, despite a lack of research into these compounds.

Notwithstanding the issues of poor safety, efficacy, and regulatory oversight, some dietary supplements and nutraceuticals have exhibited therapeutic and prophylactic potential. Some have been linked with reduced immunopathology, antiviral and anti-inflammatory activities, or even the prevention of acute respiratory distress syndrome (ARDS) ([1767](#), [1786](#), [1787](#)). A host of potential candidates have been highlighted in the literature that target various aspects of the COVID-19 viral pathology, while others are thought to prime the host immune system. These candidates include vitamins and minerals along with extracts and omega-3 polyunsaturated fatty acids (n-3 PUFA) ([1788](#)). *In vitro* and *in vivo* studies suggest that nutraceuticals containing phycocyanobilin, N-acetylcysteine, glucosamine, selenium or phase 2 inductive nutraceuticals (e.g. ferulic acid, lipoic acid, or sulforaphane) can prevent or modulate RNA virus infections via amplification of the signaling activity of mitochondrial antiviral-signaling protein (MAVS) and activation of Toll-like receptor 7 ([1789](#)). Phase 2 inductive molecules used in the production of nutraceuticals are known to activate nuclear factor erythroid 2-related factor 2 (Nrf2), which is a protein regulator of antioxidant enzymes that leads to the induction of several antioxidant enzymes, such as gamma-glutamylcysteine synthetase. While promising, further animal and human studies are required to assess the therapeutic potential of these various nutrients and nutraceuticals against COVID-19. For the purpose of this review, we have highlighted some of the main dietary supplements and nutraceuticals that are currently under investigation for their potential prophylactic and therapeutic applications. These include n-3 PUFA, zinc, vitamins C and D, and probiotics.

10.4 n-3 PUFA

One category of supplements that has been explored for beneficial effects against various viral infections are the n-3 PUFAs ([1788](#)), commonly referred to as omega-3 fatty acids, which include eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA). EPA and DHA intake can come from a diet high in fish or through dietary supplementation with fish oils or purified oils ([1790](#)). Other, more sustainable sources of EPA and DHA include algae ([1791](#), [1792](#)), which can also be exploited for their rich abundance of other bioactive compounds such as angiotensin converting enzyme inhibitor peptides and

antiviral agents including phycobiliproteins, sulfated polysaccharides, and calcium-spirulan ([1793](#)). n-3 PUFAs have been investigated for many years for their therapeutic potential ([1794](#)). Supplementation with fish oils is generally well tolerated ([1794](#)), and intake of n-3 PUFAs through dietary sources or supplementation is specifically encouraged for vulnerable groups such as pregnant and lactating women ([1795](#), [1796](#)). As a result, these well-established compounds have drawn significant interest for their potential immune effects and therapeutic potential.

Particular interest has arisen in n-3 PUFAs as potential therapeutics against diseases associated with inflammation. n-3 PUFAs have been found to modulate inflammation by influencing processes such as leukocyte chemotaxis, adhesion molecule expression, and the production of eicosanoids ([1797](#), [1798](#)). This and other evidence indicates that n-3 PUFAs may have the capacity to modulate the adaptive immune response ([1770](#), [1790](#), [1797](#)); for example, they have been found to influence antigen presentation and the production of CD4(+) Th1 cells, among other relevant effects ([1799](#)). Certainly, preliminary evidence from banked blood samples from 100 COVID-19 patients suggests that patients with a higher omega-3 index, a measure of the amount of EPA and DHA in red blood cells, had a lower risk of death due to COVID-19 ([1800](#)). Interest has also arisen as to whether nutritional status related to n-3 PUFAs can also affect inflammation associated with severe disease, such as ARDS or sepsis ([1801](#), [1802](#)). ARDS and sepsis hold particular concern in the treatment of severe COVID-19; an analysis of 82 deceased COVID-19 patients in Wuhan during January to February 2020 reported that respiratory failure (associated with ARDS) was the cause of death in 69.5% of cases, and sepsis or multi-organ failure accounted for 28.0% of deaths ([742](#)). Research in ARDS prior to current pandemic suggests that n-3 PUFAs may hold some therapeutic potential. One study randomized 16 consecutive ARDS patients to receive either a fish oil-enriched lipid emulsion or a control lipid emulsion (comprised of 100% long-chain triglycerides) under a double-blinded design ([1803](#)). They reported a statistically significant reduction in leukotriene B4 levels in the group receiving the fish oil-enriched emulsion, suggesting that the fish oil supplementation may have reduced inflammation. However, they also reported that most of their tests were not statistically significant, and therefore it seems that additional research using larger sample sizes is required. A recent meta-analysis of 10 randomized controlled trials (RCTs) examining the effects of n-3 PUFAs on ARDS patients did not find evidence of any effect on mortality, although the effect on secondary outcomes could not be determined due to a low quality of evidence ([1804](#)). However, another meta-analysis that examined 24 RCTs studying the effects of n-3 fatty acids on sepsis, including ARDS-induced sepsis, did find support for an effect on mortality when n-3 fatty acids were administered via enteral nutrition, although a paucity of high-quality evidence again limited conclusions ([1805](#)). Therefore, despite theoretical support for an immunomodulatory effect of n-3 PUFAs in COVID-19, evidence from existing RCTs is insufficient to determine whether supplementation offers an advantage in a clinical setting that would be relevant to COVID-19.

Another potential mechanism that has led to interest in n-3 PUFAs as protective against viral infections including COVID-19 is its potential as a precursor molecule for the biosynthesis of endogenous specialized

proresolving mediators (SPM), such as protectins and resolvins, that actively resolve inflammation and infection (1806). SPM have exhibited beneficial effects against a variety of lung infections, including some caused by RNA viruses (1807, 1808). Several mechanisms for SPM have been proposed, including preventing the release of pro-inflammatory cytokines and chemokines or increasing phagocytosis of cellular debris by macrophages (1809). In influenza, SPM promote antiviral B lymphocytic activities (1810), and protectin D1 has been shown to increase survival from H1N1 viral infection in mice by affecting the viral replication machinery (1811). It has thus been hypothesized that SPM could aid in the resolution of the cytokine storm and pulmonary inflammation associated with COVID-19 (1812, 1813). Another theory is that some comorbidities, such as obesity, could lead to deficiencies of SPM, which could in turn be related to the occurrence of adverse outcomes for COVID-19 (1814). However, not all studies are in agreement that n-3 PUFAs or their resulting SPM are effective against infections (1815). At a minimum, the effectiveness of n-3 PUFAs against infections would be dependent on the dosage, timing, and the specific pathogens responsible (1816). On another level, there is still the question of whether fish oils can raise the levels of SPM levels upon ingestion and in response to acute inflammation in humans (1817). Currently, Karolinska University Hospital is running a trial that will measure the levels of SPM as a secondary outcome following intravenous supplementation of n-3 PUFAs in hospitalized COVID-19 patients to determine whether n-3 PUFAs provides therapeutic value (1818, 1819). Therefore, while this mechanism provides theoretical support for a role for n-3 PUFAs against COVID-19, experimental support is still needed.

A third possible mechanism by which n-3 PUFAs could benefit COVID-19 patients arises from the fact that some COVID-19 patients, particularly those with comorbidities, are at a significant risk of thrombotic complications including arterial and venous thrombosis (105, 1820). Therefore, the use of prophylactic and therapeutic anticoagulants and antithrombotic agents is under consideration (1821, 1822). Considering that there is significant evidence that n-3 fatty acids and other fish oil-derived lipids possess antithrombotic properties and anti-inflammatory properties (1790, 1823, 1824), they may have therapeutic value against the prothrombotic complications of COVID-19. In particular, concerns have been raised within the medical community about using investigational therapeutics on COVID-19 patients who are already on antiplatelet therapies due to pre-existing comorbidities because the introduction of such therapeutics could lead to issues with dosing and drug choice and/or negative drug-drug interactions (1821). In such cases, dietary sources of n-3 fatty acids or other nutraceuticals with antiplatelet activities could hold particular value for reducing the risk of thrombotic complications in patients already receiving pharmaceutical antiplatelet therapies. A new clinical trial (1825) is currently recruiting COVID-19 positive patients to investigate the anti-inflammatory activity of a recently developed, highly purified nutraceutical derivative of EPA known as icosapent ethyl (VascepaTM) (1826). Other randomized controlled trials that are in the preparatory stages intend to investigate the administration of EPA and other bioactive compounds to COVID-19 positive patients in order to observe whether anti-inflammatory effects or disease state improvements occur (1827, 1828). Finally, while there have been studies investigating the therapeutic value of n-3 fatty acids against ARDS in

humans, there is still limited evidence of their effectiveness ([1829](#)). It should be noted that the overall lack of human studies in this area means there is limited evidence as to whether these supplements could affect COVID-19 infection. Consequently, the clinical trials that are underway and those that have been proposed will provide valuable insight into whether the anti-inflammatory potential of n-3 PUFAs and their derivatives can be beneficial to the treatment of COVID-19. All the same, while the evidence is not present to draw conclusions about whether n-3 PUFAs will be useful in treating COVID-19, there is likely little harm associated with a diet rich in fish oils, and interest in n-3 PUFA supplementation by the general public is unlikely to have negative effects.

10.5 Zinc

Zinc is nutrient supplement that may exhibit some benefits against RNA viral infections. Zinc is a trace metal obtained from dietary sources or supplementation and is important for the maintenance of immune cells involved in adaptive and innate immunity ([1830](#)). Supplements can be administered orally as a tablet or as a lozenge and are available in many forms, such as zinc picolinate, zinc acetate, and zinc citrate. Zinc is also available from dietary sources including meat, seafood, nuts, seeds, legumes, and dairy. The role of zinc in immune function has been extensively reviewed ([1830](#)). Zinc is an important signaling molecule, and zinc levels can alter host defense systems. In inflammatory situations such as an infection, zinc can regulate leukocyte immune responses and modulate the nuclear factor kappa-light-chain-enhancer of activated B cells, thus altering cytokine production ([1831](#), [1832](#)). In particular, zinc supplementation can increase natural killer cell levels, which are important cells for host defense against viral infections ([1830](#), [1833](#)). As a result of these immune-related functions, zinc is also under consideration for possible benefits against COVID-19.

Adequate zinc intake has been associated with reduced incidence of infection ([1834](#)) and antiviral immunity ([1835](#)). A randomized, double-blind, placebo-controlled trial that administered zinc supplementation to elderly subjects over the course of a year found that zinc supplementation decreased susceptibility to infection and that zinc deficiency was associated with increased susceptibility to infection ([1834](#)). Clinical trial data supports the utility of zinc to diminish the duration and severity of symptoms associated with common colds when it is provided within 24 hours of the onset of symptoms ([1836](#), [1837](#)). An observational study showed that COVID-19 patients had significantly lower zinc levels in comparison to healthy controls and that zinc-deficient COVID-19 patients (those with levels less than 80 µg/dl) tended to have more complications (70.4% vs 30.0%, $p = 0.009$) and potentially prolonged hospital stays (7.9 vs 5.7 days, $p = 0.048$) relative to patients who were not zinc deficient ([1838](#)). In coronaviruses specifically, *in vitro* evidence has demonstrated that the combination of zinc (Zn^{2+}) and zinc ionophores (pyrithione) can interrupt the replication mechanisms of SARS-CoV-GFP (a fluorescently tagged SARS-CoV-1) and a variety of other RNA viruses ([1839](#), [1840](#)). Currently, there are over twenty clinical trials registered with the intention to use zinc in a preventative or therapeutic manner for COVID-19. However, many of these trials proposed the use of zinc in conjunction with hydroxychloroquine and azithromycin ([1841](#)–[1844](#)), and it is not known how the lack of evidence supporting the use of

hydroxychloroquine will affect investigation of zinc. One retrospective observational study of New York University Langone hospitals in New York compared outcomes among hospitalized COVID-19 patients administered hydroxychloroquine and azithromycin with zinc sulfate (n = 411) versus hydroxychloroquine and azithromycin alone (n = 521). Notably, zinc is the only treatment that was used in this trial that is still under consideration as a therapeutic agent due to the lack of efficacy and potential adverse events associated with hydroxychloroquine and azithromycin against COVID-19 ([1845–1847](#)). While the addition of zinc sulfate did not affect the duration of hospitalization, the length of ICU stays or patient ventilation duration, univariate analyses indicated that zinc did increase the frequency of patients discharged and decreased the requirement for ventilation, referrals to the ICU, and mortality ([1848](#)). However, a smaller retrospective study at Hoboken University Medical Center New Jersey failed to find an association between zinc supplementation and survival of hospitalized patients ([1849](#)). Therefore, whether zinc contributes to COVID-19 recovery remains unclear. Other trials are now investigating zinc in conjunction with other supplements such as vitamin C or n-3 PUFA ([1828](#), [1850](#)). Though there is, overall, encouraging data for zinc supplementation against the common cold and viral infections, there is currently limited evidence to suggest zinc supplementation has any beneficial effects against the current novel COVID-19; thus, the clinical trials that are currently underway will provide vital information on the efficacious use of zinc in COVID-19 prevention and/or treatment. However, given the limited risk and the potential association between zinc deficiency and illness, maintaining a healthy diet to ensure an adequate zinc status may be advisable for individuals seeking to reduce their likelihood of infection.

10.6 Vitamin C

Vitamins B, C, D, and E have also been suggested as potential nutrient supplement interventions for COVID-19 ([1788](#), [1851](#)). In particular vitamin C has been proposed as a potential therapeutic agent against COVID-19 due to its long history of use against the common cold and other respiratory infections ([1852](#), [1853](#)). Vitamin C can be obtained via dietary sources such as fruits and vegetables or via supplementation. Vitamin C plays a significant role in promoting immune function due to its effects on various immune cells. It affects inflammation by modulating cytokine production, decreasing histamine levels, enhancing the differentiation and proliferation of T- and B-lymphocytes, increasing antibody levels, and protecting against the negative effects of reactive oxygen species, among other effects related to COVID-19 pathology ([1854–1856](#)). Vitamin C is utilized by the body during viral infections, as evinced by lower concentrations in leukocytes and lower concentrations of urinary vitamin C. Post-infection, these levels return to baseline ranges ([1857–1861](#)). It has been shown that as little as 0.1 g/d of vitamin C can maintain normal plasma levels of vitamin C in healthy individuals, but higher doses of at least 1-3 g/d are required for critically ill patients in ICUs ([1862](#)). Indeed, vitamin C deficiency appears to be common among COVID-19 patients ([1863](#), [1864](#)). COVID-19 is also associated with the formation of microthrombi and coagulopathy ([107](#)) that contribute to its characteristic lung pathology ([1865](#)), but these symptoms can be ameliorated by early infusions of vitamin C to inhibit endothelial surface P-selectin expression and platelet-endothelial adhesion ([1866](#)). Intravenous vitamin C also reduced D-dimer levels in a case study of 17 COVID-19 patients ([1867](#)).

D-dimer levels are an important indicator of thrombus formation and breakdown and are notably elevated in COVID-19 patients ([103](#), [104](#)). There is therefore preliminary evidence suggesting that vitamin C status and vitamin C administration may be relevant to COVID-19 outcomes.

Larger-scale studies of vitamin C, however, have provided mixed results. A recent meta-analysis found consistent support for regular vitamin C supplementation reducing the duration of the common cold, but that supplementation with vitamin C (> 200 mg) failed to reduce the incidence of colds ([1868](#)). Individual studies have found Vitamin C to reduce the susceptibility of patients to lower respiratory tract infections, such as pneumonia ([1869](#)). Another meta-analysis demonstrated that in twelve trials, vitamin C supplementation reduced the length of stay of patients in intensive care units (ICUs) by 7.8% (95% CI: 4.2% to 11.2%; $p = 0.00003$). Furthermore, high doses (1-3 g/day) significantly reduced the length of an ICU stay by 8.6% in six trials ($p = 0.003$). Vitamin C also shortened the duration of mechanical ventilation by 18.2% in three trials in which patients required intervention for over 24 hours (95% CI 7.7% to 27%; $p = 0.001$) ([1862](#)). Despite these findings, an RCT of 167 patients known as CITRUS ALI failed to show a benefit of a 96-hour infusion of vitamin C to treat ARDS ([1870](#)). Clinical trials specifically investigating vitamin C in the context of COVID-19 have now begun, as highlighted by Carr et al. ([1853](#)). These trials intend to investigate the use of intravenous vitamin C in hospitalized COVID-19 patients. The first trial to report initial results took place in Wuhan, China ([1871](#)). These initial results indicated that the administration of 12 g/12 hr of intravenous vitamin C for 7 days in 56 critically ill COVID-19 patients resulted in a promising reduction of 28-day mortality ($p = 0.06$) in univariate survival analysis ([1872](#)). Indeed, the same study reported a significant decrease in IL-6 levels by day 7 of vitamin C infusion ($p = 0.04$) ([1873](#)). Additional studies that are being conducted in Canada, China, Iran, and the USA will provide additional insight into whether vitamin C supplementation affects COVID-19 outcomes on a larger scale.

Even though evidence supporting the use of vitamin C is beginning to emerge, we will not know how effective vitamin C is as a therapeutic for quite some time. Currently (as of January 2021) over fifteen trials are registered with [clinicaltrials.gov](#) that are either recruiting, active or are currently in preparation. When completed, these trials will provide crucial evidence on the efficacy of vitamin C as a therapeutic for COVID-19 infection. However, the majority of supplementation studies investigate the intravenous infusion of vitamin C in severe patients. Therefore, there is a lack of studies investigating the potential prophylactic administration of vitamin C via oral supplementation for healthy individuals or potentially asymptomatic SARS-CoV-2 positive patients. Once again, vitamin C intake is part of a healthy diet and the vitamin likely presents minimal risk, but its potential prophylactic or therapeutic effects against COVID-19 are yet to be determined. To maintain vitamin C status, it would be prudent for individuals to ensure that they consume the recommended dietary allowance of vitamin C to maintain a healthy immune system ([1757](#)). The recommended dietary allowance according to the FDA is 75-90 mg/d, whereas EFSA recommends 110 mg/d ([1874](#)).

10.7 Vitamin D

Of all of the supplements currently under investigation, vitamin D has become a leading prophylactic and therapeutic candidate against SARS-CoV-2. Vitamin D can modulate both the adaptive and innate immune system and is associated with various aspects of immune health and antiviral defense ([1875–1879](#)). Vitamin D can be sourced through diet or supplementation, but it is mainly biosynthesized by the body on exposure to ultraviolet light (UVB) from sunlight. Vitamin D deficiency is associated with an increased susceptibility to infection ([1880](#)). In particular, vitamin D deficient patients are at risk of developing acute respiratory infections ([1881](#)) and ARDS ([1881](#)). 1,25-dihydroxyvitamin D3 is the active form of vitamin D that is involved in adaptive and innate responses; however, due to its low concentration and a short half life of a few hours, vitamin D levels are typically measured by the longer lasting and more abundant precursor 25-hydroxyvitamin D. The vitamin D receptor is expressed in various immune cells, and vitamin D is an immunomodulator of antigen presenting cells, dendritic cells, macrophages, monocytes, and T- and B-lymphocytes ([1880](#), [1882](#)). Due to its potential immunomodulating properties, vitamin D supplementation may be advantageous to maintain a healthy immune system.

Early in the pandemic it was postulated that an individual's vitamin D status could significantly affect their risk of developing COVID-19 ([1883](#)). This hypothesis was derived from the fact that the current pandemic emerged in Wuhan China during winter, when 25-hydroxyvitamin D concentrations are at their lowest due to a lack of sunlight, whereas in the Southern Hemisphere, where it was nearing the end of the summer and higher 25-hydroxyvitamin D concentrations would be higher, the number of cases was low. This led researchers to question whether there was a seasonal component to the SARS-CoV-2 pandemic and whether vitamin D levels might play a role ([1883–1886](#)). Though it is assumed that COVID-19 is seasonal, multiple other factors that can affect vitamin D levels should also be considered. These factors include an individual's nutritional status, their age, their occupation, skin pigmentation, potential comorbidities, and the variation of exposure to sunlight due to latitude amongst others. Indeed, it has been estimated that each degree of latitude north of 28 degrees corresponded to a 4.4% increase of COVID-19 mortality, indirectly linking a persons vitamin D levels via exposure to UVB light to COVID-19 mortality ([1884](#)).

As the pandemic has evolved, additional research of varying quality has investigated some of the potential links identified early in the pandemic ([1883](#)) between vitamin D and COVID-19. Indeed, studies are beginning to investigate whether there is any prophylactic and/or therapeutic relationship between vitamin D and COVID-19. A study in Switzerland demonstrated that 27 SARS-CoV-2 positive patients exhibited 25-hydroxyvitamin D plasma concentrations that were significantly lower (11.1 ng/ml) than those of SARS-CoV-2 negative patients (24.6 ng/ml; $p = 0.004$), an association that held when stratifying patients greater than 70 years old ([1887](#)). These findings seem to be supported by a Belgian observational study of 186 SARS-CoV-2 positive patients exhibiting symptoms of pneumonia, where 25-hydroxyvitamin D plasma concentrations were measured and CT scans of the lungs were obtained upon hospitalization ([1888](#)). A significant difference in 25-hydroxyvitamin D levels was observed between the SARS-CoV-2 patients and 2,717 season-matched hospitalized controls. It is not clear from the study which diseases caused the control subjects to be admitted at the time of

their 25-hydroxyvitamin D measurement, which makes it difficult to assess the observations reported. Both female and male patients possessed lower median 25-hydroxyvitamin D concentrations than the control group as a whole (18.6 ng/ml versus 21.5 ng/ml; $p = 0.0016$) and a higher rate of vitamin D deficiency (58.6% versus 42.5%). However, when comparisons were stratified by sex, evidence of sexual dimorphism became apparent, as female patients had equivalent levels of 25-hydroxyvitamin D to females in the control group, whereas male patients were deficient in 25-hydroxyvitamin D relative to male controls (67% versus 49%; $p = 0.0006$). Notably, vitamin D deficiency was progressively lower in males with advancing radiological disease stages ($p = 0.001$). However, these studies are supported by several others that indicate that vitamin D status may be an independent risk factor for the severity of COVID-19 ([1889–1892](#)) and in COVID-19 patients relative to population-based controls ([1893](#)). Indeed, serum concentrations of 25-hydroxyvitamin D above 30 ng/ml, which indicate vitamin D sufficiency, seems to be associated with a reduction in serum C-reactive protein, an inflammatory marker, along with increased lymphocyte levels, which suggests that vitamin D levels may modulate the immune response by reducing risk for cytokine storm in response to SARS-CoV-2 infection ([1893](#)). A study in India determined that COVID-19 fatality was higher in patients with severe COVID-19 and low serum 25-hydroxyvitamin D (mean level 6.2 ng/ml; 97% vitamin D deficient) levels versus asymptomatic non-severe patients with higher levels of vitamin D (mean level 27.9 ng/ml; 33% vitamin D deficient) ([1894](#)). In the same study, vitamin D deficiency was associated with higher levels of inflammatory markers including IL-6, ferritin, and tumor necrosis factor α . Collectively, these studies add to a multitude of observational studies reporting potential associations between low levels of 25-hydroxyvitamin D and COVID-19 incidence and severity ([1887](#), [1892](#), [1893](#), [1895–1901](#)).

Despite the large number of studies establishing a link between vitamin D status and COVID-19 severity, an examination of data from the UK Biobank did not support this thesis ([1902](#), [1903](#)). These analyses examined 25-hydroxyvitamin D concentrations alongside SARS-CoV-2 positivity and COVID-19 mortality in over 340,000 UK Biobank participants. However, these studies have caused considerable debate that will likely be settled following further studies ([1904](#), [1905](#)). Overall, while the evidence suggests that there is likely an association between low serum 25-hydroxyvitamin D and COVID-19 incidence, these studies must be interpreted with caution, as there is the potential for reverse causality, bias, and other confounding factors including that vitamin D deficiency is also associated with numerous pre-existing conditions and risk factors that can increase the risk for severe COVID-19 ([1757](#), [1884](#), [1906](#), [1907](#)).

While these studies inform us of the potential importance of vitamin D sufficiency and the risk of SARS-CoV-2 infection and severe COVID-19, they fail to conclusively determine whether vitamin D supplementation can therapeutically affect the clinical course of COVID-19. In one study, 40 vitamin D deficient asymptomatic or mildly symptomatic participants were either randomized to receive 60,000 IU of cholecalciferol daily for at least 7 days ($n = 16$) or a placebo ($n = 24$) with a target serum 25-hydroxyvitamin D level >50 ng/ml. At day 7, 10 patients achieved >50 ng/ml, followed by another 2 by day 14. By the end of the study, the treatment group had a

greater proportion of vitamin D-deficient participants that tested negative for SARS-CoV-2 RNA, and they had a significantly lower fibrinogen levels, potentially indicating a beneficial effect (1908). A pilot study in Spain determined that early administration of high dose calcifediol (~21,000 IU days 1-2 and ~11,000 IU days 3-7 of hospital admission) with hydroxychloroquine and azithromycin to 50 hospitalized COVID-19 patients significantly reduced ICU admissions and may have reduced disease severity versus hydroxychloroquine and azithromycin alone (1909). Although this study received significant criticism from the National Institute for Health and Care Excellence (NICE) in the UK (1910), an independent follow-up statistical analysis supported the findings of the study with respect to the results of cholecalciferol treatment (1911). Another trial of 986 patients hospitalized for COVID-19 in three UK hospitals administered cholecalciferol ($\geq 280,000$ IU in a time period of 7 weeks) to 151 patients and found an association with a reduced risk of COVID-19 mortality, regardless of baseline 25-hydroxyvitamin D levels (1912). However, a double-blind, randomized, placebo-controlled trial of 240 hospitalized COVID-19 patients in São Paulo, Brazil administered a single 200,000 IU oral dose of vitamin D. At the end of the study, there was a 24 ng/mL difference of 25-hydroxyvitamin D levels in the treatment group versus the placebo group ($p = 0.001$), and 87% of the treatment group were vitamin D sufficient versus ~11% in the placebo group. Supplementation was well tolerated. However, there was no reduction in the length of hospital stay or mortality, and no change to any other relevant secondary outcomes were reported (1913). These early findings are thus still inconclusive with regards to the therapeutic value of vitamin D supplementation. However, other trials are underway, including one trial that is investigating the utility of vitamin D as an immune-modulating agent by monitoring whether administration of vitamin D precipitates an improvement of health status in non-severe symptomatic COVID-19 patients and whether vitamin D prevents patient deterioration (1914). Other trials are examining various factors including mortality, symptom recovery, severity of disease, rates of ventilation, inflammatory markers such as C-reactive protein and IL-6, blood cell counts, and the prophylactic capacity of vitamin D administration (1914–1917). Concomitant administration of vitamin D with pharmaceuticals such as aspirin (1918) and bioactive molecules such as resveratrol (1919) is also under investigation.

The effectiveness of vitamin D supplementation against COVID-19 remains open for debate. All the same, there is no doubt that vitamin D deficiency is a widespread issue and should be addressed not only because of its potential link to SARS-CoV-2 incidence (1920), but also due to its importance for overall health. There is a possibility that safe exposure to sunlight could improve endogenous synthesis of vitamin D, potentially strengthening the immune system. However, sun exposure is not sufficient on its own, particularly in the winter months. Indeed, while the possible link between vitamin D status and COVID-19 is further investigated, preemptive supplementation of vitamin D and encouraging people to maintain a healthy diet for optimum vitamin D status is likely to raise serum levels of 25-hydroxyvitamin D while being unlikely to carry major health risks. These principles seem to be the basis of a number of guidelines issued by some countries and scientific organizations that have advised supplementation of vitamin D during the pandemic. The Académie Nationale de Médecine in France recommends rapid testing of 25-hydroxyvitamin D for people over 60 years old to identify those most at risk

of vitamin D deficiency and advises them to obtain a bolus dose of 50,000 to 100,000 IU vitamin D to limit respiratory complications. It has also recommended that those under 60 years old should take 800 to 1,000 IU daily if they receive a SARS-CoV-2 positive test ([1921](#)/). In Slovenia, doctors have been advised to provide nursing home patients with vitamin D ([1922](#)). Both Public Health England and Public Health Scotland have advised members of the Black, Asian, and minority ethnic communities to supplement for vitamin D in light of evidence that they may be at higher risk for vitamin D deficiency along with other COVID-19 risk factors, a trend that has also been observed in the United States ([1923](#), [1924](#)). However, other UK scientific bodies including the NICE recommend that individuals supplement for vitamin D as per usual UK government advice but warn that people should not supplement for vitamin D solely to prevent COVID-19. All the same, the NICE has provided guidelines for research to investigate the supplementation of vitamin D in the context of COVID-19 ([1925](#)). Despite vitamin D deficiency being a widespread issue in the United States ([1926](#)), the National Institutes of Health have stated that there is “insufficient data to recommend either for or against the use of vitamin D for the prevention or treatment of COVID-19” ([1927](#)/). These are just some examples of how public health guidance has responded to the emerging evidence regarding vitamin D and COVID-19. Outside of official recommendations, there is also evidence that individuals may be paying increased attention to their vitamin D levels, as a survey of Polish consumers showed that 56% of respondents used vitamin D during the pandemic ([1928](#)). However, some companies have used the emerging evidence surrounding vitamin D to sell products that claim to prevent and treat COVID-19, which in one incident required a federal court to intervene and issue an injunction barring the sale of vitamin-D-related products due to the lack of clinical data supporting these claims ([1929](#)). It is clear that further studies and clinical trials are required to conclusively determine the prophylactic and therapeutic potential of vitamin D supplementation against COVID-19. Until such time that sufficient evidence emerges, individuals should follow their national guidelines surrounding vitamin D intake to achieve vitamin D sufficiency.

10.8 Probiotics

Probiotics are “live microorganisms that, when administered in adequate amounts, confer a health benefit on the host” ([1930](#)). Some studies suggest that probiotics are beneficial against common viral infections, and there is modest evidence to suggest that they can modulate the immune response ([1931](#), [1932](#)). As a result, it has been hypothesized that probiotics may have therapeutic value worthy of investigation against SARS-CoV-2 ([1933](#)). Probiotics and next-generation probiotics, which are more akin to pharmacological-grade supplements, have been associated with multiple potential beneficial effects for allergies, digestive tract disorders, and even metabolic diseases through their anti-inflammatory and immunomodulatory effects ([1934](#), [1935](#)). However, the mechanisms by which probiotics affect these various conditions would likely differ among strains, with the ultimate effect of the probiotic depending on the heterogeneous set of bacteria present ([1935](#)). Some of the beneficial effects of probiotics include reducing inflammation by promoting the expression of anti-inflammatory mediators, inhibiting Toll-like receptors 2 and 4, competing directly with pathogens, synthesizing antimicrobial substances or other metabolites, improving

intestinal barrier function, and/or favorably altering the gut microbiota and the brain-gut axis ([1935–1937](#)). It is also thought that lactobacilli such as *Lactobacillus paracasei*, *Lactobacillus plantarum* and *Lactobacillus rhamnosus* have the capacity to bind to and inactivate some viruses via adsorptive and/or trapping mechanisms ([1938](#)). Other probiotic lactobacilli and even non-viable bacterium-like particles have been shown to reduce both viral attachment to host cells and viral titers, along with reducing cytokine synthesis, enhancing the antiviral IFN- α response, and inducing various other antiviral mechanisms ([1938–1946](#)). These antiviral and immunobiotic mechanisms and others have been reviewed in detail elsewhere ([1787](#), [1933](#), [1947](#)). However, there is also a bi-directional relationship between the lungs and gut microbiota known as the gut-lung axis ([1948](#)), whereby gut microbial metabolites and endotoxins may affect the lungs via the circulatory system and the lung microbiota in return may affect the gut ([1949](#)). Therefore, the gut-lung axis may play role in our future understanding of COVID-19 pathogenesis and become a target for probiotic treatments ([1950](#)). Moreover, as microbial dysbiosis of the respiratory tract and gut may play a role in some viral infections, it has been suggested that SARS-CoV-2 may interact with our commensal microbiota [([1787](#)); ([1951](#)); 10.3389/fmicb.2020.01840] and that the lung microbiome could play a role in developing immunity to viral infections ([1952](#)). These postulations, if correct, could lead to the development of novel probiotic and prebiotic treatments. However, significant research is required to confirm these associations and their relevance to patient care, if any.

Probiotic therapies and prophylactics may also confer some advantages for managing symptoms of COVID-19 or risks associated with its treatment. Probiotics have tentatively been associated with the reduction of risk and duration of viral upper respiratory tract infections ([1953–1955](#)). Some meta-analyses that have assessed the efficacy of probiotics in viral respiratory infections have reported moderate reductions in the incidence and duration of infection ([1954](#), [1956](#)). Indeed, randomized controlled trials have shown that administering *Bacillus subtilis* and *Enterococcus faecalis* ([1957](#)), *Lactobacillus rhamnosus* GG ([1958](#)), or *Lactobacillus casei* and *Bifidobacterium breve* with galactooligosaccharides ([1959](#)) via the nasogastric tube to ventilated patients reduced the occurrence of ventilator-associated pneumonia in comparison to the respective control groups in studies of viral infections and sepsis. These findings were also supported by a recent meta-analysis ([1960](#)). Additionally, COVID-19 patients carry a significant risk of ventilator-associated bacterial pneumonia ([1961](#)), but it can be challenging for clinicians to diagnose this infection due to the fact that severe COVID-19 infection presents with the symptoms of pneumonia ([1962](#)). Therefore, an effective prophylactic therapy for ventilator-associated pneumonia in severe COVID-19 patients would carry significant therapeutic value. Additionally, in recent years, probiotics have become almost synonymous with the treatment of gastrointestinal issues due to their supposed anti-inflammatory and immunomodulatory effects ([1963](#)). Notably, gastrointestinal symptoms commonly occur in COVID-19 patients ([1964](#)), and angiotensin-converting enzyme 2, the portal by which SARS-CoV-2 enters human cells, is highly expressed in enterocytes of the ileum and colon, suggesting that these organs may be a potential route of infection ([1965](#), [1966](#)). Indeed, SARS-CoV-2 viral RNA has been detected in human feces ([79](#), [540](#)), and fecal-oral transmission of the virus has not yet been ruled out ([1967](#)). Rectal swabs of

some SARS-CoV-2 positive pediatric patients persistently tested positive for several days despite negative nasopharyngeal tests, indicating the potential for fecal viral shedding (1968). However, there is conflicting evidence for the therapeutic value of various probiotics against the incidence or severity of gastrointestinal symptoms in viral or bacterial infections such as gastroenteritis (1969, 1970). Nevertheless, it has been proposed that the administration of probiotics to COVID-19 patients and healthcare workers may prevent or ameliorate the gastrointestinal symptoms of COVID-19, a hypothesis that several clinical trials are now preparing to investigate (1971, 1972). Other studies are investigating whether probiotics may affect patient outcomes following SARS-CoV-2 infection (1973).

Generally, the efficacy of probiotic use is a controversial topic among scientists. In Europe, EFSA has banned the term probiotics on products labels, which has elicited either criticism for EFSA or support for probiotics from researchers in the field (1930, 1974, 1975). This regulation is due to the hyperbolic claims placed on the labels of various probiotic products, which lack rigorous scientific data to support their efficacy. Overall, the data supporting probiotics in the treatment or prevention of many different disorders and diseases is not conclusive, as the quality of the evidence is generally considered low (1953). However, in the case of probiotics and respiratory infections, the evidence seems to be supportive of their potential therapeutic value. Consequently, several investigations are underway to investigate the prophylactic and therapeutic potential of probiotics for COVID-19. The blind use of conventional probiotics for COVID-19 is currently cautioned against until the pathogenesis of SARS-CoV-2 can be further established (1976). Until clinical trials investigating the prophylactic and therapeutic potential of probiotics for COVID-19 are complete, it is not possible to provide an evidence-based recommendation for their use. Despite these concerns, complementary use of probiotics as an adjuvant therapeutic has been proposed by the Chinese National Health Commission and National Administration of Traditional Chinese Medicine (80). While supply issues prevented the probiotics market from showing the same rapid response to the COVID-19 as some other supplements, many suppliers are reporting growth during the pandemic (1977). Therefore, the public response once again seems to have adopted supplements promoted as bolstering the immune response despite a lack of evidence suggesting they are beneficial for preventing or mitigating COVID-19.

10.9 Discussion

In this review, we report the findings to date of analyses of several dietary supplements and nutraceuticals. While existing evidence suggests potential benefits of n-3 PUFA and probiotic supplementation for COVID-19 treatment and prophylaxis, clinical data is still lacking, although trials are underway. Both zinc and vitamin C supplementation in hospitalized patients seem to be associated with positive outcomes; however, further clinical trials are required. In any case, vitamin C and zinc intake are part of a healthy diet and likely present minimal risk when supplemented, though their potential prophylactic or therapeutic effects against COVID-19 are yet to be determined. On the other hand, mounting evidence from observational studies indicates that there is an association between vitamin D deficiency and COVID-19 incidence has also been supported by meta-analysis (1978).

Indeed, scientists are working to confirm these findings and to determine whether a patient's serum 25-hydroxyvitamin D levels are also associated with COVID-19 severity. Clinical trials are required to determine whether preemptive vitamin D supplementation may mitigate against severe COVID-19. In terms of the therapeutic potential of vitamin D, initial evidence from clinical trials is conflicting but seems to indicate that vitamin D supplementation may reduce COVID-19 severity ([1909](#)). The various clinical trials currently underway will be imperative to provide information on the efficacious use of vitamin D supplementation for COVID-19 prevention and/or treatment.

The purported prophylactic and therapeutic benefits of dietary supplements and nutraceuticals for multiple disorders, diseases, and infections has been the subject of significant research and debate for the last few decades. Inevitably, scientists are also investigating the potential for these various products to treat or prevent COVID-19. This interest also extends to consumers, which led to a remarkable increase of sales of dietary supplements and nutraceuticals throughout the pandemic due to a desire to obtain additional protections from infection and disease. The nutraceuticals discussed in this review, namely vitamin C, vitamin D, n-3 PUFA, zinc, and probiotics, were selected because of potential biological mechanisms that could beneficially affect viral and respiratory infections and because they are currently under clinical investigation. Specifically, these compounds have all been found to influence cellular processes related to inflammation. Inflammation is particularly relevant to COVID-19 because of the negative outcomes (often death) observed in a large number of patients whose immune response becomes hyperactive in response to SARS-CoV-2, leading to severe outcomes such as ARDS and sepsis ([744](#)). Additionally, there is a well-established link between diet and inflammation ([1979](#)), potentially mediated in part by the microbiome ([1980](#)). Thus, the idea that dietary modifications or supplementation could be used to modify the inflammatory response is tied to a broader view of how diet and the immune system are interconnected. The supplements and nutraceuticals discussed here therefore lie in sharp contrast to other alleged nutraceutical or dietary supplements that have attracted during the pandemic, such as colloidal silver ([1981](#)), which have no known nutritional function and can be harmful. Importantly, while little clinical evidence is available about the effects of any supplements against COVID-19, the risks associated with those discussed above are likely to be low, and in some cases, they can be obtained from dietary sources alone.

There are various other products and molecules that have garnered scientific interest and could merit further investigation. These include polyphenols, lipid extracts, and tomato-based nutraceuticals, all of which have been suggested for the potential prevention of cardiovascular complications of COVID-19 such as thrombosis ([1787](#), [1822](#)). Melatonin is another supplement that has been identified as a potential antiviral agent against SARS-CoV-2 using computational methods ([1982](#)), and it has also been highlighted as a potential therapeutic agent for COVID-19 due to its documented antioxidant, anti-apoptotic, immunomodulatory, and anti-inflammatory effects ([1822](#), [1983](#), [1984](#)). Notably, melatonin, vitamin D and zinc have attracted public attention because they were included in the treatment plan of the former President of the United States upon his hospitalization due to COVID-19

(1985). These are just some of the many substances and supplements that are currently under investigation but as of yet lack evidence to support their use for the prevention or treatment of COVID-19. While there is plenty of skepticism put forward by physicians and scientists surrounding the use of supplements, these statements have not stopped consumers from purchasing these products, with one study reporting that online searches for dietary supplements in Poland began trending with the start of the pandemic (1928). Additionally, supplement usage increased between the first and second wave of the pandemic. Participants reported various reasons for their use of supplements, including to improve immunity (60%), to improve overall health (57%), and to fill nutrient gaps in their diet (53%). Other efforts to collect large datasets regarding such behavior have also sought to explore a possible association between vitamin or supplement consumption and COVID-19. An observational analysis of survey responses from 327,720 users of the COVID Symptom Study App found that the consumption of n-3 PUFA supplements, probiotics, multivitamins, and vitamin D was associated with a lower risk of SARS-CoV-2 infection in women but not men after adjusting for potential confounders (1986). According to the authors, the sexual dimorphism observed may in part be because supplements may better support females due to known differences between the male and female immune systems, or it could be due to behavioral and health consciousness differences between the sexes (1986). Certainly, randomized controlled trials are required to investigate these findings further.

Finally, it is known that a patient's nutritional status affects health outcomes in various infectious diseases (1761), and COVID-19 is no different (1759, 1987, 1988). Some of the main risk factors for severe COVID-19, which also happen to be linked to poor nutritional status, include obesity, hypertension, cardiovascular diseases, type II diabetes mellitus, and indeed age-related malnutrition (1757, 1759, 1989). Although not the main focus of this review, it is important to consider the nutritional challenges associated with severe COVID-19 patients. Hospitalized COVID-19 patients tend to report an unusually high loss of appetite preceding admission, some suffer diarrhea and gastrointestinal symptoms that result in significantly lower food intake, and patients with poorer nutritional status were more likely to have worse outcomes and require nutrition therapy (1990). Dysphagia also seems to be a significant problem in pediatric patients that suffered multisystem inflammatory syndrome (1991) and rehabilitating COVID-19 patients, potentially contributing to poor nutritional status (1992). Almost two-thirds of discharged COVID-19 ICU patients exhibit significant weight loss, of which 26% had weight loss greater than 10% (1988). As investigated in this review, hospitalized patients also tend to exhibit vitamin D deficiency or insufficiency, which may be associated with greater disease severity (1978). Therefore, further research is required to determine how dietary supplements and nutraceuticals may contribute to the treatment of severely ill and rehabilitating patients, who often rely on enteral nutrition.

10.10 Conclusions

Despite all the potential benefits of nutraceutical and dietary supplement interventions presented, currently there is a paucity of clinical evidence to support their use for the prevention or mitigation of COVID-19 infection. Nevertheless, optimal nutritional status can prime an individual's immune

system to protect against the effects of acute respiratory viral infections by supporting normal maintenance of the immune system ([1757](#), [1761](#)). Nutritional strategies can also play a role in the treatment of hospitalized patients, as malnutrition is a risk to COVID-19 patients ([1992](#)). Overall, supplementation of vitamin C, vitamin D, and zinc may be an effective method of ensuring their adequate intake to maintain optimal immune function, which may also convey beneficial effects against viral infections due to their immunomodulatory effects. Individuals should pay attention to their nutritional status, particularly their intake of vitamin D, considering that vitamin D deficiency is widespread. The prevailing evidence seems to indicate an association between vitamin D deficiency with COVID-19 incidence and, potentially, severity ([1884](#)). As a result, some international authorities have advised the general public, particularly those at high risk of infection, to consider vitamin D supplementation. However, further well-controlled clinical trials are required to confirm these observations.

Many supplements and nutraceuticals designed for various ailments that are available in the United States and beyond are not strictly regulated ([1993](#)). Consequently, there can be safety and efficacy concerns associated with many of these products. Often, the vulnerable members of society can be exploited in this regard and, unfortunately, the COVID-19 pandemic has proven no different. As mentioned above, the FDA has issued warnings to several companies for advertising falsified claims in relation to the preventative and therapeutic capabilities of their products against COVID-19 ([1994](#)). Further intensive investigation is required to establish the effects of these nutraceuticals, if any, against COVID-19. Until more effective therapeutics are established, the most effective mitigation strategies consist of encouraging standard public health practices such as regular hand washing with soap, wearing a face mask, and covering a cough with your elbow ([1995](#)), along with following social distancing measures, “stay at home” guidelines, expansive testing, and contact tracing ([1996](#), [1997](#)). Indeed, in light of this review, it would also be pertinent to adopt a healthy diet and lifestyle following national guidelines in order to maintain optimal immune health. Because of the broad public appeal of dietary supplements and nutraceuticals, it is important to evaluate the evidence regarding the use of such products. We will continue to update this review as more findings become available.

11 Social Factors Influencing COVID-19 Exposure and Outcomes

11.1 Social Factors Influencing COVID-19 Outcomes

In addition to understanding the fundamental biology of the SARS-CoV-2 virus and COVID-19, it is critical to consider how the broader environment can influence both COVID-19 outcomes and efforts to develop and implement treatments for the disease. The evidence clearly indicates that social environmental factors are critical determinants of individuals' and communities' risks related to COVID-19. There are distinct components to

COVID-19 susceptibility, and an individual's risk can be elevated at one or all stages from exposure to recovery/mortality: an individual may be more likely to be exposed to the virus, more likely to get infected once exposed, more likely to have serious complications once infected, and be less likely to receive adequate care once they are seriously ill. The fact that differences in survival between Black and white patients were no longer significant after controlling for comorbidities and socioeconomic status (type of insurance, neighborhood deprivation score, and hospital where treatment was received) in addition to sex and age ([1998](#)) underscores the relevance of social factors to understanding mortality differences between racial and ethnic groups. Moreover, the Black patients were younger and more likely to be female than white patients, yet still had a higher mortality rate without correction for the other variables ([1998](#)). Here, we outline a few systemic reasons that may exacerbate the COVID-19 pandemic in communities of color.

11.2 Factors Observed to be Associated with Susceptibility

As COVID-19 has spread into communities around the globe, it has become clear that the risks associated with this disease are not equally shared by all individuals or all communities. Significant disparities in outcomes have led to interest in the demographic, biomedical, and social factors that influence COVID-19 severity. Untangling the factors influencing COVID-19 susceptibility is a complex undertaking. Among patients who are admitted to the hospital, outcomes have generally been poor, with rates of admission to the intensive care unit (ICU) upwards of 15% in both Wuhan, China and Italy ([82](#), [1999](#), [2000](#)). However, hospitalization rates vary by location ([2001](#)). This variation may be influenced by demographic (e.g., average age in the area), medical (e.g., the prevalence of comorbid conditions such as diabetes), and social (e.g., income or healthcare availability) factors that vary geographically. Additionally, some of the same factors may influence an individual's probability of exposure to SARS-CoV-2, their risk of developing a more serious case of COVID-19 that would require hospitalization, and their access to medical support. As a result, quantifying or comparing susceptibility among individuals, communities, or other groups requires consideration of a number of complex phenomena that intersect across many disciplines of research. In this section, the term "risk factor" is used to refer to variables that are statistically associated with more severe COVID-19 outcomes. Some are intrinsic characteristics that have been observed to carry an association with variation in outcomes, whereas others may be more functionally linked to the pathophysiology of COVID-19.

11.2.1 Patient Traits Associated with Increased Risk

Two traits that have been consistently associated with more severe COVID-19 outcomes are male sex and advanced age (typically defined as 60 or older, with the greatest risk among those 85 and older ([2002](#))). In the United States, males and older individuals diagnosed with COVID-19 were found to be more likely to require hospitalization ([2003](#), [2004](#)). A retrospective study of hospitalized Chinese patients ([83](#)) found that a higher probability of mortality was associated with older age, and world-wide, population age structure has been found to be an important variable for explaining differences in

outbreak severity ([2005](#)). The CFR for adults over 80 has been estimated upwards of 14% or even 20% ([2006](#)). Male sex has also been identified as a risk factor for severe COVID-19 outcomes, including death ([2007](#), [2008](#), [2009](#)/). Early reports from China and Europe indicated that even though the case rates were similar across males and females, males were at elevated risk for hospital admission, ICU admission, and death ([2008](#)), although data from some US states indicates more cases among females, potentially due to gender representation in care-taking professions ([2010](#)). In older age groups (e.g., age 60 and older), comparable absolute numbers of male and female cases actually suggests a higher rate of occurrence in males, due to increased skew in the sex ratio ([2008](#)). Current estimates based on worldwide data suggest that, compared to females, males may be 30% more likely to be hospitalized, 80% more likely to be admitted to the ICU, and 40% more likely to die as a result of COVID-19 ([2009](#)/). There also may be a compounding effect of advanced age and male sex, with differences time to recovery worst for males over 60 years old relative to female members of their age cohort ([2011](#)).

Both of these risk factors can be approached through the lens of biology. The biological basis for greater susceptibility with age is likely linked to the prevalence of extenuating health conditions such as heart failure or diabetes ([2006](#)). Several hypotheses have been proposed to account for differences in severity between males and females. For example, some evidence suggests that female sex hormones may be protective ([2008](#), [2010](#)). ACE2 expression in the kidneys of male mice was observed to be twice as high as that of females, and a regulatory effect of estradiol on ACE2 expression was demonstrated by removing the gonads and then supplementing with estradiol ([2010](#), [2012](#)). Other work in mice has shown an inverse association between mortality due to SARS-CoV-1 and estradiol, suggesting a protective role for the sex hormone ([2010](#)). Similarly, evidence suggests that similar patterns might be found in other tissues. A preliminary analysis identified higher levels of ACE2 expression in the myocardium of male patients with aortic valve stenosis showed than female patients, although this pattern was not found in controls ([2008](#)). Additionally, research has indicated that females respond to lower doses than males of heart medications that act on the Renin angiotensin aldosterone system (RAAS) pathway, which is shared with ACE2 ([2008](#)). Additionally, several components of the immune response, including the inflammatory response, may differ in intensity and timing between males and females ([2010](#), [2012](#)). This hypothesis is supported by some preliminary evidence showing that female patients who recovered from severe COVID-19 had higher antibody titers than males ([2010](#)). Sex steroids can also bind to immune cell receptors to influence cytokine production ([2008](#)). Additionally, social factors may influence risks related to both age and sex: for example, older adults are more likely to live in care facilities, which have been a source for a large number of outbreaks ([2013](#)), and gender roles may also influence exposure and/or susceptibility due to differences in care-taking and/or risky behaviors (e.g., caring for elder relatives and smoking, respectively) ([2008](#)) among men and women (however, it should be noted that both transgender men and women are suspected to be at heightened risk ([2014](#)).)

11.2.2 Comorbid Health Conditions

A number of pre-existing or comorbid conditions have repeatedly been identified as risk factors for more severe COVID-19 outcomes. Several underlying health conditions were identified at high prevalence among hospitalized patients, including obesity, diabetes, hypertension, lung disease, and cardiovascular disease (2001). Higher Sequential Organ Failure Assessment (SOFA) scores have been associated with a higher probability of mortality (83), and comorbid conditions such as cardiovascular and lung disease as well as obesity were also associated with an increased risk of hospitalization and death, even when correcting for age and sex (2007). Diabetes may increase the risk of lengthy hospitalization (2015) or of death (2015, 2016). (2017) and (2018) discuss possible ways in which COVID-19 and diabetes may interact. Obesity also appears to be associated with higher risk of severe outcomes from SARS-CoV-2 (2019, 2020). Obesity is considered an underlying risk factor for other health problems, and the mechanism for its contributions to COVID-19 hospitalization or mortality is not yet clear (2021). Dementia and cancer were also associated with the risk of death in an analysis of a large number (more than 20,000) COVID-19 patients in the United Kingdom (2007). It should be noted that comorbid conditions are inextricably tied to age, as conditions tend to be accumulated over time, but that the prevalence of individual comorbidities or of population health overall can vary regionally (2022). Several comorbidities that are highly prevalent in older adults, such as COPD, hypertension, cardiovascular disease, and diabetes, have been associated with CFRs upwards of 8% compared to an estimate of 1.4% in people without comorbidities (2006, 2023). Therefore, both age and health are important considerations when predicting the impact of COVID-19 on a population (2022). However, other associations may exist, such as patients with sepsis having higher SOFA scores – in fact, SOFA was developed for the assessment of organ failure in the context of sepsis, and the acronym originally stood for Sepsis-Related Organ Failure Assessment (2024, 2025). Additionally, certain conditions are likely to be more prevalent under or exacerbated by social conditions, especially poverty, as is discussed further below.

11.2.3 Ancestry

A number of studies have suggested associations between individuals' racial and ethnic backgrounds and their COVID-19 risk. In particular, Black Americans are consistently identified as carrying a higher burden of COVID-19 than white Americans (2003, 2004), with differences in the rates of kidney complications from COVID-19 particularly pronounced (89). Statistics from a number of cities indicate significant discrepancies between the proportion of COVID-19 cases and deaths in Black Americans relative to their representation in the general population (2026). In addition to Black Americans, disproportionate harm and mortality from COVID-19 has also been noted in Latino/Hispanic Americans and in Native American and Alaskan Native communities, including the Navajo nation [(2027); (2028); (2029); <https://www.nytimes.com/2020/04/09/us/coronavirus-navajo-nation.html?searchResultPosition=10>; (2030); (2031); (2032)]. In Brazil, indigenous communities likewise carry an increased burden of COVID-19 (2033). In the United Kingdom, nonwhite ethnicity (principally Black or South Asian) was one of several factors found to be associated with a higher risk of death from COVID-19 (2034).

From a genetic standpoint, it is highly unlikely that ancestry itself predisposes individuals to contracting COVID-19 or to experiencing severe COVID-19 outcomes. Examining human genetic diversity indicates variation over a geographic continuum, and that most human genetic variation is associated with the African continent ([2035](#)). African-Americans are also a more genetically diverse group relative to European-Americans, with a large number of rare alleles and a much smaller fraction of common alleles identified in African-Americans ([2036](#)). Therefore, the idea that African ancestry (at the continent level) might convey some sort of genetic risk for severe COVID-19 contrasts with what is known about worldwide human genetic diversity ([2037](#)). The possibility for genetic variants that confer some risk or some protection remains possible, but has not been widely explored, especially at a global level. Research in Beijing of a small number (n=80) hospitalized COVID-19 patients revealed an association between severe COVID-19 outcomes and homozygosity for an allele in the interferon-induced transmembrane protein 3 (IFITM3) gene, which was selected as a candidate because it was previously found to be associated with influenza outcomes in Chinese patients ([435](#)). Genetic factors may also play a role in the risk of respiratory failure for COVID-19 ([464](#), [2038](#), [2039](#)). However, genetic variants associated with outcomes within ancestral groups are far less surprising than genetic variants explaining outcomes between groups. Alleles in *ACE2* and *TMPRSS2* have been identified that vary in frequency among ancestral groups ([2040](#)), but whether these variants are associated with COVID-19 susceptibility has not been explored.

Instead, examining patterns of COVID-19 susceptibility on a global scale that suggest that social factors are of primary importance in predicting mortality. Reports from several sub-Saharan African countries have indicated that the effects of the COVID-19 pandemic have been less severe than expected based on the outbreaks in China and Italy. In Kenya, for example, estimates of national prevalence based on testing blood donors for SARS-CoV-2 antibodies were consistent with 5% of Kenyan adults having recovered from COVID-19 ([2041](#)). This high seroprevalence of antibodies lies in sharp contrast to the low number of COVID-19 fatalities in Kenya, which at the time was 71 out of 2093 known cases ([2041](#)). Likewise, a serosurvey of health care workers in Blantyre City, Malawi reported an adjusted antibody prevalence of 12.3%, suggesting that the virus had been circulating more widely than thought and that the death rate was up eight times lower than models had predicted ([2042](#)). While several possible hypotheses for the apparent reduced impact of COVID-19 on the African continent are being explored, such as young demographics in many places ([2043](#)), these reports present a stark contrast to the severity of COVID-19 in Americans and Europeans of African descent. Additionally, ethnic minorities in the United Kingdom also tend to be younger than white British living in the same areas, yet the burden of COVID-19 is still more serious for minorities, especially people of Black Caribbean ancestry, both in absolute numbers and when controlling for age and location ([2044](#)). Furthermore, the groups in the United States and United Kingdom that have been identified as carrying elevated COVID-19 burden, namely Black American, indigenous American, and Black and South Asian British, are quite distinct in their position on the human ancestral tree. What is shared across these groups is instead a history of disenfranchisement under colonialism and ongoing systematic racism. A large analysis of over 11,000 COVID-19 patients hospitalized in 92 hospitals

across U.S. states revealed that Black patients were younger, more often female, more likely to be on Medicaid, more likely to have comorbidities, and came from neighborhoods identified as more economically deprived than white patients ([1998](#)). This study reported that when these factors were accounted for, the differences in mortality between Black and white patients were no longer significant. Thus, the current evidence suggests that the apparent correlations between ancestry and health outcomes must be examined in the appropriate social context.

11.3 Environmental Influences on Susceptibility

11.3.1 Exposure to COVID-19

Social distancing has emerged as one of the main social policies used to manage the COVID-19 epidemic in many countries. Many governments issued stay-at-home orders, especially in the initial months of the crisis. However, data clearly indicates that these orders impacted different socioeconomic groups differently. In U.S. counties with and without stay-at-home orders, smartphone tracking indicated a significant decrease in the general population's mobility in April relative to February through March of 2020 (-52.3% and -60.8%, respectively) ([2045](#)). A linear relationship was observed between counties' reduction in mobility and their wealth and health, as measured by access to health care, food security, income, space, and other factors ([2045](#)). Counties with greater reductions in mobility were also found to have much lower child poverty and household crowding and to be more racially segregated, and to have fewer youth and more elderly residents ([2045](#)). Similar associations between wealth and decreased mobility were observed in cellphone GPS data from Colombia, Indonesia, and Mexico collected between January and May 2020 ([2046](#)), as well as in a very large data set from several US cities ([2047](#)). These disparities in mobility are likely to be related to the role that essential workers have played during the pandemic. Essential workers are disproportionately likely to be female, people of color, immigrants, and to have an income below 200% of the poverty line ([2048](#)). Black Americans in particular are over-represented among front-line workers and in professions where social distancing is infeasible ([2049](#)). Health care work in particular presents an increased risk of exposure to SARS-CoV-2 ([2049–2053](#)). In the United Kingdom, (South) Asians are more likely than their white counterparts to be medical professionals ([2044](#)), although BAME medical professionals are still disproportionately represented in the proportion of National Health Service staff deaths ([2054](#)). Similar trends have been reported for nurses, especially nurses of color, in the United States ([2055](#) [/-/files/graphics/0920_Covid19_SinsOfOmission_Data_Report.pdf](#)). Furthermore, beyond the risks associated with work itself, use of public transportation may also impact COVID-19 risk ([2056](#)). The socioeconomic and racial/ethnic gaps in who is working on the front lines of the pandemic make it clear that socioeconomic privilege is likely to decrease the probability of exposure to SARS-CoV-2.

Increased risk of exposure can also arise outside the workplace. Nursing homes and skilled nursing facilities received attention early on as high-risk locations for COVID-19 outbreaks ([2057](#)). Prisons and detention centers also

confer a high risk of exposure or infection ([2058](#), [2059](#)). Populations in care facilities are largely older adults, and in the United States, incarcerated people are more likely to be male and persons of color, especially Black ([2060](#)). Additionally, multi-generational households are less common among non-Hispanic white Americans than people of other racial and ethnic backgrounds ([2061](#)), increasing the risk of exposure for more susceptible family members. Analysis suggests that household crowding may also be associated with increased risk of COVID-19 exposure ([2045](#)), and household crowding is associated with poverty ([2062](#)). Forms of economic insecurity like housing insecurity, which is associated with poverty and more pronounced in communities subjected to racism ([2063](#), [2064](#)), would be likely to increase household crowding and other possible sources of exposure. As a result, facets of systemic inequality such as mass incarceration of Black Americans and poverty are likely to increase the risk of exposure outside of the workplace.

11.3.2 Severity of COVID-19 Following Exposure

Following exposure to SARS-CoV-2, the likelihood that an individual develops COVID-19 and the severity of the disease presentation can be influenced by a number of social factors. As discussed above, a number of patient characteristics are associated with the likelihood of severe COVID-19 symptoms. In some cases, these trends run counter to those expected given rates of exposure: for example, although women are more likely to be exposed, men are more likely to be diagnosed with, hospitalized from, or die from COVID-19 ([2010](#)). In the case of comorbid conditions and racial/ethnic demographics, however, social factors are highly likely to modulate or at least influence the apparent association between these traits and the increased risk from COVID-19. In particular, the comorbidities and racial/ethnic correlates of severe COVID-19 outcomes suggest that poverty confers additional risk for COVID-19.

In order to explore the relationship between poverty and COVID-19 outcomes, it is necessary to consider how poverty impacts biology. In particular, we focus on the United States and the United Kingdom. Comorbidities that increase risk for COVID-19, including obesity, type II diabetes, hypertension, and cardiovascular disease, are known to be intercorrelated ([2065](#)). Metabolic conditions related to heightened inflammation, like obesity, type II diabetes, and hypertension, are more strongly associated with negative COVID-19 outcomes than other comorbid conditions, such as chronic heart disease ([2066](#)). As discussed above, dysregulated inflammation characteristic of cytokine release syndrome is one of the greatest concerns for COVID-19-related death. Therefore, it is possible that chronic inflammation characteristic of these metabolic conditions predisposes patients to COVID-19-related death ([2066](#)). The association between these diseases and severe COVID-19 outcomes is a concern from a health equity perspective because poverty exposes people to “obesogenic” conditions ([2067](#)) and is therefore unsurprisingly associated with higher incidence of obesity and associated disorders ([2068](#)). Furthermore, cell phone GPS data suggests that lower socioeconomic status may also be associated with decreased access to healthy food choices during the COVID-19 pandemic ([2069](#), [2070](#)), suggesting that health-related risk factors for COVID-19 may be exacerbated as the pandemic continues ([2071](#)). Chronic

inflammation is a known outcome of chronic stress (e.g., [\(2072–2075\)](#)). Therefore, the chronic stress of poverty is likely to influence health broadly (as summarized in [\(2076\)](#)) and especially during the stress of the ongoing pandemic.

A preprint [\(2077\)](#) provided observational evidence that geographical areas in the United States that suffer from worse air pollution by fine particulate matter have also suffered more COVID-19 deaths per capita, after adjusting for demographic covariates. Although lack of individual-level exposure data and the impossibility of randomization make it difficult to elucidate the exact causal mechanism, this finding would be consistent with similar findings for all-cause mortality (e.g., [\(2078\)](#)). Exposure to air pollution is associated with both poverty (e.g., [\(2079\)](#)) and chronic inflammation [\(2080\)](#). Other outcomes of environmental racism, such as the proximity of abandoned uranium mines to Navajo land, can also cause respiratory illnesses and other health issues [\(2032\)](#). Similarly, preliminary findings indicate that nutritional status (e.g., vitamin D deficiency [\(1893\)](#)) may be associated with COVID-19 outcomes, and reduced access to grocery stores and fresh food often co-occurs with environmental racism [\(2032, 2081\)](#). Taken together, the evidence suggests that low-income workers who face greater exposure to SARS-CoV-2 due to their home or work conditions are also more likely to face environmental and social stressors associated with increased inflammation, and therefore with increased risk from COVID-19. In particular, structural racism can play an important role on disease severity after SARS-CoV-2 exposure, due to consequences of racism which include an increased likelihood of poverty and its associated food and housing instability. COVID-19 can thus be considered a “syndemic”, or a synergistic interaction between several epidemics [\(2082\)](#). As a result, it is not surprising that people from minoritized backgrounds and/or with certain pre-existing conditions are more likely to suffer severe effects of COVID-19, but these “risk factors” are likely to be causally linked to poverty [\(2083\)](#).

11.3.3 Access to Treatment

Finally, COVID-19 outcomes can be influenced by access to healthcare. Receiving care for COVID-19 can, but does not always, include receiving a positive test for the SARS-CoV-2 virus. For example, it is common to see treatment guidelines for suspected cases regardless of whether the presence of SARS-CoV-2 has been confirmed (e.g., [\(2084\)](#)). Whether and where a patient is diagnosed can depend on their access to testing, which can vary both between and within countries. In the United States, it is not always clear whether an individual will have access to free testing [\(2085, 2086\)](#). The concern has been raised that more economic privilege is likely to correspond to increased access to testing, at least within the United States [\(2087\)](#). This is supported by the fact that African Americans seem to be more likely to be diagnosed in the hospital, while individuals from other groups were more likely to have been diagnosed in ambulatory settings in the community [\(2003\)](#). Any delays in treatment are a cause for concern [\(2087\)](#), which could potentially be increased by an inability to acquire testing because in the United States, insurance coverage for care received can depend on a positive test [\(2088\)](#).

Another important question is whether patients with moderate to severe cases are able to access hospital facilities and treatments, to the extent that they have been identified. Early findings from China as of February 2020 suggested the COVID-19 mortality rate to be much lower in the most developed regions of the country ([2089](#)), although reported mortality is generally an estimate of CFR, which is dependent on rates of testing. Efforts to make treatment accessible for all confirmed and suspected cases of COVID-19 in China are credited with expanding care to people with fewer economic resources ([2090](#)). In the United States, access to healthcare varies widely, with certain sectors of the workforce less likely to have health insurance; many essential workers in transportation, food service, and other frontline fields are among those likely to be uninsured or underinsured ([2087](#)). As of 2018, Hispanic Americans of all races were much less likely to have health insurance than people from non-Hispanic backgrounds ([2091](#)). Therefore, access to diagnostics and care prior to the development of severe COVID-19 is likely to vary depending on socioeconomic and social factors, many of which overlap with the risks of exposure and of developing more severe COVID-19 symptoms. This discrepancy ties into concerns about broad infrastructural challenges imposed by COVID-19. A major concern in many countries has been the saturation of healthcare systems due to the volume of COVID-19 hospitalizations (e.g., [320](#)). Similarly, there have been shortages of supplies such as ventilators that are critical to the survival of many COVID-19 patients, leading to extensive ethical discussions about how to allocate limited resources among patients ([2092–2095](#)). Although it is generally considered unethical to consider demographic factors such as age, sex, race, or ethnicity while making such decisions, and ideally this information would not be shared with triage teams tasked with allocating limited resources among patients ([2096](#)), there are substantial concerns about implicit and explicit biases against older adults ([2097](#)), premature infants ([2098](#)), and people with disabilities or comorbidities ([2096](#), [2099](#), [2100](#)). Because of the greater burden of chronic disease in populations subjected to systemic racism, algorithms intended to be blind to race and ethnicity could, in fact, reinforce systemic inequalities caused by structural racism ([2101–2103](#)). Because of this inequality, it has been argued that groups facing health disparities should be prioritized by these algorithms ([2104](#)). This approach would carry its own ethical concerns, including the fact that many resources that need to be distributed do not have well-established risks and benefits ([2104](#)).

As the pandemic has progressed, it has become clear that ICU beds and ventilators are not the only limited resources that needs to be allocated, and, in fact, the survival rate for patients who receive mechanical ventilation is lower than these discussions would suggest ([2105](#)). Allocation of interventions that may reduce suffering, including palliative care, has become critically important ([2105](#), [2106](#)). The ambiguities surrounding the risks and benefits associated with therapeutics that have been approved under emergency use authorizations also present ethical concerns related to the distribution of resources ([2104](#)). For example, remdesivir, discussed above, is currently available for the treatment of COVID-19 under compassionate use guidelines and through expanded access programs, and in many cases has been donated to hospitals by Gilead ([2107](#), [2108](#)). Regulations guiding the distribution of drugs in situations like these typically do not address how to determine which patients receive them ([2108](#)).

Prioritizing marginalized groups for treatment with a drug like remdesivir would also be unethical because it would entail disproportionately exposing these groups to a therapeutic that may or not be beneficial (2104). On the other hand, given that the drug is one of the most promising treatments available for many patients, using a framework that tacitly feeds into structural biases would also be unethical. At present, the report prepared for the Director of the CDC by Ethics Subcommittee of the CDC fails to address the complexity of this ethical question given the state of structural racism in the United States, instead stating that “prioritizing individuals according to their chances for short-term survival also avoids ethically irrelevant considerations, such as race or socioeconomic status” (2109). In many cases, experimental therapeutics are made available only through participation in clinical trials (2110). However, given the history of medical trials abusing minority communities, especially Black Americans, there is a history of unequal representation in clinical trial enrollment (2110). As a result, the standard practice of requiring enrollment in a clinical trial in order to receive experimental treatment may also reinforce patterns established by systemic racism.

11.3.4 Access to and Representation in Clinical Trials

Experimental treatments are often made available to patients primarily or even exclusively through clinical trials. The advantage of this approach is that clinical trials are designed to collect rigorous data about the effects of a treatment on patients. The disadvantage is that access to clinical trials is not equal among all people who suffer from a disease. Two important considerations that can impact an individual's access to clinical trials are geography and social perceptions of clinical trials. For the first, the geographic distribution of trial recruitment efforts are typically bounded and can vary widely among different locations, and for the second, the social context of medical interactions can impact strategies for and the success of outreach to different communities. Differential access to clinical trials raises concerns because it introduces biases that can influence scientific and medical research on therapeutics and prophylactics broadly. Concerns about bias in clinical trials need to address both trial recruitment and operation. In the present crisis, such biases are particularly salient because COVID-19 is a disease of global concern. Treatment is needed by people all over the world, and clinical research that characterizes treatment outcomes in a variety of populations is critically important.

Global representation in clinical trials is important to ensuring that experimental treatments are available equally to COVID-19 patients who may need them. The advantage to a patient of participation in a clinical trial is that they may receive an experimental treatment they would not have been able to access otherwise. The potential downsides of participation include that the efficacy and side effects of such treatments are often poorly characterized and that patients who enroll in clinical trials will in some cases run the risk of being assigned to a placebo condition where they do not receive the treatment but miss out on opportunities to receive other treatments. The benefits and burdens of clinical trials therefore need to be weighed carefully to ensure that they don't reinforce existing health disparities. The WHO Director-General Tedros Adhanom Ghebreyesus stated his condemnation of utilizing low and middle income countries as test subjects for clinical trials,

yet having highly developed countries as the majority of clinical trial representation is also not the answer (2111). Figure 13 showcases two choropleths detailing COVID-19 clinical trial recruitment by country. China, the United States, and France are among the countries with the most clinical trial recruiting for trials with single-country enrollment. Many countries have little to no clinical trial recruiting, with the continents of Africa and South America much less represented than Asia, Europe, and North America. Trials that recruit across multiple countries do appear to broaden geographic representation, but these trials seem to be heavily dominated by the United States and European Union.

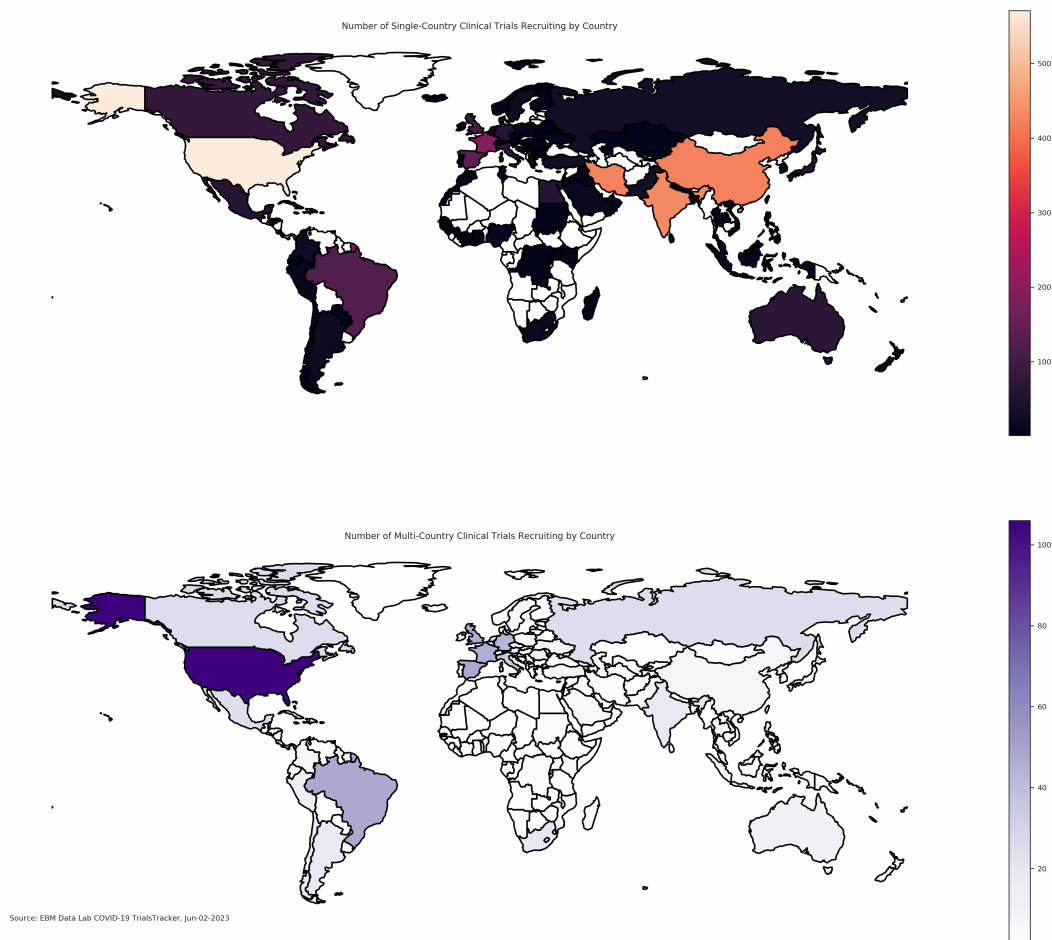


Figure 13: Geographic distribution of COVID-19 clinical trials. The density of clinical trials is reported at the country level. As of December 31, 2020, there are 6,987 trials in the University of Oxford Evidence-Based Medicine Data Lab's COVID-19 TrialsTracker (739), of which 3,962 are interventional. The top figure demonstrates the density of interventional trials recruiting only from a singular country, while the bottom shows the distribution of recruitment for interventional trials that involve more than one country.

A few different concerns arise from this skewed geographic representation in clinical trial recruitment. First, treatments such as remdesivir that are promising but primarily available to clinical trial participants are unlikely to be accessible by people in many countries. Second, it raises the concern that the findings of clinical trials will be based on participants from many of the wealthiest countries, which may lead to ambiguity in whether the findings can be extrapolated to COVID-19 patients elsewhere. Especially with the global nature of COVID-19, equitable access to therapeutics and vaccines has been a concern at the forefront of many discussions about policy (e.g., (2112), yet data like that shown in Figure 13 demonstrates that accessibility is likely to be a significant issue. Another concern with the heterogeneous

international distribution of clinical trials is that the governments of countries leading these clinical trials might prioritize their own populations once vaccines are developed, causing unequal health outcomes (2113). Additionally, even within a single state in the United States (Maryland), geography was found to influence the likelihood of being recruited into or enrolled in a clinical trial, with patients in under-served rural areas less likely to enroll (2114). Thus, geography both on the global and local levels may influence when treatments and vaccines are available and who is able to access them. Efforts such as the African Union's efforts to coordinate and promote vaccine development (2115) are therefore critical to promoting equity in the COVID-19 response.

Even when patients are located within the geographic recruitment area of clinical trials, however, there can still be demographic inequalities in enrollment. When efforts are made to ensure equal opportunity to participate in clinical trials, there is no significant difference in participation among racial/ethnic groups (2116). However, within the United States, real clinical trial recruitment numbers have indicated for many years that racial minorities, especially African-Americans, tend to be under-represented (e.g., (2117–2120)). This trend is especially concerning given the disproportionate impact of COVID-19 on African-Americans. Early evidence suggests that the proportion of Black, Latinx, and Native American participants in clinical trials for drugs such as remdesivir is much lower than the representation of these groups among COVID-19 patients (2121).

One proposed explanation for differences among racial and ethnic groups in clinical trial enrollment refers to different experiences in healthcare settings. While some plausible reasons for the disparity in communication between physicians and patients could be a lack of awareness and education, mistrust in healthcare professionals, and a lack of health insurance (2116), a major concern is that patients from certain racial and ethnic groups are marginalized even while seeking healthcare. In the United States, many patients experience “othering” from physicians and other medical professionals due to their race or other external characteristics such as gender (e.g., (2122)). Many studies have sought to characterize implicit biases in healthcare providers and whether they affect their perceptions or treatment of patients. A systematic review that examined 37 such studies reported that most (31) identified racial and/or ethnic biases in healthcare providers in many different roles, although the evidence about whether these biases translated to different attitudes towards patients was mixed (2123), with similar findings reported by a second systematic review (2124). However, data about real-world patient outcomes are very limited, with most studies relying on clinical vignette-based exercises (2123), and other analyses suggest that physician implicit bias could impact the patient's perception of the negativity/positivity of the interaction regardless of the physician's explicit behavior towards the patient (2125). Because racism is a common factor in both, negative patient experiences with medical professionals are likely to compound other issues of systemic inequality, such as a lack of access to adequate care, a lack of insurance, or increased exposure to SARS-CoV-2 (2126). Furthermore, the experience of being othered is not only expected to impact patients' trust in and comfort with their provider, but also may directly impact whether or not the patient is offered the opportunity to participate in a clinical trial at all. Some studies suggest communication

between physicians and patients impacts whether or not a physician offers a patient participation in a clinical trial. For example, researchers utilized a linguistic analysis to assess mean word count of phrases related to clinical trial enrollment, such as voluntary participation, clinical trial, etc. (2116). The data indicated that the mean word count of the entire visit was 1.5 times more for white patients in comparison to Black patients. In addition, the greatest disparity between white and Black patients' experience was the discussion of risks, with over 2 times as many risk-related words spoken with white patients than Black patients (2116). The trends observed for other clinical trials raise the concern that COVID-19 clinical trial information may not be discussed as thoroughly or as often with Black patients compared to white patients.

These discrepancies are especially concerning given that many COVID-19 treatments are being or are considered being made available to patients prior to FDA approval through Emergency Use Authorizations. In the past, African-Americans have been over-represented relative to national demographics in use of the FDA's Exception From Informed Consent (EFIC) pathway (2127). Through this pathway, people who are incapacitated can receive an experimental treatment even if they are not able to consent and there is not sufficient time to seek approval from an authorized representative. This pathway presents concerns, however, when it is considered in the context of a long history of systematic abuses in medical experimentation where informed consent was not obtained from people of color, such as the Tuskegee syphilis experiments (2128). While the goal of EFIC approval is to provide treatment to patients who urgently need it, the combination of the ongoing legacy of racism in medicine renders this trend concerning. With COVID-19, efforts to prioritize people who suffer from systemic racism are often designed with the goal of righting some of these inequalities (e.g., (2129)), but particular attention to informed consent will be imperative in ensuring these trials are ethical given that the benefits and risks of emerging treatments are still poorly characterized. Making a substantial effort to run inclusive clinical trials is also important because of the possibility that racism could impact how a patient responds to a treatment. For example, as discussed above, dexamethasone has been identified as a promising treatment for patients experiencing cytokine release syndrome, but the mechanism of action is tied to the stress response. A study from 2005 reported that Black asthma patients showed reduced responsiveness to dexamethasone in comparison to white patients and suggested Black patients might therefore require higher doses of the drug (2130). In the context of chronic stress caused by systemic racism, this result is not surprising: chronic stress is associated with dysregulated production of glucocorticoids (2131) and glucocorticoid receptor resistance (2132). However, it underscores the critical need for treatment guidelines to take into account differences in life experience, which would be facilitated by the recruitment of patients from a wide range of backgrounds. Attention to the social aspects of clinical trial enrollment must therefore be an essential component of the medical research community's response to COVID-19.

11.4 Conclusions and Future Directions

As the COVID-19 pandemic evolves, the scientific community's response will be critical for identifying potential pharmacological and biotechnological developments that may aid in combating the virus and the disease it causes. However, this global crisis highlights the importance of mounting a response based on collaboration among a wide variety of disciplines. Understanding the basic science of the virus and its pathogenesis is imperative for identifying and envisioning possible diagnostic and therapeutic approaches; understanding how social factors can influence outcomes and shape implementation of a response is critical to disseminating any scientific advancements. Summarizing such a complex and ever-changing topic presents a number of challenges. This review represents the effort of over 50 contributors to distill and interpret the available information. However, this text represents a dynamic and evolving document, and we welcome continued contributions from all researchers who have insights into how these topics intersect. A multidisciplinary perspective is critical to understanding this evolving crisis, and in this review we seek to use open science tools to coordinate a response among a variety of researchers. We intend to publish additional updates as the situation evolves.

12 An Open-Publishing Response to the COVID-19 Infodemic

12.1 ABSTRACT

The COVID-19 pandemic catalyzed the rapid dissemination of papers and preprints investigating the disease and its associated virus, SARS-CoV-2. The multifaceted nature of COVID-19 demands a multidisciplinary approach, but the urgency of the crisis combined with the need for social distancing measures present unique challenges to collaborative science. We applied a massive online open publishing approach to this problem using Manubot. Through GitHub, collaborators summarized and critiqued COVID-19 literature, creating a review manuscript. Manubot automatically compiled citation information for referenced preprints, journal publications, websites, and clinical trials. Continuous integration workflows retrieved up-to-date data from online sources nightly, regenerating some of the manuscript's figures and statistics. Manubot rendered the manuscript into PDF, HTML, LaTeX, and DOCX outputs, immediately updating the version available online upon the integration of new content. Through this effort, we organized over 50 scientists from a range of backgrounds who evaluated over 1,500 sources and developed seven literature reviews. While many efforts from the computational community have focused on mining COVID-19 literature, our project illustrates the power of open publishing to organize both technical and non-technical scientists to aggregate and disseminate information in response to an evolving crisis.

KEYWORDS

COVID-19, living document, open publishing, open-source, data integration, manubot

12.2 INTRODUCTION

Coronavirus Disease 2019 (COVID-19) caused a worldwide public health crisis that has reshaped many aspects of society. The scientific community has, in turn, devoted significant attention and resources towards COVID-19 and the associated virus, SARS-CoV-2, resulting in the release of data and publications at a rate and scale never previously seen for a single topic. Over 20,000 articles about COVID-19 were released in the first four months of the pandemic (2133), causing an “infodemic” (2133, 2134). The COVID-19 Open Research Dataset (CORD-19) (2135), which was developed in part with the goal of training machine learning algorithms on COVID-19-related text, illustrates the growth of related scholarly literature (Figure 14). This resource was developed by querying several sources for terms related to SARS-CoV-2 and COVID-19, as well as the coronaviruses SARS-CoV-1 and MERS-CoV and their associated diseases (2135). CORD-19 contained 1056660 manuscripts as of 2022-06-02. Additional curation by CoronaCentral (2136) has produced, at present, a set of over 180,000 publications particularly relevant to COVID-19 and closely related viruses. Despite many advances in understanding the virus and the disease, there are also downsides to the availability of so much information. “Excessive publication” has been recognized as a concern for over forty years (2137) and has been discussed with respect to the COVID-19 literature (992). Any effort to synthesize, summarize, and contextualize COVID-19 research will face a vast corpus of potentially relevant material.

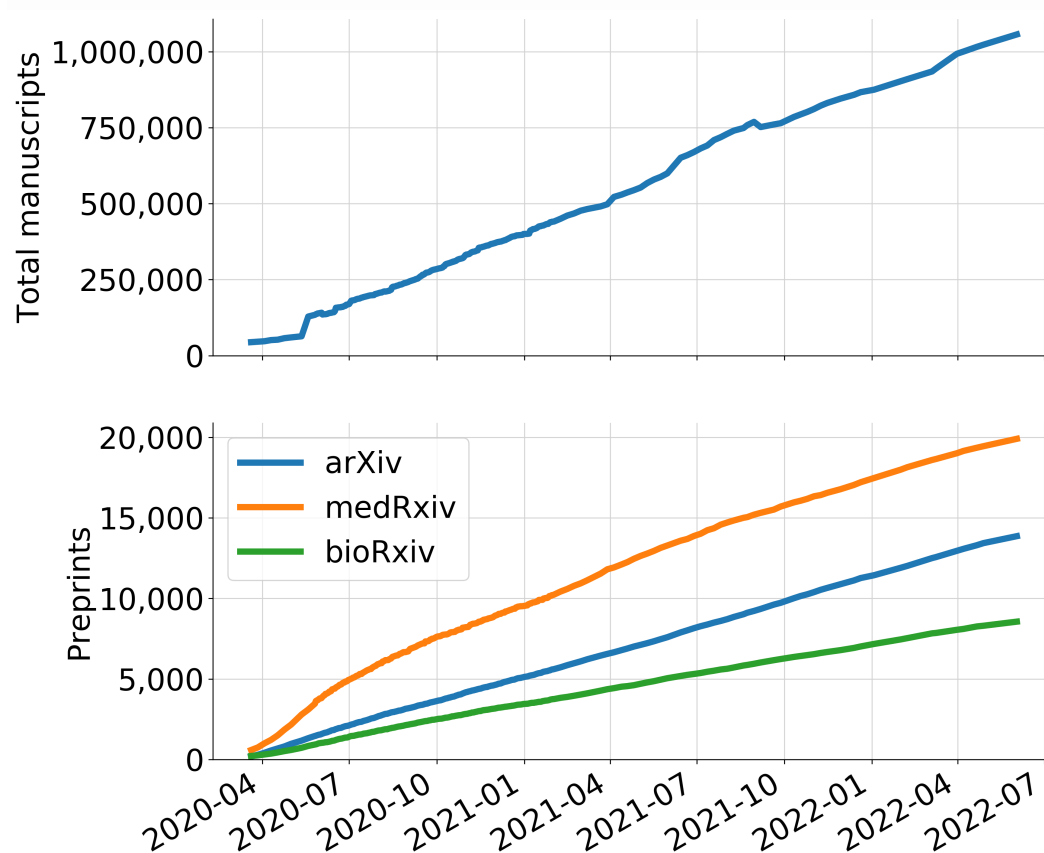


Figure 14: Growth of the CORD-19 dataset. The number of articles has proliferated, with both traditional and preprint manuscripts in the corpus. The first release (March 16, 2020) contained 28,000 documents (2135). As of 2022-06-02, this had increased to 1056660 articles. Of these, 42343 are preprints from *arXiv*, *medRxiv*, and *bioRxiv*.

Information was released rapidly by both traditional publishers and preprint servers, and many papers faced subsequent scrutiny. The number of COVID-19 papers retracted may be higher, and potentially much higher, than is typical, although a thorough investigation of this question requires more time to elapse (2138, 2139). Many papers and preprints are also associated with corrections or expressions of concern¹ (2139). Preprints are released prior to peer review, but some traditional publishing venues have fast-tracked COVID-19 papers through peer review, leading to questions about whether they are held to typical standards (2140). Therefore, evaluating the COVID-19 literature requires not only digesting available information but also monitoring subsequent changes.

Because of the fast-moving nature of the topic, many efforts to summarize and synthesize the COVID-19 literature have been undertaken. These efforts include newsletters² (2141), web portals³ (2142) or the now-defunct <http://covidpreprints.com>⁴, comments on preprint servers⁵ (2143), and even a journal⁶. However, the explosive rate of publication presents challenges for such efforts, many of which are no longer active. Similarly, many literature reviews have been written on the available COVID-19 literature (2144–2148), but static reviews quickly become outdated as new research is released or existing research is retracted or superseded. One example is a review of topics in COVID-19 research including vaccine development (2148). This review was published on July 10, 2020, four days before Moderna released the surprisingly promising results of their phase 1 trial (1119) that changed expectations surrounding vaccines. Therefore, the COVID-19 publishing climate presented a challenge where curation of the literature by a diverse group of experts in a format that could respond quickly to high-volume, high-velocity information was desirable.

We therefore sought to develop a platform for scientific discussion and collaboration around COVID-19 by adapting open publishing infrastructure to accommodate the scale of COVID-19 publishing. Recent advances in open publishing have created an infrastructure that facilitates distributed, version-controlled collaboration on manuscripts (2149). Manubot (2149) is a collaborative framework developed to adapt open-source software development techniques and version control for manuscript writing. With Manubot, manuscripts are managed and maintained using GitHub, a popular, online version control interface. We selected Manubot because it offers several advantages over comparable collaborative writing platforms such as Authorea, Overleaf, Google Docs, Word Online, or wikis (2149). Citation-by-identifier ensures consistent reference metadata standards that would be difficult to maintain manually in a manuscript with dozens of authors and over 1,500 citations. Manubot's pull request-based contribution model balances the goals of making the project open to everyone and maintaining scientific accuracy. All contributions are reviewed, discussed, and formally approved on GitHub before text updates appear in the public-facing manuscript⁷. Continuous integration (CI) seamlessly combines author-produced text and figures with automatically generated and updated statistics and figures derived from external data sources and the manuscript's own content. In addition, the authors who initially launched this project included Manubot developers who had prior successes using Manubot for massively open and traditional manuscripts.

Collaboration via massively open online papers has been identified as a strategy for promoting inclusion and interdisciplinary thought ([2150](#)). However, the Manubot workflow can be intimidating to contributors who are not well-versed in git ([2150](#)). The synthesis and discussion of the emerging literature by biomedical scientists and clinicians is imperative to a robust interpretation of COVID-19 research. Such efforts in biology often rely on What You See Is What You Get tools such as Google Docs, despite the significant limitations of these platforms in the face of excessive publication. We recognized that the problem of synthesizing the COVID-19 literature lent itself well to the Manubot platform, but that the potential technical expertise required to work with Manubot presented a barrier to domain experts.

Here, we describe the adaptation of Manubot to facilitate collaboration in the extreme case of the COVID-19 infodemic, with the objective of developing a centralized platform for summarizing and synthesizing a massive amount of preprints, news stories, journal publications, and data. Unlike prior collaborations built on Manubot, most contributors to the COVID-19 collaborative literature review came from biology or medicine. The members of the COVID-19 Review Consortium consolidated information about the virus in the context of related viruses and to synthesize rapidly emerging literature. Manubot provided the infrastructure to manage contributions from the community and create a living, scholarly document integrating data from multiple sources. Its back-end allowed biomedical scientists to sort and distill informative content out of the overwhelming flood of information ([2151](#)) in order to provide a resource that would be useful to the broader scientific community. This case study demonstrates the value of open collaborative writing tools such as Manubot to emerging challenges. Because it is open source software, we were able to adapt and customize Manubot to flexibly meet the needs of COVID-19 review. Recording the evolution of information over time and assembling a resource that auto-updated in response to the evolving crisis revealed the particular value that Manubot holds for managing rapid changes in scientific thought.

12.3 METHODS

12.3.1 Contributor Recruitment and Roles

First, it was necessary to establish Manubot as a platform accessible to researchers with limited experience working version control, given that this is not typically emphasized in biology and medicine ([2152–2154](#)). Contributors were recruited primarily by word of mouth and on Twitter, and we also collaborated with existing efforts to train early-career researchers. We invited potential collaborators to contribute a short introduction on a GitHub issue in order to collect information about participants and provide an introduction to working with GitHub issues. Interested participants were encouraged to contribute in several ways. One option was to catalog articles of interest as issues. We developed a standardized set of questions for contributors to consider when evaluating an article following a framework often used for assessing medical literature. This approach emphasizes examining the methods used, assignment (whether the study was observational or randomized), assessment, results, interpretation, and how well the study extrapolates ([2155](#)). Contributors were also invited to

contribute or edit text using GitHub's pull request system. These contributions were not strictly defined and could range from minor corrections to punctuation and grammar to large-scale additions of text. Finally, a small number of contributors (the authors of this paper) contributed technical expertise, either through the development of standardized approaches to the evaluation of papers based on the MAARIE Framework (2156), the writing of code to generate manuscript figures, or the addition of features to Manubot. All of these additions were also submitted as pull requests, either to the COVID-19 review repository or to an external repository, as appropriate.

Each pull request was reviewed and approved by at least one other contributor before being merged into the main branch. We tagged potential reviewers based on the introductions they had contributed in order to encourage participation. Authorship was determined based on the Contributor Roles Taxonomy⁸. Due to the permeability of ideas among different sections, contributors to a specific manuscript were recognized with masthead authorship, while all contributors to the project were recognized with consortium authorship on all papers. Emphasizing the use of issues and pull requests was designed to encourage authors with and without git experience to discuss papers and provide feedback (both formal and informal) on proposed text additions or changes. We also used the Gitter chat platform⁹ to promote informal questions and sharing of information among collaborators.

12.3.2 Utilization and Expansion of Manubot

Applying Manubot's existing capabilities allowed us to confront several challenges common in large-scale collaborations, such as maintaining a record of contributions that allowed us to allocate credit appropriately or to contact the original author if questions arose. Additionally, an up-to-date version of the content was available at all times online in HTML¹⁰ or PDF format¹¹. This approach also allowed us to minimize the demand on authors to curate and sync bibliographic resources. Manubot provides the functionality to create a bibliography using digital object identifiers (DOIs), website URLs, or other identifiers such as PubMed identifiers and arXiv IDs. The author can insert a citation in-line using a format such as `[@doi:10.1371/journal.pcbi.1007128]`. Manubot then obtains reference metadata, exports the citations as Citation Style Language JSON Data Items, and renders the bibliographic information needed to generate the references section (2149). This approach allows multiple authors to work on a piece of text without needing to make manual adjustments to the reference lists.

Due to the needs of this project, several new features were implemented in Manubot. Because of the ever-evolving nature of the COVID-19 crisis, figures and statistics in the text quickly became outdated. To address this concern, Manubot and GitHub's CI features were used to create figures that integrated online data sources and to dynamically update information, such as the current number of active COVID-19 clinical trials (3), within the text of the manuscripts (Figure 15). GitHub Actions runs a nightly workflow to update these external data and regenerate the statistics and figures for the manuscript. The workflow uses the GitHub API to detect and save the latest commit of the external data sources that are GitHub repositories¹². It then

downloads versioned data from that snapshot of the external repositories and runs bash and Python scripts to calculate the desired statistics and produce the summary figures using Matplotlib (2157). The statistics are stored in JSON files that are accessed by Manubot to populate the values of placeholder template variables dynamically every time the manuscript is built. For instance, the template variable `{{ebm_trials_results}}` in the manuscript is replaced by the actual number of clinical trials with results, 98. The template variables also include versioned URLs to the dynamically updated figures. The JSON files and figures are stored in the `external-resources` branch of the GitHub repository, providing versioned storage. The GitHub Actions workflow automatically adds and commits the new JSON files and figures to the `external-resources` branch every time it runs, and Manubot uses the latest version of these resources when it builds the manuscript. The GitHub Actions workflow file is available online¹³, as are the scripts¹⁴. The Python package versions are also available¹⁵.

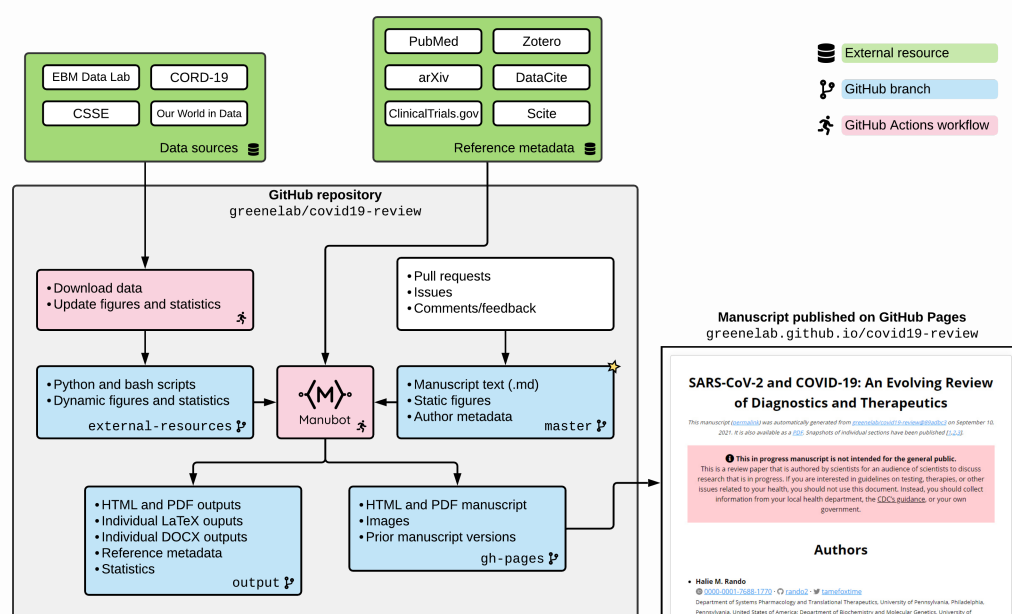


Figure 15: COVID-19 review GitHub repository organization and workflows. Manubot uses CI to combine author-contributed content with automatically updated information from outside sources. A nightly workflow updates figures and statistics derived from external resources. Authors write text and add figures to the `master` branch (starred) via GitHub pull requests. Manubot generates updated manuscript outputs for each new git commit, integrating the static text and figures with the dynamic statistics and figures and automatically-extracted citation information. GitHub Pages hosts the latest HTML and PDF versions of the manuscript along with permanent links to prior versions.

Another issue identified was the need for standardized citation to clinical trials. Other researchers identified the same need¹⁶. Trials that are registered with clinicaltrials.gov receive a unique clinical trial identifier, or “NCT ID.” Because clinical trials are registered long before results are published, referencing clinical trial identifiers was a priority. Manubot uses the Zotero translation server¹⁷ to extract citation metadata for some types of citations. However, Zotero did not support clinical trial identifiers and could not extract relevant metadata from their URLs. In order to pull clinical trial metadata associated into Manubot, we added Zotero support for these identifiers. To achieve this, we query clinicaltrials.gov to retrieve XML metadata associated with each identifier using JavaScript¹⁸. This extension enables citing a trial as `@clinicaltrials:NCT04280705` instead of the URL.

Then, when Manubot requests clinical trial metadata from the Zotero translation server, the response includes the trial sponsors, responsible investigators, title, and summary. Manubot now supports directly citing hundreds of registered Compact Uniform Resource Identifiers¹⁹, beyond just the `clinicaltrials` identifier.

Because of the large number of citations used in this manuscript and the fast-moving nature of COVID-19 research, keeping track of retractions, corrections, and notices of concern also became a challenge. We implemented a new Manubot plugin to support “smart citations” in the HTML build of manuscripts. The plugin uses the scite (²¹⁵⁸) service to display a badge below any citation with a DOI. The badge contains a set of icons and numbers that indicate how many times that source has been mentioned, supported, or disputed and whether there have been any important editorial notices. We were thus able to identify references that needed to be reevaluated by an expert. This addition was invaluable given the nature of the project, where we were disseminating rapidly evolving information of great consequence from over 1,500 different sources. The badges also allow readers to ascertain a rough approximation of the reliability of cited sources at a glance.

Because most collaborators were writing and editing text through the GitHub website rather than in a local text editor, we also needed to add spell-checking functionalities to Manubot. We integrated an existing Pandoc²⁰ spell-check extension with AppVeyor CI to automatically post spelling errors as comments in a GitHub pull request. The comment reported both unique misspelled tokens and all locations where the token was detected. Project maintainers managed a custom dictionary to allow over 1,500 scientific and technical terms that were not common English words. Spell-checking also helped standardize the writing style across dozens of authors by detecting features such as British versus American English spellings. The actual spell-checking was implemented using GNU Aspell²¹ and the Pandoc spellcheck filter²². The filter enables checking only the manuscript text, ignoring URLs and formatting.

Manubot can render a manuscript in several formats that serve different purposes. Prior to this project, Manubot could use Pandoc to convert the markdown-formatted manuscript to HTML, PDF, and DOCX formats. We expanded this functionality to export individual sections of the manuscript as separate DOCX files while still rendering the complete manuscript in HTML and PDF formats. This development was necessary because the manuscript grew so large that it needed to be split into seven separate papers for journal submission while still maintaining shared GitHub discussion across topics. When exporting an individual section, Manubot customizes the manuscript title, authors, and author contributions to pertain to that specific section. In addition, we expanded the export formats to include partial LaTeX support via Pandoc. Pandoc converts the markdown content for an individual section to TeX and the Citation Style Language JSON, which contains reference metadata generated by Manubot, to BibTeX. We customized a LaTeX template and reformatted the Manubot metadata, such as authors and their affiliations, for the LaTeX template. The exported TeX file requires manual refinement but contains all manuscript content and most of the formatting. Because LaTeX is required for manuscript submission in many fields,

automating most of the process of converting markdown to a submission-friendly format expands Manubot's potential user base. Manubot users can write in the simple markdown format, render the manuscript in continuously-updated PDF or interactive HTML formats, and export the manuscript in DOCX or TeX and BibTeX for submission to traditional publishers, taking full advantage of Pandoc's powerful document conversion capabilities and Manubot's automation.

12.4 RESULTS

12.4.1 Recruitment and Manuscript Development

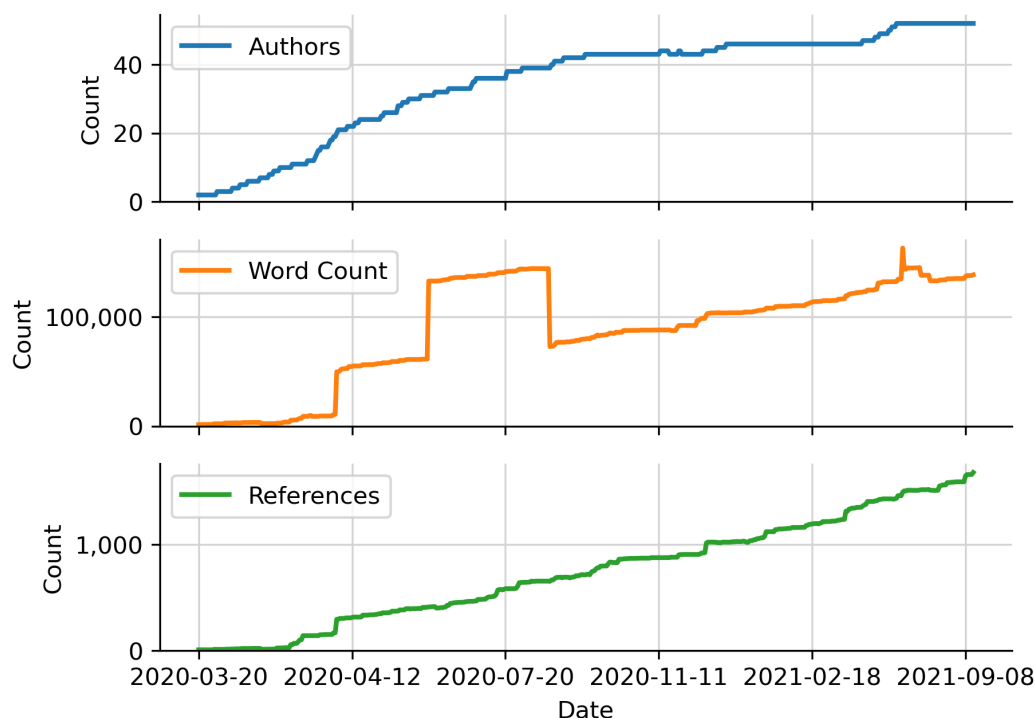


Figure 16: Project growth over time. The number of authors, word count, and number of references have all grown dramatically from when the project began on March 20, 2020. As of September 10, 2021, there were 52 authors (including consortia), 1676 references, and 138213 words. The spike in word count during summer 2020 was caused by erroneous duplication and subsequent removal of a large appendix.

Coverage by *Nature Toolbox* ([2159](#)) and an associated tweet^{[23](#)} about the project on April 1, 2020 attracted the interest of the scientific community (Figure [16](#)). Because the GitHub issues and comment systems are similar to other common web commenting systems, authors learned these tools quickly. The Gitter chat also presented a low barrier to entry. The manuscript continued to grow throughout the first year and a half of the project in both word count and the number of references (Figure [16](#)). Though only a fraction of potential contributors contributed to the text included in the manuscripts (Figure [16](#)), many contributors remained engaged over the long term (Figure [17](#)). Additionally, new contributors continued to join even into the second year of the project.



Figure 17: User contributions to the manuscript text over time. The dot size indicates the number of words added or edited each month since March 2020. The figure does not depict other types of author contributions such as literature summaries, pull request review, visualization, or software.

In order to make the project more accessible, we developed resources explaining how to use GitHub's web interface to develop and edit text for Manubot assuming no prior experience working with version control. These tutorials explained how to open an issue, open a pull request, and review a pull request²⁴. Additionally, the framework for evaluating literature was converted into issue templates to simplify the review of new articles. Articles were classified as *diagnostic*, *therapeutic*, or *other*, with an associated template developed to guide the review of papers and preprints in each category. A total of 285 new paper issues had been opened as of September 13, 2021.

The manuscripts produced by the consortium (excluding this one) will be submitted to *mSystems* as part of a special issue that provides support for continuous updates as more information becomes available. One has been published and two are available as preprints. This approach allows for a version of record to be maintained alongside the most recent version, which is always available through GitHub. These manuscripts cover a wide range of topics including the fundamental biology of SARS-CoV-2 (pathogenesis (1) and evolution), biomedical advances in responding to the virus and COVID-19 (pharmaceuticals (3), nutraceuticals (2), vaccines, and diagnostic technologies), and biological and social factors influencing disease transmission and outcomes. To date, 52 authors are associated with the consortium (Figure 16).

More formal recruitment efforts to integrate with existing projects providing support for undergraduate students during COVID-19 were also successful. We incorporated summaries written by the students, post-docs, and faculty of the Immunology Institute at the Mount Sinai School of Medicine²⁵ (2143). Additionally, two of the consortium authors were undergraduate students recruited through the American Physician Scientist Association's Virtual Summer Research Program. Thus, the consortium was successful in providing a venue for researchers across all career stages to continue investigating and publishing at a time when many biomedical researchers were unable to access their laboratory facilities.

12.4.2 Integrating Data

We integrated data into the manuscripts from several sources (Figure 15). Worldwide cases and deaths were tracked by the COVID-19 Data Repository by the Center for Systems Science and Engineering at Johns Hopkins University²⁶. The clinical trials statistics and figure were generated based on data from the University of Oxford Evidence-Based Medicine Data Lab's COVID-19 TrialsTracker (739). Information about vaccine distribution was extracted from Our World In Data²⁷ (2160). (Figure 14) integrates data from the COVID-19 dataset (2135).

Manubot's bibliographic management capabilities were critical because the amount of relevant literature published far outstripped what we had anticipated at the beginning of the project. As of September 10, 2021, there were 1676 references (Figure 16). The scite plugin provided a way to visually inspect the reference list to identify possible references of concern. This and the other new features required for the COVID-19 project are now included in Manubot's rootstock, which is the template GitHub repository for creating a new manuscript. Using CI, Manubot now checks that the manuscript was built correctly, runs spell-checking, and cross-references the manuscripts cited in this review. In addition, Manubot rootstock now supports citing clinical trial identifiers such as `clinicaltrials:NCT04292899` (809).

12.5 DISCUSSION

The current project was based in the GitHub repository `greeneLab/covid19-review` using Manubot (2149) to continuously generate the manuscript. The Manubot framework facilitated a massive collaborative review on an urgent

topic. We demonstrated the utility of Manubot to a project where many contributors lacked expertise or even experience working with version control. This effort has produced not only seven literature reviews on topics relevant to the COVID-19 pandemic, but has also generated cyberinfrastructure for training novice users in GitHub. We also extended the functionalities of Manubot to provide more of the benefits of What You See Is What You Get platforms such as Google Docs (Table 6). Open publishing thus allowed us to harness the domain expertise of a large group of non-technical users to respond to the flood of COVID-19 publications.

Several existing and new features in Manubot aid in responding to the challenges posed by the infodemic. Manuscripts are written in markdown and can be rendered in several formats providing different advantages to users. For example, beyond building just a PDF, Manubot also renders the manuscript in HTML, DOCX, and now, LaTeX (in a more limited capacity). The interactive HTML manuscript format offers several advantages over a static PDF to harmonize available resources and address specific problems related to COVID-19. The integration of scite into the HTML build makes references more manageable by visually indicating whether their results are contested or whether they have been corrected or retracted. Cross-referencing different pieces of the manuscript, such as cited preprints with reviews stored in an appendix, is another interactive option presented by HTML. The DOCX format was preferred by most non-technical users for reviewing the final version of the manuscript and was useful for creating submissions to a biological journal. Additionally, because of the heavy emphasis on Word processing in biology, Manubot's ability to generate DOCX outputs was expanded to allow users to generate DOCX files containing only a section of the manuscript. In our case, where the full project is nearly 150,000 words, this allows individual pieces to be shared more easily. Finally, the preliminary addition of LaTeX output is useful for researchers from computational fields who submit papers in TeX format and removes the step of reformatting markdown prior to submission.

Table 6: Manubot extensions for the COVID-19 review.

Type	Description
CI	Regularly download external data sources, generate new figures and statistics, and read them when Manubot builds the latest manuscript
CI	Post spell-checking reports as pull request comments
Citations	Zotero extension to report more relevant clinical trial metadata from https://clinicaltrials.gov
Citations	Cite any Compact Uniform Resource Identifier, such as <code>clinicaltrials</code> or <code>ncbigene</code>
Citations	scite badges to track retractions, corrections, and notices of concern
Outputs	Improved support for Pandoc's LaTeX output
Outputs	Build complete manuscript alongside individual sections as standalone documents

The COVID-19 Review Consortium provided a platform for researchers to engage in scientific investigation early in the pandemic when many biological scientists were unable to access their research spaces. In turn, by seeking to adapt Manubot to allow for broader participation, we made a number of improvements that are expected to increase its appeal to researchers from all backgrounds. Manubot provided a way for contributors from a variety of backgrounds, including early-career researchers, to join a massive collaborative project while demonstrating their individual contributions to the larger work and gaining experience with version control. The licensing and infrastructure also provide the basis for individuals to adapt from this project to create their own snapshots of the COVID-19 literature that derive from, but are not wholly identical to, the primary versions of these reviews. This project suggests that massive online open publishing efforts can indeed advance scholarship through inclusion ([2150](#)), including during the extreme challenges presented by the COVID-19 pandemic.

Some challenges did arise in efforts to include an academically diverse set of authors. The barriers to entry posed by git and GitHub likely still reduced participation from individuals who might have otherwise been interested. Using pull requests as a tool for writing text is also unfamiliar to many or most scientists, and the review process can be slow, which might cause interested contributors to lose interest. Additionally, the pull request model may limit people from providing general feedback on the manuscript or a section of the manuscript, unless there is an open pull request. As a result, some feedback came through email or comments on the DOCX outputs that were then translated into issues or pull requests by the project managers. Given that our approach hinged on these version control tools, it is likely that our group of contributors was biased towards those who were interested in or experienced with computational tools. The trajectory of the pandemic itself also likely influenced participation: engagement waned over the course of the pandemic as labs opened back up and researchers were able to return to their work, and we recruited very few senior clinicians to the project, which is unsurprising given the load on medical professionals during this time. Engagement that waxes and wanes is, however, typical when writing massively open online papers ([2150](#)). Adding features such as spell-check did improve usability, and additional features such as automatically checking the formatting of citations could further improve the usability of this tool. In the future, a formal study of participation could allow for quantification of these biases and improved efforts to foster inclusion.

Additional limitations are challenges associated with massively open online papers in general. With such a large amount of text, it is not possible to keep all sections of the manuscript up to date at all times. Readers are not able to distinguish when each section was updated. Even GitHub's blame functionality does not distinguish minor changes from substantive updates to the text. While much of the data and statistics update automatically, the text itself required updating by human experts. This asynchronicity could potentially introduce incompatibility between the figures and the surrounding text. Similarly, in line with the collaboration-related challenges of the project, some authors returned to update their text, while others did not. As a result, the lead authors of each paper often spent several weeks prior to journal submission updating the text to reflect new developments in each area. In the future, it may be possible to streamline this process

through integration with a tool such as CoronaCentral ([2136](#)) to automatically identify relevant, high-impact papers that need to be included, although expertise would still be required to incorporate them. Another challenge involves tracking preprints as they are reviewed or critiqued, revised, and potentially published. While updating the content of the manuscript would likely fall to human contributors, automatic detection of published versions of preprints ([2161](#)) could be integrated in the future. These challenges are exacerbated by the scale of the infodemic, but developing solutions would benefit future projects tracking more typical trends in publication. Similarly, outputting machine readable summaries of key information in the COVID-19 review manuscripts could reduce their contribution to the infodemic. As it stands, the integration of Compact Uniform Resource Identifier does make a step in this direction. Formal identifiers could be used to extract relationships among clinical trials, genes, publications, and other entities. Thus, the experience of using Manubot for a massive project has laid the foundation for future additions to enhance user experience and inclusivity.

12.6 CONCLUSION

With the worldwide scientific community uniting during 2020 and 2021 to investigate COVID-19 from a wide range of perspectives, findings from many disciplines are relevant on a rapid timescale to a broad scientific audience. As many other efforts have described, the publishing rate of formal manuscripts and preprints about COVID-19 has been unprecedented ([2133](#)), and efforts to review the body of COVID-19 literature are faced with an ever-expanding corpus to evaluate. In the case of the seven manuscripts produced by the COVID-19 Review Consortium, Manubot allows for continuous updating of the manuscripts as the pandemic enters its second year and the landscape shifts with the emergence of promising therapeutics and vaccines ([3](#)). These manuscripts pull data from external sources and update information and visualizations daily using CI. By off-loading some updates to computational pipelines, domain experts can focus on the broader implications of new information as it emerges. Centralizing, summarizing, and critiquing data and literature broadly relevant to COVID-19 can expedite the interdisciplinary scientific process that is currently happening at an advanced pace. As of September 13, 2021, 2886 commits have been made to the manuscript across 575 merged pull requests. The efforts of the COVID-19 Review Consortium illustrate the value of including open source tools, including those focused on open publishing, in these efforts. By facilitating the versioning of text, such platforms also allow for documentation of the evolution of thought in an evolving area and formal analysis of a collaborative project. This application of version control holds the potential to improve scientific publishing in a range of disciplines, including those outside of traditional computational fields. While Manubot is a technologically complex tool, this project demonstrates that it can be applied to a variety of projects. Future work can address remaining limitations and continue to advance Manubot as an inclusive tool for open publishing projects.

13 Additional Items

13.1 Competing Interests

Author	Competing Interests	Last Reviewed
Halie M. Rando	None	2021-01-20
Casey S. Greene	None	2021-01-20
Michael P. Robson	None	2020-11-12
Simina M. Boca	Currently an employee at AstraZeneca, Gaithersburg, MD, USA, may own stock or stock options.	2021-07-01
Nils Wellhausen	None	2020-11-03
Ronan Lordan	None	2020-11-03
Christian Brueffer	Shareholder of SAGA Diagnostics AB.	2023-03-06
Sandipan Ray	None	2020-11-11
Lucy D'Agostino McGowan	Received consulting fees from Acelyt and Sanofi in the past five years	2020-11-10
Anthony Gitter	Inventor of patent US-11410440-B2 assigned to the Wisconsin Alumni Research Foundation related to classifying activated T cells.	2023-01-11
Anna Ada Dattoli	None	2020-03-26
Ryan Velazquez	None	2020-11-10
John P. Barton	None	2020-11-11
Jeffrey M. Field	None	2020-11-12
Bharath Ramsundar	None	2020-11-11
Adam L. MacLean	None	2021-02-23
Alexandra J. Lee	None	2020-11-09
Immunology Institute of the Icahn School of Medicine	None	2020-04-07
Fengling Hu	None	2020-04-08
Nafisa M. Jadavji	None	2020-11-11
Elizabeth Sell	None	2020-11-11
Vincent Rubinetti	None	2021-04-29
Jinhui Wang	None	2021-01-21
Diane N. Rafizadeh	None	2020-11-11
Ashwin N. Skelly	None	2020-11-11

Author	Competing Interests	Last Reviewed
Marouen Ben Guebila	None	2021-08-02
Likhitha Kolla	None	2020-11-16
David Manheim	None	2022-03-15
Soumita Ghosh	None	2020-11-09
James Brian Byrd	Funded by FastGrants to conduct a COVID-19-related clinical trial	2020-11-12
YoSon Park	YoSon Park is affiliated with Pfizer Worldwide Research. The author has no financial interests to declare and contributed as an author prior to joining Pfizer, and the work was not part of a Pfizer collaboration nor was it funded by Pfizer.	2020-01-22
Vikas Bansal	None	2021-01-25
Stephen Capone	None	2020-11-11
John J. Dziak	None	2020-11-11
Yuchen Sun	None	2020-11-11
Yanjun Qi	None	2020-07-09
Lamonica Shinholster	None	2020-11-11
Temitayo Lukan	None	2020-11-10
Sergey Knyazev	None	2020-11-11
Dimitri Perrin	None	2020-11-11
Serghei Mangul	None	2020-11-11
Shikta Das	None	2020-08-13
Gregory L Szeto	None	2020-11-16
Tiago Lubiana	None	2020-11-11
David Mai	None	2021-01-08
COVID-19 Review Consortium	None	2021-01-16
Rishi Raj Goel	None	2021-01-20
Joel D Boerckel	None	2021-03-26
Amruta Naik	None	2021-04-05
Yusha Sun	None	2021-04-10
Daniel S. Himmelstein	None	2021-04-30
Jeremy P. Kamil	None	2022-10-11
Jesse G. Meyer	None	2022-01-06
Ariel I. Mundo	None	2021-12-19

13.2 Author Contributions

Author	Contributions
Halie M. Rando	D, E, Project Administration, Software, Visualization, Writing - Original Draft, Writing - Review & Editing
Casey S. Greene	Conceptualization, Project Administration, Software, Supervision, Writing - Original Draft, Writing - Review & Editing
Michael P. Robson	Methodology, Software, Supervision
Simina M. Boca	Methodology, Project Administration, Writing - Review & Editing
Nils Wellhausen	Project Administration, Visualization, Writing - Original Draft, Writing - Review & Editing
Ronan Lordan	Conceptualization, Project Administration, Visualization, Writing - Original Draft, Writing - Review & Editing
Christian Brueffer	Project Administration, Writing - Original Draft, Writing - Review & Editing
Sandipan Ray	Writing - Original Draft, Writing - Review & Editing
Lucy D'Agostino McGowan	Methodology, Writing - Original Draft, Writing - Review & Editing
Anthony Gitter	Methodology, Project Administration, Software, Visualization, Writing - Original Draft, Writing - Review & Editing
Anna Ada Dattoli	Writing - Original Draft
Ryan Velazquez	Methodology, Software, Writing - Review & Editing
John P. Barton	Writing - Original Draft, Writing - Review & Editing
Jeffrey M. Field	Writing - Original Draft, Writing - Review & Editing
Bharath Ramsundar	Writing - Review & Editing
Adam L. MacLean	Writing - Original Draft, Writing - Review & Editing
Alexandra J. Lee	Writing - Original Draft, Writing - Review & Editing
Immunology Institute of the Icahn School of Medicine	Data Curation
Fengling Hu	Writing - Original Draft, Writing - Review & Editing
Nafisa M. Jadavji	Supervision, Writing - Original Draft, Writing - Review & Editing
Elizabeth Sell	Writing - Original Draft, Writing - Review & Editing

Author	Contributions
Vincent Rubinetti	Software, Visualization, Writing - Original Draft
Jinhui Wang	Writing - Original Draft, Writing - Review & Editing
Diane N. Rafizadeh	Project Administration, Writing - Original Draft, Writing - Review & Editing
Ashwin N. Skelly	Writing - Original Draft, Writing - Review & Editing
Marouen Ben Guebila	Writing - Original Draft, Writing - Review & Editing
Likhitha Kolla	Writing - Original Draft, Writing - Review & Editing
David Manheim	Writing - Original Draft, Writing - Review & Editing
Soumita Ghosh	Writing - Original Draft
James Brian Byrd	Writing - Original Draft, Writing - Review & Editing
YoSon Park	Writing - Original Draft, Writing - Review & Editing
Vikas Bansal	Writing - Original Draft, Writing - Review & Editing
Stephen Capone	Writing - Original Draft, Writing - Review & Editing
John J. Dziak	Writing - Original Draft, Writing - Review & Editing
Yuchen Sun	Visualization
Yanjun Qi	Visualization
Lamonica Shinholster	Writing - Original Draft
Temitayo Lukan	Investigation, Writing - Original Draft
Sergey Knyazev	Writing - Original Draft, Writing - Review & Editing
Dimitri Perrin	Writing - Original Draft, Writing - Review & Editing
Serghei Mangul	Writing - Review & Editing
Shikta Das	Writing - Review & Editing
Gregory L Szeto	Writing - Review & Editing
Tiago Lubiana	Writing - Review & Editing
David Mai	Writing - Original Draft, Writing - Review & Editing
COVID-19 Review Consortium	Project Administration
Rishi Raj Goel	Writing - Original Draft, Writing - Review & Editing
Joel D Boerckel	Writing - Review & Editing

Author	Contributions
Amruta Naik	Writing - Original Draft, Writing - Review & Editing
Yusha Sun	Writing - Review & Editing
Daniel S. Himmelstein	Software
Jeremy P. Kamil	Writing - Review & Editing
Jesse G. Meyer	Writing - Original Draft, Writing - Review & Editing
Ariel I. Mundo	Writing - Original Draft, Writing - Review & Editing

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13.4 References

1. Rando HM, MacLean AL, Lee AJ, Lordan R, Ray S, Bansal V, Skelly AN, Sell E, Dziak JJ, Shinholster L, D'Agostino McGowan L, Ben Guebila M, Wellhausen N, Knyazev S, Boca SM, Capone S, Qi Y, Park Y, Mai D, Sun Y, Boerckel JD, Brueffer C, Byrd JB, Kamil JP, Wang J, Velazquez R, Szeto GL, Barton JP, Goel RR, Mangul S, Lubiana T, COVID-19 Review Consortium Vikas Bansal, John P. Barton, Simina M. Boca, Joel D. Boerckel, Christian Brueffer, James Brian Byrd, Stephen Capone, Shikta Das, Anna Ada Dattoli, John J. Dziak, Jeffrey M. Field, Soumita Ghosh, Anthony Gitter, Rishi Raj Goel, Casey S. Greene, Marouen Ben Guebila, Daniel S. Himmelstein, Fengling Hu, Nafisa M. Jadavji, Jeremy P. Kamil, Sergey Knyazev, Likhitha Kolla, Alexandra J. Lee, Ronan Lordan, Tiago Lubiana, Temitayo Lukan, Adam L. MacLean, David Mai, Serghei Mangul, David Manheim, Lucy D'Agostino McGowan, Amruta Naik, YoSon Park, Dimitri Perrin, Yanjun Qi, Diane N. Rafizadeh, Bharath Ramsundar, Halie M. Rando, Sandipan Ray, Michael P. Robson, Vincent Rubinetti, Elizabeth Sell, Lamonica Shinholster, Ashwin N. Skelly, Yuchen Sun, Yusha Sun, Gregory L. Szeto, Ryan Velazquez, Jinhui Wang, Nils Wellhausen, Gitter A, Greene CS. 2021. [Pathogenesis, Symptomatology, and Transmission of SARS-CoV-2 through Analysis of Viral Genomics and Structure](#). mSystems 6:e0009521.
2. Lordan R, Rando HM, COVID-19 Review Consortium, Greene CS. 2021. [Dietary Supplements and Nutraceuticals under Investigation for COVID-19 Prevention and Treatment](#). mSystems 6:e00122–21.
3. Rando HM, Wellhausen N, Ghosh S, Lee AJ, Dattoli AA, Hu F, Byrd JB, Rafizadeh DN, Lordan R, Qi Y, Sun Y, Brueffer C, Field JM, Ben Guebila M, Jadavji NM, Skelly AN, Ramsundar B, Wang J, Goel RR, Park Y, COVID-19 Review Consortium Vikas Bansal, John P. Barton, Simina M. Boca, Joel D. Boerckel, Christian Brueffer, James Brian Byrd, Stephen Capone, Shikta Das, Anna Ada Dattoli, John J. Dziak, Jeffrey M. Field, Soumita Ghosh, Anthony Gitter, Rishi Raj Goel, Casey S. Greene, Marouen Ben Guebila, Daniel S. Himmelstein, Fengling Hu, Nafisa M. Jadavji, Jeremy P. Kamil, Sergey Knyazev, Likhitha Kolla, Alexandra J. Lee, Ronan Lordan, Tiago Lubiana, Temitayo Lukan, Adam L. MacLean, David Mai, Serghei Mangul, David Manheim, Lucy D'Agostino McGowan, Amruta Naik, YoSon Park, Dimitri Perrin, Yanjun Qi, Diane N. Rafizadeh, Bharath Ramsundar, Halie M. Rando, Sandipan Ray, Michael P. Robson, Vincent Rubinetti, Elizabeth Sell, Lamonica Shinholster, Ashwin N. Skelly, Yuchen Sun, Yusha Sun, Gregory L. Szeto, Ryan Velazquez, Jinhui Wang, Nils Wellhausen, Boca SM, Gitter A, Greene CS. 2021. [Identification and Development of Therapeutics for COVID-19](#). mSystems 6:e0023321.
4. Rando HM, Boca SM, D'Agostino McGowan L, Himmelstein DS, Robson MP, Rubinetti V, Velazquez R, Greene CS, Gitter A. 2021. [An Open-Publishing Response to the COVID-19 Infodemic](#). Proceedings of the Workshop on Digital Infrastructures for Scholarly Content Objects (DISCO 2021) 2976:29–38.

5. Rando HM, Lordan R, Lee AJ, Naik A, Wellhausen N, Sell E, Kolla L, COVID-19 Review Consortium, Gitter A, Greene CS. 2023. [Application of Traditional Vaccine Development Strategies to SARS-CoV-2](#). mSystems 8:e0092722.
6. Rando HM, Lordan R, Kolla L, Sell E, Lee AJ, Wellhausen N, Naik A, Kamil JP, COVID-19 Review Consortium, Gitter A, Greene CS. 2023. [The Coming of Age of Nucleic Acid Vaccines during COVID-19](#). mSystems 8:e0092822.
7. Rando HM, Brueffer C, Lordan R, Dattoli AA, Manheim D, Meyer JG, Mundo AI, Perrin D, Mai D, Wellhausen N, Consortium CR, Gitter A, Greene CS. 2022. [Molecular and Serologic Diagnostic Technologies for SARS-CoV-2](#). 2204.12598arXiv. arXiv.
8. CDC. 2020. Coronavirus Disease 2019 (COVID-19) – Symptoms. Centers for Disease Control and Prevention. <https://www.cdc.gov/coronavirus/2019-ncov/symptoms-testing/symptoms.html>. Retrieved 8 February 2021.
9. Cucinotta D, Vanelli M. 2020. [WHO Declares COVID-19 a Pandemic](#). Acta Bio Medica Atenei Parmensis 91:157–160.
10. Li L, Huang Q, Wang DC, Ingbar DH, Wang X. 2020. [Acute lung injury in patients with COVID-19 infection](#). Clinical and Translational Medicine 10:20–27.
11. Tan W, Zhao X, Ma X, Wang W, Niu P, Xu W, F. Gao G, Wu G, MHC Key Laboratory of Biosafety, National Institute for Viral Disease Control and Prevention, China CDC, Beijing, China, Center for Biosafety Mega-Science, Chinese Academy of Sciences, Beijing, China. 2020. [A Novel Coronavirus Genome Identified in a Cluster of Pneumonia Cases — Wuhan, China 2019–2020](#). China CDC Weekly 2:61–62.
12. Li F. 2016. [Structure, Function, and Evolution of Coronavirus Spike Proteins](#). Annual Review of Virology 3:237–261.
13. Chan JF-W, Yuan S, Kok K-H, To KK-W, Chu H, Yang J, Xing F, Liu J, Yip CC-Y, Poon RW-S, Tsoi H-W, Lo SK-F, Chan K-H, Poon VK-M, Chan W-M, Ip JD, Cai J-P, Cheng VC-C, Chen H, Hui CK-M, Yuen K-Y. 2020. [A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: a study of a family cluster](#). The Lancet 395:514–523.
14. 2007. Fields virology 5th ed. Wolters Kluwer Health/Lippincott Williams & Wilkins, Philadelphia.
15. Broer R, Boson B, Spaan W, Cosset F-L, Corver J. 2006. [Important Role for the Transmembrane Domain of Severe Acute Respiratory Syndrome Coronavirus Spike Protein during Entry](#). Journal of Virology 80:1302–1310.
16. Wrapp D, Wang N, Corbett KS, Goldsmith JA, Hsieh C-L, Abiona O, Graham BS, McLellan JS. 2020. [Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation](#). Science 367:1260–1263.

17. 1996. Medical microbiology 4th ed. University of Texas Medical Branch at Galveston, Galveston, Tex.
18. Fehr AR, Perlman S. 2015. [Coronaviruses: An Overview of Their Replication and Pathogenesis](#) Methods in Molecular Biology. Springer Science and Business Media LLC.
19. Lu R, Zhao X, Li J, Niu P, Yang B, Wu H, Wang W, Song H, Huang B, Zhu N, Bi Y, Ma X, Zhan F, Wang L, Hu T, Zhou H, Hu Z, Zhou W, Zhao L, Chen J, Meng Y, Wang J, Lin Y, Yuan J, Xie Z, Ma J, Liu WJ, Wang D, Xu W, Holmes EC, Gao GF, Wu G, Chen W, Shi W, Tan W. 2020. [Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding](#). The Lancet 395:565–574.
20. Chen Y, Liu Q, Guo D. 2020. [Emerging coronaviruses: Genome structure, replication, and pathogenesis](#). Journal of Medical Virology 92:418–423.
21. Wang H, Yang P, Liu K, Guo F, Zhang Y, Zhang G, Jiang C. 2008. [SARS coronavirus entry into host cells through a novel clathrin- and caveolae-independent endocytic pathway](#). Cell Research 18:290–301.
22. Mercer J, Schelhaas M, Helenius A. 2010. [Virus Entry by Endocytosis](#). Annual Review of Biochemistry 79:803–833.
23. Belouzard S, Millet JK, Licitra BN, Whittaker GR. 2012. [Mechanisms of Coronavirus Cell Entry Mediated by the Viral Spike Protein](#). Viruses 4:1011–1033.
24. Jaimes JA, André NM, Chappie JS, Millet JK, Whittaker GR. 2020. [Phylogenetic Analysis and Structural Modeling of SARS-CoV-2 Spike Protein Reveals an Evolutionary Distinct and Proteolytically Sensitive Activation Loop](#). Journal of Molecular Biology 432:3309–3325.
25. Walls AC, Park Y-J, Tortorici MA, Wall A, McGuire AT, Veesler D. 2020. [Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein](#). Cell 181:281–292.e6.
26. Zhang H, Penninger JM, Li Y, Zhong N, Slutsky AS. 2020. [Angiotensin-converting enzyme 2 \(ACE2\) as a SARS-CoV-2 receptor: molecular mechanisms and potential therapeutic target](#). Intensive Care Medicine 46:586–590.
27. Jia HP, Look DC, Hickey M, Shi L, Pewe L, Netland J, Farzan M, Wohlford-Lenane C, Perlman S, McCray PB. 2006. [Infection of Human Airway Epithelia by Sars Coronavirus is Associated with ACE2 Expression and Localization](#) Advances in Experimental Medicine and Biology. Springer Science and Business Media LLC.
28. Hikmet F, Méar L, Edvinsson Å, Micke P, Uhlén M, Lindskog C. 2020. [The protein expression profile of ACE2 in human tissues](#). Molecular Systems Biology 16.
29. Li F. 2015. [Receptor Recognition Mechanisms of Coronaviruses: a Decade of Structural Studies](#). Journal of Virology 89:1954–1964.

30. Du L, He Y, Zhou Y, Liu S, Zheng B-J, Jiang S. 2009. [The spike protein of SARS-CoV — a target for vaccine and therapeutic development](#). *Nature Reviews Microbiology* 7:226–236.
31. de Haan CAM, Rottier PJM. 2005. [Molecular Interactions in the Assembly of Coronaviruses](#) *Advances in Virus Research*. Elsevier BV.
32. Tang T, Bidon M, Jaimes JA, Whittaker GR, Daniel S. 2020. [Coronavirus membrane fusion mechanism offers a potential target for antiviral development](#). *Antiviral Research* 178:104792.
33. Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, Schiergens TS, Herrler G, Wu N-H, Nitsche A, Müller MA, Drosten C, Pöhlmann S. 2020. [SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor](#). *Cell* 181:271–280.e8.
34. Ou X, Liu Y, Lei X, Li P, Mi D, Ren L, Guo L, Guo R, Chen T, Hu J, Xiang Z, Mu Z, Chen X, Chen J, Hu K, Jin Q, Wang J, Qian Z. 2020. [Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV](#). *Nature Communications* 11:1620.
35. Coutard B, Valle C, de Lamballerie X, Canard B, Seidah NG, Decroly E. 2020. [The spike glycoprotein of the new coronavirus 2019-nCoV contains a furin-like cleavage site absent in CoV of the same clade](#). *Antiviral Research* 176:104742.
36. Xia S, Lan Q, Su S, Wang X, Xu W, Liu Z, Zhu Y, Wang Q, Lu L, Jiang S. 2020. [The role of furin cleavage site in SARS-CoV-2 spike protein-mediated membrane fusion in the presence or absence of trypsin](#). *Sig Transduct Target Ther* 5.
37. Johnson BA, Xie X, Kalveram B, Lokugamage KG, Muruato A, Zou J, Zhang X, Juelich T, Smith JK, Zhang L, Bopp N, Schindewolf C, Vu M, Vanderheiden A, Swetnam D, Plante JA, Aguilar P, Plante KS, Lee B, Weaver SC, Suthar MS, Routh AL, Ren P, Ku Z, An Z, Debbink K, Shi PY, Freiberg AN, Menachery VD. 2020. [Furin Cleavage Site Is Key to SARS-CoV-2 Pathogenesis](#). Cold Spring Harbor Laboratory.
38. Johnson BA, Xie X, Bailey AL, Kalveram B, Lokugamage KG, Muruato A, Zou J, Zhang X, Juelich T, Smith JK, Zhang L, Bopp N, Schindewolf C, Vu M, Vanderheiden A, Winkler ES, Swetnam D, Plante JA, Aguilar P, Plante KS, Popov V, Lee B, Weaver SC, Suthar MS, Routh AL, Ren P, Ku Z, An Z, Debbink K, Diamond MS, Shi P-Y, Freiberg AN, Menachery VD. 2021. [Loss of furin cleavage site attenuates SARS-CoV-2 pathogenesis](#). *Nature* 591:293–299.
39. Peacock TP, Goldhill DH, Zhou J, Baillon L, Frise R, Swann OC, Kugathasan R, Penn R, Brown JC, Sanchez-David RY, Braga L, Williamson MK, Hassard JA, Staller E, Hanley B, Osborn M, Giacca M, Davidson AD, Matthews DA, Barclay WS. 2021. [The furin cleavage site in the SARS-CoV-2 spike protein is required for transmission in ferrets](#). *Nat Microbiol* 6:899–909.

40. Cheng Y-W, Chao T-L, Li C-L, Chiu M-F, Kao H-C, Wang S-H, Pang Y-H, Lin C-H, Tsai Y-M, Lee W-H, Tao M-H, Ho T-C, Wu P-Y, Jang L-T, Chen P-J, Chang S-Y, Yeh S-H. 2020. [Furin Inhibitors Block SARS-CoV-2 Spike Protein Cleavage to Suppress Virus Production and Cytopathic Effects](#). Cell Reports 33:108254.
41. Coronaviridae ~ ViralZone. <https://viralzone.expasy.org/30>. Retrieved 5 December 2022.
42. Jaimes JA, Millet JK, Whittaker GR. 2020. [Proteolytic Cleavage of the SARS-CoV-2 Spike Protein and the Role of the Novel S1/S2 Site](#). iScience 23:101212.
43. Dyer L, Patterson C. 2010. [Development of the Endothelium: An Emphasis on Heterogeneity](#). Semin Thromb Hemost 36:227–235.
44. Thorgeirsson G, Robertson AL. 1978. [The vascular endothelium-pathobiologic significance](#). Am J Pathol 93:803–48.
45. Reitsma S, Slaaf DW, Vink H, van Zandvoort MAMJ, oude Egbrink MGA. 2007. [The endothelial glycocalyx: composition, functions, and visualization](#). Pflugers Arch - Eur J Physiol 454:345–359.
46. Clausen TM, Sandoval DR, Spliid CB, Pihl J, Perrett HR, Painter CD, Narayanan A, Majowicz SA, Kwong EM, McVicar RN, Thacker BE, Glass CA, Yang Z, Torres JL, Golden GJ, Bartels PL, Porell RN, Garretson AF, Laubach L, Feldman J, Yin X, Pu Y, Hauser BM, Caradonna TM, Kellman BP, Martino C, Gordts PLSM, Chanda SK, Schmidt AG, Godula K, Leibel SL, Jose J, Corbett KD, Ward AB, Carlin AF, Esko JD. 2020. [SARS-CoV-2 Infection Depends on Cellular Heparan Sulfate and ACE2](#). Cell 183:1043–1057.e15.
47. Ihrcke NS, Wrenshall LE, Lindman BJ, Platt JL. 1993. [Role of heparan sulfate in immune system-blood vessel interactions](#). Immunology Today 14:500–505.
48. Liu L, Chopra P, Li X, Bouwman KM, Tompkins SM, Wolfert MA, de Vries RP, Boons G-J. 2020. [Heparan sulfate proteoglycans as attachment factor for SARS-CoV-2](#). Cold Spring Harbor Laboratory.
49. Shriver Z, Capila I, Venkataraman G, Sasisekharan R. 2011. [Heparin and Heparan Sulfate: Analyzing Structure and Microheterogeneity](#), p. 159–176. In Heparin - A Century of Progress. Springer Berlin Heidelberg.
50. LaRivière WB, Schmidt EP. 2018. [The Pulmonary Endothelial Glycocalyx in ARDS: A Critical Role for Heparan Sulfate](#), p. 33–52. In Current Topics in Membranes. Elsevier.
51. Zhang Q, Chen CZ, Swaroop M, Xu M, Wang L, Lee J, Wang AQ, Pradhan M, Hagen N, Chen L, Shen M, Luo Z, Xu X, Xu Y, Huang W, Zheng W, Ye Y. 2020. [Heparan sulfate assists SARS-CoV-2 in cell entry and can be targeted by approved drugs in vitro](#). Cell Discov 6.
52. Buijsers B, Yanginlar C, Maciej-Hulme ML, de Mast Q, van der Vlag J. 2020. [Beneficial non-anticoagulant mechanisms underlying heparin treatment of COVID-19 patients](#). eBioMedicine 59:102969.

53. Salah HM, Naser JA, Calcaterra G, Bassareo PP, Mehta JL. 2020. [The Effect of Anticoagulation Use on Mortality in COVID-19 Infection](#). The American Journal of Cardiology 134:155–157.
54. Salaris C, Scarpa M, Elli M, Bertolini A, Guglielmetti S, Pregliasco F, Blandizzi C, Brun P, Castagliuolo I. 2021. [Protective Effects of Lactoferrin against SARS-CoV-2 Infection In Vitro](#). Nutrients 13:328.
55. Wadowski PP, Jilma B, Kopp CW, Ertl S, Gremmel T, Koppensteiner R. 2021. [Glycocalyx as Possible Limiting Factor in COVID-19](#). Front Immunol 12.
56. Liu L, Wei Q, Nishiura K, Peng J, Wang H, Midkiff C, Alvarez X, Qin C, Lackner A, Chen Z. 2015. [Spatiotemporal interplay of severe acute respiratory syndrome coronavirus and respiratory mucosal cells drives viral dissemination in rhesus macaques](#). Mucosal Immunology 9:1089–1101.
57. Poynter SJ, DeWitte-Orr SJ. 2018. [Understanding Viral dsRNA-Mediated Innate Immune Responses at the Cellular Level Using a Rainbow Trout Model](#). Frontiers in Immunology 9:829.
58. Snijder EJ, van der Meer Y, Zevenhoven-Dobbe J, Onderwater JJM, van der Meulen J, Koerten HK, Mommaas AM. 2006. [Ultrastructure and Origin of Membrane Vesicles Associated with the Severe Acute Respiratory Syndrome Coronavirus Replication Complex](#). Journal of Virology 80:5927–5940.
59. Li X, Geng M, Peng Y, Meng L, Lu S. 2020. [Molecular immune pathogenesis and diagnosis of COVID-19](#). Journal of Pharmaceutical Analysis 10:102–108.
60. Thoms M, Buschauer R, Ameisemeier M, Koepke L, Denk T, Hirschenberger M, Kratzat H, Hayn M, Mackens-Kiani T, Cheng J, Straub JH, Stürzel CM, Fröhlich T, Berninghausen O, Becker T, Kirchhoff F, Sparrer KMJ, Beckmann R. 2020. [Structural basis for translational shutdown and immune evasion by the Nsp1 protein of SARS-CoV-2](#). Science 369:1249–1255.
61. 2020. Immune responses in COVID-19 and potential vaccines: Lessons learned from SARS and MERS epidemic. Asian Pacific Journal of Allergy and Immunology <https://doi.org/10.12932/ap-200220-0772>.
62. Li W, Moore MJ, Vasilieva N, Sui J, Wong SK, Berne MA, Somasundaran M, Sullivan JL, Luzuriaga K, Greenough TC, Choe H, Farzan M. 2003. [Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus](#). Nature 426:450–454.
63. Matsuyama S, Nagata N, Shirato K, Kawase M, Takeda M, Taguchi F. 2010. [Efficient Activation of the Severe Acute Respiratory Syndrome Coronavirus Spike Protein by the Transmembrane Protease TMPRSS2](#). Journal of Virology 84:12658–12664.
64. Glowacka I, Bertram S, Muller MA, Allen P, Soilleux E, Pfefferle S, Steffen I, Tsegaye TS, He Y, Gnirss K, Niemeyer D, Schneider H, Drosten C, Pohlmann S. 2011. [Evidence that TMPRSS2 Activates the Severe](#)

[Acute Respiratory Syndrome Coronavirus Spike Protein for Membrane Fusion and Reduces Viral Control by the Humoral Immune Response.](#)
Journal of Virology 85:4122–4134.

65. Zhuang M, Cheng Y, Zhang J, Jiang X, Wang L, Deng J, Wang P. 2020. [Increasing host cellular receptor—angiotensin-converting enzyme 2 expression by coronavirus may facilitate 2019-nCoV \(or SARS-CoV-2\) infection.](#) J Med Virol 92:2693–2701.
66. Li Y, Zhou W, Yang L, You R. 2020. [Physiological and pathological regulation of ACE2, the SARS-CoV-2 receptor.](#) Pharmacological Research 157:104833.
67. Hamming I, Timens W, Bulthuis M, Lely A, Navis G, van Goor H. 2004. [Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis.](#) The Journal of Pathology 203:631–637.
68. Foote MB, White JR, Jee J, Argilés G, Wan JCM, Rousseau B, Pessin MS, Diaz LA Jr. 2021. [Association of Antineoplastic Therapy With Decreased SARS-CoV-2 Infection Rates in Patients With Cancer.](#) JAMA Oncol 7:1686.
69. Thomas G. 2002. [Furin at the cutting edge: From protein traffic to embryogenesis and disease.](#) Nat Rev Mol Cell Biol 3:753–766.
70. Hoffmann M, Kleine-Weber H, Pöhlmann S. 2020. [A Multibasic Cleavage Site in the Spike Protein of SARS-CoV-2 Is Essential for Infection of Human Lung Cells.](#) Molecular Cell 78:779–784.e5.
71. Wang W, Xu Y, Gao R, Lu R, Han K, Wu G, Tan W. 2020. Detection of SARS-CoV-2 in Different Types of Clinical Specimens. JAMA <https://doi.org/10.1001/jama.2020.3786>.
72. Young BE, Ong SWX, Kalimuddin S, Low JG, Tan SY, Loh J, Ng O-T, Marimuthu K, Ang LW, Mak TM, Lau SK, Anderson DE, Chan KS, Tan TY, Ng TY, Cui L, Said Z, Kurupatham L, Chen MI-C, Chan M, Vasoo S, Wang L-F, Tan BH, Lin RTP, Lee VJM, Leo Y-S, Lye DC, for the Singapore 2019 Novel Coronavirus Outbreak Research Team. 2020. [Epidemiologic Features and Clinical Course of Patients Infected With SARS-CoV-2 in Singapore.](#) JAMA 323:1488.
73. Ling Y, Xu S-B, Lin Y-X, Tian D, Zhu Z-Q, Dai F-H, Wu F, Song Z-G, Huang W, Chen J, Hu B-J, Wang S, Mao E-Q, Zhu L, Zhang W-H, Lu H-Z. 2020. [Persistence and clearance of viral RNA in 2019 novel coronavirus disease rehabilitation patients.](#) Chinese Medical Journal 133:1039–1043.
74. Menter T, Haslbauer JD, Nienhold R, Savic S, Hopfer H, Deigendesch N, Frank S, Turek D, Willi N, Pargger H, Bassetti S, Leuppi JD, Cathomas G, Tolnay M, Mertz KD, Tzankov A. 2020. [Postmortem examination of COVID-19 patients reveals diffuse alveolar damage with severe capillary congestion and variegated findings in lungs and other organs suggesting vascular dysfunction.](#) Histopathology 77:198–209.

75. Shi S, Qin M, Shen B, Cai Y, Liu T, Yang F, Gong W, Liu X, Liang J, Zhao Q, Huang H, Yang B, Huang C. 2020. [Association of Cardiac Injury With Mortality in Hospitalized Patients With COVID-19 in Wuhan, China](#). JAMA Cardiology 5:802.
76. Wang S, Zhou X, Zhang T, Wang Z. 2020. [The need for urogenital tract monitoring in COVID-19](#). Nature Reviews Urology 17:314–315.
77. Fanelli V, Fiorentino M, Cantaluppi V, Gesualdo L, Stallone G, Ronco C, Castellano G. 2020. [Acute kidney injury in SARS-CoV-2 infected patients](#). Critical Care 24:155.
78. Zhang C, Shi L, Wang F-S. 2020. [Liver injury in COVID-19: management and challenges](#). The Lancet Gastroenterology & Hepatology 5:428–430.
79. Xiao F, Tang M, Zheng X, Liu Y, Li X, Shan H. 2020. [Evidence for Gastrointestinal Infection of SARS-CoV-2](#). Gastroenterology 158:1831–1833.e3.
80. Gao QY, Chen YX, Fang JY. 2020. [2019 Novel coronavirus infection and gastrointestinal tract](#). Journal of Digestive Diseases 21:125–126.
81. Agarwal A, Rochwerf B, Lamontagne F, Siemieniuk RA, Agoritsas T, Askie L, Lytvyn L, Leo Y-S, Macdonald H, Zeng L, Amin W, da Silva ARA, Aryal D, Barragan FAJ, Bausch FJ, Burhan E, Calfee CS, Cecconi M, Chacko B, Chanda D, Dat VQ, De Sutter A, Du B, Freedman S, Geduld H, Gee P, Gotte M, Harley N, Hashmi M, Hunt B, Jehan F, Kabra SK, Kanda S, Kim Y-J, Kissoon N, Krishna S, Kuppalli K, Kwizera A, Castro-Rial ML, Lisboa T, Lodha R, Mahaka I, Manai H, Mendelson M, Battista Migliori G, Mino G, Nsutebu E, Preller J, Pshenichnaya N, Qadir N, Relan P, Sabzwari S, Sarin R, Shankar-Hari M, Sharland M, Shen Y, Ranganathan SS, Souza JP, Stegeman M, Swanstrom R, Ugarte S, Uyeki T, Venkatapuram S, Vuyiseka D, Wijewickrama A, Tran L, Zeraatkar D, Bartoszko JJ, Ge L, Brignardello-Petersen R, Owen A, Guyatt G, Diaz J, Kawano-Dourado L, Jacobs M, Vandvik PO. 2020. [A living WHO guideline on drugs for covid-19](#). BMJ m3379.
82. Guan W, Ni Z, Hu Y, Liang W, Ou C, He J, Liu L, Shan H, Lei C, Hui DSC, Du B, Li L, Zeng G, Yuen K-Y, Chen R, Tang C, Wang T, Chen P, Xiang J, Li S, Wang J, Liang Z, Peng Y, Wei L, Liu Y, Hu Y, Peng P, Wang J, Liu J, Chen Z, Li G, Zheng Z, Qiu S, Luo J, Ye C, Zhu S, Zhong N. 2020. [Clinical Characteristics of Coronavirus Disease 2019 in China](#). New England Journal of Medicine 382:1708–1720.
83. Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, Xiang J, Wang Y, Song B, Gu X, Guan L, Wei Y, Li H, Wu X, Xu J, Tu S, Zhang Y, Chen H, Cao B. 2020. [Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study](#). The Lancet 395:1054–1062.
84. Richardson S, Hirsch JS, Narasimhan M, Crawford JM, McGinn T, Davidson KW, Barnaby DP, Becker LB, Chelico JD, Cohen SL, Cookingham J, Coppa K, Diefenbach MA, Dominello AJ, Duer-Hefele J, Falzon L, Gitlin J, Hajizadeh N, Harvin TG, Hirschwerk DA, Kim EJ, Kozel ZM, Marrast LM, Mogavero JN, Osorio GA, Qiu M, Zanos TP, and the

Northwell COVID-19 Research Consortium. 2020. [Presenting Characteristics, Comorbidities, and Outcomes Among 5700 Patients Hospitalized With COVID-19 in the New York City Area](#). JAMA 323:2052.

85. Goyal P, Choi JJ, Pinheiro LC, Schenck EJ, Chen R, Jabri A, Satlin MJ, Campion TR, Nahid M, Ringel JB, Hoffman KL, Alshak MN, Li HA, Wehmeyer GT, Rajan M, Reshetnyak E, Hupert N, Horn EM, Martinez FJ, Gulick RM, Safford MM. 2020. [Clinical Characteristics of Covid-19 in New York City](#). New England Journal of Medicine 382:2372–2374.
86. Burke RM, Killerby ME, Newton S, Ashworth CE, Berns AL, Brennan S, Bressler JM, Bye E, Crawford R, Harduar Morano L, Lewis NM, Markus TM, Read JS, Rissman T, Taylor J, Tate JE, Midgley CM, Balachandran N, Dahl RM, Dott M, Gilani Z, Grober A, Leung J, O'Hegarty M, Person J, Ricaldi JN, Roth NM, Sejvar JJ, Shimabukuro T, Tran CH, Watson JT, Whitham H, Chiou H, Clogher P, Duca LM, Dratch A, Feldpausch A, Fill M-M, Ghinai I, Holshue M, Scott S, Westergaard R, Case Investigation Form Working Group, Case Investigation Form Working Group. 2020. [Symptom Profiles of a Convenience Sample of Patients with COVID-19 — United States, January–April 2020](#). MMWR Morbidity and Mortality Weekly Report 69:904–908.
87. Allen WE, Altae-Tran H, Briggs J, Jin X, McGee G, Shi A, Raghavan R, Kamariza M, Nova N, Pereta A, Danford C, Kamel A, Gothe P, Milam E, Aurambault J, Primke T, Li W, Inkenbrandt J, Huynh T, Chen E, Lee C, Croatto M, Bentley H, Lu W, Murray R, Travassos M, Coull BA, Openshaw J, Greene CS, Shalem O, King G, Probasco R, Cheng DR, Silbermann B, Zhang F, Lin X. 2020. [Population-scale longitudinal mapping of COVID-19 symptoms, behaviour and testing](#). Nat Hum Behav 4:972–982.
88. Gupta A, Madhavan MV, Sehgal K, Nair N, Mahajan S, Sehrawat TS, Bikdeli B, Ahluwalia N, Ausiello JC, Wan EY, Freedberg DE, Kirtane AJ, Parikh SA, Maurer MS, Nordvig AS, Accili D, Bathon JM, Mohan S, Bauer KA, Leon MB, Krumholz HM, Uriel N, Mehra MR, Elkind MSV, Stone GW, Schwartz A, Ho DD, Bilezikian JP, Landry DW. 2020. [Extrapulmonary manifestations of COVID-19](#). Nature Medicine 26:1017–1032.
89. Hirsch JS, Ng JH, Ross DW, Sharma P, Shah HH, Barnett RL, Hazzan AD, Fishbane S, Jhaveri KD, Abate M, Andrade HP, Barnett RL, Bellucci A, Bhaskaran MC, Corona AG, Chang BF, Finger M, Fishbane S, Gitman M, Halinski C, Hasan S, Hazzan AD, Hirsch JS, Hong S, Jhaveri KD, Khanin Y, Kuan A, Madireddy V, Malieckal D, Muzib A, Nair G, Nair VV, Ng JH, Parikh R, Ross DW, Sakhiya V, Sachdeva M, Schwarz R, Shah HH, Sharma P, Singhal PC, Uppal NN, Wanchoo R, Bessy Suyin Flores Chang, Ng JHwei. 2020. [Acute kidney injury in patients hospitalized with COVID-19](#). Kidney International 98:209–218.
90. Velez JCQ, Caza T, Larsen CP. 2020. [COVAN is the new HIVAN: the re-emergence of collapsing glomerulopathy with COVID-19](#). Nat Rev Nephrol 16:565–567.
91. Wu Y, Xu X, Chen Z, Duan J, Hashimoto K, Yang L, Liu C, Yang C. 2020. [Nervous system involvement after infection with COVID-19 and other coronaviruses](#). Brain, Behavior, and Immunity 87:18–22.

92. Ellul MA, Benjamin L, Singh B, Lant S, Michael BD, Easton A, Kneen R, Defres S, Sejvar J, Solomon T. 2020. [Neurological associations of COVID-19](#). The Lancet Neurology 19:767–783.
93. Boldrini M, Canoll PD, Klein RS. 2021. [How COVID-19 Affects the Brain](#). JAMA Psychiatry 78:682.
94. Finsterer J, Stollberger C. 2020. [Update on the neurology of COVID-19](#). Journal of Medical Virology 92:2316–2318.
95. Koralnik IJ, Tyler KL. 2020. [COVID-19: A Global Threat to the Nervous System](#). Annals of Neurology 88:1–11.
96. Meinhardt J, Radke J, Dittmayer C, Franz J, Thomas C, Mothes R, Laue M, Schneider J, Brünink S, Greuel S, Lehmann M, Hassan O, Aschman T, Schumann E, Chua RL, Conrad C, Eils R, Stenzel W, Windgassen M, Rößler L, Goebel H-H, Gelderblom HR, Martin H, Nitsche A, Schulz-Schaeffer WJ, Hakrrouch S, Winkler MS, Tampe B, Scheibe F, Körtvélyessy P, Reinhold D, Siegmund B, Kühl AA, Elezkurtaj S, Horst D, Oesterhelweg L, Tsokos M, Ingold-Heppner B, Stadelmann C, Drosten C, Corman VM, Radbruch H, Heppner FL. 2020. [Olfactory transmucosal SARS-CoV-2 invasion as a port of central nervous system entry in individuals with COVID-19](#). Nature Neuroscience 24:168–175.
97. Thakur KT, Miller EH, Glendinning MD, Al-Dalahmah O, Banu MA, Boehme AK, Boubour AL, Bruce SS, Chong AM, Claassen J, Faust PL, Hargus G, Hickman RA, Jambawalikar S, Khandji AG, Kim CY, Klein RS, Lignelli-Dipple A, Lin C-C, Liu Y, Miller ML, Moonis G, Nordvig AS, Overdevest JB, Prust ML, Przedborski S, Roth WH, Soung A, Tanji K, Teich AF, Agalliu D, Uhlemann A-C, Goldman JE, Canoll P. 2021. [COVID-19 neuropathology at Columbia University Irving Medical Center/New York Presbyterian Hospital](#). Brain 144:2696–2708.
98. Oxley TJ, Mocco J, Majidi S, Kellner CP, Shoirah H, Singh IP, De Leacy RA, Shigematsu T, Ladner TR, Yaeger KA, Skliut M, Weinberger J, Dangayach NS, Bederson JB, Tuhim S, Fifi JT. 2020. [Large-Vessel Stroke as a Presenting Feature of Covid-19 in the Young](#). New England Journal of Medicine 382:e60.
99. Klok FA, Kruip MJHA, van der Meer NJM, Arbous MS, Gommers DAMPJ, Kant KM, Kaptein FHJ, van Paassen J, Stals MAM, Huisman MV, Endeman H. 2020. [Incidence of thrombotic complications in critically ill ICU patients with COVID-19](#). Thrombosis Research 191:145–147.
100. Zhang Y, Xiao M, Zhang S, Xia P, Cao W, Jiang W, Chen H, Ding X, Zhao H, Zhang H, Wang C, Zhao J, Sun X, Tian R, Wu W, Wu D, Ma J, Chen Y, Zhang D, Xie J, Yan X, Zhou X, Liu Z, Wang J, Du B, Qin Y, Gao P, Qin X, Xu Y, Zhang W, Li T, Zhang F, Zhao Y, Li Y, Zhang S. 2020. [Coagulopathy and Antiphospholipid Antibodies in Patients with Covid-19](#). New England Journal of Medicine 382:e38.
101. Tang N, Li D, Wang X, Sun Z. 2020. [Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia](#). Journal of Thrombosis and Haemostasis 18:844–847.

102. Goeijenbier M, van Wissen M, van de Weg C, Jong E, Gerdes VEA, Meijers JCM, Brandjes DPM, van Gorp ECM. 2012. [Review: Viral infections and mechanisms of thrombosis and bleeding](#). Journal of Medical Virology 84:1680–1696.
103. McGonagle D, O'Donnell JS, Sharif K, Emery P, Bridgewood C. 2020. [Immune mechanisms of pulmonary intravascular coagulopathy in COVID-19 pneumonia](#). The Lancet Rheumatology 2:e437–e445.
104. Guglielmetti G, Quaglia M, Sainaghi PP, Castello LM, Vaschetto R, Pirisi M, Corte FD, Avanzi GC, Stratta P, Cantaluppi V. 2020. [“War to the knife” against thromboinflammation to protect endothelial function of COVID-19 patients](#). Critical Care 24:365.
105. Becker RC. 2020. [COVID-19 update: Covid-19-associated coagulopathy](#). Journal of Thrombosis and Thrombolysis 50:54–67.
106. Java A, Apicelli AJ, Liszewski MK, Coler-Reilly A, Atkinson JP, Kim AHJ, Kulkarni HS. 2020. [The complement system in COVID-19: friend and foe?](#) JCI Insight 5:e140711.
107. Song W-C, FitzGerald GA. 2020. [COVID-19, microangiopathy, hemostatic activation, and complement](#). Journal of Clinical Investigation 10.1172/JCI140183.
108. Nalbandian A, Sehgal K, Gupta A, Madhavan MV, McGroder C, Stevens JS, Cook JR, Nordvig AS, Shalev D, Sehrawat TS, Ahluwalia N, Bikdeli B, Dietz D, Der-Nigoghossian C, Liyanage-Don N, Rosner GF, Bernstein EJ, Mohan S, Beckley AA, Seres DS, Choueiri TK, Uriel N, Ausiello JC, Accili D, Freedberg DE, Baldwin M, Schwartz A, Brodie D, Garcia CK, Elkind MSV, Connors JM, Bilezikian JP, Landry DW, Wan EY. 2021. [Post-acute COVID-19 syndrome](#). Nat Med 27:601–615.
109. Horwitz LI, Garry K, Prete AM, Sharma S, Mendoza F, Kahan T, Karpel H, Duan E, Hochman KA, Weerahandi H. 2021. [Six-Month Outcomes in Patients Hospitalized with Severe COVID-19](#). J GEN INTERN MED 36:3772–3777.
110. Buonsenso D, Munblit D, De Rose C, Sinatti D, Ricchiuto A, Carfi A, Valentini P. 2021. [Preliminary evidence on long COVID in children](#). Acta Paediatr 110:2208–2211.
111. Davis HE, Assaf GS, McCorkell L, Wei H, Low RJ, Re'em Y, Redfield S, Austin JP, Akrami A. 2021. [Characterizing long COVID in an international cohort: 7 months of symptoms and their impact](#). EClinicalMedicine 38:101019.
112. Huang C, Huang L, Wang Y, Li X, Ren L, Gu X, Kang L, Guo L, Liu M, Zhou X, Luo J, Huang Z, Tu S, Zhao Y, Chen L, Xu D, Li Y, Li C, Peng L, Li Y, Xie W, Cui D, Shang L, Fan G, Xu J, Wang G, Wang Y, Zhong J, Wang C, Wang J, Zhang D, Cao B. 2021. [6-month consequences of COVID-19 in patients discharged from hospital: a cohort study](#). The Lancet 397:220–232.
113. Rando HM, Bennett TD, Byrd JB, Bramante C, Callahan TJ, Chute CG, Davis HE, Deer R, Gagnier J, Koraishy FM, Liu F, McMurry JA, Moffitt RA,

- Pfaff ER, Reese JT, Relevo R, Robinson PN, Saltz JH, Solomonides A, Sule A, Topaloglu U, Haendel MA. 2021. [Challenges in defining Long COVID: Striking differences across literature, Electronic Health Records, and patient-reported information](#). Cold Spring Harbor Laboratory.
114. Deer RR, Rock MA, Vasilevsky N, Carmody L, Rando H, Anzalone AJ, Callahan TJ, Bramante CT, Chute CG, Greene CS, Gagnier J, Chu H, Koraishy FM, Liang C, Liu F, Madlock-Brown CR, Mazzotti DR, McNair DS, Parker AM, Coleman BD, Davis HE, Perry MA, Reese JT, Saltz J, Solomonides AE, Sule AA, Stein GS, Köhler S, Monteith TS, Madhira V, Kimble WD, Kavuluru R, Hillegass WB, Chan LE, Byrd JB, Boudreau EA, Liu H, McMurphy JA, Pfaff E, Matentzoglou N, Relevo R, Moffitt RA, Schuff RA, Solway J, Spratt H, Bergquist T, Bennett TD, Basson MD, Topaloglu U, Wang L, Haendel MA, Robinson PN. 2021. [Characterizing Long COVID: Deep Phenotype of a Complex Condition](#). Cold Spring Harbor Laboratory.
115. Fontanet A, Grant R, Tondeur L, Madec Y, Grzelak L, Cailleau I, Ungeheuer M-N, Renaudat C, Pellerin SF, Kuhmel L, Staropoli I, Anna F, Charneau P, Demeret C, Bruel T, Schwartz O, Hoen B. 2020. SARS-CoV-2 infection in primary schools in northern France: A retrospective cohort study in an area of high transmission. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.06.25.20140178>.
116. Lu X, Zhang L, Du H, Zhang J, Li YY, Qu J, Zhang W, Wang Y, Bao S, Li Y, Wu C, Liu H, Liu D, Shao J, Peng X, Yang Y, Liu Z, Xiang Y, Zhang F, Silva RM, Pinkerton KE, Shen K, Xiao H, Xu S, Wong GWK. 2020. [SARS-CoV-2 Infection in Children](#). New England Journal of Medicine 382:1663–1665.
117. Ludvigsson JF. 2020. [Systematic review of COVID-19 in children shows milder cases and a better prognosis than adults](#). Acta Paediatrica 109:1088–1095.
118. Lordan R, FitzGerald GA, Grosser T. 2020. [Reopening schools during COVID-19](#). Science 369:1146–1146.
119. Castagnoli R, Votto M, Licari A, Brambilla I, Bruno R, Perlini S, Rovida F, Baldanti F, Marseglia GL. 2020. [Severe Acute Respiratory Syndrome Coronavirus 2 \(SARS-CoV-2\) Infection in Children and Adolescents](#). JAMA Pediatrics 174:882.
120. Abdel-Mannan O, Eyre M, Löbel U, Bamford A, Eltze C, Hameed B, Hemingway C, Hachon Y. 2020. [Neurologic and Radiographic Findings Associated With COVID-19 Infection in Children](#). JAMA Neurology 77:1440.
121. Parri N, Lenge M, Buonsenso D. 2020. [Children with Covid-19 in Pediatric Emergency Departments in Italy](#). New England Journal of Medicine 383:187–190.
122. CDC. 2020. COVID Data Tracker. Centers for Disease Control and Prevention. <https://covid.cdc.gov/covid-data-tracker>. Retrieved 5 December 2022.
123. Mahase E. 2021. [Delta variant: What is happening with transmission, hospital admissions, and restrictions?](#) BMJ n1513.

124. Anthes E. 2021. [The Delta Variant Is Sending More Children to the Hospital. Are They Sicker, Too?](#) The New York Times.
125. Hoang A, Chorath K, Moreira A, Evans M, Burmeister-Morton F, Burmeister F, Naqvi R, Petershack M, Moreira A. 2020. [COVID-19 in 7780 pediatric patients: A systematic review](#). EClinicalMedicine 24:100433.
126. Koskela U, Helve O, Sarvikivi E, Helminen M, Nieminen T, Peltola V, Renko M, Saxén H, Pasma H, Pokka T, Honkila M, Tapiainen T. 2021. [Multi-inflammatory syndrome and Kawasaki disease in children during the COVID-19 pandemic: A nationwide register-based study and time series analysis](#). Acta Paediatr 110:3063–3068.
127. Chiotos K, Bassiri H, Behrens EM, Blatz AM, Chang J, Diorio C, Fitzgerald JC, Topjian A, John ARO. 2020. [Multisystem Inflammatory Syndrome in Children During the Coronavirus 2019 Pandemic: A Case Series](#). Journal of the Pediatric Infectious Diseases Society 9:393–398.
128. Whittaker E, Bamford A, Kenny J, Kaforou M, Jones CE, Shah P, Ramnarayan P, Fraisse A, Miller O, Davies P, Kucera F, Brierley J, McDougall M, Carter M, Tremoulet A, Shimizu C, Herberg J, Burns JC, Lyall H, Levin M, for the PIMS-TS Study Group and EUCLIDS and PERFORM Consortia. 2020. [Clinical Characteristics of 58 Children With a Pediatric Inflammatory Multisystem Syndrome Temporally Associated With SARS-CoV-2](#). JAMA 324:259.
129. Greene AG, Saleh M, Roseman E, Sinert R. 2020. [Toxic shock-like syndrome and COVID-19: Multisystem inflammatory syndrome in children \(MIS-C\)](#). The American Journal of Emergency Medicine 38:2492.e5–2492.e6.
130. Diorio C, Henrickson SE, Vella LA, McNerney KO, Chase J, Burudpakdee C, Lee JH, Jasen C, Balamuth F, Barrett DM, Banwell BL, Bernt KM, Blatz AM, Chiotos K, Fisher BT, Fitzgerald JC, Gerber JS, Gollomp K, Gray C, Grupp SA, Harris RM, Kilbaugh TJ, John ARO, Lambert M, Liebling EJ, Paessler ME, Petrosa W, Phillips C, Reilly AF, Romberg ND, Seif A, Sesok-Pizzini DA, Sullivan KE, Vardaro J, Behrens EM, Teachey DT, Bassiri H. 2020. [Multisystem inflammatory syndrome in children and COVID-19 are distinct presentations of SARS-CoV-2](#). Journal of Clinical Investigation 130:5967–5975.
131. Consiglio CR, Cotugno N, Sardh F, Pou C, Amodio D, Rodriguez L, Tan Z, Zicari S, Ruggiero A, Pascucci GR, Santilli V, Campbell T, Bryceson Y, Eriksson D, Wang J, Marchesi A, Lakshmikanth T, Campana A, Villani A, Rossi P, Landegren N, Palma P, Brodin P. 2020. [The Immunology of Multisystem Inflammatory Syndrome in Children with COVID-19](#). Cell 183:968–981.e7.
132. Belhadjer Z, Méot M, Bajolle F, Khraiche D, Legendre A, Abakka S, Auriau J, Grimaud M, Oualha M, Beghetti M, Wacker J, Ovaert C, Hascoet S, Selegny M, Malekzadeh-Milani S, Maltret A, Bosser G, Giroux N, Bonnemaïns L, Bordet J, Di Filippo S, Mauran P, Falcon-Eicher S, Thambo J-B, Lefort B, Moceri P, Houyel L, Renolleau S, Bonnet D. 2020. [Acute Heart Failure in Multisystem Inflammatory Syndrome in](#)

[Children in the Context of Global SARS-CoV-2 Pandemic](#). Circulation 142:429–436.

133. Shaigany S, Gnirke M, Guttman A, Chong H, Meehan S, Raabe V, Louie E, Solitar B, Femia A. 2020. [An adult with Kawasaki-like multisystem inflammatory syndrome associated with COVID-19](#). The Lancet 396:e8–e10.
134. Nune A, Iyengar KP, Goddard C, Ahmed AE. 2021. [Multisystem inflammatory syndrome in an adult following the SARS-CoV-2 vaccine \(MIS-V\)](#). BMJ Case Rep 14:e243888.
135. Sokolovsky S, Soni P, Hoffman T, Kahn P, Scheers-Masters J. 2021. [COVID-19 associated Kawasaki-like multisystem inflammatory disease in an adult](#). The American Journal of Emergency Medicine 39:253.e1–253.e2.
136. Boudhabhay I, Rabant M, Roumenina LT, Couprie L-M, Poillerat V, Marchal A, Frémeaux-Bacchi V, El Karoui K, Monchi M, Pourcine F. 2021. [Case Report: Adult Post-COVID-19 Multisystem Inflammatory Syndrome and Thrombotic Microangiopathy](#). Front Immunol 12.
137. Feldstein LR, Tenforde MW, Friedman KG, Newhams M, Rose EB, Dapul H, Soma VL, Maddux AB, Mourani PM, Bowens C, Maamari M, Hall MW, Riggs BJ, Giuliano JS Jr, Singh AR, Li S, Kong M, Schuster JE, McLaughlin GE, Schwartz SP, Walker TC, Loftis LL, Hobbs CV, Halasa NB, Doymaz S, Babbitt CJ, Hume JR, Gertz SJ, Irby K, Clouser KN, Cvijanovich NZ, Bradford TT, Smith LS, Heidemann SM, Zackai SP, Wellnitz K, Nofziger RA, Horwitz SM, Carroll RW, Rowan CM, Tarquinio KM, Mack EH, Fitzgerald JC, Coates BM, Jackson AM, Young CC, Son MBF, Patel MM, Newburger JW, Randolph AG. 2021. [Characteristics and Outcomes of US Children and Adolescents With Multisystem Inflammatory Syndrome in Children \(MIS-C\) Compared With Severe Acute COVID-19](#). JAMA 325:1074.
138. Buonsenso D, Munblit D, De Rose C, Sinatti D, Ricchiuto A, Carfi A, Valentini P. 2021. [Preliminary Evidence on Long COVID in children](#). Cold Spring Harbor Laboratory.
139. Brackel CLH, Lap CR, Buddingh EP, van Houten MA, van der Sande LJT, Langereis EJ, Bannier MAGE, Pijnenburg MWH, Hashimoto S, Terheggen-Lagro SWJ. 2021. [Pediatric long-COVID: An overlooked phenomenon?](#) Pediatric Pulmonology 56:2495–2502.
140. Say D, Crawford N, McNab S, Wurzel D, Steer A, Tosif S. 2021. [Post-acute COVID-19 outcomes in children with mild and asymptomatic disease](#). The Lancet Child & Adolescent Health 5:e22–e23.
141. Radtke T, Ulyte A, Puhan MA, Kriemler S. 2021. [Long-term symptoms after SARS-CoV-2 infection in school children: population-based cohort with 6-months follow-up](#). Cold Spring Harbor Laboratory.
142. Radtke T, Ulyte A, Puhan MA, Kriemler S. 2021. [Long-term Symptoms After SARS-CoV-2 Infection in Children and Adolescents](#). JAMA 326:869.

143. Mallapaty S. 2021. [Kids and COVID: why young immune systems are still on top](#). Nature 597:166–168.
144. Cicchese JM, Evans S, Hult C, Joslyn LR, Wessler T, Millar JA, Marino S, Cilfone NA, Mattila JT, Linderman JJ, Kirschner DE. 2018. [Dynamic balance of pro- and anti-inflammatory signals controls disease and limits pathology](#). Immunological Reviews 285:147–167.
145. Elenkov IJ, Iezzoni DG, Daly A, Harris AG, Chrousos GP. 2005. [Cytokine Dysregulation, Inflammation and Well-Being](#). Neuroimmunomodulation 12:255–269.
146. Chen L, Deng H, Cui H, Fang J, Zuo Z, Deng J, Li Y, Wang X, Zhao L. 2017. [Inflammatory responses and inflammation-associated diseases in organs](#). Oncotarget 9:7204–7218.
147. 2002. Molecular biology of the cell 4th ed. Garland Science, New York.
148. Widmaier EP, Raff H, Strang KT. 2008. Vander's human physiology: the mechanisms of body function. McGraw-Hill Higher Education, Boston.
149. McKechnie JL, Blish CA. 2020. [The Innate Immune System: Fighting on the Front Lines or Fanning the Flames of COVID-19?](#) Cell Host & Microbe 27:863–869.
150. Tisoncik JR, Korth MJ, Simmons CP, Farrar J, Martin TR, Katze MG. 2012. [Into the Eye of the Cytokine Storm](#). Microbiology and Molecular Biology Reviews 76:16–32.
151. Hall MJ, Williams SN, DeFrances CJ, Golosinskiy A. 2011. [Inpatient care for septicemia or sepsis: a challenge for patients and hospitals](#). NCHS Data Brief 1–8.
152. Gu X, Zhou F, Wang Y, Fan G, Cao B. 2020. [Respiratory viral sepsis: epidemiology, pathophysiology, diagnosis and treatment](#). Eur Respir Rev 29:200038.
153. Li H, Liu L, Zhang D, Xu J, Dai H, Tang N, Su X, Cao B. 2020. [SARS-CoV-2 and viral sepsis: observations and hypotheses](#). The Lancet 395:1517–1520.
154. PARK WILLIAM Y, GOODMAN RICHARD B, STEINBERG KENNETH P, RUZINSKI JOHN T, RADELLA F, PARK DAVID R, PUGIN J, SKERRETT SHAWN J, HUDSON LEONARD D, MARTIN THOMAS R. 2001. [Cytokine Balance in the Lungs of Patients with Acute Respiratory Distress Syndrome](#). American Journal of Respiratory and Critical Care Medicine 164:1896–1903.
155. Shimabukuro-Vornhagen A, Gödel P, Subklewe M, Stemmler HJ, Schlößer HA, Schlaak M, Kochanek M, Böll B, von Bergwelt-Baildon MS. 2018. [Cytokine release syndrome](#). Journal for ImmunoTherapy of Cancer 6:56.
156. He L, Ding Y, Zhang Q, Che X, He Y, Shen H, Wang H, Li Z, Zhao L, Geng J, Deng Y, Yang L, Li J, Cai J, Qiu L, Wen K, Xu X, Jiang S. 2006. [Expression of elevated levels of pro-inflammatory cytokines in SARS-CoV-infected](#)

[ACE2 cells in SARS patients: relation to the acute lung injury and pathogenesis of SARS](#). The Journal of Pathology 210:288–297.

157. Wang W, Ye L, Ye L, Li B, Gao B, Zeng Y, Kong L, Fang X, Zheng H, Wu Z, She Y. 2007. [Up-regulation of IL-6 and TNF- \$\alpha\$ induced by SARS-coronavirus spike protein in murine macrophages via NF- \$\kappa\$ B pathway](#). Virus Research 128:1–8.
158. Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ. 2020. [COVID-19: consider cytokine storm syndromes and immunosuppression](#). The Lancet 395:1033–1034.
159. Mangalmurti N, Hunter CA. 2020. [Cytokine Storms: Understanding COVID-19](#). Immunity 53:19–25.
160. Kellum JA. 2007. [Understanding the Inflammatory Cytokine Response in Pneumonia and Sepsis](#). Archives of Internal Medicine 167:1655.
161. Scheller J, Chalaris A, Schmidt-Arras D, Rose-John S. 2011. [The pro- and anti-inflammatory properties of the cytokine interleukin-6](#). Biochimica et Biophysica Acta (BBA) - Molecular Cell Research 1813:878–888.
162. Velazquez-Salinas L, Verdugo-Rodriguez A, Rodriguez LL, Borca MV. 2019. [The Role of Interleukin 6 During Viral Infections](#). Frontiers in Microbiology 10:1057.
163. Sinha P, Matthay MA, Calfee CS. 2020. [Is a “Cytokine Storm” Relevant to COVID-19?](#) JAMA Internal Medicine 180:1152.
164. Liu B, Li M, Zhou Z, Guan X, Xiang Y. 2020. [Can we use interleukin-6 \(IL-6\) blockade for coronavirus disease 2019 \(COVID-19\)-induced cytokine release syndrome \(CRS\)?](#) Journal of Autoimmunity 111:102452.
165. Eckhardt M, Hultquist JF, Kaake RM, Hüttenhain R, Krogan NJ. 2020. [A systems approach to infectious disease](#). Nature Reviews Genetics 21:339–354.
166. Ray S, Patel SK, Kumar V, Damahe J, Srivastava S. 2014. [Differential expression of serum/plasma proteins in various infectious diseases: Specific or nonspecific signatures](#). PROTEOMICS - Clinical Applications 8:53–72.
167. Blanco-Melo D, Nilsson-Payant BE, Liu W-C, Uhl S, Hoagland D, Möller R, Jordan TX, Oishi K, Panis M, Sachs D, Wang TT, Schwartz RE, Lim JK, Albrecht RA, tenOever BR. 2020. [Imbalanced Host Response to SARS-CoV-2 Drives Development of COVID-19](#). Cell 181:1036–1045.e9.
168. Versteeg GA, García-Sastre A. 2010. [Viral tricks to grid-lock the type I interferon system](#). Current Opinion in Microbiology 13:508–516.
169. Stewart CE, Randall RE, Adamson CS. 2014. [Inhibitors of the Interferon Response Enhance Virus Replication In Vitro](#). PLoS ONE 9:e112014.
170. Kopecky-Bromberg SA, Martínez-Sobrido L, Frieman M, Baric RA, Palese P. 2007. [Severe Acute Respiratory Syndrome Coronavirus Open](#)

[Reading Frame \(ORF\) 3b, ORF 6, and Nucleocapsid Proteins Function as Interferon Antagonists.](#) J Virol 81:548–557.

171. Niemeyer D, Zillinger T, Muth D, Zielecki F, Horvath G, Suliman T, Barchet W, Weber F, Drosten C, Müller MA. 2013. [Middle East Respiratory Syndrome Coronavirus Accessory Protein 4a Is a Type I Interferon Antagonist.](#) J Virol 87:12489–12495.
172. Wang P-H, Cheng Y. 2020. Increasing Host Cellular Receptor—Angiotensin-Converting Enzyme 2 (ACE2) Expression by Coronavirus may Facilitate 2019-nCoV Infection. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.24.963348>.
173. Yuen C-K, Lam J-Y, Wong W-M, Mak L-F, Wang X, Chu H, Cai J-P, Jin D-Y, To KK-W, Chan JF-W, Yuen K-Y, Kok K-H. 2020. [SARS-CoV-2 nsp13, nsp14, nsp15 and orf6 function as potent interferon antagonists.](#) Emerging Microbes & Infections 9:1418–1428.
174. Konno Y, Kimura I, Uriu K, Fukushi M, Irie T, Koyanagi Y, Sauter D, Gifford RJ, Nakagawa S, Sato K. 2020. [SARS-CoV-2 ORF3b Is a Potent Interferon Antagonist Whose Activity Is Increased by a Naturally Occurring Elongation Variant.](#) Cell Reports 32:108185.
175. Emanuel W, Kirstin M, Vedran F, Asija D, Theresa GL, Roberto A, Filippos K, David K, Salah A, Christopher B, Anja R, Ivano L, Andranik I, Tommaso M, Simone DG, Patrick PJ, Alexander MM, Daniela N, Matthias S, Altuna A, Nikolaus R, Christian D, Markus L. 2020. Bulk and single-cell gene expression profiling of SARS-CoV-2 infected human cell lines identifies molecular targets for therapeutic intervention. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.05.05.079194>.
176. Harcourt J, Tamin A, Lu X, Kamili S, Sakthivel SKumar, Murray J, Queen K, Tao Y, Paden CR, Zhang J, Li Y, Uehara A, Wang H, Goldsmith C, Bullock HA, Wang L, Whitaker B, Lynch B, Gautam R, Schindewolf C, Lokugamage KG, Scharton D, Plante JA, Mirchandani D, Widen SG, Narayanan K, Makino S, Ksiazek TG, Plante KS, Weaver SC, Lindstrom S, Tong S, Menachery VD, Thornburg NJ. 2020. Isolation and characterization of SARS-CoV-2 from the first US COVID-19 patient. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.02.972935>.
177. Aschenbrenner AC, Mouktaroudi M, Krämer B, Oestreich M, Antonakos N, Nuesch-Germano M, Gkizeli K, Bonaguro L, Reusch N, Baßler K, Saridaki M, Knoll R, Pecht T, Kapellos TS, Doulou S, Kröger C, Herbert M, Holsten L, Horne A, Gemünd ID, Rovina N, Agrawal S, Dahm K, van Uelft M, Drews A, Lenkeit L, Bruse N, Gerretsen J, Gierlich J, Becker M, Händler K, Kraut M, Theis H, Mengiste S, De Domenico E, Schulte-Schrepping J, Seep L, Raabe J, Hoffmeister C, ToVinh M, Keitel V, Rieke G, Talevi V, Skowasch D, Aziz NA, Pickkers P, van de Veerdonk FL, Netea MG, Schultze JL, Kox M, Breteler MMB, Nattermann J, Koutsoukou A, Giamarellos-Bourboulis EJ, Ulas T. 2021. [Disease severity-specific neutrophil signatures in blood transcriptomes stratify COVID-19 patients.](#) Genome Med 13.

178. Bernardes JP, Mishra N, Tran F, Bahmer T, Best L, Blase JI, Bordoni D, Franzenburg J, Geisen U, Josephs-Spaulling J, Köhler P, Künstner A, Rosati E, Aschenbrenner AC, Bacher P, Baran N, Boysen T, Brandt B, Bruse N, Dörr J, Dräger A, Elke G, Ellinghaus D, Fischer J, Forster M, Franke A, Franzenburg S, Frey N, Friedrichs A, Fuß J, Glück A, Hamm J, Hinrichsen F, Hoepfner MP, Imm S, Junker R, Kaiser S, Kan YH, Knoll R, Lange C, Laue G, Lier C, Lindner M, Marinos G, Markewitz R, Nattermann J, Noth R, Pickkers P, Rabe KF, Renz A, Röcken C, Rupp J, Schaffarzyk A, Scheffold A, Schulte-Schrepping J, Schunk D, Skowasch D, Ulas T, Wandinger K-P, Wittig M, Zimmermann J, Busch H, Hoyer BF, Kaleta C, Heyckendorf J, Kox M, Rybníček J, Schreiber S, Schultze JL, Rosenstiel P, Banovich NE, Desai T, Eickelberg O, Haniffa M, Horvath P, Kropski JA, Lafyatis R, Lundeborg J, Meyer K, Nawijn MC, Nikolic M, Ordoñas Montañes J, Pe'er D, Tata PR, Rawlins E, Regev A, Reyfman P, Samakovlis C, Schultze J, Shalek A, Shepherd D, Spence J, Teichmann S, Theis F, Tsankov A, van den Berge M, von Papen M, Whitsett J, Zaragosi LE, Angelov A, Bals R, Bartholomäus A, Becker A, Bezdan D, Bonifacio E, Bork P, Clavel T, Colme-Tatche M, Diefenbach A, Dilthey A, Fischer N, Förstner K, Frick J-S, Gagneur J, Goesmann A, Hain T, Hummel M, Janssen S, Kalinowski J, Kallies R, Kehr B, Keller A, Kim-Hellmuth S, Klein C, Kohlbacher O, Korbel JO, Kurth I, Landthaler M, Li Y, Ludwig K, Makarewicz O, Marz M, McHardy A, Mertes C, Nöthen M, Nürnberg P, Ohler U, Ossowski S, Overmann J, Peter S, Pfeffer K, Poetsch AR, Pühler A, Rajewsky N, Ralser M, Rieß O, Ripke S, Nunes da Rocha U, Rosenstiel P, Saliba A-E, Sander LE, Sawitzki B, Schiffer P, Schulte E-C, Schultze JL, Sczyrba A, Stegle O, Stoye J, Theis F, Vehreschild J, Vogel J, von Kleist M, Walker A, Walter J, Wieczorek D, Ziebuhr J. 2020. [Longitudinal Multi-omics Analyses Identify Responses of Megakaryocytes, Erythroid Cells, and Plasmablasts as Hallmarks of Severe COVID-19](#). Immunity 53:1296–1314.e9.
179. Xiong Y, Liu Y, Cao L, Wang D, Guo M, Jiang A, Guo D, Hu W, Yang J, Tang Z, Wu H, Lin Y, Zhang M, Zhang Q, Shi M, Liu Y, Zhou Y, Lan K, Chen Y. 2020. [Transcriptomic characteristics of bronchoalveolar lavage fluid and peripheral blood mononuclear cells in COVID-19 patients](#). Emerging Microbes & Infections 9:761–770.
180. Liu C, Martins AJ, Lau WW, Rachmaninoff N, Chen J, Imberti L, Mostaghimi D, Fink DL, Burbelo PD, Dobbs K, Delmonte OM, Bansal N, Failla L, Sottini A, Quiros-Roldan E, Han KL, Sellers BA, Cheung F, Sparks R, Chun T-W, Moir S, Lionakis MS, Rossi C, Su HC, Kuhns DB, Cohen JI, Notarangelo LD, Tsang JS, Abers MS, Apps R, Bosticardo M, Milanez-Almeida P, Mulè MP, Shaw E, Zhang Y, Castelli F, Muiesan ML, Tomasoni G, Scolari F, Tucci A. 2021. [Time-resolved systems immunology reveals a late juncture linked to fatal COVID-19](#). Cell 184:1836–1857.e22.
181. Prokop JW, Hartog NL, Chesla D, Faber W, Love CP, Karam R, Abualkheir N, Feldmann B, Teng L, McBride T, Leimanis ML, English BK, Holsworth A, Frisch A, Bauss J, Kalpage N, Derbedrossian A, Pinti RM, Hale N, Mills J, Eby A, VanSickle EA, Pageau SC, Shankar R, Chen B, Carcillo JA, Sanfilippo D, Olivero R, Bupp CP, Rajasekaran S. 2021. [High-Density Blood Transcriptomics Reveals Precision Immune Signatures of SARS-CoV-2 Infection in Hospitalized Individuals](#). Front Immunol 12.

182. Arunachalam PS, Wimmers F, Mok CKP, Perera RAPM, Scott M, Hagan T, Sigal N, Feng Y, Bristow L, Tak-Yin Tsang O, Wagh D, Collier J, Pellegrini KL, Kazmin D, Alaaeddine G, Leung WS, Chan JMC, Chik TSH, Choi CYC, Huerta C, Paine McCullough M, Lv H, Anderson E, Edupuganti S, Upadhyay AA, Bosinger SE, Maecker HT, Khatri P, Rouphael N, Peiris M, Pulendran B. 2020. [Systems biological assessment of immunity to mild versus severe COVID-19 infection in humans](#). *Science* 369:1210–1220.
183. Cheemarla NR, Watkins TA, Mihaylova VT, Wang B, Zhao D, Wang G, Landry ML, Foxman EF. 2021. [Dynamic innate immune response determines susceptibility to SARS-CoV-2 infection and early replication kinetics](#). *Journal of Experimental Medicine* 218.
184. Yang AC, Kern F, Losada PM, Agam MR, Maat CA, Schmartz GP, Fehlmann T, Stein JA, Schaum N, Lee DP, Calcuttawala K, Vest RT, Berdnik D, Lu N, Hahn O, Gate D, McNERney MW, Channappa D, Cobos I, Ludwig N, Schulz-Schaeffer WJ, Keller A, Wyss-Coray T. 2021. [Dysregulation of brain and choroid plexus cell types in severe COVID-19](#). *Nature* 595:565–571.
185. Bojkova D, Klann K, Koch B, Widera M, Krause D, Ciesek S, Cinatl J, Münch C. 2020. [Proteomics of SARS-CoV-2-infected host cells reveals therapy targets](#). *Nature* 583:469–472.
186. Ju B, Zhang Q, Ge X, Wang R, Yu J, Shan S, Zhou B, Song S, Tang X, Yu J, Ge J, Lan J, Yuan J, Wang H, Zhao J, Zhang S, Wang Y, Shi X, Liu L, Wang X, Zhang Z, Zhang L. 2020. Potent human neutralizing antibodies elicited by SARS-CoV-2 infection. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.21.990770>.
187. Chen J-H, Chang Y-W, Yao C-W, Chiueh T-S, Huang S-C, Chien K-Y, Chen A, Chang F-Y, Wong C-H, Chen Y-J. 2004. [Plasma proteome of severe acute respiratory syndrome analyzed by two-dimensional gel electrophoresis and mass spectrometry](#). *Proceedings of the National Academy of Sciences* 101:17039–17044.
188. He R, Dobie F, Ballantine M, Leeson A, Li Y, Bastien N, Cutts T, Andonov A, Cao J, Booth TF, Plummer FA, Tyler S, Baker L, Li X. 2004. [Analysis of multimerization of the SARS coronavirus nucleocapsid protein](#). *Biochemical and Biophysical Research Communications* 316:476–483.
189. The UniProt Consortium. 2019. [UniProt: a worldwide hub of protein knowledge](#). *Nucleic Acids Research* 47:D506–D515.
190. Home - Genome - NCBI. <https://www.ncbi.nlm.nih.gov/genome>. Retrieved 8 February 2021.
191. Vita R, Mahajan S, Overton JA, Dhanda SK, Martini S, Cantrell JR, Wheeler DK, Sette A, Peters B. 2019. [The Immune Epitope Database \(IEDB\): 2018 update](#). *Nucleic Acids Research* 47:D339–D343.
192. Pickett BE, Sadat EL, Zhang Y, Noronha JM, Squires RB, Hunt V, Liu M, Kumar S, Zaremba S, Gu Z, Zhou L, Larson CN, Dietrich J, Klem EB, Scheuermann RH. 2012. [ViPR: an open bioinformatics database and](#)

[analysis resource for virology research](#). Nucleic Acids Research 40:D593–D598.

193. Gordon DE, Jang GM, Bouhaddou M, Xu J, Obernier K, White KM, O'Meara MJ, Rezelj VV, Guo JZ, Swaney DL, Tummino TA, Hüttenhain R, Kaake RM, Richards AL, Tutuncuoglu B, Foussard H, Batra J, Haas K, Modak M, Kim M, Haas P, Polacco BJ, Braberg H, Fabius JM, Eckhardt M, Soucheray M, Bennett MJ, Cakir M, McGregor MJ, Li Q, Meyer B, Roesch F, Vallet T, Mac Kain A, Miorin L, Moreno E, Naing ZZC, Zhou Y, Peng S, Shi Y, Zhang Z, Shen W, Kirby IT, Melnyk JE, Chorba JS, Lou K, Dai SA, Barrio-Hernandez I, Memon D, Hernandez-Armenta C, Lyu J, Mathy CJP, Perica T, Pilla KB, Ganesan SJ, Saltzberg DJ, Rakesh R, Liu X, Rosenthal SB, Calviello L, Venkataramanan S, Liboy-Lugo J, Lin Y, Huang X-P, Liu Y, Wankowicz SA, Bohn M, Safari M, Ugur FS, Koh C, Savar NS, Tran QD, Shengjuler D, Fletcher SJ, O'Neal MC, Cai Y, Chang JC, Broadhurst DJ, Klippsten S, Sharp PP, Wenzell NA, Kuzuoglu-Ozturk D, Wang H-Y, Trenker R, Young JM, Cavero DA, Hiatt J, Roth TL, Rathore U, Subramanian A, Noack J, Hubert M, Stroud RM, Frankel AD, Rosenberg OS, Verba KA, Agard DA, Ott M, Emerman M, Jura N, von Zastrow M, Verdin E, Ashworth A, Schwartz O, d'Enfert C, Mukherjee S, Jacobson M, Malik HS, Fujimori DG, Ideker T, Craik CS, Floor SN, Fraser JS, Gross JD, Sali A, Roth BL, Ruggero D, Taunton J, Kortemme T, Beltrao P, Vignuzzi M, García-Sastre A, Shokat KM, Shoichet BK, Krogan NJ. 2020. [A SARS-CoV-2 protein interaction map reveals targets for drug repurposing](#). Nature 583:459–468.
194. Sobocińska J, Roszczenko-Jasińska P, Ciesielska A, Kwiatkowska K. 2018. [Protein Palmitoylation and Its Role in Bacterial and Viral Infections](#). Frontiers in Immunology 8:2003.
195. Li J, Guo M, Tian X, Liu C, Wang X, Yang X, Wu P, Xiao Z, Qu Y, Yin Y, Fu J, Zhu Z, Liu Z, Peng C, Zhu T, Liang Q. 2020. Virus-host interactome and proteomic survey of PMBCs from COVID-19 patients reveal potential virulence factors influencing SARS-CoV-2 pathogenesis. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.31.019216>.
196. Lawrence T. 2009. [The Nuclear Factor NF- B Pathway in Inflammation](#). Cold Spring Harbor Perspectives in Biology 1:a001651–a001651.
197. Navratil V, Lionnard L, Longhi S, Hardwick JM, Combet C, Aouacheria A. 2020. The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) envelope (E) protein harbors a conserved BH3-like sequence. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.04.09.033522>.
198. Overmyer KA, Shishkova E, Miller IJ, Balnis J, Bernstein MN, Peters-Clarke TM, Meyer JG, Quan Q, Muehlbauer LK, Trujillo EA, He Y, Chopra A, Chieng HC, Tiwari A, Judson MA, Paulson B, Brademan DR, Zhu Y, Serrano LR, Linke V, Drake LA, Adam AP, Schwartz BS, Singer HA, Swanson S, Mosher DF, Stewart R, Coon JJ, Jaitovich A. 2021. [Large-Scale Multi-omic Analysis of COVID-19 Severity](#). Cell Systems 12:23–40.e7.
199. Geyer PE, Arend FM, Doll S, Louiset M, Virreira Winter S, Müller-Reif JB, Torun FM, Weigand M, Eichhorn P, Bruegel M, Strauss MT, Holdt LM, Mann M, Teupser D. 2021. [High-resolution serum proteome](#)

[trajectories in COVID-19 reveal patient-specific seroconversion](#). EMBO Mol Med 13.

200. Li F. 2005. [Structure of SARS Coronavirus Spike Receptor-Binding Domain Complexed with Receptor](#). Science 309:1864–1868.
201. Yan R, Zhang Y, Li Y, Xia L, Guo Y, Zhou Q. 2020. [Structural basis for the recognition of SARS-CoV-2 by full-length human ACE2](#). Science 367:1444–1448.
202. Shang J, Ye G, Shi K, Wan Y, Luo C, Aihara H, Geng Q, Auerbach A, Li F. 2020. [Structural basis of receptor recognition by SARS-CoV-2](#). Nature 581:221–224.
203. Lan J, Ge J, Yu J, Shan S, Zhou H, Fan S, Zhang Q, Shi X, Wang Q, Zhang L, Wang X. 2020. Crystal structure of the 2019-nCoV spike receptor-binding domain bound with the ACE2 receptor. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.19.956235>.
204. Wang Q, Zhang Y, Wu L, Niu S, Song C, Zhang Z, Lu G, Qiao C, Hu Y, Yuen K-Y, Wang Q, Zhou H, Yan J, Qi J. 2020. [Structural and Functional Basis of SARS-CoV-2 Entry by Using Human ACE2](#). Cell 181:894–904.e9.
205. Wan Y, Shang J, Graham R, Baric RS, Li F. 2020. [Receptor Recognition by the Novel Coronavirus from Wuhan: an Analysis Based on Decade-Long Structural Studies of SARS Coronavirus](#). Journal of Virology 94:e00127–20.
206. Wrobel AG, Benton DJ, Xu P, Roustan C, Martin SR, Rosenthal PB, Skehel JJ, Gamblin SJ. 2020. [SARS-CoV-2 and bat RaTG13 spike glycoprotein structures inform on virus evolution and furin-cleavage effects](#). Nat Struct Mol Biol 27:763–767.
207. Zhou P, Yang X-L, Wang X-G, Hu B, Zhang L, Zhang W, Si H-R, Zhu Y, Li B, Huang C-L, Chen H-D, Chen J, Luo Y, Guo H, Jiang R-D, Liu M-Q, Chen Y, Shen X-R, Wang X, Zheng X-S, Zhao K, Chen Q-J, Deng F, Liu L-L, Yan B, Zhan F-X, Wang Y-Y, Xiao G-F, Shi Z-L. 2020. [A pneumonia outbreak associated with a new coronavirus of probable bat origin](#). Nature 579:270–273.
208. Chen J, Lee KH, Steinhauer DA, Stevens DJ, Skehel JJ, Wiley DC. 1998. [Structure of the Hemagglutinin Precursor Cleavage Site, a Determinant of Influenza Pathogenicity and the Origin of the Labile Conformation](#). Cell 95:409–417.
209. Steinhauer DA. 1999. [Role of Hemagglutinin Cleavage for the Pathogenicity of Influenza Virus](#). Virology 258:1–20.
210. Pachetti M, Marini B, Benedetti F, Giudici F, Mauro E, Storici P, Masciovecchio C, Angeletti S, Ciccozzi M, Gallo RC, Zella D, Ippodrino R. 2020. [Emerging SARS-CoV-2 mutation hot spots include a novel RNA-dependent-RNA polymerase variant](#). Journal of Translational Medicine 18:179.
211. Tang X, Wu C, Li X, Song Y, Yao X, Wu X, Duan Y, Zhang H, Wang Y, Qian Z, Cui J, Lu J. 2020. [On the origin and continuing evolution of SARS-CoV-](#)

212. van Dorp L, Acman M, Richard D, Shaw LP, Ford CE, Ormond L, Owen CJ, Pang J, Tan CCS, Boshier FAT, Ortiz AT, Balloux F. 2020. [Emergence of genomic diversity and recurrent mutations in SARS-CoV-2](#). *Infection, Genetics and Evolution* 83:104351.
213. Lu J, du Plessis L, Liu Z, Hill V, Kang M, Lin H, Sun J, François S, Kraemer MUG, Faria NR, McCrone JT, Peng J, Xiong Q, Yuan R, Zeng L, Zhou P, Liang C, Yi L, Liu J, Xiao J, Hu J, Liu T, Ma W, Li W, Su J, Zheng H, Peng B, Fang S, Su W, Li K, Sun R, Bai R, Tang X, Liang M, Quick J, Song T, Rambaut A, Loman N, Raghwani J, Pybus OG, Ke C. 2020. [Genomic Epidemiology of SARS-CoV-2 in Guangdong Province, China](#). *Cell* 181:997–1003.e9.
214. 2020. [An integrated national scale SARS-CoV-2 genomic surveillance network](#). *The Lancet Microbe* 1:e99–e100.
215. CDC. 2020. Cases, Data, and Surveillance. Centers for Disease Control and Prevention. <https://www.cdc.gov/coronavirus/2019-ncov/variants/spheres.html>. Retrieved 5 December 2022.
216. Fauver JR, Petrone ME, Hodcroft EB, Shioda K, Ehrlich HY, Watts AG, Vogels CBF, Brito AF, Alpert T, Muyombwe A, Razeq J, Downing R, Cheemarla NR, Wyllie AL, Kalinich CC, Ott IM, Quick J, Loman NJ, Neugebauer KM, Greninger AL, Jerome KR, Roychoudhury P, Xie H, Shrestha L, Huang M-L, Pitzer VE, Iwasaki A, Omer SB, Khan K, Bogoch II, Martinello RA, Foxman EF, Landry ML, Neher RA, Ko AI, Grubaugh ND. 2020. [Coast-to-Coast Spread of SARS-CoV-2 during the Early Epidemic in the United States](#). *Cell* 181:990–996.e5.
217. Gonzalez-Reiche AS, Hernandez MM, Sullivan MJ, Ciferri B, Alshammary H, Obla A, Fabre S, Kleiner G, Polanco J, Khan Z, Alburquerque B, van de Guchte A, Dutta J, Francoeur N, Melo BS, Oussenko I, Deikus G, Soto J, Sridhar SH, Wang Y-C, Twyman K, Kasarskis A, Altman DR, Smith M, Sebra R, Aberg J, Krammer F, García-Sastre A, Luksza M, Patel G, Paniz-Mondolfi A, Gitman M, Sordillo EM, Simon V, van Bakel H. 2020. [Introductions and early spread of SARS-CoV-2 in the New York City area](#). *Science* eabc1917.
218. Gudbjartsson DF, Helgason A, Jonsson H, Magnusson OT, Melsted P, Norddahl GL, Saemundsdottir J, Sigurdsson A, Sulem P, Agustsdottir AB, Eirisdottir B, Fridriksdottir R, Gardarsdottir EE, Georgsson G, Gretarsdottir OS, Gudmundsson KR, Gunnarsdottir TR, Gylfason A, Holm H, Jensson BO, Jonasdottir A, Jonsson F, Josefsdottir KS, Kristjansson T, Magnusdottir DN, le Roux L, Sigmundsdottir G, Sveinbjornsson G, Sveinsdottir KE, Sveinsdottir M, Thorarensen EA, Thorbjornsson B, Löve A, Masson G, Jonsdottir I, Möller AD, Gudnason T, Kristinsson KG, Thorsteinsdottir U, Stefansson K. 2020. [Spread of SARS-CoV-2 in the Icelandic Population](#). *New England Journal of Medicine* 382:2302–2315.
219. GISAID - Initiative. <https://www.gisaid.org/>. Retrieved 8 February 2021.

220. NCBI SARS-CoV-2 Resources. <https://www.ncbi.nlm.nih.gov/sars-cov-2/>. Retrieved 8 February 2021.
221. COVID-19 Data Portal - accelerating scientific research through data. <https://www.covid19dataportal.org/>. Retrieved 8 February 2021.
222. Korber B, Fischer WM, Gnanakaran S, Yoon H, Theiler J, Abfalterer W, Hengartner N, Giorgi EE, Bhattacharya T, Foley B, Hastie KM, Parker MD, Partridge DG, Evans CM, Freeman TM, de Silva TI, McDanal C, Perez LG, Tang H, Moon-Walker A, Whelan SP, LaBranche CC, Saphire EO, Montefiori DC, Angyal A, Brown RL, Carrilero L, Green LR, Groves DC, Johnson KJ, Keeley AJ, Lindsey BB, Parsons PJ, Raza M, Rowland-Jones S, Smith N, Tucker RM, Wang D, Wyles MD. 2020. [Tracking Changes in SARS-CoV-2 Spike: Evidence that D614G Increases Infectivity of the COVID-19 Virus](#). Cell 182:812–827.e19.
223. Yurkovetskiy L, Wang X, Pascal KE, Tomkins-Tinch C, Nyalile TP, Wang Y, Baum A, Diehl WE, Dauphin A, Carbone C, Veinotte K, Egri SB, Schaffner SF, Lemieux JE, Munro JB, Rafique A, Barve A, Sabeti PC, Kyratsous CA, Dudkina NV, Shen K, Luban J. 2020. [Structural and Functional Analysis of the D614G SARS-CoV-2 Spike Protein Variant](#). Cell 183:739–751.e8.
224. Plante JA, Liu Y, Liu J, Xia H, Johnson BA, Lokugamage KG, Zhang X, Muruato AE, Zou J, Fontes-Garfias CR, Mirchandani D, Scharton D, Bilello JP, Ku Z, An Z, Kalveram B, Freiberg AN, Menachery VD, Xie X, Plante KS, Weaver SC, Shi P-Y. 2020. [Spike mutation D614G alters SARS-CoV-2 fitness](#). Nature 592:116–121.
225. Thomson EC, Rosen LE, Shepherd JG, Spreafico R, da Silva Filipe A, Wojcechowskyj JA, Davis C, Piccoli L, Pascall DJ, Dillen J, Lytras S, Czudnochowski N, Shah R, Meury M, Jesudason N, De Marco A, Li K, Bassi J, O'Toole A, Pinto D, Colquhoun RM, Culap K, Jackson B, Zatta F, Rambaut A, Jaconi S, Sreenu VB, Nix J, Zhang I, Jarrett RF, Glass WG, Beltramello M, Nomikou K, Pizzuto M, Tong L, Cameroni E, Croll TI, Johnson N, Di Iulio J, Wickenhagen A, Ceschi A, Harbison AM, Mair D, Ferrari P, Smollett K, Sallusto F, Carmichael S, Garzoni C, Nichols J, Galli M, Hughes J, Riva A, Ho A, Schiuma M, Semple MG, Openshaw PJM, Fadda E, Baillie JK, Chodera JD, Rihn SJ, Lycett SJ, Virgin HW, Telenti A, Corti D, Robertson DL, Snell G. 2021. [Circulating SARS-CoV-2 spike N439K variants maintain fitness while evading antibody-mediated immunity](#). Cell 184:1171–1187.e20.
226. Young BE, Fong S-W, Chan Y-H, Mak T-M, Ang LW, Anderson DE, Lee CY-P, Amrun SN, Lee B, Goh YS, Su YCF, Wei WE, Kalimuddin S, Chai LYA, Pada S, Tan SY, Sun L, Parthasarathy P, Chen YYC, Barkham T, Lin RTP, Maurer-Stroh S, Leo Y-S, Wang L-F, Renia L, Lee VJ, Smith GJD, Lye DC, Ng LFP. 2020. [Effects of a major deletion in the SARS-CoV-2 genome on the severity of infection and the inflammatory response: an observational cohort study](#). The Lancet 396:603–611.
227. Liu Z, Zheng H, Lin H, Li M, Yuan R, Peng J, Xiong Q, Sun J, Li B, Wu J, Yi L, Peng X, Zhang H, Zhang W, Hulswit RJG, Loman N, Rambaut A, Ke C, Bowden TA, Pybus OG, Lu J. 2020. [Identification of Common Deletions](#)

[in the Spike Protein of Severe Acute Respiratory Syndrome Coronavirus 2](#). J Virol 94.

228. Tao K, Tzou PL, Nouhin J, Gupta RK, de Oliveira T, Kosakovsky Pond SL, Fera D, Shafer RW. 2021. [The biological and clinical significance of emerging SARS-CoV-2 variants](#). Nat Rev Genet 22:757–773.
229. Grabowski F, Preibisch G, Giziński S, Kochańczyk M, Lipniacki T. 2021. [SARS-CoV-2 Variant of Concern 20212/01 Has about Twofold Replicative Advantage and Acquires Concerning Mutations](#). Viruses 13:392.
230. Kemp S, Harvey W, Datir R, Collier D, Ferreira I, Meng B, Carabelii A, Robertson DL, Gupta RK, COVID-19 Genomics UK (COG-UK) consortium. 2021. Recurrent emergence and transmission of a SARS-CoV-2 Spike deletion H69/V70. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.12.14.422555>.
231. B.1.351 report. https://cov-lineages.org/global_report_B.1.351.html. Retrieved 8 February 2021.
232. Ramanathan M, Ferguson ID, Miao W, Khavari PA. 2021. [SARS-CoV-2 B.1.1.7 and B.1.351 spike variants bind human ACE2 with increased affinity](#). The Lancet Infectious Diseases 21:1070.
233. Spratt AN, Kannan SR, Woods LT, Weisman GA, Quinn TP, Lorson CL, Sönnernborg A, Byrareddy SN, Singh K. 2021. [Evolution, correlation, structural impact and dynamics of emerging SARS-CoV-2 variants](#). Computational and Structural Biotechnology Journal 19:3799–3809.
234. Volz E, Mishra S, Chand M, Barrett JC, Johnson R, Geidelberg L, Hinsley WR, Laydon DJ, Dabrera G, O'Toole Á, Amato R, Ragonnet-Cronin M, Harrison I, Jackson B, Ariani CV, Boyd O, Loman NJ, McCrone JT, Gonçalves S, Jorgensen D, Myers R, Hill V, Jackson DK, Gaythorpe K, Groves N, Sillitoe J, Kwiatkowski DP, Flaxman S, Ratmann O, Bhatt S, Hopkins S, Gandy A, Rambaut A, Ferguson NM, The COVID-19 Genomics UK (COG-UK) consortium. 2021. Transmission of SARS-CoV-2 Lineage B.1.1.7 in England: Insights from linking epidemiological and genetic data. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.12.30.20249034>.
235. Laffeber C, de Koning K, Kanaar R, Lebbink JHG. 2021. [Experimental Evidence for Enhanced Receptor Binding by Rapidly Spreading SARS-CoV-2 Variants](#). Journal of Molecular Biology 433:167058.
236. Ali F, Kasry A, Amin M. 2021. [The new SARS-CoV-2 strain shows a stronger binding affinity to ACE2 due to N501Y mutant](#). Medicine in Drug Discovery 10:100086.
237. Hoffmann M, Arora P, Groß R, Seidel A, Hörnich BF, Hahn AS, Krüger N, Graichen L, Hofmann-Winkler H, Kempf A, Winkler MS, Schulz S, Jäck H-M, Jahrsdörfer B, Schrezenmeier H, Müller M, Kleger A, Münch J, Pöhlmann S. 2021. [SARS-CoV-2 variants B.1.351 and P.1 escape from neutralizing antibodies](#). Cell 184:2384–2393.e12.

238. Tracking SARS-CoV-2 variants. <https://www.who.int/activities/tracking-SARS-CoV-2-variants>. Retrieved 5 December 2022.
239. Saito A, Nasser H, Uriu K, Kosugi Y, Irie T, Shirakawa K, Sadamasu K, Kimura I, Ito J, Wu J, Ozono S, Tokunaga K, Butlertanaka EP, Tanaka YL, Shimizu R, Shimizu K, Fukuhara T, Kawabata R, Sakaguchi T, Yoshida I, Asakura H, Nagashima M, Yoshimura K, Kazuma Y, Nomura R, Horisawa Y, Takaori-Kondo A, Nakagawa S, Ikeda T, Sato K. 2021. [SARS-CoV-2 spike P681R mutation enhances and accelerates viral fusion](#). Cold Spring Harbor Laboratory.
240. Singanayagam A, Patel M, Charlett A, Lopez Bernal J, Saliba V, Ellis J, Ladhani S, Zambon M, Gopal R. 2020. [Duration of infectiousness and correlation with RT-PCR cycle threshold values in cases of COVID-19, England, January to May 2020](#). Eurosurveillance 25.
241. La Rosa G, Fratini M, Della Libera S, Iaconelli M, Muscillo M. 2013. [Viral infections acquired indoors through airborne, droplet or contact transmission](#). Ann Ist Super Sanita 49:124–32.
242. Shiu EYC, Leung NHL, Cowling BJ. 2019. [Controversy around airborne versus droplet transmission of respiratory viruses](#). Current Opinion in Infectious Diseases 32:372–379.
243. Tang JW, Bahnfleth WP, Bluyssen PM, Buonanno G, Jimenez JL, Kurnitski J, Li Y, Miller S, Sekhar C, Morawska L, Marr LC, Melikov AK, Nazaroff WW, Nielsen PV, Tellier R, Wargocki P, Dancer SJ. 2021. [Dismantling myths on the airborne transmission of severe acute respiratory syndrome coronavirus-2 \(SARS-CoV-2\)](#). Journal of Hospital Infection 110:89–96.
244. Randall K, Ewing ET, Marr L, Jimenez J, Bourouiba L. 2021. [How Did We Get Here: What Are Droplets and Aerosols and How Far Do They Go? A Historical Perspective on the Transmission of Respiratory Infectious Diseases](#). SSRN.
245. 2020. Transmission of SARS-CoV-2: implications for infection prevention precautions. <https://www.who.int/news-room/commentaries/detail/transmission-of-sars-cov-2-implications-for-infection-prevention-precautions>. Retrieved 8 February 2021.
246. Lemieux C, Brankston G, Gitterman L, Hirji Z, Gardam M. 2007. [Questioning Aerosol Transmission of Influenza](#). Emerging Infectious Diseases 13:173–175.
247. Smieszek T, Lazzari G, Salathé M. 2019. [Assessing the Dynamics and Control of Droplet- and Aerosol-Transmitted Influenza Using an Indoor Positioning System](#). Scientific Reports 9:2185.
248. Richard M, Fouchier RAM. 2016. [Influenza A virus transmission via respiratory aerosols or droplets as it relates to pandemic potential](#). FEMS Microbiology Reviews 40:68–85.
249. Weiss SR, Leibowitz JL. 2011. [Coronavirus Pathogenesis](#)Advances in Virus Research. Elsevier BV.

250. de Wit E, van Doremalen N, Falzarano D, Munster VJ. 2016. [SARS and MERS: recent insights into emerging coronaviruses](#). Nature Reviews Microbiology 14:523–534.
251. Li Y, Huang X, Yu ITS, Wong TW, Qian H. 2005. [Role of air distribution in SARS transmission during the largest nosocomial outbreak in Hong Kong](#). Indoor Air 15:83–95.
252. Booth TF, Kournikakis B, Bastien N, Ho J, Kobasa D, Stadnyk L, Li Y, Spence M, Paton S, Henry B, Mederski B, White D, Low DE, McGeer A, Simor A, Vearncombe M, Downey J, Jamieson FB, Tang P, Plummer F. 2005. [Detection of Airborne Severe Acute Respiratory Syndrome \(SARS\) Coronavirus and Environmental Contamination in SARS Outbreak Units](#). The Journal of Infectious Diseases 191:1472–1477.
253. Xiao S, Li Y, Wong T, Hui DSC. 2017. [Role of fomites in SARS transmission during the largest hospital outbreak in Hong Kong](#). PLOS ONE 12:e0181558.
254. van Doremalen N, Bushmaker T, Munster VJ. 2013. [Stability of Middle East respiratory syndrome coronavirus \(MERS-CoV\) under different environmental conditions](#). Eurosurveillance 18.
255. Mackay IM, Arden KE. 2015. [MERS coronavirus: diagnostics, epidemiology and transmission](#). Virology Journal 12:222.
256. van Doremalen N, Bushmaker T, Morris DH, Holbrook MG, Gamble A, Williamson BN, Tamin A, Harcourt JL, Thornburg NJ, Gerber SI, Lloyd-Smith JO, de Wit E, Munster VJ. 2020. [Aerosol and Surface Stability of SARS-CoV-2 as Compared with SARS-CoV-1](#). New England Journal of Medicine 382:1564–1567.
257. Peng X, Xu X, Li Y, Cheng L, Zhou X, Ren B. 2020. [Transmission routes of 2019-nCoV and controls in dental practice](#). International Journal of Oral Science 12:9.
258. Klompas M, Baker MA, Rhee C. 2020. [Airborne Transmission of SARS-CoV-2](#). JAMA 324:441.
259. Goldman E. 2020. [Exaggerated risk of transmission of COVID-19 by fomites](#). The Lancet Infectious Diseases 20:892–893.
260. Prather KA, Wang CC, Schooley RT. 2020. [Reducing transmission of SARS-CoV-2](#). Science 368:1422–1424.
261. Morawska L, Milton DK. 2020. [It Is Time to Address Airborne Transmission of Coronavirus Disease 2019 \(COVID-19\)](#). Clinical Infectious Diseases ciaa939.
262. Greenhalgh T, Jimenez JL, Prather KA, Tufekci Z, Fisman D, Schooley R. 2021. [Ten scientific reasons in support of airborne transmission of SARS-CoV-2](#). The Lancet 397:1603–1605.
263. Liu Y, Ning Z, Chen Y, Guo M, Liu Y, Gali NK, Sun L, Duan Y, Cai J, Westerdahl D, Liu X, Xu K, Ho K, Kan H, Fu Q, Lan K. 2020. [Aerodynamic analysis of SARS-CoV-2 in two Wuhan hospitals](#). Nature 582:557–560.

264. Lednicky JA, Lauzardo M, Fan ZH, Jutla A, Tilly TB, Gangwar M, Usmani M, Shankar SN, Mohamed K, Eiguren-Fernandez A, Stephenson CJ, Alam MdM, Elbadry MA, Loeb JC, Subramaniam K, Waltzek TB, Cherabuddi K, Morris JG Jr., Wu C-Y. 2020. [Viable SARS-CoV-2 in the air of a hospital room with COVID-19 patients](#). International Journal of Infectious Diseases 100:476–482.
265. Santarpia JL, Herrera VL, Rivera DN, Ratnesar-Shumate S, Reid StP, Denton PW, Martens JWS, Fang Y, Conoan N, Callahan MV, Lawler JV, Brett-Major DM, Lowe JJ. 2020. [The Infectious Nature of Patient-Generated SARS-CoV-2 Aerosol](#). Cold Spring Harbor Laboratory.
266. Santarpia JL, Rivera DN, Herrera VL, Morwitzer MJ, Creager HM, Santarpia GW, Crown KK, Brett-Major DM, Schnaubelt ER, Broadhurst MJ, Lawler JV, Reid StP, Lowe JJ. 2020. [Aerosol and surface contamination of SARS-CoV-2 observed in quarantine and isolation care](#). Sci Rep 10.
267. Zhou J, Otter JA, Price JR, Cimpeanu C, Meno Garcia D, Kinross J, Boshier PR, Mason S, Bolt F, Holmes AH, Barclay WS. 2020. [Investigating Severe Acute Respiratory Syndrome Coronavirus 2 \(SARS-CoV-2\) Surface and Air Contamination in an Acute Healthcare Setting During the Peak of the Coronavirus Disease 2019 \(COVID-19\) Pandemic in London](#). Clinical Infectious Diseases 73:e1870–e1877.
268. Comber L, O Murchu E, Drummond L, Carty PG, Walsh KA, De Gascun CF, Connolly MA, Smith SM, O'Neill M, Ryan M, Harrington P. 2020. [Airborne transmission of SARS-CoV-2 via aerosols](#). Rev Med Virol 31.
269. Bourouiba L. 2020. Turbulent Gas Clouds and Respiratory Pathogen Emissions. JAMA <https://doi.org/10.1001/jama.2020.4756>.
270. Hu Z, Song C, Xu C, Jin G, Chen Y, Xu X, Ma H, Chen W, Lin Y, Zheng Y, Wang J, Hu Z, Yi Y, Shen H. 2020. [Clinical characteristics of 24 asymptomatic infections with COVID-19 screened among close contacts in Nanjing, China](#). Science China Life Sciences 63:706–711.
271. Tindale LC, Stockdale JE, Coombe M, Garlock ES, Lau WYV, Saraswat M, Zhang L, Chen D, Wallinga J, Colijn C. 2020. [Evidence for transmission of COVID-19 prior to symptom onset](#). eLife 9:e57149.
272. Chang D, Mo G, Yuan X, Tao Y, Peng X, Wang F-S, Xie L, Sharma L, Dela Cruz CS, Qin E. 2020. [Time Kinetics of Viral Clearance and Resolution of Symptoms in Novel Coronavirus Infection](#). American Journal of Respiratory and Critical Care Medicine 201:1150–1152.
273. He X, Lau EHY, Wu P, Deng X, Wang J, Hao X, Lau YC, Wong JY, Guan Y, Tan X, Mo X, Chen Y, Liao B, Chen W, Hu F, Zhang Q, Zhong M, Wu Y, Zhao L, Zhang F, Cowling BJ, Li F, Leung GM. 2020. [Temporal dynamics in viral shedding and transmissibility of COVID-19](#). Nature Medicine 26:672–675.
274. CDC. 2020. COVID-19 and Your Health. Centers for Disease Control and Prevention. <https://www.cdc.gov/coronavirus/2019-ncov/prevent-getting-sick/how-covid-spreads.html>. Retrieved 8 February 2021.

275. Wölfel R, Corman VM, Guggemos W, Seilmaier M, Zange S, Müller MA, Niemeyer D, Jones TC, Vollmar P, Rothe C, Hoelscher M, Bleicker T, Brünink S, Schneider J, Ehmann R, Zwirgmaier K, Drosten C, Wendtner C. 2020. [Virological assessment of hospitalized patients with COVID-2019](#). *Nature* 581:465–469.
276. Corsini Campioli C, Cano Cevallos E, Assi M, Patel R, Binnicker MJ, O'Horo JC. 2020. [Clinical predictors and timing of cessation of viral RNA shedding in patients with COVID-19](#). *Journal of Clinical Virology* 130:104577.
277. Arons MM, Hatfield KM, Reddy SC, Kimball A, James A, Jacobs JR, Taylor J, Spicer K, Bardossy AC, Oakley LP, Tanwar S, Dyal JW, Harney J, Chisty Z, Bell JM, Methner M, Paul P, Carlson CM, McLaughlin HP, Thornburg N, Tong S, Tamin A, Tao Y, Uehara A, Harcourt J, Clark S, Brostrom-Smith C, Page LC, Kay M, Lewis J, Montgomery P, Stone ND, Clark TA, Honein MA, Duchin JS, Jernigan JA. 2020. [Presymptomatic SARS-CoV-2 Infections and Transmission in a Skilled Nursing Facility](#). *New England Journal of Medicine* 382:2081–2090.
278. Baggett TP, Keyes H, Sporn N, Gaeta JM. 2020. [Prevalence of SARS-CoV-2 Infection in Residents of a Large Homeless Shelter in Boston](#). *JAMA* 323:2191.
279. Bai Y, Yao L, Wei T, Tian F, Jin D-Y, Chen L, Wang M. 2020. [Presumed Asymptomatic Carrier Transmission of COVID-19](#). *JAMA* 323:1406.
280. Li P, Fu J-B, Li K-F, Liu J-N, Wang H-L, Liu L-J, Chen Y, Zhang Y-L, Liu S-L, Tang A, Tong Z-D, Yan J-B. 2020. [Transmission of COVID-19 in the terminal stages of the incubation period: A familial cluster](#). *International Journal of Infectious Diseases* 96:452–453.
281. Zhang P, Tian F, Wan Y, Cai J, Qian Z, Wu R, Zhang Y, Zhang S, Li H, Li M, Trevathan E, Lin H. 2020. A Cohort of SARS-CoV-2 Infected Asymptomatic and Pre-Symptomatic Contacts from COVID-19 Contact Tracing in Hubei Province, China: Short-Term Outcomes. SSRN Electronic Journal <https://doi.org/10.2139/ssrn.3678556>.
282. Mizumoto K, Kagaya K, Zarebski A, Chowell G. 2020. [Estimating the asymptomatic proportion of coronavirus disease 2019 \(COVID-19\) cases on board the Diamond Princess cruise ship, Yokohama, Japan, 2020](#). *Eurosurveillance* 25.
283. Zhang K, Tong W, Wang X, Lau JY-N. 2020. [Estimated prevalence and viral transmissibility in subjects with asymptomatic SARS-CoV-2 infections in Wuhan, China](#). *Precision Clinical Medicine* 3:301–305.
284. Ladhani SN, Chow JY, Janarthanan R, Fok J, Crawley-Boevey E, Vusirikala A, Fernandez E, Perez MS, Tang S, Dun-Campbell K, Evans EW-, Bell A, Patel B, Amin-Chowdhury Z, Aiano F, Paranthaman K, Ma T, Saavedra-Campos M, Myers R, Ellis J, Lackenby A, Gopal R, Patel M, Brown C, Chand M, Brown K, Ramsay ME, Hopkins S, Shetty N, Zambon M. 2020. [Investigation of SARS-CoV-2 outbreaks in six care homes in London, April 2020](#). *EClinicalMedicine* 26:100533.

285. Long Q-X, Tang X-J, Shi Q-L, Li Q, Deng H-J, Yuan J, Hu J-L, Xu W, Zhang Y, Lv F-J, Su K, Zhang F, Gong J, Wu B, Liu X-M, Li J-J, Qiu J-F, Chen J, Huang A-L. 2020. [Clinical and immunological assessment of asymptomatic SARS-CoV-2 infections](#). *Nature Medicine* 26:1200–1204.
286. Lavezzo E, Franchin E, Ciavarella C, Cuomo-Dannenburg G, Barzon L, Del Vecchio C, Rossi L, Manganelli R, Loregian A, Navarin N, Abate D, Sciro M, Merigliano S, De Canale E, Vanuzzo MC, Besutti V, Saluzzo F, Onelia F, Pacenti M, Parisi SG, Carretta G, Donato D, Flor L, Cocchio S, Masi G, Sperduti A, Cattarino L, Salvador R, Nicoletti M, Caldart F, Castelli G, Nieddu E, Labella B, Fava L, Drigo M, Gaythorpe KAM, Brazzale AR, Toppo S, Trevisan M, Baldo V, Donnelly CA, Ferguson NM, Dorigatti I, Crisanti A, Imperial College COVID-19 Response Team, Imperial College COVID-19 Response Team. 2020. [Suppression of a SARS-CoV-2 outbreak in the Italian municipality of Vo'](#). *Nature* 584:425–429.
287. Meyerowitz-Katz G, Merone L. 2020. [A systematic review and meta-analysis of published research data on COVID-19 infection fatality rates](#). *International Journal of Infectious Diseases* 101:138–148.
288. Global Covid-19 Case Fatality Rates. The Centre for Evidence-Based Medicine. <https://www.cebm.net/covid-19/global-covid-19-case-fatality-rates/>. Retrieved 8 February 2021.
289. Grewelle R, De Leo G. 2020. Estimating the Global Infection Fatality Rate of COVID-19. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.05.11.20098780>.
290. Stadlbauer D, Tan J, Jiang K, Hernandez MM, Fabre S, Amanat F, Teo C, Arunkumar GA, McMahon M, Capuano C, Twyman K, Jhang J, Nowak MD, Simon V, Sordillo EM, van Bakel H, Krammer F. 2020. [Repeated cross-sectional sero-monitoring of SARS-CoV-2 in New York City](#). *Nature* 590:146–150.
291. What do we know about the risk of dying from COVID-19? Our World in Data. <https://ourworldindata.org/covid-mortality-risk>. Retrieved 8 February 2021.
292. Heesterbeek JAP, Dietz K. 1996. [The concept of \$R_0\$ in epidemic theory](#). *Statistica Neerlandica* 50:89–110.
293. Keeling MJ, Rohani P. 2008. Modeling infectious diseases in humans and animals. Princeton University Press, Princeton.
294. 1997. [A contribution to the mathematical theory of epidemics](#). *Proceedings of the Royal Society of London Series A, Containing Papers of a Mathematical and Physical Character* 115:700–721.
295. Anderson RM, May RM. 1979. [Population biology of infectious diseases: Part I](#). *Nature* 280:361–367.
296. Cobey S. 2020. [Modeling infectious disease dynamics](#). *Science* 368:713–714.

297. 2007. Theoretical ecology: principles and applications. Oxford University Press, Oxford ; New York.
298. Heng K, Althaus CL. 2020. [The approximately universal shapes of epidemic curves in the Susceptible-Exposed-Infectious-Recovered \(SEIR\) model](#). Sci Rep 10.
299. Wu JT, Leung K, Leung GM. 2020. [Nowcasting and forecasting the potential domestic and international spread of the 2019-nCoV outbreak originating in Wuhan, China: a modelling study](#). The Lancet 395:689–697.
300. Liu Y, Gayle AA, Wilder-Smith A, Rocklöv J. 2020. [The reproductive number of COVID-19 is higher compared to SARS coronavirus](#). Journal of Travel Medicine 27:taaa021.
301. Li R, Pei S, Chen B, Song Y, Zhang T, Yang W, Shaman J. 2020. [Substantial undocumented infection facilitates the rapid dissemination of novel coronavirus \(SARS-CoV-2\)](#). Science 368:489–493.
302. Ma S, Zhang J, Zeng M, Yun Q, Guo W, Zheng Y, Zhao S, Wang MH, Yang Z. 2020. Epidemiological parameters of coronavirus disease 2019: a pooled analysis of publicly reported individual data of 1155 cases from seven countries. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.21.20040329>.
303. Majumder M, Mandl KD. 2020. Early Transmissibility Assessment of a Novel Coronavirus in Wuhan, China. SSRN Electronic Journal <https://doi.org/10.2139/ssrn.3524675>.
304. Liu T, Hu J, Xiao J, He G, Kang M, Rong Z, Lin L, Zhong H, Huang Q, Deng A, Zeng W, Tan X, Zeng S, Zhu Z, Li J, Gong D, Wan D, Chen S, Guo L, Li Y, Sun L, Liang W, Song T, He J, Ma W. 2020. Time-varying transmission dynamics of Novel Coronavirus Pneumonia in China. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.01.25.919787>.
305. Zhang S, Diao M, Yu W, Pei L, Lin Z, Chen D. 2020. [Estimation of the reproductive number of novel coronavirus \(COVID-19\) and the probable outbreak size on the Diamond Princess cruise ship: A data-driven analysis](#). International Journal of Infectious Diseases 93:201–204.
306. Tang B, Wang X, Li Q, Bragazzi NL, Tang S, Xiao Y, Wu J. 2020. [Estimation of the Transmission Risk of the 2019-nCoV and Its Implication for Public Health Interventions](#). Journal of Clinical Medicine 9:462.
307. Cao Z, Zhang Q, Lu X, Pfeiffer D, Jia Z, Song H, Zeng DD. 2020. [Estimating the effective reproduction number of the 2019-nCoV in China](#). medRxiv 2020.01.27.20018952.
308. Shen M, Peng Z, Xiao Y, Zhang L. 2020. Modelling the epidemic trend of the 2019 novel coronavirus outbreak in China. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.01.23.916726>.

309. Read JM, Bridgen JRE, Cummings DAT, Ho A, Jewell CP. 2020. Novel coronavirus 2019-nCoV: early estimation of epidemiological parameters and epidemic predictions. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.01.23.20018549>.
310. Roques L, Klein E, Papaix J, Sar A, Soubeyrand S. 2020. Using early data to estimate the actual infection fatality ratio from COVID-19 in France. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.22.20040915>.
311. Park SW, Sun K, Viboud C, Grenfell BT, Dushoff J. 2020. [Potential Role of Social Distancing in Mitigating Spread of Coronavirus Disease, South Korea](#). Emerg Infect Dis 26:2697–2700.
312. Kucharski AJ, Russell TW, Diamond C, Liu Y, Edmunds J, Funk S, Eggo RM, Sun F, Jit M, Munday JD, Davies N, Gimma A, van Zandvoort K, Gibbs H, Hellewell J, Jarvis CI, Clifford S, Quilty BJ, Bosse NI, Abbott S, Klepac P, Flasche S. 2020. [Early dynamics of transmission and control of COVID-19: a mathematical modelling study](#). The Lancet Infectious Diseases 20:553–558.
313. Sahafizadeh E, Sartoli S. 2020. Estimating the reproduction number of COVID-19 in Iran using epidemic modeling. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.20.20038422>.
314. Flaxman S, Mishra S, Gandy A, Unwin H, Coupland H, Mellan T, Zhu H, Berah T, Eaton J, Perez Guzman P, Schmit N, Cilloni L, Ainslie K, Baguelin M, Blake I, Boonyasiri A, Boyd O, Cattarino L, Ciavarella C, Cooper L, Cucunuba Perez Z, Cuomo-Dannenburg G, Dighe A, Djaafara A, Dorigatti I, Van Elsland S, Fitzjohn R, Fu H, Gaythorpe K, Geidelberg L, Grassly N, Green W, Hallett T, Hamlet A, Hinsley W, Jeffrey B, Jorgensen D, Knock E, Laydon D, Nedjati Gilani G, Nouvellet P, Parag K, Siveroni I, Thompson H, Verity R, Volz E, Walters C, Wang H, Wang Y, Watson O, Winskill P, Xi X, Whittaker C, Walker P, Ghani A, Donnelly C, Riley S, Okell L, Vollmer M, Ferguson N, Bhatt S. 2020. [Report 13: Estimating the number of infections and the impact of non-pharmaceutical interventions on COVID-19 in 11 European countries](#). Imperial College London.
315. Campbell F, Archer B, Laurenson-Schafer H, Jinnai Y, Konings F, Batra N, Pavlin B, Vandemaele K, Van Kerkhove MD, Jombart T, Morgan O, le Polain de Waroux O. 2021. [Increased transmissibility and global spread of SARS-CoV-2 variants of concern as at June 2021](#). Eurosurveillance 26.
316. Washington NL, Gangavarapu K, Zeller M, Bolze A, Cirulli ET, Barrett KMS, Larsen BB, Anderson C, White S, Cassens T, Jacobs S, Levan G, Nguyen J, Ramirez JM III, Rivera-Garcia C, Sandoval E, Wang X, Wong D, Spencer E, Robles-Sikisaka R, Kurzban E, Hughes LD, Deng X, Wang C, Servellita V, Valentine H, De Hoff P, Seaver P, Sathe S, Gietzen K, Sickler B, Antico J, Hoon K, Liu J, Harding A, Bakhtar O, Basler T, Austin B, Isaksson M, Febbo PG, Becker D, Laurent M, McDonald E, Yeo GW, Knight R, Laurent LC, de Feo E, Worobey M, Chiu C, Suchard MA, Lu JT, Lee W, Andersen KG. 2021. [Genomic epidemiology identifies emergence and rapid transmission of SARS-CoV-2 B.1.1.7 in the United States](#). Cold Spring Harbor Laboratory.

317. Davies NG, Abbott S, Barnard RC, Jarvis CI, Kucharski AJ, Munday JD, Pearson CAB, Russell TW, Tully DC, Washburne AD, Wenseleers T, Gimma A, Waites W, Wong KLM, van Zandvoort K, Silverman JD, Diaz-Ordaz K, Keogh R, Eggo RM, Funk S, Jit M, Atkins KE, Edmunds WJ. 2021. [Estimated transmissibility and impact of SARS-CoV-2 lineage B.1.1.7 in England](#). Science 372.
318. Liu Y, Rocklöv J. 2021. [The reproductive number of the Delta variant of SARS-CoV-2 is far higher compared to the ancestral SARS-CoV-2 virus](#). Journal of Travel Medicine 28.
319. Ito K, Piantham C, Nishiura H. 2021. [Predicted dominance of variant Delta of SARS-CoV-2 before Tokyo Olympic Games, Japan, July 2021](#). Eurosurveillance 26.
320. Moghadas SM, Shoukat A, Fitzpatrick MC, Wells CR, Sah P, Pandey A, Sachs JD, Wang Z, Meyers LA, Singer BH, Galvani AP. 2020. [Projecting hospital utilization during the COVID-19 outbreaks in the United States](#). Proceedings of the National Academy of Sciences 117:9122–9126.
321. Prem K, Liu Y, Russell TW, Kucharski AJ, Eggo RM, Davies N, Jit M, Klepac P, Flasche S, Clifford S, Pearson CAB, Munday JD, Abbott S, Gibbs H, Rosello A, Quilty BJ, Jombart T, Sun F, Diamond C, Gimma A, van Zandvoort K, Funk S, Jarvis CI, Edmunds WJ, Bosse NI, Hellewell J. 2020. [The effect of control strategies to reduce social mixing on outcomes of the COVID-19 epidemic in Wuhan, China: a modelling study](#). The Lancet Public Health 5:e261–e270.
322. Gatto M, Bertuzzo E, Mari L, Miccoli S, Carraro L, Casagrandi R, Rinaldo A. 2020. [Spread and dynamics of the COVID-19 epidemic in Italy: Effects of emergency containment measures](#). Proceedings of the National Academy of Sciences 117:10484–10491.
323. EpiForecasts and the CMMID Covid working group. Covid-19: Temporal variation in transmission during the COVID-19 outbreak. <https://epiforecasts.io/covid/>. Retrieved 27 July 2020.
324. Systrom K, Vladeck T, Krieger M. Rt COVID-19. <https://rt.live/>. Retrieved 27 July 2020.
325. Zhou R, Li F, Chen F, Liu H, Zheng J, Lei C, Wu X. 2020. [Viral dynamics in asymptomatic patients with COVID-19](#). International Journal of Infectious Diseases 96:288–290.
326. Hasanoglu I, Korukluoglu G, Asilturk D, Cosgun Y, Kalem AK, Altas AB, Kayaaslan B, Eser F, Kuzucu EA, Guner R. 2020. [Higher viral loads in asymptomatic COVID-19 patients might be the invisible part of the iceberg](#). Infection 49:117–126.
327. Tsukagoshi H, Shinoda D, Saito M, Okayama K, Sada M, Kimura H, Saruki N. 2021. [Relationships between Viral Load and the Clinical Course of COVID-19](#). Viruses 13:304.
328. Byambasuren O, Cardona M, Bell K, Clark J, McLaws M-L, Glasziou P. 2020. [Estimating the extent of asymptomatic COVID-19 and its](#)

[potential for community transmission: Systematic review and meta-analysis](#). Official Journal of the Association of Medical Microbiology and Infectious Disease Canada 5:223–234.

329. Keehner J, Horton LE, Pfeffer MA, Longhurst CA, Schooley RT, Currier JS, Abeles SR, Torriani FJ. 2021. [SARS-CoV-2 Infection after Vaccination in Health Care Workers in California](#). N Engl J Med 384:1774–1775.
330. Tande AJ, Pollock BD, Shah ND, Farrugia G, Virk A, Swift M, Breeher L, Binnicker M, Berbari EF. 2021. [Impact of the Coronavirus Disease 2019 \(COVID-19\) Vaccine on Asymptomatic Infection Among Patients Undergoing Preprocedural COVID-19 Molecular Screening](#). Clinical Infectious Diseases 74:59–65.
331. Angel Y, Spitzer A, Henig O, Saiag E, Sprecher E, Padova H, Ben-Ami R. 2021. [Association Between Vaccination With BNT162b2 and Incidence of Symptomatic and Asymptomatic SARS-CoV-2 Infections Among Health Care Workers](#). JAMA 325:2457.
332. Thompson MG, Burgess JL, Naleway AL, Tyner H, Yoon SK, Meece J, Olsho LEW, Caban-Martinez AJ, Fowlkes AL, Lutrick K, Groom HC, Dunnigan K, Odean MJ, Hegmann K, Stefanski E, Edwards LJ, Schaefer-Solle N, Grant L, Ellingson K, Kuntz JL, Zunie T, Thiese MS, Ivacic L, Wesley MG, Mayo Lamberte J, Sun X, Smith ME, Phillips AL, Groover KD, Yoo YM, Gerald J, Brown RT, Herring MK, Joseph G, Beitel S, Morrill TC, Mak J, Rivers P, Poe BP, Lynch B, Zhou Y, Zhang J, Kelleher A, Li Y, Dickerson M, Hanson E, Guenther K, Tong S, Bateman A, Reisdorf E, Barnes J, Azziz-Baumgartner E, Hunt DR, Arvay ML, Kutty P, Fry AM, Gaglani M. 2021. [Prevention and Attenuation of Covid-19 with the BNT162b2 and mRNA-1273 Vaccines](#). N Engl J Med 385:320–329.
333. Levine-Tiefenbrun M, Yelin I, Katz R, Herzel E, Golan Z, Schreiber L, Wolf T, Nadler V, Ben-Tov A, Kuint J, Gazit S, Patalon T, Chodick G, Kishony R. 2021. [Initial report of decreased SARS-CoV-2 viral load after inoculation with the BNT162b2 vaccine](#). Nat Med 27:790–792.
334. Chia PY, Xiang Ong SW, Chiew CJ, Ang LW, Chavatte J-M, Mak T-M, Cui L, Kalimuddin S, Chia WN, Tan CW, Ann Chai LY, Tan SY, Zheng S, Pin Lin RT, Wang L, Leo Y-S, Lee VJ, Lye DC, Young BE. 2021. [Virological and serological kinetics of SARS-CoV-2 Delta variant vaccine-breakthrough infections: a multi-center cohort study](#). Cold Spring Harbor Laboratory.
335. Riemersma KK, Haddock LA III, Wilson NA, Minor N, Eickhoff J, Grogan BE, Kita-Yarbro A, Halfmann PJ, Segaloff HE, Kocharian A, Florek KR, Westergaard R, Bateman A, Jeppson GE, Kawaoka Y, O'Connor DH, Friedrich TC, Grande KM. 2021. [Shedding of Infectious SARS-CoV-2 Despite Vaccination](#). Cold Spring Harbor Laboratory.
336. Acharya CB, Schrom J, Mitchell AM, Coil DA, Marquez C, Rojas S, Wang CY, Liu J, Pilarowski G, Solis L, Georgian E, Petersen M, DeRisi J, Micheltore R, Havlir D. 2021. [No Significant Difference in Viral Load Between Vaccinated and Unvaccinated, Asymptomatic and Symptomatic Groups When Infected with SARS-CoV-2 Delta Variant](#). Cold Spring Harbor Laboratory.

337. Anderson RM, Fraser C, Ghani AC, Donnelly CA, Riley S, Ferguson NM, Leung GM, Lam TH, Hedley AJ. 2004. [Epidemiology, transmission dynamics and control of SARS: the 2002–2003 epidemic](#). Philosophical Transactions of the Royal Society of London Series B: Biological Sciences 359:1091–1105.
338. 2021. A timeline of the CDC's advice on face masks. Los Angeles Times. <https://www.latimes.com/science/story/2021-07-27/timeline-cdc-mask-guidance-during-covid-19-pandemic>. Retrieved 5 December 2022.
339. COVID-19 Review Consortium. 2021. [Social Factors Influencing COVID-19 Exposure and Outcomes](#). Manubot.
340. Guarner J. 2020. [Three Emerging Coronaviruses in Two Decades](#). American Journal of Clinical Pathology 153:420–421.
341. COVID-19 Review Consortium. 2021. [Evolutionary and Genomic Analysis of SARS-CoV-2](#). Manubot.
342. Cui J, Li F, Shi Z-L. 2018. [Origin and evolution of pathogenic coronaviruses](#). Nature Reviews Microbiology 17:181–192.
343. Lim Y, Ng Y, Tam J, Liu D. 2016. [Human Coronaviruses: A Review of Virus–Host Interactions](#). Diseases 4:26.
344. Killerby ME, Biggs HM, Haynes A, Dahl RM, Mustaquim D, Gerber SI, Watson JT. 2018. [Human coronavirus circulation in the United States 2014–2017](#). Journal of Clinical Virology 101:52–56.
345. Hamre D, Procknow JJ. 1966. [A New Virus Isolated from the Human Respiratory Tract](#). Experimental Biology and Medicine 121:190–193.
346. McIntosh K, Dees JH, Becker WB, Kapikian AZ, Chanock RM. 1967. [Recovery in tracheal organ cultures of novel viruses from patients with respiratory disease](#). Proceedings of the National Academy of Sciences 57:933–940.
347. Pyrc K, Jebbink MF, Berkhout B, van der Hoek L. 2004. [:\(unav\)](#). Virology Journal 1:7.
348. Abdul-Rasool S, Fielding BC. 2010. [Understanding Human Coronavirus HCoV-NL63](#). Open Virol J 4:76–84.
349. Luo X, Zhou G-Z, Zhang Y, Peng L-H, Zou L-P, Yang Y-S. 2020. [Coronaviruses and gastrointestinal diseases](#). Military Medical Research 7:49.
350. Vabret A, Dina J, Gouarin S, Petitjean J, Tripey V, Brouard J, Freymuth F. 2008. [Human \(non-severe acute respiratory syndrome\) coronavirus infections in hospitalised children in France](#). Journal of Paediatrics and Child Health 44:176–181.
351. van der Hoek L, Pyrc K, Jebbink MF, Vermeulen-Oost W, Berkhout RJM, Wolthers KC, Wertheim-van Dillen PME, Kaandorp J, Spaargaren J, Berkhout B. 2004. [Identification of a new human coronavirus](#). Nature Medicine 10:368–373.

352. Woo PCY, Lau SKP, Chu C, Chan K, Tsoi H, Huang Y, Wong BHL, Poon RWS, Cai JJ, Luk W, Poon LLM, Wong SSY, Guan Y, Peiris JSM, Yuen K. 2005. [Characterization and Complete Genome Sequence of a Novel Coronavirus, Coronavirus HKU1, from Patients with Pneumonia](#). Journal of Virology 79:884–895.
353. Pene F, Merlat A, Vabret A, Rozenberg F, Buzyn A, Dreyfus F, Cariou A, Freymuth F, Lebon P. 2003. [Coronavirus 229E-Related Pneumonia in Immunocompromised Patients](#). Clinical Infectious Diseases 37:929–932.
354. Corman VM, Muth D, Niemeyer D, Drosten C. 2018. [Hosts and Sources of Endemic Human Coronaviruses](#)Advances in Virus Research. Elsevier BV.
355. Vlasova AN, Diaz A, Damtie D, Xiu L, Toh T-H, Lee JS-Y, Saif LJ, Gray GC. 2021. [Novel Canine Coronavirus Isolated from a Hospitalized Patient With Pneumonia in East Malaysia](#). Clinical Infectious Diseases 74:446–454.
356. Wu F, Zhao S, Yu B, Chen Y-M, Wang W, Song Z-G, Hu Y, Tao Z-W, Tian J-H, Pei Y-Y, Yuan M-L, Zhang Y-L, Dai F-H, Liu Y, Wang Q-M, Zheng J-J, Xu L, Holmes EC, Zhang Y-Z. 2020. [A new coronavirus associated with human respiratory disease in China](#). Nature 579:265–269.
357. Taylor LH, Latham SM, woolhouse MEJ. 2001. [Risk factors for human disease emergence](#). Phil Trans R Soc Lond B 356:983–989.
358. Han BA, Kramer AM, Drake JM. 2016. [Global Patterns of Zoonotic Disease in Mammals](#). Trends in Parasitology 32:565–577.
359. Wolfe ND, Daszak P, Kilpatrick AM, Burke DS. 2005. [Bushmeat Hunting, Deforestation, and Prediction of Zoonotic Disease](#). Emerg Infect Dis 11:1822–1827.
360. Guan Y, Zheng BJ, He YQ, Liu XL, Zhuang ZX, Cheung CL, Luo SW, Li PH, Zhang LJ, Guan YJ, Butt KM, Wong KL, Chan KW, Lim W, Shortridge KF, Yuen KY, Peiris JSM, Poon LLM. 2003. [Isolation and Characterization of Viruses Related to the SARS Coronavirus from Animals in Southern China](#). Science 302:276–278.
361. Ye Z-W, Yuan S, Yuen K-S, Fung S-Y, Chan C-P, Jin D-Y. 2020. [Zoonotic origins of human coronaviruses](#). Int J Biol Sci 16:1686–1697.
362. Lau SKP, Woo PCY, Li KSM, Huang Y, Tsoi H-W, Wong BHL, Wong SSY, Leung S-Y, Chan K-H, Yuen K-Y. 2005. [Severe acute respiratory syndrome coronavirus-like virus in Chinese horseshoe bats](#). Proc Natl Acad Sci USA 102:14040–14045.
363. Cyranoski D. 2020. [Mystery deepens over animal source of coronavirus](#). Nature 579:18–19.
364. Ill husband and wife add to Wuhan riddle - The Standard. <https://www.thestandard.com.hk/sections-news-print/215457/III-husband-and-wife>. Retrieved 5 December 2022.

365. Xiao X, Newman C, Buesching CD, Macdonald DW, Zhou Z-M. 2021. [Animal sales from Wuhan wet markets immediately prior to the COVID-19 pandemic](#). Sci Rep 11.
366. Brook CE, Dobson AP. 2015. [Bats as 'special' reservoirs for emerging zoonotic pathogens](#). Trends in Microbiology 23:172–180.
367. Han H-J, Wen H, Zhou C-M, Chen F-F, Luo L-M, Liu J, Yu X-J. 2015. [Bats as reservoirs of severe emerging infectious diseases](#). Virus Research 205:1–6.
368. Calisher CH, Childs JE, Field HE, Holmes KV, Schountz T. 2006. [Bats: Important Reservoir Hosts of Emerging Viruses](#). Clin Microbiol Rev 19:531–545.
369. Zhang T, Wu Q, Zhang Z. 2020. [Probable Pangolin Origin of SARS-CoV-2 Associated with the COVID-19 Outbreak](#). Current Biology 30:1346–1351.e2.
370. Lam TT-Y, Jia N, Zhang Y-W, Shum MH-H, Jiang J-F, Zhu H-C, Tong Y-G, Shi Y-X, Ni X-B, Liao Y-S, Li W-J, Jiang B-G, Wei W, Yuan T-T, Zheng K, Cui X-M, Li J, Pei G-Q, Qiang X, Cheung WY-M, Li L-F, Sun F-F, Qin S, Huang J-C, Leung GM, Holmes EC, Hu Y-L, Guan Y, Cao W-C. 2020. [Identifying SARS-CoV-2-related coronaviruses in Malayan pangolins](#). Nature 583:282–285.
371. Andersen KG, Rambaut A, Lipkin WI, Holmes EC, Garry RF. 2020. [The proximal origin of SARS-CoV-2](#). Nature Medicine 26:450–452.
372. Li X, Giorgi EE, Marichannelgowda MH, Foley B, Xiao C, Kong X-P, Chen Y, Gnanakaran S, Korber B, Gao F. 2020. [Emergence of SARS-CoV-2 through recombination and strong purifying selection](#). Science Advances 6:eabb9153.
373. Banerjee A, Doxey AC, Mossman K, Irving AT. 2021. [Unraveling the Zoonotic Origin and Transmission of SARS-CoV-2](#). Trends in Ecology & Evolution 36:180–184.
374. Bloom JD, Chan YA, Baric RS, Bjorkman PJ, Cobey S, Deverman BE, Fisman DN, Gupta R, Iwasaki A, Lipsitch M, Medzhitov R, Neher RA, Nielsen R, Patterson N, Stearns T, van Nimwegen E, Worobey M, Relman DA. 2021. [Investigate the origins of COVID-19](#). Science 372:694–694.
375. Maxmen A. 2021. [Divisive COVID 'lab leak' debate prompts dire warnings from researchers](#). Nature 594:15–16.
376. Bender JB, Shulman SA, __. 2004. [Reports of zoonotic disease outbreaks associated with animal exhibits and availability of recommendations for preventing zoonotic disease transmission from animals to people in such settings](#). JAMA 292:1105–1109.
377. Saif LJ. 1993. [Coronavirus immunogens](#). Veterinary Microbiology 37:285–297.

378. Fermin G. 2018. [Host Range, Host-Virus Interactions, and Virus Transmission](#), p. 101–134. *In* Viruses. Elsevier.
379. Kim J, Farré M, Auvil L, Capitanu B, Larkin DM, Ma J, Lewin HA. 2017. [Reconstruction and evolutionary history of eutherian chromosomes](#). Proc Natl Acad Sci USA 114.
380. Tsagkogeorga G, Parker J, Stupka E, Cotton James A, Rossiter Stephen J. 2013. [Phylogenomic Analyses Elucidate the Evolutionary Relationships of Bats](#). Current Biology 23:2262–2267.
381. Nishihara H, Hasegawa M, Okada N. 2006. [Pegasoferae, an unexpected mammalian clade revealed by tracking ancient retroposon insertions](#). Proc Natl Acad Sci USA 103:9929–9934.
382. Murphy WJ, Pringle TH, Crider TA, Springer MS, Miller W. 2007. [Using genomic data to unravel the root of the placental mammal phylogeny](#). Genome Res 17:413–421.
383. Murphy WJ, Eizirik E. 2009. [Placental mammals \(Eutheria\)](#), p. 471–474. *In* The Timetree of Life. Oxford University Press.
384. McNamara T, Richt JA, Glickman L. 2020. [A Critical Needs Assessment for Research in Companion Animals and Livestock Following the Pandemic of COVID-19 in Humans](#). Vector-Borne and Zoonotic Diseases 20:393–405.
385. Li R, Qiao S, Zhang G. 2020. [Analysis of angiotensin-converting enzyme 2 \(ACE2\) from different species sheds some light on cross-species receptor usage of a novel coronavirus 2019-nCoV](#). Journal of Infection 80:469–496.
386. Damas J, Hughes GM, Keough KC, Painter CA, Persky NS, Corbo M, Hiller M, Koepfli K-P, Pfenning AR, Zhao H, Genereux DP, Swofford R, Pollard KS, Ryder OA, Nweeia MT, Lindblad-Toh K, Teeling EC, Karlsson EK, Lewin HA. 2020. [Broad host range of SARS-CoV-2 predicted by comparative and structural analysis of ACE2 in vertebrates](#). Proc Natl Acad Sci USA 117:22311–22322.
387. Opriessnig T, Huang Y. 2020. [Update on possible animal sources for COVID-19 in humans](#). Xenotransplantation 27.
388. Hosie MJ, Hofmann-Lehmann R, Hartmann K, Egberink H, Truyen U, Addie DD, Belák S, Boucraut-Baralon C, Frymus T, Lloret A, Lutz H, Marsilio F, Pennisi MG, Tasker S, Thiry E, Möstl K. 2021. [Anthropogenic Infection of Cats during the 2020 COVID-19 Pandemic](#). Viruses 13:185.
389. Sit THC, Brackman CJ, Ip SM, Tam KWS, Law PYT, To EMW, Yu VYT, Sims LD, Tsang DNC, Chu DKW, Perera RAPM, Poon LLM, Peiris M. 2020. [Infection of dogs with SARS-CoV-2](#). Nature 586:776–778.
390. Decaro N, Vaccari G, Lorusso A, Lorusso E, De Sabato L, Patterson EI, Di Bartolo I, Hughes GL, Teodori L, Desario C, Colitti B, Ricci D, Buonavoglia D, Rosati S, Martella V, Cammà C, Agrimi U, Elia G. 2021. [Possible Human-to-Dog Transmission of SARS-CoV-2, Italy, 2020](#). Emerg Infect Dis 27:1981–1984.

391. Patterson EI, Elia G, Grassi A, Giordano A, Desario C, Medardo M, Smith SL, Anderson ER, Prince T, Patterson GT, Lorusso E, Lucente MS, Lanave G, Lauzi S, Bonfanti U, Stranieri A, Martella V, Solari Basano F, Barrs VR, Radford AD, Agrimi U, Hughes GL, Paltrinieri S, Decaro N. 2020. [Evidence of exposure to SARS-CoV-2 in cats and dogs from households in Italy](#). Nat Commun 11.
392. Mohebbali M, Hassanpour G, Zainali M, Gouya MM, Khayatzadeh S, Parsaei M, Sarafriz N, Hassanzadeh M, Azarm A, Salehi-Vaziri M, Sasani F, Heidari Z, Jalali T, Pouriayeali MH, Shoja Z, Ahmadi Z, Sadjadi M, Tavakoli M, Azad-Manjiri S, Karami C, Zarei Z. 2022. [SARS-CoV-2 in domestic cats \(Felis catus\) in the northwest of Iran: Evidence for SARS-CoV-2 circulating between human and cats](#). Virus Research 310:198673.
393. Bosco-Lauth AM, Hartwig AE, Porter SM, Gordy PW, Nehring M, Byas AD, VandeWoude S, Ragan IK, Maison RM, Bowen RA. 2020. [Experimental infection of domestic dogs and cats with SARS-CoV-2: Pathogenesis, transmission, and response to reexposure in cats](#). Proc Natl Acad Sci USA 117:26382–26388.
394. 2020. Exclusive: Buddy, first dog to test positive for COVID-19 in the U.S., has died. Animals. <https://www.nationalgeographic.com/animals/article/first-dog-to-test-positive-for-covid-in-us-dies>. Retrieved 5 December 2022.
395. Connecticut puppy that died unexpectedly tested positive for coronavirus, UConn researchers say. Hartford Courant. <https://www.courant.com/coronavirus/hc-news-coronavirus-uconn-dog-infection-20210413-yrbg6icd6bcirk6ylwnzkxi6di-story.html>. Retrieved 5 December 2022.
396. North Carolina dog that died after 'acute' illness tests positive for coronavirus. NBC News. <https://www.nbcnews.com/news/us-news/north-carolina-dog-died-after-acute-illness-tests-positive-coronavirus-n1236477>. Retrieved 5 December 2022.
397. Hart R. Kitten Dies After Catching Covid As Study Uncovers More Evidence Of Human-To-Cat Transmission. Forbes. <https://www.forbes.com/sites/roberthart/2021/04/23/kitten-dies-after-catching-covid-as-study-uncovers-more-evidence-of-human-to-cat-transmission/>. Retrieved 5 December 2022.
398. Hosie MJ, Epifano I, Herder V, Orton RJ, Stevenson A, Johnson N, MacDonald E, Dunbar D, McDonald M, Howie F, Tennant B, Herrity D, Da Silva Filipe A, Streicker DG, Willett BJ, Murcia PR, Jarrett RF, Robertson DL, Weir W. 2021. [Detection of SARS-CoV-2 in respiratory samples from cats in the UK associated with human-to-cat transmission](#). Veterinary Record 188.
399. Collins A. Investigation into cat's death, cat had COVID. <https://www.wbrc.com>. <https://www.wbrc.com/2020/10/09/investigation-into-cats-death-cat-had-covid/>. Retrieved 5 December 2022.

400. do Vale B, Lopes AP, Fontes M da C, Silvestre M, Cardoso L, Coelho AC. 2021. [Bats, pangolins, minks and other animals - villains or victims of SARS-CoV-2?](#) Vet Res Commun 45:1–19.
401. Bosco-Lauth AM, Walker A, Guilbert L, Porter S, Hartwig A, McVicker E, Bielefeldt-Ohmann H, Bowen RA. 2021. [Susceptibility of livestock to SARS-CoV-2 infection](#). Emerging Microbes & Infections 10:2199–2201.
402. Gaudreault NN, Cool K, Trujillo JD, Morozov I, Meekins DA, McDowell C, Bold D, Carossino M, Balaraman V, Mitzel D, Kwon T, Madden DW, Artiaga BL, Pogradichnyi RM, Roman-Sosa G, Wilson WC, Balasuriya UBR, García-Sastre A, Richt JA. 2022. [Susceptibility of sheep to experimental co-infection with the ancestral lineage of SARS-CoV-2 and its alpha variant](#). Emerging Microbes & Infections 11:662–675.
403. Villanueva-Saz S, Giner J, Fernández A, Lacasta D, Ortín A, Ramos JJ, Ferrer LM, Ruiz de Arcaute M, Tobajas AP, Pérez MD, Verde M, Marteles D, Hurtado-Guerrero R, Pardo J, Santiago L, González-Ramírez AM, Macías-León J, García-García A, Taleb V, Lira-Navarrete E, Paño-Pardo JR, Ruíz H. 2021. [Absence of SARS-CoV-2 Antibodies in Natural Environment Exposure in Sheep in Close Contact with Humans](#). Animals 11:1984.
404. Taylor CA, Boulos C, Almond D. 2020. [Livestock plants and COVID-19 transmission](#). Proc Natl Acad Sci USA 117:31706–31715.
405. Marchant-Forde JN, Boyle LA. 2020. [COVID-19 Effects on Livestock Production: A One Welfare Issue](#). Front Vet Sci 7.
406. Oreshkova N, Molenaar RJ, Vreman S, Harders F, Oude Munnink BB, Hakze-van der Honing RW, Gerhards N, Tolsma P, Bouwstra R, Sikkema RS, Tacken MG, de Rooij MM, Weesendorp E, Engelsma MY, Bruschke CJ, Smit LA, Koopmans M, van der Poel WH, Stegeman A. 2020. [SARS-CoV-2 infection in farmed minks, the Netherlands, April and May 2020](#). Eurosurveillance 25.
407. Hammer AS, Quaade ML, Rasmussen TB, Fonager J, Rasmussen M, Mundbjerg K, Lohse L, Strandbygaard B, Jørgensen CS, Alfaro-Núñez A, Rosenstjerne MW, Boklund A, Halasa T, Fomsgaard A, Belsham GJ, Bøtner A. 2021. [SARS-CoV-2 Transmission between Mink \(<i>Neovison vison</i>\) and Humans, Denmark](#). Emerg Infect Dis 27:547–551.
408. Eckstrand CD, Baldwin TJ, Rood KA, Clayton MJ, Lott JK, Wolking RM, Bradway DS, Baszler T. 2021. [An outbreak of SARS-CoV-2 with high mortality in mink \(Neovison vison\) on multiple Utah farms](#). PLoS Pathog 17:e1009952.
409. Larsen HD, Fonager J, Lomholt FK, Dalby T, Benedetti G, Kristensen B, Urth TR, Rasmussen M, Lassaunière R, Rasmussen TB, Strandbygaard B, Lohse L, Chaîne M, Møller KL, Berthelsen A-SN, Nørgaard SK, Sönksen UW, Boklund AE, Hammer AS, Belsham GJ, Krause TG, Mortensen S, Bøtner A, Fomsgaard A, Mølbak K. 2021. [Preliminary report of an outbreak of SARS-CoV-2 in mink and mink farmers](#)

[associated with community spread, Denmark, June to November 2020.](#)
Eurosurveillance 26.

410. Hoffmann M, Zhang L, Krüger N, Graichen L, Kleine-Weber H, Hofmann-Winkler H, Kempf A, Nessler S, Riggert J, Winkler MS, Schulz S, Jäck H-M, Pöhlmann S. 2021. [SARS-CoV-2 mutations acquired in mink reduce antibody-mediated neutralization.](#) Cell Reports 35:109017.
411. Fernández-Bellón H, Rodon J, Fernández-Bastit L, Almagro V, Padilla-Solé P, Lorca-Oró C, Valle R, Roca N, Grazioli S, Trogu T, Bensaid A, Carrillo J, Izquierdo-Useros N, Blanco J, Parera M, Noguera-Julián M, Clotet B, Moreno A, Segalés J, Vergara-Alert J. 2021. [Monitoring Natural SARS-CoV-2 Infection in Lions \(Panthera leo\) at the Barcelona Zoo: Viral Dynamics and Host Responses.](#) Viruses 13:1683.
412. Frutos R, Devaux CA. 2020. [Mass culling of minks to protect the COVID-19 vaccines: is it rational?](#) New Microbes and New Infections 38:100816.
413. Enserink M. 2020. [Coronavirus rips through Dutch mink farms, triggering culls.](#) Science 368:1169–1169.
414. Harrington LA, Díez-León M, Gómez A, Harrington A, Macdonald DW, Maran T, Pödra M, Roy S. 2021. [Wild American mink \(*Neovison vison* \) may pose a COVID-19 threat.](#) Front Ecol Environ 19:266–267.
415. Aguiló-Gisbert J, Padilla-Blanco M, Lizana V, Maiques E, Muñoz-Baquero M, Chillida-Martínez E, Cardells J, Rubio-Guerri C. 2021. [First Description of SARS-CoV-2 Infection in Two Feral American Mink \(Neovison vison\) Caught in the Wild.](#) Animals 11:1422.
416. USDA APHIS | Confirmation of COVID-19 in Gorillas at a California Zoo. https://www.aphis.usda.gov/aphis/newsroom/stakeholder-info/sa_by_date/sa-2021/sa-01/ca-gorillas-sars-cov-2. Retrieved 5 December 2022.
417. 2022. Dallas Zoo says 5 gorillas have tested positive for COVID-19. wfaacom. <https://www.wfaa.com/article/news/local/dallas-zoo-5-gorillas-tested-positive-covid-19/287-901a235f-b120-49ff-9fbe-1a1a6dfbadb5>. Retrieved 5 December 2022.
418. 2021. Nearly all gorillas at Atlanta's zoo have contracted COVID-19. CTVNews. <https://www.ctvnews.ca/health/coronavirus/nearly-all-gorillas-at-atlanta-s-zoo-have-contracted-covid-19-1.5586112>. Retrieved 5 December 2022.
419. Staff K9N. 2021. Kansas City Zoo gorillas recovering from COVID-19. KMBC. <https://www.kmbc.com/article/kc-zoo-gorillas-recovering-from-covid/37873956>. Retrieved 5 December 2022.
420. McAloose D, Laverack M, Wang L, Killian ML, Caserta LC, Yuan F, Mitchell PK, Queen K, Mauldin MR, Cronk BD, Bartlett SL, Sykes JM, Zec S, Stokol T, Ingerman K, Delaney MA, Fredrickson R, Ivančić M, Jenkins-Moore M, Mzingo K, Franzen K, Bergeson NH, Goodman L, Wang H, Fang Y, Olmstead C, McCann C, Thomas P, Goodrich E, Elvinger F, Smith DC, Tong S, Slavinski S, Calle PP, Terio K, Torchetti MK, Diel DG.

2020. [From People to *Panthera* : Natural SARS-CoV-2 Infection in Tigers and Lions at the Bronx Zoo](#). mBio 11.

421. 2022. A snow leopard at Miller Park Zoo dies from COVID-induced pneumonia. WGLT. <https://www.wgl.org/local-news/2022-01-06/a-snow-leopard-at-miller-park-zoo-is-mclean-countys-latest-death-from-covid-19>. Retrieved 5 December 2022.
422. Thebault R. 2021. [A zoo's three 'beloved' snow leopards die of covid-19](#). Washington Post.
423. Anthes E. 2022. [When People Take Pandemic Precautions, Gorillas Breathe Easier](#). The New York Times.
424. Cranfield MR. 2008. [Mountain gorilla research: the risk of disease transmission relative to the benefit from the perspective of ecosystem health](#). Am J Primatol 70:751–754.
425. Firozi P. 2021. [A group of gorillas is being treated for covid. The great apes will soon get their shots, too, zoo says](#). Washington Post.
426. Hoyte A, Webster M, Ameiss K, Conlee DA, Hainer N, Hutchinson K, Burakova Y, Dominowski PJ, Baima ET, King VL, Rosey EL, Hardham JM, Millership J, Kumar M. 2022. [Experimental veterinary SARS-CoV-2 vaccine cross neutralization of the Delta \(B.1.617.2\) variant virus in cats](#). Veterinary Microbiology 268:109395.
427. COVID-19 Animal Vaccines. Zoetis. <https://www.zoetis.com/news-and-insights/featured-stories/zoetis-emerging-infectious-disease-capabilities-support-covid-19-solutions-for-great-apes-and-minks>. Retrieved 5 December 2022.
428. Sharun K, Tiwari R, Saied AA, Dhama K. 2021. [SARS-CoV-2 vaccine for domestic and captive animals: An effort to counter COVID-19 pandemic at the human-animal interface](#). Vaccine 39:7119–7122.
429. Do we need to have a Covid vaccine for domestic animals? The Irish Times. <https://www.irishtimes.com/life-and-style/do-we-need-to-have-a-covid-vaccine-for-domestic-animals-1.4736360>. Retrieved 5 December 2022.
430. Chavda VP, Feehan J, Apostolopoulos V. 2021. [A Veterinary Vaccine for SARS-CoV-2: The First COVID-19 Vaccine for Animals](#). Vaccines 9:631.
431. Nguyen C, Campos R, Horn • • M. Cats and Dogs Top List of COVID-19 Infected Animals in US. NBC Bay Area. <https://www.nbcbayarea.com/investigations/cats-and-dogs-top-list-of-covid-19-infected-animals-in-u-s/2625085/>. Retrieved 5 December 2022.
432. Liu Y, Hu G, Wang Y, Ren W, Zhao X, Ji F, Zhu Y, Feng F, Gong M, Ju X, Zhu Y, Cai X, Lan J, Guo J, Xie M, Dong L, Zhu Z, Na J, Wu J, Lan X, Xie Y, Wang X, Yuan Z, Zhang R, Ding Q. 2021. [Functional and genetic analysis of viral receptor ACE2 orthologs reveals a broad potential host range of SARS-CoV-2](#). Proc Natl Acad Sci USA 118.

433. Conceicao C, Thakur N, Human S, Kelly JT, Logan L, Bialy D, Bhat S, Stevenson-Leggett P, Zagrajek AK, Hollinghurst P, Varga M, Tsirigoti C, Tully M, Chiu C, Moffat K, Silesian AP, Hammond JA, Maier HJ, Bickerton E, Shelton H, Dietrich I, Graham SC, Bailey D. 2020. [The SARS-CoV-2 Spike protein has a broad tropism for mammalian ACE2 proteins](#). PLoS Biol 18:e3001016.
434. Sullivan PF. 2007. [Spurious Genetic Associations](#). Biological Psychiatry 61:1121–1126.
435. Zhang Y, Qin L, Zhao Y, Zhang P, Xu B, Li K, Liang L, Zhang C, Dai Y, Feng Y, Sun J, Hu Z, Xiang H, Knight JC, Dong T, Jin R. 2020. [Interferon-Induced Transmembrane Protein 3 Genetic Variant rs12252-C Associated With Disease Severity in Coronavirus Disease 2019](#). The Journal of Infectious Diseases 222:34–37.
436. Zhang Y-H, Zhao Y, Li N, Peng Y-C, Giannoulatou E, Jin R-H, Yan H-P, Wu H, Liu J-H, Liu N, Wang D-Y, Shu Y-L, Ho L-P, Kellam P, McMichael A, Dong T. 2013. [Interferon-induced transmembrane protein-3 genetic variant rs12252-C is associated with severe influenza in Chinese individuals](#). Nat Commun 4.
437. Kim Y-C, Jeong B-H. 2020. [Ethnic variation in risk genotypes based on single nucleotide polymorphisms \(SNPs\) of the interferon-inducible transmembrane 3 \(IFITM3\) gene, a susceptibility factor for pandemic 2009 H1N1 influenza A virus](#). Immunogenetics 72:447–453.
438. Kim Y-C, Jeong B-H. 2020. [Strong Correlation between the Case Fatality Rate of COVID-19 and the rs6598045 Single Nucleotide Polymorphism \(SNP\) of the Interferon-Induced Transmembrane Protein 3 \(IFITM3\) Gene at the Population-Level](#). Genes 12:42.
439. Nikoloudis D, Kountouras D, Hiona A. 2020. [The frequency of combined IFITM3 haplotype involving the reference alleles of both rs12252 and rs34481144 is in line with COVID-19 standardized mortality ratio of ethnic groups in England](#). PeerJ 8:e10402.
440. Shi G, Kenney AD, Kudryashova E, Zani A, Zhang L, Lai KK, Hall-Stoodley L, Robinson RT, Kudryashov DS, Compton AA, Yount JS. 2020. [Opposing activities of IFITM proteins in SARS-CoV-2 infection](#). EMBO J 40.
441. Zheng M, Zhao X, Zheng S, Chen D, Du P, Li X, Jiang D, Guo J-T, Zeng H, Lin H. 2020. [Bat SARS-Like WIV1 coronavirus uses the ACE2 of multiple animal species as receptor and evades IFITM3 restriction via TMPRSS2 activation of membrane fusion](#). Emerging Microbes & Infections 9:1567–1579.
442. Tabor HK, Risch NJ, Myers RM. 2002. [Candidate-gene approaches for studying complex genetic traits: practical considerations](#). Nat Rev Genet 3:391–397.
443. , Niemi MEK, Karjalainen J, Liao RG, Neale BM, Daly M, Ganna A, Pathak GA, Andrews SJ, Kanai M, Veerapen K, Fernandez-Cadenas I, Schulte EC, Striano P, Marttila M, Minica C, Marouli E, Karim MA, Wendt FR, Savage J, Sloofman L, Butler-Laporte G, Kim H-N, Kanoni S, Okada Y, Byun J, Han Y, Uddin MJ, Smith GD, Willer CJ, Buxbaum JD, Mehtonen J,

Finucane H, Cordioli M, Martin AR, Zhou W, Pasaniuc B, Julianne H, Aschard H, Shi H, Yengo L, Polimanti R, Ghoussaini M, Schwartzentruber J, Dunham I, Chwialkowska K, Francescatto M, Trankiem A, Balaconis MK, Davis L, Lee S, Priest J, Renieri A, Sankaran VG, van Heel D, Deelen P, Richards JB, Nakanishi T, Biesecker L, Kerchberger VE, Baillie JK, Mari F, Bernasconi A, Baillie SC, Canakoglu A, Wolford B, Faucon A, Dutta AK, Schurmann C, Harry E, Birney E, Nguyen H, Nasir J, Kaunisto M, Solomonson M, Dueker N, Vadgama N, Limou S, Rahmouni S, Mbarek H, Darwish D, Uddin MM, Albertos R, Pérez-Tur J, Li R, Folkersen L, Moltke I, Koelling N, Teumer A, Kousathanas A, Utrilla A, Verdugo RA, Zárata R, Medina-Gómez C, Gómez-Cabrero D, Carnero-Montoro E, Cadilla CL, Moreno-Estrada A, Garmendia A, Moya L, Sedaghati-Khayat B, Boua PR, Favé M-J, Francioli L, Lemaçon A, Migeotte I, Patel S, Varnai R, Szentpeteri JL, Sipeky C, Colombo F, von Hohenstaufen K, Lio P, Vallerga C, Wang Q, Tanigawa Y, Im H, Han C, Song H, Lim J, Lee Y, Kim S, Im S, Atanasovska B, Ahmad HF, Boer C, Jansen P, Franke L, Kaja E, Pasko D, Kennis-Szilagyi I, Kornilov SA, Prijatelj V, Prokić I, Sivanadhan I, Perumal S, Esmaeeli S, Pearson NM, Auton A, Shelton JF, Shastri AJ, Filshtein-Sonmez T, Coker D, Symons A, Esparza-Gordillo J, Aslibekyan S, O'Connell J, Ye C, Weldon CH, Perera M, O'Leary K, Tuck M, O'Brien T, Meltzer D, O'Donnell P, Nutescu E, Yang G, Alarcon C, Herrmann S, Mazurek S, Banagan J, Hamidi Z, Barbour A, Raffat N, Moreno D, Friedman P, Ferwerda B, van de Beek D, Brouwer MC, Vlaar APJ, Wiersinga WJ, Posthuma D, Tissink E, Koos Zwinderman AH, Uffelman E, van Agtmael M, Algera AG, van Baarle F, Bax D, Beudel M, Jan Bogaard H, Bomers M, Bonta PI, Bos L, Botta M, de Brabander J, de Bree G, de Bruin S, Bugiani M, Bulle E, Chouchane O, Cloherty A, Dongelmans D, Elbers P, Fleuren L, Geerlings S, Geerts B, Geijtenbeek T, Girbes A, Goorhuis B, Grobusch MP, Hafkamp F, Hagens L, Hamann J, Harris V, Hemke R, Hermans SM, Heunks L, Hollmann M, Horn J, Hovius JW, de Jong MD, Koning R, van Mourik N, Nellen J, Nossent EJ, Paulus F, Peters E, van der Poll T, Preckel B, Prins JM, Raasveld J, Reijnders T, Schinkel M, Schultz MJ, Schuurman A, Sigaloff K, Smit M, Stijnis CS, Stilma W, Teunissen C, Thorald P, Tsonas A, van der Valk M, Veelo D, de Vries H, van Vugt M, Wouters D, Minnaar RP, Kromhout A, van Uffelen KWJ, Wolterman RA, Roberts G, Park D, Ball CA, Coignet M, McCurdy S, Knight S, Partha R, Rhead B, Zhang M, Berkowitz N, Gaddis M, Noto K, Ruiz L, Pavlovic M, Hong EL, Rand K, Girshick A, Guturu H, Baltzell AH, Rahmouni S, Guntz J, Beguin Y, Pigazzini S, Nkambule L, Bouysran Y, Busson A, Peyrassol X, Wilkin F, Pichon B, Smits G, Vandernoot I, Goffard J-C, Georges M, Moutschen M, Misset B, Darcis G, Guiot J, Jadot L, Azarzar S, Dellot P, Gofflot S, Claassen S, Bertrand A, Parzibut G, Clarinval M, Moermans C, Malaise O, El Kandoussi K, Thonon R, Huynen P, Mesdagh A, Melo S, Jacques N, Di Valentin E, Giroule F, Collignon A, Radermecker C, Lebrun M, Perée H, Latour S, Barada O, Sanchez J, Josse C, Boujemla B, Meunier M, Mariavelle E, Anania S, Gazon H, Juszcak D, Fadeur M, Camby S, Meuris C, Thys M, Jacques J, Henket M, Léonard P, Fripiat F, Giot J-B, Sauvage A-S, Von Freckell C, Mni M, Wéry M, Staderoli A, Belhaj Y, Lambermont B, Morrison DR, Mooser V, Forgetta V, Li R, Ghosh B, Laurent L, Belisle A, Henry D, Abdullah T, Adeleye O, Mamlouk N, Kimchi N, Afrasiabi Z, Rezk N, Vulesevic B, Bouab M, Guzman C, Petitjean L, Tselios C, Xue X, Afilalo J, Afilalo M, Oliveira M, Brenner B, Brassard N, Durand M, Schurr E, Lepage P, Ragoussis J, Auld

D, Chassé M, Kaufmann DE, Lathrop GM, Adra D, Davis LK, Cox NJ, Below JE, Sealock JM, Faucon AB, Shuey MM, Polikowsky HG, Petty LE, Shaw DM, Chen H-H, Zhu W, Ludwig KU, Schröder J, Maj C, Rolker S, Nöthen MM, Fazaal J, Keitel V, Jensen B-EO, Feldt T, Kurth I, Marx N, Dreher M, Pink I, Cornberg M, Illig T, Lehmann C, Schommers P, Augustin M, Rybniker J, Knopp L, Eggermann T, Volland S, Altmüller J, Berger MM, Brenner T, Hinney A, Witzke O, Bals R, Herr C, Ludwig N, Walter J, Fuchsberger C, Pattaro C, De Grandi A, Pramstaller P, Emmert D, Melotti R, Foco L, Mascalzoni D, Gögele M, Domingues F, Hicks A, Gignoux CR, Wicks SJ, Crooks K, Barnes KC, Daya M, Shortt J, Rafaels N, Chavan S, Goldstein DB, Kiryluk K, Sengupta S, Chung W, Reilly MP, Khan A, Wang C, Povysil G, Bhardwaj N, Gharavi AG, Ionita-Laza I, Shang N, O'Byrne SM, Nandakumar R, Menon A, So YS, Hod E, Pendrick D, Kim H-N, Park S-K, Kim H-L, Kang CK, Lee H-J, Song K-H, Jae Yoon K, Paik N-J, Seok W, Yoon H, Joo E-J, Chang Y, Ryu S, Park WB, Su Park J, Un Park K, Ham SY, Jung J, Kim ES, Kim HB, Ellinghaus D, Degenhardt F, Cáceres M, Juzenas S, Lenz TL, Albillos A, Julià A, Heidecker B, Garcia F, Kurth F, Tran F, Hanses F, Zoller H, Holter JC, Fernández J, Sander LE, Rosenstiel P, Koehler P, de Cid R, Asselta R, Schreiber S, Hehr U, Prati D, Baselli G, Valenti L, Bujanda L, Banales JM, Duga S, D'Amato M, Romero-Gómez M, Buti M, Invernizzi P, Franke A, Hov JR, Karlsen TH, Folseraas T, Maya-Miles D, Teles A, Azuure C, Wacker EM, Uellendahl-Werth F, ElAbd H, Arora J, Lerga-Jaso J, Wienbrandt L, Rühlemann MC, Wendorff M, Vadla MS, Lenning OB, Özer O, Myhre R, Raychaudhuri S, Tanck A, Gassner C, Hemmrich-Stanisak G, Kässens J, Figuera Basso ME, Schulzky M, Wittig M, Braun N, Wesse T, Albrecht W, Yi X, Ortiz AB, Chercoles AG, Ruiz A, Mantovani A, Holten AR, Mayer A, Cherubini A, Protti A, Aghemo A, Gerussi A, Ramirez A, Braun A, Barreira A, Lleo A, Kildal AB, Glück A, Nolla AC, Latiano A, Dyrhol-Riise AM, Muscatello A, Voza A, Rando-Segura A, Solier A, Karina B, Cortes B, Mateos B, Nafria-Jimenez B, Schaefer B, Bellinghausen C, Ferrando C, Quereda C, Skurk C, Thibeault C, Spinner CD, Lange C, Hu C, Cappadona C, Bianco C, Sancho C, Lihaug Hoff DA, Galimberti D, Jiménez D, Pestaña D, Toapanta D, Azzolini E, Scarpini E, Helbig ET, Urrechaga E, Paraboschi EM, Pontali E, Reverter E, Navas E, Arana E, Sánchez FG, Ceriotti F, Malvestiti F, Mesonero F, Pezzoli G, Lamorte G, Neb H, My I, Hernández I, de Rojas I, Galván-Femenia I, Heyckendorf J, Rybniker J, Badia JR, Schneider J, Goikoetxea J, Kraft J, Müller KE, Gaede KI, Garcia-Etxebarria K, Tonby K, Heggelund L, Izquierdo-Sanchez L, Sumoy L, Lippert LJ, Terranova L, Garbarino L, Téllez L, Roade L, Ostadreza M, Intxausti M, Kogevinas M, Gutiérrez-Stampa MA, Vehreschild MJGT, Marquié M, Castoldi M, Cecconi M, Boada M, Seilmaier MJ, Mazzocco M, Rodríguez-Gandía M, Ayo NI, Blay N, Martínez N, Cornely OA, Palmieri O, Tentorio P, Rodrigues PM, España PP, Hoffmann P, Bacher P, Suwalski P, de Pablo R, Nieto R, Badalamenti S, Ciesek S, Bombace S, Wilfling S, Brunak S, Heilmann-Heimbach S, Ripke S, Bahmer T, Landmesser U, Protzer U, Rimoldi V, Skogen V, Andrade V, Moreno V, Poller W, Farre X, Wang X, Khodamoradi Y, Karadeniz Z, de Salazar A, Palom A, Garcia-Fernandez A-E, Blanco-Grau A, Zanella A, Bandera A, Nebel A, Biondi A, Caballero-Garralda A, Gori A, Lind A, Fracanzani AL, Peschuck A, Pesenti A, de la Horra C, Milani C, Paccapelo C, Angelini C, Cea C, Muñoz-Diaz E, Sandoval E, Calderón EJ, Solligård E, Aziz F, Martinelli-Boneschi F, Peyvandi F, Blasi F, Medrano FJ, Rodríguez-Frias F, Müller F, Grasselli G, Costantino G, Cardamone G, Foti G, Matullo G,

Kurihara H, Afset JE, Damås JK, Ampuero J, Martín J, Erdmann J, Bergan J, Goerg S, Ferrusquía-Acosta J, Quero JH, Delgado J, Guerrero JM, Risnes K, Bettini LR, Moreira L, Gustad LT, Santoro L, Scudeller L, Riveiro-Barciela M, Schaefer M, Carrabba M, Valsecchi MG, Hernandez-Tejero M, Acosta-Herrera M, D'Angiò M, Baldini M, Cazzaniga M, Ciccarelli M, Bocciolone M, Miozzo M, Chueca N, Montano N, Faverio P, Preatoni P, Bonfanti P, Omodei P, Castro P, Ferrer R, Gualtierotti R, Gallego-Durán R, Morilla R, Haider S, Marsal S, Aneli S, Pelusi S, Bosari S, Aliberti S, Dudman S, Zheng T, Pumarola T, Cejudo TG, Monzani V, Friaza V, Peter W, Dopazo X, Duga S, May S, Grimsrud MM, Gudbjartsson DF, Stefansson K, Sulem P, Sveinbjornsson G, Melsted P, Norddahl G, Swerford Moore KH, Thorsteinsdottir U, Holm H, Alarcón-Riquelme ME, Bernardo D, Martínez-Bueno M, Rello SR, Magi R, Milani L, Metspalu A, Laisk T, Läll K, Lepamets M, Esko T, Reimann E, Naaber P, Laane E, Pesukova J, Peterson P, Kisand K, Tabri J, Allos R, Hensen K, Starkopf J, Ringmets I, Tamm A, Kallaste A, Alavere H, Metsalu K, Puusepp M, Kristiansson K, Koskelainen S, Perola M, Donner K, Kivinen K, Palotie A, Palotie A, Rivolta C, Bochud P-Y, Bibert S, Boillat N, Nussle SG, Albrich W, Quinodoz M, Kamdar D, Suh N, Neofytos D, Erard V, Voide C, Bochud PY, Rivolta C, Bibert S, Quinodoz M, Kamdar D, Neofytos D, Erard V, Voide C, Friolet R, Vollenweider P, Pagani JL, Oddo M, zu Bentrup FM, Conen A, Clerc O, Marchetti O, Guillet A, Guyat-Jacques C, Foucras S, Rime M, Chassot J, Jaquet M, Viollet RM, Lannepoudenx Y, Portopena L, Desgranges F, Filippidis P, Guéry B, Haeffliger D, Kampouri EE, Manuel O, Munting A, Papadimitriou-Olivgeris M, Regina J, Rochat-Stettler L, Suttels V, Tadini E, Tschopp J, Van Singer M, Viala B, Boillat-Blanco N, Brahier T, Hügli O, Meuwly JY, Pantet O, Nussle SG, Bochud M, D'Acremont V, Younes SE, Albrich WC, Suh N, Cerny A, O'Mahony L, von Mering C, Bochud PY, Frischknecht M, Kleger G-R, Filipovic M, Kahlert CR, Wozniak H, Negro TR, Pugin J, Bouras K, Knapp C, Egger T, Perret A, Montillier P, di Bartolomeo C, Barda B, de Cid R, Carreras A, Moreno V, Galván-Femenía I, Blay N, Farré X, Sumoy L, Cortés B, Mercader JM, Guindo-Martinez M, Torrents D, Garcia-Aymerich J, Castaño-Vinyals G, Dobaño C, Gori M, Renieri A, Mondelli MU, Castelli F, Vaghi M, Rusconi S, Montagnani F, Bargagli E, Franchi F, Mazzei MA, Cantarini L, Tacconi D, Feri M, Scala R, Spargi G, Nencioni C, Bandini M, Caldarelli GP, Spagnesi M, Canaccini A, Ognibene A, D'Arminio Monforte A, Girardis M, Antinori A, Francisci D, Schiaroli E, Scotton PG, Panese S, Scaggiante R, Monica MD, Capasso M, Fiorentino G, Castori M, Aucella F, Di Biagio A, Masucci L, Valente S, Mandalà M, Zucchi P, Giannattasio F, Coviello DA, Mussini C, Bosio G, Tavecchia L, Crotti L, Rizzi M, La Rovere MT, Sarzi-Braga S, Bussotti M, Ravaglia S, Artuso R, Perrella A, Romani D, Bergomi P, Catena E, Vincenti A, Ferri C, Grassi D, Pessina G, Tumbarello M, Di Pietro M, Sabrina R, Luchi S, Barbieri C, Acquilini D, Andreucci E, Paciosi F, Segala FV, Tiseo G, Falcone M, Lista M, Poscente M, De Vivo O, Petrocelli P, Guarnaccia A, Baroni S, Perticaroli V, Furini S, Dei S, Benetti E, Picchiotti N, Sanarico M, Ceri S, Pinoli P, Raimondi F, Biscarini F, Stella A, Bergomi M, Zguro K, Capitani K, Tanfoni M, Fallerini C, Daga S, Baldassarri M, Fava F, Frullanti E, Valentino F, Doddato G, Giliberti A, Tita R, Amitrano S, Bruttini M, Croci S, Meloni I, Mencarelli MA, Lo Rizzo C, Pinto AM, Beligni G, Tommasi A, Di Sarno L, Palmieri M, Carriero ML, Alaverdian D, Iuso N, Inchingolo G, Busani S, Bruno R, Vecchia M, Belli MA, Mantovani S, Ludovisi S, Quiros-Roldan E, Antoni MD, Zanella I, Siano

M, Emiliozzi A, Fabbiani M, Rossetti B, Zanelli G, Bergantini L, D'Alessandro M, Cameli P, Bennet D, Anedda F, Marcantonio S, Scolletta S, Guerrini S, Conticini E, Frediani B, Spertilli C, Donati A, Guidelli L, Corridi M, Croci L, Piacentini P, Desanctis E, Cappelli S, Verzuri A, Anemoli V, Pancrazi A, Lorubbio M, Merlini E, Miraglia FG, Venturelli S, Cossarizza A, Vergori A, Gabrieli A, Riva A, Paciosi F, Andretta F, Gatti F, Parisi SG, Baratti S, Piscopo C, Russo R, Andolfo I, Iolascon A, Carella M, Merla G, Squeo GM, Raggi P, Marciano C, Perna R, Bassetti M, Sanguinetti M, Giorli A, Salerni L, Parravicini P, Menatti E, Trotta T, Coiro G, Lena F, Martinelli E, Mancarella S, Gabbi C, Maggiolo F, Ripamonti D, Bachetti T, Suardi C, Parati G, Bottà G, Di Domenico P, Rancan I, Bianchi F, Colombo R, van Heel DA, Hunt KA, Trembath RC, Huang QQ, Martin HC, Mason D, Trivedi B, Wright J, Finer S, Griffiths CJ, Akhtar S, Anwar M, Arciero E, Ashraf S, Breen G, Chung R, Curtis CJ, Chowdhury M, Colligan G, Deloukas P, Durham C, Finer S, Griffiths C, Huang QQ, Hurles M, Hunt KA, Hussain S, Islam K, Khan A, Khan A, Lavery C, Lee SH, Lerner R, MacArthur D, MacLaughlin B, Martin H, Mason D, Miah S, Newman B, Safa N, Tahmasebi F, Trembath RC, Trivedi B, van Heel DA, Wright J, Smith AV, Boughton AP, Li KW, LeFaive J, Annis A, Jannes CE, Krieger JE, Pereira AC, Velho M, Marques E, Lima IR, Tada MT, Valino K, McCarthy M, Rosenberger C, Lee JE, Chang D, Hammer C, Hunkapiller J, Mahajan A, Pendergrass S, Sucheston-Campbell L, Yaspan B, Lee HS, Shin E, Jang HY, Kim S, Kym S, Kim Y-S, Jeong H, Kwon KT, Kim S-W, Kim JY, Jang YR, Kim H ah, Lee JY, Lee JE, Lee S, Choe K-W, Kang YM, Jee SH, Jung KJ, Parikh V, Ashley E, Wheeler M, Rivas M, Bustamante C, Pinksy B, Febbo P, Farh K, Schroth GP, deSouza F, Dalton K, Christle J, Deboever C, Szalma S, Tanigawa Y, Rubinacci S, Delaneau O, Gorzynski J, de Jong H, Sutton S, Youlton N, Joshi R, Jimenez-Morales D, Hughes C, Amar D, Ioannidis A, Hershman S, Kirillova A, Seo K, Huang Y, Shoura M, Hammond N, Watson N, Raja A, Huang C, Sahoo M, Wang H, Zhen J, Rakitko A, Ilinsky V, Yermakovich D, Popov I, Chernitsov A, Kovalenko E, Krasnenko A, Plotnikov N, Stetsenko I, Kim A, Cirulli ET, Schiabor Barrett KM, Bolze A, White S, Washington NL, Lu JT, Riffle S, Tanudjaja F, Wang X, Ramirez JM III, Leonetti N, Sandoval E, Neveux I, Dabe S, Grzymiski JJ, Esteban Miñano JI, Aguirre LA, López-Collazo E, de la Mata Pazos M, Cerrato L, Folkersen L, Lozano-Rodríguez R, Avendaño-Ortiz J, Arcos VT, Montalbán-Hernández KM, Quiroga JV, Pascual-Iglesias A, Maroun-Eid C, Martín-Quirós A, Namkoong H, Okada Y, Imoto S, Katayama K, Fukunaga K, Kitagawa Y, Sato T, Hasegawa N, Kumanogoh A, Kimura A, Ai M, Tokunaga K, Kanai T, Miyano S, Ogawa S, Edahiro R, Sonehara K, Shirai Y, Kanai M, Ishii M, Kabata H, Masaki K, Kamata H, Ikemura S, Chubachi S, Okamori S, Terai H, Tanaka H, Morita A, Lee H, Asakura T, Sasaki J, Morisaki H, Uwamino Y, Nanki K, Mikami Y, Tomono K, Kato K, Matsuda F, Takahashi M, Hizawa N, Takeda Y, Hirata H, Shiroyama T, Miyawaki S, Suzuki K, Maeda Y, Nii T, Noda Y, Niitsu T, Adachi Y, Enomoto T, Amiya S, Hara R, Takahashi K, Anzai T, Hasegawa T, Ito S, Koike R, Endo A, Uchimura Y, Miyazaki Y, Honda T, Tateishi T, Tohda S, Ichimura N, Sonobe K, Sassa C, Nakajima J, Nannya Y, Omae Y, Takahashi K, Harada N, Hiki M, Takagi H, Nakamura A, Tagaya E, Kawana M, Arimura K, Ishiguro T, Takayanagi N, Isono T, Takaku Y, Takano K, Anan R, Nakajima Y, Nakano Y, Nishio K, Ueda S, Hayashi R, Tateno H, Hase I, Yoshida S, Suzuki S, Mitamura K, Saito F, Ueda T, Azuma M, Nagasaki T, Yasui Y, Hasegawa Y, Mutoh Y, Yoshiyama T,

Shoko T, Kojima M, Adachi T, Ishikawa M, Takahashi K, Watanabe K, Manabe T, Ito F, Fukui T, Funatsu Y, Koh H, Hirai Y, Kawashima H, Narita A, Niwa K, Sekikawa Y, Saito F, Yoshiya K, Yoshihara T, Suzuki Y, Nakayama S, Masuzawa K, Nishi K, Nishitsuji M, Tani M, Inoue T, Hirano T, Kobayashi K, Miyazawa N, Kimura Y, Sado R, Ogura T, Kitamura H, Murohashi K, Nakachi I, Baba R, Arai D, Fuke S, Saito H, Kuwahara N, Fujiwara A, Okada T, Baba T, Noda J, Mashimo S, Yagi K, Shiomi T, Hashiguchi M, Odani T, Mochimaru T, Oyamada Y, Mori N, Izumi N, Nagata K, Taki R, Murakami K, Yamada M, Sugiura H, Hayashi K, Shimizu T, Gon Y, Fujitani S, Tsuchida T, Yoshida T, Kagaya T, Kita T, Sakagami S, Kimizuka Y, Kawana A, Nakamura Y, Ishikura H, Takata T, Kikuchi T, Taniyama D, Nakamura M, Kodama N, Kaneyama Y, Maeda S, Nagasaki Y, Okamoto M, Ishihara S, Ito A, Chihara Y, Takeuchi M, Onoi K, Hashimoto N, Wakahara K, Ando A, Masuda M, Wakabayashi A, Watanabe H, Sageshima H, Nakada T-A, Abe R, Shimada T, Kawamura K, Ichikado K, Nishiyama K, Yamasaki M, Hashimoto S, Kusaka Y, Ohba T, Isogai S, Takada M, Kanda H, Komase Y, Sano F, Asano K, Oguma T, Harada M, Takahashi T, Shibusawa T, Abe S, Kono Y, Togashi Y, Izumo T, Inomata M, Awano N, Ogawa S, Ogata T, Ishihara S, Kanehiro A, Ozaki S, Fuchimoto Y, Kitagawa Y, Yoshida S, Ogura S, Nishiyama K, Yoshida K, Beppu S, Fukuyama S, Eriguchi Y, Yonekawa A, Inoue Y, Yamagata K, Chiba S, Narumoto O, Nagai H, Ooshima N, Motegi M, Sagara H, Tanaka A, Ohta S, Shibata Y, Tanino Y, Sato Y, Yamada Y, Hashino T, Shinoki M, Iwagoe H, Imamura T, Umeda A, Shimada H, Endo M, Hayashi S, Takahashi M, Nakano S, Yatomi M, Maeno T, Ishii T, Utsugi M, Ono A, Kanaoka K, Ihara S, Komuta K, Franke L, Boezen M, Claringbould A, Lopera E, Warmerdam R, Vonk Judith M, van Blokland I, Lanting P, Ori APS, Obeidat M, Hernández Cordero AI, Sin DD, Bossé Y, Joubert P, Hao K, Nickle D, Timens W, van den Berge M, Feng Y-CA, Mercader J, Weiss ST, Karlson EW, Smoller JW, Murphy SN, Meigs JB, Woolley AE, Green RC, Perez EF, Wolford B, Zöllner S, Wang J, Beck A, Sloofman LG, Ascolillo S, Sebra RP, Collins BL, Levy T, Sealfon SC, Jordan DM, Thompson RC, Gettler K, Chaudhary K, Belbin GM, Preuss M, Hoggart C, Choi S, Underwood SJ, Salib I, Britvan B, Keller K, Tang L, Peruggia M, Hiester LL, Niblo K, Aksentijevich A, Labkowsky A, Karp A, Zlatopolsky M, Zyndorf M, Charney AW, Beckmann ND, Schadt EE, Abul-Husn NS, Cho JH, Itan Y, Kenny EE, Loos RJF, Nadkarni GN, Do R, O'Reilly P, Huckins LM, Ferreira MAR, Abecasis GR, Leader JB, Cantor MN, Justice AE, Carey DJ, Chittoor G, Josyula NS, Kosmicki JA, Horowitz JE, Baras A, Gass MC, Yadav A, Mirshahi T, Jan Hottenga J, Bartels M, de Geus EJC, Nivard MG, Verma A, Ritchie MD, Rader D, Li B, Verma SS, Lucas A, Bradford Y, Zara F, Salpietro V, Scala M, Iacomino M, Scudieri P, Boccardi R, Minetti C, Riva A, Vari MS, Rahier J-F, Giorgio E, Carli D, Louis E, Bulik CM, Landén M, Brusco A, Ferrero GB, Madia F, Fundin B, Ismail SI, Saad C, Al-Sarraj Y, Badji RM, Al-Muftah W, Al Thani A, Afifi N, Klovins J, Rovite V, Rescenko R, Peculis R, Ustinova M, Zeberg H, Frithiof R, Hultström M, Lipcsey M, Johnson R, Geschwind DH, Freimer N, Butte MJ, Geschwind DH, Pasaniuc B, Ding Y, Chiu A, Chang TS, Boutros P, Moutsianas L, Caulfield MJ, Scott RH, Walker S, Stuckey A, Odhams CA, Rhodes D, Fowler T, Rendon A, Chan G, Arumugam P, Karczewski KJ, Wilson DJ, Spencer CA, Crook DW, Wyllie DH, O'Connell AM, Atkinson EG, Kanai M, Tsuo K, Baya N, Turley P, Gupta R, Walters RK, Palmer DS, Sarma G, Solomonson M, Cheng N, Lu W, Churchhouse C, Goldstein JL, King D, Seed C, Daly MJ, Neale BM, Bryant S, Satterstrom FK, Band G,

Earle SG, Lin S-K, Arning N, Armstrong J, Rudkin JK, Callier S, Bryant S, Cusick C, Soranzo N, Zhao JH, Danesh J, Di Angelantonio E, Butterworth AS, Sun YV, Huffman JE, Cho K, O'Donnell CJ, Tsao P, Gaziano JM, Peloso G, Ho Y-L, Mian M, Scaggiante F, Chang X, Glessner JR, Hakonarson H, McGuigan PJ, Prockter Moore LS, Vizcaychipi MP, Hall K, Campbell A, Nichol A, Ward G, Page VJ, Semple MG, Adeniji K, Agranoff D, Agwuh K, Ail D, Aldera EL, Alegria A, Angus B, Ashish A, Atkinson D, Bari S, Barlow G, Barnass S, Barrett N, Bassford C, Basude S, Baxter D, Beadsworth M, Bernatoniene J, Berridge J, Best N, Bothma P, Chadwick D, Brittain-Long R, Bulteel N, Burden T, Burtenshaw A, Caruth V, Chambler D, Chee N, Child J, Chukkambotla S, Clark T, Collini P, Cosgrove C, Cupitt J, Cutino-Moguel M-T, Dark P, Dawson C, Dervisevic S, Donnison P, Douthwaite S, Drummond A, DuRand I, Dushianthan A, Dyer T, Evans C, Eziefula C, Fegan C, Finn A, Fullerton D, Garg S, Garg A, Gkrania-Klotsas E, Godden J, Goldsmith A, Graham C, Hardy E, Hartshorn S, Harvey D, Havalda P, Hawcutt DB, Hobrok M, Hodgson L, Hormis A, Jacobs M, Jain S, Jennings P, Kaliappan A, Kasipandian V, Kegg S, Kelsey M, Kendall J, Kerrison C, Kerslake I, Koch O, Koduri G, Koshy G, Laha S, Laird S, Larkin S, Leiner T, Lillie P, Limb J, Linnett V, Little J, Lyttle M, MacMahon M, MacNaughton E, Mankregod R, Masson H, Matovu E, McCullough K, McEwen R, Meda M, Mills GH, Minton J, Ward K, Mirfenderesky M, Mohandas K, Mok Q, Moon J, Moore E, Morgan P, Morris C, Mortimore K, Moses S, Mpenge M, Mulla R, Murphy M, Nagel M, Nagarajan T, Nelson M, O'Shea MK, Otahal I, Ostermann M, Pais M, Panchatsharam S, Papakonstantinou D, Paraiso H, Patel B, Pattison N, Pepperell J, Peters M, Phull M, Pintus S, Pooni JS, Post F, Price D, Prout R, Rae N, Reschreiter H, Reynolds T, Richardson N, Roberts M, Roberts D, Rose A, Rousseau G, Ryan B, Saluja T, Shah A, Shanmuga P, Sharma A, Shawcross A, Sizer J, Shankar-Hari M, Smith R, Snelson C, Spittle N, Staines N, Stambach T, Stewart R, Subudhi P, Szakmany T, Tatham K, Thomas J, Thompson C, Thompson R, Tridente A, Tupper-Carey D, Twagira M, Ustianowski A, Vallotton N, Vincent-Smith L, Visuvanathan S, Vuylsteke A, Waddy S, Wake R, Walden A, Welters I, Whitehouse T, Whittaker P, Whittington A, Papineni P, Wijesinghe M, Williams M, Wilson L, Cole S, Winchester S, Wiselka M, Wolverson A, Wooton DG, Workman A, Yates B, Young P, Beale R, Bretherick AD, Clohisey S, Fourman MH, Furniss J, Gountouna E, Grimes G, Haley C, Harrison D, Hayward C, Keating S, Klaric L, Klenerman P, Law A, Meynert AM, Millar J, Pairo-Castineira E, Parkinson N, Ponting CP, Porteous DJ, Rawlik K, Richmond A, Rowan K, Russell CD, Scott RH, Shen X, Shih B, Tenesa A, Vitart V, Wang B, Wilson JF, Wu Y, Yang J, Yang Z, Zechner M, Zhai R, Zheng C, Norman L, Pius R, Drake TM, Fairfield CJ, Knight SR, Mclean KA, Murphy D, Shaw CA, Dalton J, Girvan M, Saviciute E, Roberts S, Harrison J, Marsh L, Connor M, Halpin S, Jackson C, Gamble C, Leeming G, Law A, Wham M, Hendry R, Scott-Brown J, Begg C, Hinds C, Wai Ho AY, Horby PW, Knight J, Ling L, Maslove D, McAuley D, Montgomery H, Nichol A, Openshaw PJM, Semple MG, Shankar-Hari M, Summers C, Walsh T, Armstrong L, Bates H, Dooks E, Farquhar F, Hairsine B, McParland C, Packham S, Alldis Z, Astin-Chamberlain R, Bibi F, Biddle J, Blow S, Bolton M, Borra C, Bowles R, Burton M, Choudhury Y, Collier D, Cox A, Easthope A, Ebano P, Fotiadis S, Gurasashvili J, Halls R, Hartridge P, Kallon D, Kassam J, Lancoma-Malcolm I, Matharu M, May P, Mitchelmore O, Newman T, Patel M, Pheby J, Pinzuti I, Prime Z, Pryszazhna O, Shiel J, Taylor M, Tierney C, Wood S, Zak A, Zongo O,

Forsey M, Kaliappan A, Nicholson A, Riches J, Vertue M, Wasson C, Finn S, Green J, Collins E, King B, Grauslyte L, Hussain M, Phull M, Pogreban T, Rosaroso L, Salciute E, Franke G, Wong J, George A, Akeroyd L, Bano S, Bromley M, Gurr L, Lawton T, Morgan J, Sellick K, Warren D, Wilkinson B, McGowan J, Ledgerd C, Stacey A, Pye K, Bellwood R, Bentley M, Hobrok M, Loosley R, McGuinness H, Tench H, Wolf-Roberts R, Gibson S, Lyle A, McNeela F, Radhakrishnan J, Hughes A, Ali A, Brady M, Dale S, Dance A, Gledhill L, Greig J, Hanson K, Holdroyd K, Home M, Kelly D, Kitson R, Matapure L, Melia D, Mellor S, Nortcliffe T, Pinnell J, Robinson M, Shaw L, Shaw R, Thomis L, Wilson A, Wood T, Bayo L-A, Merwaha E, Ishaq T, Hanley S, Antcliffe D, Banach D, Brett S, Coghlan P, Fernandez Z, Gordon A, Rojo R, Arias SS, Templeton M, Jha R, Krishnamurthy V, Lim L, Bi R, Scholefield B, Ashton L, Williams A, Cheyne C, Saunderson A, Allan A, Anderson F, Kaye C, Liew J, Medhora J, Scott T, Trumper E, Botello A, Polgarova P, Stroud K, Meaney E, Jones M, Ng A, Agrawal S, Pathan N, White D, Daubney E, Elston K, Parker R, Reddy A, Turner-Bone I, Wilding L, Harding P, Jacob R, Jones C, Denmade C, Croft M, White I, Lim L, Griffin D, Muchenje N, Mupudzi M, Partridge R, Conyngham J-A, Thomas R, Wright M, Corral MA, Bastion V, Clarke D, David B, Kent H, Lorusso R, Lubimbi G, Murdoch S, Penacerrada M, Thomas A, Valentine J, Vochin A, Wulandari R, Djeugam B, Dawson J, Garrioch S, Tolson M, Aldridge J, de Almeida Martins LG, Carungcong J, Beavis S, Dale K, Gascoyne R, Hawes J, Pritchard K, Stevenson L, Whileman A, Cowley A, Highgate J, Crawley R, Crew A, Cunningham M, Daniels A, Harrison L, Hope S, Inweregbu K, Jones S, Lancaster N, Matthews J, Nicholson A, Wray G, Benham L, Bradshaw Z, Brown J, Caswell M, Cupitt J, Melling S, Preston S, Slawson N, Stoddard E, Warden S, Combes E, Joefield T, Monnery S, Beech V, Trotman S, Hopkins B, Scriven J, Thrasyvoulou L, Willis H, Anderson S, Birch J, Collins E, Hammerton K, O'Leary R, Abernathy C, Foster L, Gratrix A, Martinson V, Parkinson P, Stones E, Carbral-Ortega L, Kapoor R, Loader D, Castle K, Brandwood C, Smith L, Clark R, Birchall K, Kolakaluri L, Baines D, Sukumaran A, Mapfunde I, Meredith M, Morris L, Ryan L, Clark A, Sampson J, Peters C, Dent M, Langley M, Ashraf S, Wei S, Andrew A, Chablani M, Kirkby A, Netherton K, Bates M, Dasgin J, Gill J, Nilsson A, Scriven J, Apetri E, Basikolo C, Blackledge B, Catlow L, Charles B, Dark P, Doonan R, Harris J, Harvey A, Horner D, Knowles K, Lee S, Lomas D, Lyons C, Marsden T, McLaughlan D, McMorrow L, Pendlebury J, Perez J, Poulaka M, Proudfoot N, Slaughter M, Slevin K, Taylor M, Thomas V, Walker D, Michael A, Collis M, Clark M, Coulding M, Jude E, McCormick J, Mercer O, Potla D, Rehman H, Savill H, Turner V, Davey M, Golden D, Seaman R, Hunt J, Dearden J, Dobson E, Mulcahy M, Munt S, O'Connor G, Philbin J, Rishton C, Tully R, Winnard S, Cagova L, Fofano A, Garner L, Holcombe H, Mephram S, Mitchell AM, Mwaura L, Praman K, Vuylsteke A, Zamikula J, Bercades G, Brealey D, Hass I, MacCallum N, Martir G, Raith E, Reyes A, Smyth D, Taylor A, Hughes RA, Thomas H, Rees A, Duskova M, Phipps J, Brooks S, Edwards M, Alexander P, Allen S, Bradley-Potts J, Brantwood C, Egan J, Felton T, Padden G, Ward L, Moss S, Glasgow S, Beesley K, Board S, Kubisz-Pudelko A, Lewis A, Perry J, Pippard L, Wood D, Buckley C, Brown A, Gregory J, O'Connell S, Smith T, Belagodu Z, Fuller B, Gherman A, Olufuwa O, Paramsothy R, Stuart C, Oakley N, Kamundi C, Tyl D, Collins K, Silva P, Taylor J, King L, Coates C, Crowley M, Wakefield P, Beadle J, Johnson L, Sargeant J, Anderson M, Jardine C, Williams D, Parris V,

Quaid S, Watson E, Melville J, Naisbitt J, Joseph R, Lazo M, Walton O, Neal A, Hill M, Kannan T, Wild L, Allan E, Darlington K, Davies F, Easton J, Kumar S, Lean R, Menzies D, Pugh R, Qiu X, Davies L, Williams H, Scanlon J, Davies G, Mackay C, Lewis J, Rees S, Coetzee S, Gales A, Otahal I, Raj M, Sell C, Langton H, Prout R, Watters M, Novis C, Arbane G, Bociek A, Campos S, Grau N, Jones TO, Lim R, Marotti M, Ostermann M, Shankar-Hari M, Whitton C, Barron A, Collins C, Kaul S, Passmore H, Prendergast C, Reed A, Rogers P, Shokkar R, Woodruff M, Middleton H, Polgar O, Nolan C, Thwaites V, Mahay K, Sri-Chandana C, Scherewode J, Stephenson L, Marsh S, Bancroft H, Bellamy M, Carmody M, Daglish J, Moore F, Rhodes J, Sangombe M, Kadiri S, Scriven J, Ayers A, Harrison W, North J, Cavazza A, Cockrell M, Corcoran E, Depante M, Finney C, Jerome E, McPhail M, Nayak M, Noble H, O'Reilly K, Pappa E, Saha R, Saha S, Smith J, Knighton A, Gill M, Paul P, Ratnam V, Shelton S, Wynter I, Baptista D, Crowe R, Fernandes R, Herdman-Grant R, Joseph A, Loveridge A, McKenley I, Morino E, Naranjo A, Simms R, Sollesta K, Swain A, Venkatesh H, Khera J, Fox J, Barber R, Hewitt C, Hilldrith A, Jackson-Lawrence K, Shepardson S, Wills M, Butler S, Tavares S, Cunningham A, Hindale J, Arif S, George L, Twiss S, Wright D, Holland M, Keenan N, Lyons M, Wassall H, Marsh C, Mahenthiran M, Carter E, Kong T, Adanini O, Bhatia N, Msiska M, Mew L, Mwaura E, Stewart R, Williams F, Wren L, Sutherland S-B, Battle C, Brinkworth E, Harford R, Murphy C, Newey L, Rees T, Williams M, Arnold S, Brealey D, Hardy J, Houlden H, Moncur E, Raith E, Tariq A, Tucci A, Convery K, Fottrell-Gould D, Hudig L, Keshet-Price J, Randell G, Stammers K, Abdelrazik M, Bakthavatsalam D, Elhassan M, Ganesan A, Haldeos A, Moreno-Cuesta J, Purohit D, Vincent R, Xavier K, Rohit K, Alasdair F, Saleem M, David C, Jenkins S, Lamond Z, Wall A, Yates B, Reynolds J, Campbell H, Thompsom M, Dodds S, Duffy S, Butcher D, O'Sullivan S, Butterworth-Cowin N, Deacon B, Hibbert M, Potheary C, Tetla D, Woodford C, Durga L, Kennard-Holden G, de Gordo LO-R, Peasgood E, Phillips C, Skinner D, Gaylard J, Mullan D, Newman J, Davies E, Roche L, Sathe S, Brimfield L, Daly Z, Pogson D, Rose S, Collins A, Khaliq W, Gude ET, Allen L, Beranova E, Crisp N, Deery J, Hazelton T, Knight A, Price C, Tilbey S, Turki S, Turney S, Giles J, Booth S, Bell G, English K, Katary A, Wilcox L, Campbell R, Clarke N, Whiteside J, Mascarenhas M, Donaldson A, Matheson J, Barrett F, O'Hara M, O'Keefe L, Bradley C, Collier D, Hormis A, Walker R, Maynard V, Patel T, Smith M, Chukkambotla S, Kazi A, Hartley J, Dykes J, Hijazi M, Keith S, Khan M, Ryan-Smith J, Springle P, Thomas J, Truman N, Saad S, Coleman D, Fine C, Matt R, Gay B, Dalziel J, Ali S, Goodchild D, Harling R, Bhatteejee R, Goddard W, Davison C, Duberly S, Hargreaves J, Bolton R, Laha S, Verlander M, Williams A, Blackman H, Creagh-Brown B, Donlon S, Michalak-Glinska N, Mtuwa S, Pristopan V, Salberg A, Smith E, Stone S, Piercy C, Verula J, Burda D, Montaser R, Harden L, Mayangao I, Marriott C, Bradley P, Harris C, Cooper J, Finch C, Liderth S, Quinn A, Waddington N, Fidler K, Tagliavini E, Donnelly K, Abel L, Brett M, Digby B, Gemmell L, Hornsby J, MacGoey P, O'Neil P, Price R, Rodden N, Rooney K, Sundaram R, Thomson N, Flanagan R, Hughes G, Latham S, McKenna E, Anderson J, Hull R, Rhead K, Branney D, Frankham J, Pitts S, White N, Cristiano D, Dormand N, Farzad Z, Gummadi M, Liyanage K, Patel BV, Salmi S, Sloane G, Thwaites V, Varghese M, Zborowski AC, Bean S, Burt K, Spivey M, Eastgate-Jackson C, Filipe H, Martin D, Maharajh A, Garcia SM, De Neef M, Deacon B, Lynch C, Potheary C,

Roche L, Howe GS, Singh J, Turner K, Ellis H, Stroud N, Cherian S, Cutler S, Heron AE, Roynon-Reed A, Szakmany T, Williams G, Richards O, Cheema Y, Ahmad N, Barker J, Bauchmuller K, Bird S, Cawthron K, Harrington K, Jackson Y, Kibutu F, Lenagh B, Masuko S, Mills GH, Raithatha A, Wiles M, Willson J, Newell H, Lye A, Nwafor L, Jarman C, Rowland-Jones S, Foote D, Cole J, Thompson R, Watson J, Hesseldon L, Macharia I, Chetam L, Smith J, Ford A, Anderson S, Birchall K, Housley K, Walker S, Milner L, Hanratty H, Trower H, Phillips P, Oxspring S, Donne B, Bevan E, Martin J, Trodd D, Watson G, Brown CW, Bunni L, Jennings C, Latif M, Marshall R, Subramanian G, Bandla N, Gellamucho M, Davies M, Thompson C, Trim F, Eapen B, Ahmed C, Baines B, Clamp S, Colley J, Haq R, Hayes A, Hulme J, Hussain S, Joseph S, Kumar R, Maqsood Z, Purewal M, Chandler B, Elliott K, Mallinson J, Turnbull A, Dent K, Horsley E, Akhtar MN, Pearson S, Potoczna D, Spencer S, Blakemore H, Borislavova B, Faulkner B, Gendall E, Goff E, Hayes K, Thomas M, Worner R, Smith K, Stephens D, Delgado CC, Dawson D, Ding L, Durrant G, Ezeobu O, Farnell-Ward S, Harrison A, Kanu R, Leaver S, Maccacari E, Manna S, Saluzzio RP, Queiroz J, Samakomva T, Sicat C, Texeira J, Da Gloria EF, Lisboa A, Rawlins J, Mathew J, Kinch A, Hurt WJ, Shah N, Clark V, Thanasi M, Yun N, Patel K, Crickmore V, Debreceni G, Wilkins J, Nicol L, Burn I, Hambrook G, Manso K, Penn R, Shanmugasundaram P, Tebbutt J, Thornton D, Rostron A, Roy A, Woods L, Cornell S, Wakinshaw F, Rogerson K, Jarmain J, Anderson P, Archer K, Austin K, Davis C, Durie A, Kelsall O, Thrush J, Vigurs C, Wild L, Wood H-L, Tranter H, Harrison A, Cowley N, McAlindon M, Burtenshaw A, Digby S, Low E, Morgan A, Cother N, Rankin T, Clayton S, McCurdy A, Allibone S, Mary-Genetu R, Kasipandian V, Patel A, Mac A, Murphy A, Mahjoob P, Nazari R, Worsley L, Fagan A, Mohamed Ali IA, Beaumont K, Blunt M, Coton Z, Curgenvin H, Elsaadany M, Fernandes K, Ally SM, Rangarajan H, Sarathy V, Selvanayagam S, Vedage D, White M, Fernandez-Roman J, Hamilton DO, Johnson E, Johnston B, Martinez ML, Mulla S, Shaw D, Waite AAC, Waugh V, Welters ID, Williams K, Bemand T, Black E, Rosa AD, Howle R, Jhanji S, Baikady RR, Tatham KC, Thomas B, Halkes M, Mercer P, Thornton L, West J, Baird T, Ruddy J, Reece-Anthony R, Birt M, Cowton A, Kay A, Kent M, Potts K, Wilkinson A, Naylor S, Brown E, Clark M, Purvis S, Cole J, Davies M, Davies R, Duffin D, Hill H, Player B, Thomas E, Williams A, Beith CM, Black K, Clements S, Morrison A, Strachan D, Taylor M, Clarkson M, D'Sylva S, Norman K, Coventry T, Fowler S, MacMahon M, McGregor A, Brady A, Chan R, Little J, Mclvor S, Prady H, Whittle H, Mathew B, Clapham M, Harper R, Poultney U, Rice P, Smith T, Mutch R, Baird Y, Butler A, Chadbourn I, Folkes L, Fox H, Gardner A, Gomez R, Hobden G, Hodgson L, King K, Margaron M, Martindale T, Meadows E, Raynard D, Thirlwall Y, Helm D, Margalef J, Greer S, Shuker K, Tridente A, Smuts S, Duffield J, Smith O, Mallon L, Claire W, Birkinshaw I, Carter J, Howard K, Ingham J, Joy R, Pearson H, Roche S, Scott Z, Knights E, Price A, Thomas A, Thorpe C, Abraheem A, Bamford P, Cawley K, Dunmore C, Faulkner M, Girach R, Jeffrey H, Jones R, London E, Nagra I, Nasir F, Sainsbury H, Smedley C, Khade R, Sundar A, Tsinaslanidis G, Behan T, Burnett C, Hatton J, Heeney E, Mitra A, Newton M, Pollard R, Stead R, Birch J, Bough L, Goodsell J, Tutton R, Williams P, Williams S, Winter-Goodwin B, Auld F, Donnachie J, Edmond I, Prentice L, Runciman N, Salutous D, Symon L, Todd A, Turner P, Short A, Sweeney L, Murdoch E, Senaratne D, Burns K, Higham A, Anderson T, Hawcutt D, O'Malley L, Rad L, Rogers N,

Saunderson P, Allison KS, Afolabi D, Whitbread J, Jones D, Dore R, Lankester L, Nikitas N, Wells C, Stowe B, Spencer K, Cathcart S, Duffy K, Puxty A, Puxty K, Turner L, Ireland J, Semple G, Barry P, Hilltout P, Evitts J, Tyler A, Waldron J, Irvine V, Shelley B, Akinkugbe O, Bamford A, Beech E, Belfield H, Bell M, Davies C, Jones GAL, McHugh T, Meghari H, O'Neill L, Peters MJ, Ray S, Tomas AL, Easthope A, Gorman C, Gupta A, Timlick E, Brady R, Bonner S, Hugill K, Jones J, Liggett S, Bashyal A, Davidson N, Hutton P, McKechnie S, Wilson J, Flint N, Rekha P, Hales D, Cruz C, Pattison N, Gopal S, Harris N, Lake V, Methereil S, Radford E, Clement I, Patel B, Gulati A, Hays C, Webster K, Hudson A, Webster A, Stephenson E, McCormack L, Slater V, Nixon R, Hanson H, Fearby M, Kelly S, Bridgett V, Robinson P, Almaden-Boyle C, Austin P, Cabrelli L, Cole S, Casey M, Chapman S, Whyte C, Brayne A, Fisher E, Hunt J, Jackson P, Kaye D, Love N, Parkin J, Tuckey V, van Koutrik L, Carter S, Andrew B, Findlay L, Adams K, Bruce M, Connolly K, Duncan T, T.-Michael H, Lindergard G, Hey S, Fox C, Alfonso J, Durrans LJ, Guerin J, Blackledge B, Harris J, Hruska M, Eltayeb A, Lamb T, Hodgkiss T, Cooper L, Rothwell J, Dennis C, McGregor A, Parris V, Srikanan S, Sukha A, Davies K, O'Brien L, Omar Z, Otahal I, Perkins E, Lewis T, Sutherland I, Brooke H, Buckley S, Suarez JC, Charlesworth R, Hansson K, Norris J, Poole A, Rose A, Sandhu R, Sloan B, Smithson E, Thirumaran M, Wagstaff V, Metcalfe A, Camsooksai J, Humphrey C, Jenkins S, Reschreiter H, Wadams B, DeAth Y, Adams C, Agasou A, Arden T, Bowes A, Boyle P, Beekes M, Button H, Capps N, Carnahan M, Carter A, Childs D, Donaldson D, Hard K, Hurford F, Hussain Y, Javaid A, Jones J, Jose S, Leigh M, Martin T, Millward H, Motherwell N, Rikunenko R, Stickley J, Summers J, Ting L, Tivenan H, Tonks L, Wilcox R, Bokhari M, Linnett V, Lucas R, McCormick W, Ritzema J, Sanderson A, Wild H, Baxter N, Henderson S, Kennedy-Hay S, McParland C, Rooney L, Sim M, McCreath G, Brunton M, Caterson J, Coles H, Frise M, Rai SG, Jacques N, Keating L, Tilney E, Bartley S, Bhuie P, Downes C, Holding K, Riches K, Hilton M, Hayman M, Subramanian D, Daniel P, Zitter L, Benyon S, Marriott S, Park L, Keenan S, Gordon E, Quinn H, Baines K, Andrew G, Baillie JK, Barclay L, Callaghan M, Campbell R, Clark S, Hope D, Marshall L, McCulloch C, Briton K, Singleton J, Birch S, Higham A, Simpson K, Craig J, Demetriou C, Eckbad C, Hierons S, Howie L, Mitchard S, Ramos L, Serrano-Ruiz A, White K, Kelly F, Amin V, Anastasescu E, Anumakonda V, Karthik K, Kausar R, Reid K, Smith J, Imeson-Wood J, Bellini A, Bryant J, Mayer A, Pickard A, Roe N, Sowter J, Howlett A, Criste K, Cusack R, Golder K, Golding H, Jones O, Leggett S, Male M, Marani M, Prager K, Williams T, Roberts B, Salmon K, Gondo P, Hadebe B, Kayani A, Masunda B, Ahmed A, Morris A, Jakkula S, Long K, Whiteley S, Wilby E, Ogg B, Moultrie S, Odam M, Bewley J, Garland Z, Grimmer L, Gumbrell B, Johnson R, Sweet K, Webster D, Efford G, Bennett S, Goodwin E, Jackson M, Kent A, Tibke C, Woodyatt W, Zaki A, Daniel A, Finn J, Saha R, Staines N, Easthope A, Bremmer P, Allan J, Geary T, Houston G, Meikle A, O'Brien P, Bell D, Boyle R, Douglas K, Glass L, Lee E, Lennon L, Rattray A, Charnock R, McFarland D, Cosgrove D, Attwood B, Parsons P, Carmody S, Oblak M, Popescu M, Thankachen M, Baruah R, Morris S, Ferguson S, Shepherd A, Altabaibeh A, Alvaro A, Gilbert K, Ma L, Mostoles L, Parmar C, Simpson K, Jetha C, Booker L, Pratley A, Cosier T, Millen G, Richardson N, Schumacher N, Weston H, Rand J, Alex B, Bach B, Barclay WS, Bogaert D, Chand M, Cooke GS, Docherty AB, Dunning J, da Silva Filipe A, Fletcher T, Green CA, Harrison EM,

Hiscox JA, Ijaz S, Khoo S, Klenerman P, Lim WS, Mentzer AJ, Merson L, Noursadeghi M, Moore SC, Palmarini M, Paxton WA, Pollakis G, Price N, Rambaut A, Robertson DL, Russell CD, Sancho-Shimizu V, Scott JT, de Silva T, Sigfrid L, Solomon T, Srisakandan S, Stuart D, Tedder RS, Thomson EC, Roger Thompson AA, Thwaites RS, Turtle LCW, Gupta RK, Palmieri C, Swann OV, Zambon M, Dumas M-E, Griffin JL, Takats Z, Chechi K, Andrikopoulos P, Osagie A, Olanipekun M, Liggi S, Lewis MR, Correia G dos S, Sands CJ, Takis P, Maslen L, Greenhalf W, Shaw V, McDonald SE, Keating S, Ahmed KA, Armstrong JA, Ashworth M, Asiimwe IG, Bakshi S, Barlow SL, Booth L, Brennan B, Bullock K, Catterall BWA, Clark JJ, Clarke EA, Cooper L, Cox H, Davis C, Dincarslan O, Dunn C, Dyer P, Elliott A, Evans A, Finch L, Fisher LWS, Foster T, Garcia-Dorival I, Greenhalf W, Gunning P, Hartley C, Jensen RL, Jones CB, Jones TR, Khandaker S, King K, Kiy RT, Koukorava C, Lake A, Lant S, Latawiec D, Lavelle-Langham L, Lefteri D, Lett L, Livoti LA, Mancini M, McDonald S, McEvoy L, McLauchlan J, Metelmann S, Miah NS, Middleton J, Mitchell J, Moore SC, Murphy EG, Penrice-Randal R, Pilgrim J, Prince T, Reynolds W, Ridley PM, Sales D, Shaw VE, Shears RK, Small B, Subramaniam KS, Szemiel A, Taggart A, Tanianis-Hughes J, Thomas J, Trochu E, van Tonder L, Wilcock E, Zhang JE, Flaherty L, Maziere N, Cass E, Carracedo AD, Carlucci N, Holmes A, Massey H, Murphy L, Wrobel N, McCafferty S, Morrice K, MacLean A, Armstrong R, Boz C, Brown A, Clark R, Coutts A, Cullum L, Day N, Donnelly L, Duncan E, Fawkes A, Finernan P, Gilchrist T, Golightly A, Hafezi K, Law D, Law R, Law S, Macgillivray L, Maclean A, Mal H, McCafferty S, McMaster E, Meikle J, Moore SC, Morrice K, Murphy L, Oosthuyzen W, Paterson T, Stenhouse A, Swets M, Szoor-McElhinney H, Taneski F, Wackett T, Ward M, Weaver J, Wrobel N, Coyle J, Gallagher B, Lidstone-Scott R, Hamilton D, Schon K, Furlong A, Biggs H, Griffiths F, Andrews E, Brickell K, Smyth M, Murphy L, Carson G, Hardwick H, Donohue C. 2021. [Mapping the human genetic architecture of COVID-19](#). *Nature* 600:472–477.

444. Erlich H. 2012. [HLA DNA typing: past, present, and future](#). *Tissue Antigens* 80:1–11.
445. Gourraud P-A, Khankhanian P, Cereb N, Yang SY, Feolo M, Maiers M, D. Rioux J, Hauser S, Oksenberg J. 2014. [HLA Diversity in the 1000 Genomes Dataset](#). *PLoS ONE* 9:e97282.
446. de Sousa E, Ligeiro D, Lérias JR, Zhang C, Agrati C, Osman M, El-Kafrawy SA, Azhar EI, Ippolito G, Wang F-S, Zumla A, Maeurer M. 2020. [Mortality in COVID-19 disease patients: Correlating the association of major histocompatibility complex \(MHC\) with severe acute respiratory syndrome 2 \(SARS-CoV-2\) variants](#). *International Journal of Infectious Diseases* 98:454–459.
447. Naemi FMA, Al-adwani S, Al-khatibi H, Al-nazawi A. 2021. [Association between the HLA genotype and the severity of COVID-19 infection among South Asians](#). *Journal of Medical Virology* 93:4430–4437.
448. Langton DJ, Bourke SC, Lie BA, Reiff G, Natu S, Darlay R, Burn J, Echevarria C. 2021. [The influence of HLA genotype on the severity of COVID-19 infection](#). *HLA* 98:14–22.

449. Schindler E, Dribus M, Duffy BF, Hock K, Farnsworth CW, Gragert L, Liu C. 2021. [HLA genetic polymorphism in patients with Coronavirus Disease 2019 in Midwestern United States](#). HLA 98:370–379.
450. Ben Shachar S, Barda N, Manor S, Israeli S, Dagan N, Carmi S, Balicer R, Zisser B, Louzoun Y. 2021. [MHC Haplotyping of SARS-CoV-2 Patients: HLA Subtypes Are Not Associated with the Presence and Severity of COVID-19 in the Israeli Population](#). J Clin Immunol 41:1154–1161.
451. Rubin R. 2020. [Investigating Whether Blood Type Is Linked to COVID-19 Risk](#). JAMA 324:1273.
452. Zhao J, Yang Y, Huang H, Li D, Gu D, Lu X, Zhang Z, Liu L, Liu T, Liu Y, He Y, Sun B, Wei M, Yang G, Wang X, Zhang L, Zhou X, Xing M, Wang PG. 2020. Relationship between the ABO Blood Group and the COVID-19 Susceptibility. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.11.20031096>.
453. Kerbage A, Haddad SF, Nasr L, Riachy A, Mekhael E, Nassim N, Hoyek K, Sleilaty G, Nasr F, Riachy M. 2021. [Impact of ABO and Rhesus blood groups on COVID-19 susceptibility and severity: A case-control study](#). Journal of Medical Virology 94:1162–1166.
454. Fan Q, Zhang W, Li B, Li D-J, Zhang J, Zhao F. 2020. [Association Between ABO Blood Group System and COVID-19 Susceptibility in Wuhan](#). Front Cell Infect Microbiol 10.
455. Hoiland RL, Fergusson NA, Mitra AR, Griesdale DEG, Devine DV, Stukas S, Cooper J, Thiara S, Foster D, Chen LYC, Lee AYY, Conway EM, Wellington CL, Sekhon MS. 2020. [The association of ABO blood group with indices of disease severity and multiorgan dysfunction in COVID-19](#). Blood Advances 4:4981–4989.
456. Anderson JL, May HT, Knight S, Bair TL, Muhlestein JB, Knowlton KU, Horne BD. 2021. [Association of Sociodemographic Factors and Blood Group Type With Risk of COVID-19 in a US Population](#). JAMA Netw Open 4:e217429.
457. Zhang Y, Garner R, Salehi S, La Rocca M, Duncan D. 2021. [Association between ABO blood types and coronavirus disease 2019 \(COVID-19\), genetic associations, and underlying molecular mechanisms: a literature review of 23 studies](#). Ann Hematol 100:1123–1132.
458. Wu B-B, Gu D-Z, Yu J-N, Yang J, Shen W-Q. 2020. [Association between ABO blood groups and COVID-19 infection, severity and demise: A systematic review and meta-analysis](#). Infection, Genetics and Evolution 84:104485.
459. Dai X. 2020. [ABO blood group predisposes to COVID-19 severity and cardiovascular diseases](#). Eur J Prev Cardiol 27:1436–1437.
460. Silva-Filho JC, Melo CGF de, Oliveira JL de. 2020. [The influence of ABO blood groups on COVID-19 susceptibility and severity: A molecular hypothesis based on carbohydrate-carbohydrate interactions](#). Medical Hypotheses 144:110155.

461. Marigorta UM, Rodríguez JA, Gibson G, Navarro A. 2018. [Replicability and Prediction: Lessons and Challenges from GWAS](#). Trends in Genetics 34:504–517.
462. Price AL, Zaitlen NA, Reich D, Patterson N. 2010. [New approaches to population stratification in genome-wide association studies](#). Nat Rev Genet 11:459–463.
463. Weiner J 3rd, Suwalski P, Holtgrewe M, Rakitko A, Thibeault C, Müller M, Patriki D, Quedenau C, Krüger U, Ilinsky V, Popov I, Balnis J, Jaitovich A, Helbig ET, Lippert LJ, Stubbemann P, Real LM, Macías J, Pineda JA, Fernandez-Fuertes M, Wang X, Karadeniz Z, Saccomanno J, Doehn J-M, Hübner R-H, Hinzmann B, Salvo M, Blueher A, Siemann S, Jurisic S, Beer JH, Rutishauser J, Wiggli B, Schmid H, Danninger K, Binder R, Corman VM, Mühlemann B, Arjun Arkal R, Fragiadakis GK, Mick E, COMET C, Calfee CS, Erle DJ, Hendrickson CM, Kangelaris KN, Krummel MF, Woodruff PG, Langelier CR, Venkataramani U, García F, Zyla J, Drosten C, Alice B, Jones TC, Suttorp N, Witzernath M, Hippenstiel S, Zemojtel T, Skurk C, Poller W, Borodina T, Pa-COVID SG, Ripke S, Sander LE, Beule D, Landmesser U, Guettouche T, Kurth F, Heidecker B. 2021. [Increased risk of severe clinical course of COVID-19 in carriers of HLA-C*04:01](#). EclinicalMedicine 40:101099.
464. The Severe Covid-19 GWAS Group. 2020. [Genomewide Association Study of Severe Covid-19 with Respiratory Failure](#). New England Journal of Medicine 383:1522–1534.
465. 2020. [The COVID-19 Host Genetics Initiative, a global initiative to elucidate the role of host genetic factors in susceptibility and severity of the SARS-CoV-2 virus pandemic](#). Eur J Hum Genet 28:715–718.
466. Zeberg H, Pääbo S. 2020. [The major genetic risk factor for severe COVID-19 is inherited from Neanderthals](#). Nature 587:610–612.
467. Ding Q, Hu Y, Xu S, Wang J, Jin L. 2013. [Neanderthal Introgression at Chromosome 3p21.31 Was Under Positive Natural Selection in East Asians](#). Molecular Biology and Evolution 31:683–695.
468. Zhou J, Sun Y, Huang W, Ye K. 2021. [Altered Blood Cell Traits Underlie a Major Genetic Locus of Severe COVID-19](#). The Journals of Gerontology: Series A 76:e147–e154.
469. Grubaugh ND, Hanage WP, Rasmussen AL. 2020. [Making Sense of Mutation: What D614G Means for the COVID-19 Pandemic Remains Unclear](#). Cell 182:794–795.
470. Avanzato VA, Matson MJ, Seifert SN, Pryce R, Williamson BN, Anzick SL, Barbian K, Judson SD, Fischer ER, Martens C, Bowden TA, de Wit E, Riedo FX, Munster VJ. 2020. [Case Study: Prolonged Infectious SARS-CoV-2 Shedding from an Asymptomatic Immunocompromised Individual with Cancer](#). Cell 183:1901–1912.e9.
471. Choi B, Choudhary MC, Regan J, Sparks JA, Padera RF, Qiu X, Solomon IH, Kuo H-H, Boucau J, Bowman K, Adhikari UD, Winkler ML, Mueller AA, Hsu TY-T, Desjardins M, Baden LR, Chan BT, Walker BD, Lichterfeld M, Brigl M, Kwon DS, Kanjilal S, Richardson ET, Jonsson AH, Alter G,

- Barczak AK, Hanage WP, Yu XG, Gaiha GD, Seaman MS, Cernadas M, Li JZ. 2020. [Persistence and Evolution of SARS-CoV-2 in an Immunocompromised Host](#). *New England Journal of Medicine* 383:2291–2293.
472. Andreano E, Piccini G, Licastro D, Casalino L, Johnson NV, Paciello I, Monego SD, Pantano E, Manganaro N, Manenti A, Manna R, Casa E, Hyseni I, Benincasa L, Montomoli E, Amaro RE, McLellan JS, Rappuoli R. 2020. SARS-CoV-2 escape *in vitro* from a highly neutralizing COVID-19 convalescent plasma. *Cold Spring Harbor Laboratory* <https://doi.org/10.1101/2020.12.28.424451>.
473. Luring AS, Hodcroft EB. 2021. Genetic Variants of SARS-CoV-2—What Do They Mean? *JAMA* <https://doi.org/10.1001/jama.2020.27124>.
474. Oude Munnink BB, Sikkema RS, Nieuwenhuijse DF, Molenaar RJ, Munger E, Molenkamp R, van der Spek A, Tolsma P, Rietveld A, Brouwer M, Bouwmeester-Vincken N, Harders F, Hakze-van der Honing R, Wegdam-Blans MCA, Bouwstra RJ, GeurtsvanKessel C, van der Eijk AA, Velkers FC, Smit LAM, Stegeman A, van der Poel WHM, Koopmans MPG. 2021. [Transmission of SARS-CoV-2 on mink farms between humans and mink and back to humans](#). *Science* 371:172–177.
475. Davies NG, Barnard RC, Jarvis CI, Kucharski AJ, Munday J, Pearson CAB, Russell TW, Tully DC, Abbott S, Gimma A, Waites W, Wong KL, van Zandvoort K, Eggo RM, Funk S, Jit M, Atkins KE, Edmunds WJ, CMMID COVID-19 Working Group. 2020. Estimated transmissibility and severity of novel SARS-CoV-2 Variant of Concern 202012/01 in England. *Cold Spring Harbor Laboratory* <https://doi.org/10.1101/2020.12.24.20248822>.
476. du Plessis L, McCrone JT, Zarebski AE, Hill V, Ruis C, Gutierrez B, Raghwanji J, Ashworth J, Colquhoun R, Connor TR, Faria NR, Jackson B, Loman NJ, O'Toole Á, Nicholls SM, Parag KV, Scher E, Vasylyeva TI, Volz EM, Watts A, Bogoch II, Khan K, Aanensen DM, Kraemer MUG, Rambaut A, Pybus OG, COVID-19 Genomics UK (COG-UK) Consortium†. 2021. [Establishment and lineage dynamics of the SARS-CoV-2 epidemic in the UK](#). *Science* eabf2946.
477. Hie B, Zhong ED, Berger B, Bryson B. 2021. [Learning the language of viral evolution and escape](#). *Science* 371:284–288.
478. Grubaugh ND, Petrone ME, Holmes EC. 2020. [We shouldn't worry when a virus mutates during disease outbreaks](#). *Nature Microbiology* 5:529–530.
479. [Zhao Z, Li H, Wu X, Zhong Y, Zhang K, Zhang Y-P, Boerwinkle E, Fu Y-X. 2004.. BMC Evol Biol 4:21.](#)
480. Collington E. 2020. PHE document. https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/947048/Technical_Briefing_VOC_SH_NJL2_SH2.pdf.
481. 2020. Preliminary genomic characterisation of an emergent SARS-CoV-2 lineage in the UK defined by a novel set of spike mutations.

- Virological. <https://virological.org/t/preliminary-genomic-characterisation-of-an-emergent-sars-cov-2-lineage-in-the-uk-defined-by-a-novel-set-of-spike-mutations/563>. Retrieved 8 February 2021.
482. B.1.1.7 report. https://cov-lineages.org/global_report_B.1.1.7.html. Retrieved 8 February 2021.
483. Davies NG, Jarvis CI, Edmunds WJ, Jewell NP, Diaz-Ordaz K, Keogh RH. 2021. [Increased mortality in community-tested cases of SARS-CoV-2 lineage B.1.1.7](#). Nature 593:270–274.
484. 2020. Identification of a novel SARS-CoV-2 Spike 69-70 deletion lineage circulating in the United States. Virological. <https://virological.org/t/identification-of-a-novel-sars-cov-2-spike-69-70-deletion-lineage-circulating-in-the-united-states/577>. Retrieved 8 February 2021.
485. Washington NL, White S, Barrett KMS, Cirulli ET, Bolze A, Lu JT. 2020. S gene dropout patterns in SARS-CoV-2 tests suggest spread of the H69del/V70del mutation in the US. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.12.24.20248814>.
486. 2021. Investigation of novel SARS-CoV-2 variant: Variant of Concern 202012/01. Public Health England. https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/959360/Variant_of_Concern_VOC_202012_01_Technical_Briefing_3.pdf.
487. Health C for D and R. 2021. [Genetic Variants of SARS-CoV-2 May Lead to False Negative Results with Molecular Tests for Detection of SARS-CoV-2 - Letter to Clinical Laboratory Staff and Health Care Providers](#). FDA.
488. CDC. 2020. Coronavirus Disease 2019 (COVID-19). Centers for Disease Control and Prevention. <https://www.cdc.gov/coronavirus/2019-ncov/more/science-and-research/scientific-brief-emerging-variants.html>. Retrieved 8 February 2021.
489. CDC. 2020. Coronavirus Disease 2019 (COVID-19). Centers for Disease Control and Prevention. <https://www.cdc.gov/coronavirus/2019-ncov/variants/variant-surveillance.html>. Retrieved 5 December 2022.
490. Minister Zweli Mkhize confirms 8 725 more cases of Coronavirus COVID-19 | South African Government. <https://www.gov.za/speeches/minister-zweli-mkhize-confirms-8-725-more-cases-coronavirus-covid-19-18-dec-2020-0000>. Retrieved 8 February 2021.
491. 2021. Tracking the international spread of SARS-CoV-2 lineages B.1.1.7 and B.1.351/501Y-V2. Virological. <https://virological.org/t/tracking-the-international-spread-of-sars-cov-2-lineages-b-1-1-7-and-b-1-351-501y-v2/592>. Retrieved 8 February 2021.
492. Tegally H, Wilkinson E, Giovanetti M, Iranzadeh A, Fonseca V, Giandhari J, Doolabh D, Pillay S, San EJ, Msomi N, Mlisana K, von Gottberg A, Walaza S, Allam M, Ismail A, Mohale T, Glass AJ, Engelbrecht S, Van Zyl

- G, Preiser W, Petruccione F, Sigal A, Hardie D, Marais G, Hsiao M, Korsman S, Davies M-A, Tyers L, Mudau I, York D, Maslo C, Goedhals D, Abrahams S, Laguda-Akingba O, Alisoltani-Dehkordi A, Godzik A, Wibmer CK, Sewell BT, Lourenço J, Alcantara LCJ, Pond SLK, Weaver S, Martin D, Lessells RJ, Bhiman JN, Williamson C, de Oliveira T. 2020. Emergence and rapid spread of a new severe acute respiratory syndrome-related coronavirus 2 (SARS-CoV-2) lineage with multiple spike mutations in South Africa. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.12.21.20248640>.
493. Wibmer CK, Ayres F, Hermanus T, Madzivhandila M, Kgagudi P, Oosthuysen B, Lambson BE, de Oliveira T, Vermeulen M, van der Berg K, Rossouw T, Boswell M, Ueckermann V, Meiring S, von Gottberg A, Cohen C, Morris L, Bhiman JN, Moore PL. 2021. [SARS-CoV-2 501Y.V2 escapes neutralization by South African COVID-19 donor plasma](#). Nat Med 27:622–625.
 494. Cele S, Gazy I, Jackson L, Hwa S-H, Tegally H, Lustig G, Giandhari J, Pillay S, Wilkinson E, Naidoo Y, Karim F, Ganga Y, Khan K, Bernstein M, Balazs AB, Gosnell BI, Hanekom W, Moosa M-YS, Lessells RJ, de Oliveira T, Sigal A. 2021. [Escape of SARS-CoV-2 501Y.V2 from neutralization by convalescent plasma](#). Nature 593:142–146.
 495. ECDC. 2021. Risk of spread of new SARS-CoV-2 variants of concern in the EU/EEA - first update . <https://www.ecdc.europa.eu/sites/default/files/documents/COVID-19-risk-related-to-spread-of-new-SARS-CoV-2-variants-EU-EEA-first-update.pdf>.
 496. 2021. Genomic characterisation of an emergent SARS-CoV-2 lineage in Manaus: preliminary findings. Virological. <https://virological.org/t/genomic-characterisation-of-an-emergent-sars-cov-2-lineage-in-manaus-preliminary-findings/586>. Retrieved 8 February 2021.
 497. P.1 report. https://cov-lineages.org/global_report_P.1.html. Retrieved 8 February 2021.
 498. Staff R. 2021. [UK detects 77 cases of South African COVID variant, nine of Brazilian](#). Reuters.
 499. PANGO lineages. <https://cov-lineages.org/lineages.html>. Retrieved 8 February 2021.
 500. Zhang W, Davis BD, Chen SS, Martinez JMS, Plummer JT, Vail E. 2021. Emergence of a novel SARS-CoV-2 strain in Southern California, USA. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2021.01.18.21249786>.
 501. GISAID - hCov19 Variants. <https://gisaid.org/hcov19-variants/>. Retrieved 5 December 2022.
 502. Li Q, Wu J, Nie J, Zhang L, Hao H, Liu S, Zhao C, Zhang Q, Liu H, Nie L, Qin H, Wang M, Lu Q, Li X, Sun Q, Liu J, Zhang L, Li X, Huang W, Wang Y. 2020. [The Impact of Mutations in SARS-CoV-2 Spike on Viral Infectivity and Antigenicity](#). Cell 182:1284–1294.e9.

503. Greaney AJ, Loes AN, Crawford KHD, Starr TN, Malone KD, Chu HY, Bloom JD. 2021. Comprehensive mapping of mutations to the SARS-CoV-2 receptor-binding domain that affect recognition by polyclonal human serum antibodies. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.12.31.425021>.
504. 2021. Neue Corona-Variante: 35 Fälle in Garmisch-Partenkirchen. BR24. <https://www.br.de/nachrichten/bayern/neue-coronavirus-mutation-35-faelle-in-garmisch-partenkirchen,SMQ1V6u>. Retrieved 8 February 2021.
505. PANGO lineages. https://cov-lineages.org/global_report.html. Retrieved 8 February 2021.
506. Starr TN, Greaney AJ, Hilton SK, Ellis D, Crawford KHD, Dingens AS, Navarro MJ, Bowen JE, Tortorici MA, Walls AC, King NP, Veelsler D, Bloom JD. 2020. [Deep Mutational Scanning of SARS-CoV-2 Receptor Binding Domain Reveals Constraints on Folding and ACE2 Binding](#). Cell 182:1295–1310.e20.
507. Gu H, Chen Q, Yang G, He L, Fan H, Deng Y-Q, Wang Y, Teng Y, Zhao Z, Cui Y, Li Y, Li X-F, Li J, Zhang N-N, Yang X, Chen S, Guo Y, Zhao G, Wang X, Luo D-Y, Wang H, Yang X, Li Y, Han G, He Y, Zhou X, Geng S, Sheng X, Jiang S, Sun S, Qin C-F, Zhou Y. 2020. [Adaptation of SARS-CoV-2 in BALB/c mice for testing vaccine efficacy](#). Science eabc4730.
508. Greaney AJ, Starr TN, Gilchuk P, Zost SJ, Binshtein E, Loes AN, Hilton SK, Huddleston J, Eguia R, Crawford KHD, Dingens AS, Nargi RS, Sutton RE, Suryadevara N, Rothlauf PW, Liu Z, Whelan SPJ, Carnahan RH, Crowe JE, Bloom JD. 2021. [Complete Mapping of Mutations to the SARS-CoV-2 Spike Receptor-Binding Domain that Escape Antibody Recognition](#). Cell Host & Microbe 29:44–57.e9.
509. McCarthy KR, Rennick LJ, Nambulli S, Robinson-McCarthy LR, Bain WG, Haidar G, Duprex WP. 2021. Recurrent deletions in the SARS-CoV-2 spike glycoprotein drive antibody escape. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.11.19.389916>.
510. Kupferschmidt K. 2021. Viral mutations may cause another ‘very, very bad’ COVID-19 wave, scientists warn. Science <https://doi.org/10.1126/science.abg4312>.
511. 2021. SARS-CoV-2 reinfection by the new Variant of Concern (VOC) P.1 in Amazonas, Brazil. Virological. <https://virological.org/t/sars-cov-2-reinfection-by-the-new-variant-of-concern-voc-p-1-in-amazonas-brazil/596>. Retrieved 8 February 2021.
512. Callaway E. 2021. [Fast-spreading COVID variant can elude immune responses](#). Nature 589:500–501.
513. Starr TN, Greaney AJ, Addetia A, Hannon WW, Choudhary MC, Dingens AS, Li JZ, Bloom JD. 2021. [Prospective mapping of viral mutations that escape antibodies used to treat COVID-19](#). Science eabf9302.
514. Kupferschmidt K. 2021. [New mutations raise specter of ‘immune escape’](#). Science 371:329–330.

515. Wu K, Werner AP, Moliva JI, Koch M, Choi A, Stewart-Jones GBE, Bennett H, Boyoglu-Barnum S, Shi W, Graham BS, Carfi A, Corbett KS, Seder RA, Edwards DK. 2021. mRNA-1273 vaccine induces neutralizing antibodies against spike mutants from global SARS-CoV-2 variants. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2021.01.25.427948>.
516. Polack FP, Thomas SJ, Kitchin N, Absalon J, Gurtman A, Lockhart S, Perez JL, Pérez Marc G, Moreira ED, Zerbini C, Bailey R, Swanson KA, Roychoudhury S, Koury K, Li P, Kalina WV, Cooper D, Frenck RW, Hammitt LL, Türeci Ö, Nell H, Schaefer A, Ünal S, Tresnan DB, Mather S, Dormitzer PR, Şahin U, Jansen KU, Gruber WC. 2020. [Safety and Efficacy of the BNT162b2 mRNA Covid-19 Vaccine](#). New England Journal of Medicine 383:2603–2615.
517. Walsh EE, Frenck RW, Falsey AR, Kitchin N, Absalon J, Gurtman A, Lockhart S, Neuzil K, Mulligan MJ, Bailey R, Swanson KA, Li P, Koury K, Kalina W, Cooper D, Fontes-Garfias C, Shi P-Y, Türeci Ö, Tompkins KR, Lyke KE, Raabe V, Dormitzer PR, Jansen KU, Şahin U, Gruber WC. 2020. [Safety and Immunogenicity of Two RNA-Based Covid-19 Vaccine Candidates](#). New England Journal of Medicine 383:2439–2450.
518. Xie X, Zou J, Fontes-Garfias CR, Xia H, Swanson KA, Cutler M, Cooper D, Menachery VD, Weaver S, Dormitzer PR, Shi P-Y. 2021. Neutralization of N501Y mutant SARS-CoV-2 by BNT162b2 vaccine-elicited sera. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2021.01.07.425740>.
519. Wang Z, Schmidt F, Weisblum Y, Muecksch F, Barnes CO, Finkin S, Schaefer-Babajew D, Cipolla M, Gaebler C, Lieberman JA, Oliveira TY, Yang Z, Abernathy ME, Huey-Tubman KE, Hurley A, Turroja M, West KA, Gordon K, Millard KG, Ramos V, Da Silva J, Xu J, Colbert RA, Patel R, Dizon J, Unson-O'Brien C, Shimeliovich I, Gazumyan A, Caskey M, Bjorkman PJ, Casellas R, Hatzioannou T, Bieniasz PD, Nussenzweig MC. 2021. mRNA vaccine-elicited antibodies to SARS-CoV-2 and circulating variants. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2021.01.15.426911>.
520. Shen X, Tang H, Pajon R, Smith G, Glenn GM, Shi W, Korber B, Montefiori DC. 2021. [Neutralization of SARS-CoV-2 Variants B.1.429 and B.1.351](#). N Engl J Med 384:2352–2354.
521. Jangra S, Ye C, Rathnasinghe R, Stadlbauer D, Krammer F, Simon V, Martinez-Sobrido L, García-Sastre A, Schotsaert M, Alshammary H, Amoako AA, Awawda MH, Beach KF, Bermúdez-González MC, Chernet RL, Eaker LQ, Ferreri ED, Floda DL, Gleason CR, Kleiner G, Jurczynszak D, Matthews JC, Mendez WA, Mulder LCF, Russo KT, Salimbangon A-BT, Saksena M, Shin AS, Sominsky LA, Srivastava K. 2021. SARS-CoV-2 spike E484K mutation reduces antibody neutralisation. The Lancet Microbe [https://doi.org/10.1016/s2666-5247\(21\)00068-9](https://doi.org/10.1016/s2666-5247(21)00068-9).
522. Collier DA, De Marco A, Ferreira IATM, Meng B, Datir RP, Walls AC, Kemp SA, Bassi J, Pinto D, Silacci-Fregni C, Bianchi S, Tortorici MA, Bowen J, Culap K, Jaconi S, Cameroni E, Snell G, Pizzuto MS, Pellanda AF, Garzoni C, Riva A, Baker S, Dougan G, Hess C, Kingston N, Lehner PJ, Lyons PA, Matheson NJ, Owehand WH, Saunders C, Summers C,

Thaventhiran JED, Toshner M, Weekes MP, Bucke A, Calder J, Canna L, Domingo J, Elmer A, Fuller S, Harris J, Hewitt S, Kennet J, Jose S, Kourampa J, Meadows A, O'Brien C, Price J, Publico C, Rastall R, Ribeiro C, Rowlands J, Ruffolo V, Tordesillas H, Bullman B, Dunmore BJ, Fawke S, Gräf S, Hodgson J, Huang C, Hunter K, Jones E, Legchenko E, Matará C, Martin J, Mescia F, O'Donnell C, Pointon L, Pond N, Shih J, Sutcliffe R, Tilly T, Treacy C, Tong Z, Wood J, Wylot M, Bergamaschi L, Betancourt A, Bower G, Cossetti C, De Sa A, Epping M, Grenfell R, Hinch A, Huhn O, Jackson S, Jarvis I, Lewis D, Marsden J, Nice F, Okecha G, Omarjee O, Perera M, Richoz N, Romashova V, Yarkoni NS, Sharma R, Stefanucci L, Stephens J, Strezlecki M, Turner L, De Bie EMDD, Bunclark K, Josipovic M, Mackay M, Rossi S, Selvan M, Spencer S, Yong C, Ansaripour A, Michael A, Mwaura L, Patterson C, Polwarth G, Polgarova P, di Stefano G, Fahey C, Michel R, Bong S-H, Coudert JD, Holmes E, Allison J, Butcher H, Caputo D, Clapham-Riley D, Dewhurst E, Furlong A, Graves B, Gray J, Ivers T, Kasanicki M, Le Gresley E, Linger R, Meloy S, Muldoon F, Ovington N, Papadia S, Phelan I, Stark H, Stirrups KE, Townsend P, Walker N, Webster J, Elmer A, Kingston N, Graves B, McCoy LE, Smith KGC, Bradley JR, Temperton N, Ceron-Gutierrez L, Barcenás-Morales G, Robson SC, Loman NJ, Connor TR, Golubchik T, Martínez Nunez RT, Ludden C, Corden S, Johnston I, Bonsall D, Smith CP, Awan AR, Bucca G, Torok ME, Saeed K, Prieto JA, Jackson DK, Hamilton WL, Snell LB, Moore C, Harrison EM, Gonçalves S, Fairley DJ, Loose MW, Watkins J, Livett R, Moses S, Amato R, Nicholls S, Bull M, Smith DL, Barrett J, Aanensen DM, Curran MD, Parmar S, Aggarwal D, Shepherd JG, Parker MD, Glaysher S, Bashton M, Underwood AP, Pacchiarini N, Loveson KF, Carabelli AM, Templeton KE, Langford CF, Sillitoe J, de Silva TI, Wang D, Kwiatkowski D, Rambaut A, O'Grady J, Cottrell S, Holden MTG, Thomson EC, Osman H, Andersson M, Chauhan AJ, Hassan-Ibrahim MO, Lawniczak M, Alderton A, Chand M, Constantinidou C, Unnikrishnan M, Darby AC, Hiscox JA, Paterson S, Martincorena I, Robertson DL, Volz EM, Page AJ, Pybus OG, Bassett AR, Ariani CV, Spencer Chapman MH, Li KK, Shah RN, Jesudason NG, Taha Y, McHugh MP, Dewar R, Jahun AS, McMurray C, Pandey S, McKenna JP, Nelson A, Young GR, McCann CM, Elliott S, Lowe H, Temperton B, Roy S, Price A, Rey S, Wyles M, Rooke S, Shaaban S, de Cesare M, Letchford L, Silveira S, Pelosi E, Wilson-Davies E, Hosmillo M, O'Toole Á, Hesketh AR, Stark R, du Plessis L, Ruis C, Adams H, Bourgeois Y, Michell SL, Grammatopoulos D, Edgeworth J, Breuer J, Todd JA, Fraser C, Buck D, John M, Kay GL, Palmer S, Peacock SJ, Heyburn D, Weldon D, Robinson E, McNally A, Muir P, Vipond IB, Boyes J, Sivaprakasam V, Salluja T, Dervisevic S, Meader EJ, Park NR, Oliver K, Jeffries AR, Ott S, da Silva Filipe A, Simpson DA, Williams C, Masoli JAH, Knight BA, Jones CR, Koshy C, Ash A, Casey A, Bosworth A, Ratcliffe L, Xu-McCrae L, Pymont HM, Hutchings S, Berry L, Jones K, Halstead F, Davis T, Holmes C, Iturriza-Gomara M, Lucaci AO, Randell PA, Cox A, Madona P, Harris KA, Brown JR, Mahungu TW, Irish-Tavares D, Haque T, Hart J, Witele E, Fenton ML, Liggett S, Graham C, Swindells E, Collins J, Eltringham G, Campbell S, McClure PC, Clark G, Sloan TJ, Jones C, Lynch J, Warne B, Leonard S, Durham J, Williams T, Haldenby ST, Storey N, Alikhan N-F, Holmes N, Moore C, Carlile M, Perry M, Craine N, Lyons RA, Beckett AH, Goudarzi S, Fearn C, Cook K, Dent H, Paul H, Davies R, Blane B, Girgis ST, Beale MA, Bellis KL, Dorman MJ, Drury E, Kane L, Kay S, McGuigan S, Nelson R, Prestwood L, Rajatileka S, Batra R, Williams RJ, Kristiansen M,

Green A, Justice A, Mahanama AIK, Samaraweera B, Hadjirin NF, Quick J, Poplawski R, Kermack LM, Reynolds N, Hall G, Chaudhry Y, Pinckert ML, Georgana I, Moll RJ, Thornton A, Myers R, Stockton J, Williams CA, Yew WC, Trotter AJ, Trebes A, MacIntyre-Cockett G, Birchley A, Adams A, Plimmer A, Gatica-Wilcox B, McKerr C, Hilvers E, Jones H, Asad H, Coombes J, Evans JM, Fina L, Gilbert L, Graham L, Cronin M, Kumziene-Summerhayes S, Taylor S, Jones S, Groves DC, Zhang P, Gallis M, Louka SF, Starinskij I, Jackson C, Gourtovaia M, Tonkin-Hill G, Lewis K, Tovar-Corona JM, James K, Baxter L, Alam MT, Orton RJ, Hughes J, Vattipally S, Ragonnet-Cronin M, Nascimento FF, Jorgensen D, Boyd O, Geidelberg L, Zarebski AE, Raghawani J, Kraemer MUG, Southgate J, Lindsey BB, Freeman TM, Keatley J-P, Singer JB, de Oliveira Martins L, Yeats CA, Abudahab K, Taylor BEW, Menegazzo M, Danesh J, Hogsden W, Eldirdiri S, Kenyon A, Mason J, Robinson TI, Holmes A, Price J, Hartley JA, Curran T, Mather AE, Shankar G, Jones R, Howe R, Morgan S, Wastenge E, Chapman MR, Mookerjee S, Stanley R, Smith W, Peto T, Eyre D, Crook D, Vernet G, Kitchen C, Gulliver H, Merrick I, Guest M, Munn R, Bradley DT, Wyatt T, Beaver C, Foulser L, Palmer S, Churcher CM, Brooks E, Smith KS, Galai K, McManus GM, Bolt F, Coll F, Meadows L, Attwood SW, Davies A, De Lacy E, Downing F, Edwards S, Scarlett GP, Jeremiah S, Smith N, Leek D, Sridhar S, Forrest S, Cormie C, Gill HK, Dias J, Higginson EE, Maes M, Young J, Wantoch M, Jamrozy D, Lo S, Patel M, Hill V, Bewshea CM, Ellard S, Auckland C, Harrison I, Bishop C, Chalker V, Richter A, Beggs A, Best A, Percival B, Mirza J, Megram O, Mayhew M, Crawford L, Ashcroft F, Moles-Garcia E, Cumley N, Hopes R, Asamaphan P, Niebel MO, Gunson RN, Bradley A, Maclean A, Mollett G, Blacow R, Bird P, Helmer T, Fallon K, Tang J, Hale AD, Macfarlane-Smith LR, Harper KL, Carden H, Machin NW, Jackson KA, Ahmad SSY, George RP, Turtle L, O'Toole E, Watts J, Breen C, Cowell A, Alcolea-Medina A, Charalampous T, Patel A, Levett LJ, Heaney J, Rowan A, Taylor GP, Shah D, Atkinson L, Lee JCD, Westhorpe AP, Jannoo R, Lowe HL, Karamani A, Ensell L, Chatterton W, Pusok M, Dadrah A, Symmonds A, Sluga G, Molnar Z, Baker P, Bonner S, Essex S, Barton E, Padgett D, Scott G, Greenaway J, Payne BAI, Burton-Fanning S, Waugh S, Raviprakash V, Sheriff N, Blakey V, Williams L-A, Moore J, Stonehouse S, Smith L, Davidson RK, Bedford L, Coupland L, Wright V, Chappell JG, Tsoleridis T, Ball J, Khakh M, Fleming VM, Lister MM, Howson-Wells HC, Berry L, Boswell T, Joseph A, Willingham I, Duckworth N, Walsh S, Wise E, Moore N, Mori M, Cortes N, Kidd S, Williams R, Gifford L, Bicknell K, Wyllie S, Lloyd A, Impey R, Malone CS, Cogger BJ, Levene N, Monaghan L, Keeley AJ, Partridge DG, Raza M, Evans C, Johnson K, Betteridge E, Farr BW, Goodwin S, Quail MA, Scott C, Shirley L, Thurston SAJ, Rajan D, Bronner IF, Aigrain L, Redshaw NM, Lensing SV, McCarthy S, Makunin A, Balcazar CE, Gallagher MD, Williamson KA, Stanton TD, Michelsen ML, Warwick-Dugdale J, Manley R, Farbos A, Harrison JW, Sambles CM, Studholme DJ, Lackenby A, Mbisa T, Platt S, Miah S, Bibby D, Manso C, Hubb J, Dabrera G, Ramsay M, Bradshaw D, Schaefer U, Groves N, Gallagher E, Lee D, Williams D, Ellaby N, Hartman H, Manesis N, Patel V, Ledesma J, Twohig KA, Allara E, Pearson C, Cheng JKJ, Bridgewater HE, Frost LR, Taylor-Joyce G, Brown PE, Tong L, Broos A, Mair D, Nichols J, Carmichael SN, Smollett KL, Nomikou K, Aranday-Cortes E, Johnson N, Nickbakhsh S, Vamos EE, Hughes M, Rainbow L, Eccles R, Nelson C, Whitehead M, Gregory R, Gemmell M, Wierzbicki C, Webster HJ, Fisher CL, Signell AW, Betancor G, Wilson HD, Nebbia G, Flaviani F, Cerda AC,

Merrill TV, Wilson RE, Cotic M, Bayzid N, Thompson T, Acheson E, Rushton S, O'Brien S, Baker DJ, Rudder S, Aydin A, Sang F, Debebe J, Francois S, Vasylyeva TI, Zamudio ME, Gutierrez B, Marchbank A, Maksimovic J, Spellman K, McCluggage K, Morgan M, Beer R, Afifi S, Workman T, Fuller W, Bresner C, Angyal A, Green LR, Parsons PJ, Tucker RM, Brown R, Whiteley M, Bonfield J, Puethe C, Whitwham A, Liddle J, Rowe W, Siveroni I, Le-Viet T, Gaskin A, Johnson R, Abnizova I, Aigrain L, Alderton A, Ali M, Allen L, Amato R, Anderson R, Ariani C, Austin-Guest S, Bala S, Barrett J, Bassett A, Battleday K, Beal J, Beale M, Beaver C, Bellany S, Bellerby T, Bellis K, Berger D, Berriman M, Betteridge E, Bevan P, Binley S, Bishop J, Blackburn K, Bonfield J, Boughton N, Bowker S, Brendler-Spaeth T, Bronner I, Brooklyn T, Buddenborg SK, Bush R, Caetano C, Cagan A, Carter N, Cartwright J, Monteiro TC, Chapman L, Chillingworth T-J, Clapham P, Clark R, Clarke A, Clarke C, Cole D, Cook E, Coppola M, Cornell L, Cornwell C, Corton C, Crackett A, Cranage A, Craven H, Craw S, Crawford M, Cutts T, Dabrowska M, Davies M, Davies R, Dawson J, Day C, Densem A, Dibling T, Dockree C, Dodd D, Dogga S, Dorman M, Dougan G, Dougherty M, Dove A, Drummond L, Drury E, Dudek M, Durham J, Durrant L, Easthope E, Eckert S, Ellis P, Farr B, Fenton M, Ferrero M, Flack N, Fordham H, Forsythe G, Foulser L, Francis M, Fraser A, Freeman A, Galvin A, Garcia-Casado M, Gedny A, Girgis S, Glover J, Goncalves S, Goodwin S, Gould O, Gourtovaia M, Gray A, Gray E, Griffiths C, Gu Y, Guerin F, Hamilton W, Hanks H, Harrison E, Harrott A, Harry E, Harvison J, Heath P, Hernandez-Koutoucheva A, Hobbs R, Holland D, Holmes S, Hornett G, Hough N, Huckle L, Hughes-Hallet L, Hunter A, Inglis S, Iqbal S, Jackson A, Jackson D, James K, Jamrozy D, Verdejo CJ, Johnston I, Jones M, Kallepally K, Kane L, Kay K, Kay S, Keatley J, Keith A, King A, Kitchen L, Kleanthous M, Klimekova M, Korlevic P, Krasheninnikova K, Kwiatkowski D, Lane G, Langford C, Laverack A, Law K, Lawniczak M, Lensing S, Leonard S, Letchford L, Lewis K, Lewis-Wade A, Liddle J, Lin Q, Lindsay S, Linsdell S, Livett R, Lo S, Long R, Lovell J, Lovell J, Ludden C, Mack J, Maddison M, Makunin A, Mamun I, Mansfield J, Marriott N, Martin M, Martincorena I, Mayho M, McCarthy S, McClintock J, McGuigan S, McHugh S, McMinn L, Meadows C, Mobley E, Moll R, Morra M, Morrow L, Murie K, Nash S, Nathwani C, Naydenova P, Neaverson A, Nelson R, Nerou E, Nicholson J, Nimz T, Noell GG, O'Meara S, Ohan V, Oliver K, Olney C, Ormond D, Oszlanczi A, Palmer S, Pang YF, Pardubska B, Park N, Parmar A, Patel G, Patel M, Payne M, Peacock S, Petersen A, Plowman D, Preston T, Prestwood L, Puethe C, Quail M, Rajan D, Rajatileka S, Rance R, Rawlings S, Redshaw N, Reynolds J, Reynolds M, Rice S, Richardson M, Roberts C, Robinson K, Robinson M, Robinson D, Rogers H, Rojo EM, Roopra D, Rose M, Rudd L, Sadri R, Salmon N, Saul D, Schwach F, Scott C, Seekings P, Shirley L, Sillitoe J, Simms A, Sinnott M, Sivadasan S, Siwek B, Sizer D, Skeldon K, Skelton J, Slater-Tunstall J, Sloper L, Smerdon N, Smith C, Smith C, Smith J, Smith K, Smith M, Smith S, Smith T, Sneade L, Soria CD, Sousa C, Souster E, Sparkes A, Spencer-Chapman M, Squares J, Stanley R, Steed C, Stickland T, Still I, Stratton M, Strickland M, Swann A, Swiatkowska A, Sycamore N, Swift E, Symons E, Szluha S, Taluy E, Tao N, Taylor K, Taylor S, Thompson S, Thompson M, Thomson M, Thomson N, Thurston S, Tonkin-Hill G, Toombs D, Topping B, Tovar-Corona J, Ungureanu D, Uphill J, Urbanova J, Van PJ, Vancollie V, Voak P, Walker D, Walker M, Waller M, Ward G, Weatherhogg C, Webb N, Weldon D,

- Wells A, Wells E, Westwood L, Whipp T, Whiteley T, Whitton G, Whitwham A, Widaa S, Williams M, Wilson M, Wright S, Harvey W, Virgin HW, Lanzavecchia A, Piccoli L, Doffinger R, Wills M, Velesler D, Corti D, Gupta RK. 2021. [Sensitivity of SARS-CoV-2 B.1.1.7 to mRNA vaccine-elicited antibodies](#). *Nature* 593:136–141.
523. Dearlove B, Lewitus E, Bai H, Li Y, Reeves DB, Joyce MG, Scott PT, Amare MF, Vasan S, Michael NL, Modjarrad K, Rolland M. 2020. [A SARS-CoV-2 vaccine candidate would likely match all currently circulating variants](#). *Proceedings of the National Academy of Sciences* 117:23652–23662.
 524. Sanjuán R, Nebot MR, Chirico N, Mansky LM, Belshaw R. 2010. [Viral Mutation Rates](#). *J Virol* 84:9733–9748.
 525. Manzanares-Meza LD, Medina-Contreras O. 2020. [SARS-CoV-2 and influenza: a comparative overview and treatment implications](#). *BMJ* 77.
 526. Li X, Deem MW. 2016. [Influenza evolution and H3N2 vaccine effectiveness, with application to the 2014/2015 season](#). *Protein Engineering, Design and Selection* 29:309–315.
 527. Liu Y, Liu J, Xia H, Zhang X, Fontes-Garfias CR, Swanson KA, Cai H, Sarkar R, Chen W, Cutler M, Cooper D, Weaver SC, Muik A, Sahin U, Jansen KU, Xie X, Dormitzer PR, Shi P-Y. 2021. [Neutralizing Activity of BNT162b2-Elicited Serum](#). *N Engl J Med* 384:1466–1468.
 528. 2021. Covid-19 vaccine effectiveness affected by variants. *Pharmaceutical Technology*. <https://www.pharmaceutical-technology.com/comment/covid-19-vaccine-effectiveness-affected-by-variants/>. Retrieved 5 December 2022.
 529. 2021. The effects of virus variants on COVID-19 vaccines. <https://www.who.int/news-room/feature-stories/detail/the-effects-of-virus-variants-on-covid-19-vaccines>. Retrieved 5 December 2022.
 530. Bonomo ME, Deem MW. 2018. [Predicting Influenza H3N2 Vaccine Efficacy From Evolution of the Dominant Epitope](#). *Clinical Infectious Diseases* 67:1129–1131.
 531. Kim JH, Marks F, Clemens JD. 2021. [Looking beyond COVID-19 vaccine phase 3 trials](#). *Nat Med* 27:205–211.
 532. Mathieu E, Ritchie H, Rodés-Guirao L, Appel C, Giattino C, Hasell J, Macdonald B, Dattani S, Beltekian D, Ortiz-Ospina E, Roser M. 2020. [Coronavirus Pandemic \(COVID-19\)](#). *Our World in Data*.
 533. Knyazev S, Chhugani K, Sarwal V, Ayyala R, Singh H, Karthikeyan S, Deshpande D, Baykal PI, Comarova Z, Lu A, Porozov Y, Vasylyeva TI, Wertheim JO, Tierney BT, Chiu CY, Sun R, Wu A, Abedalthagafi MS, Pak VM, Nagaraj SH, Smith AL, Skums P, Pasaniuc B, Komissarov A, Mason CE, Bortz E, Lemey P, Kondrashov F, Beerenwinkel N, Lam TT-Y, Wu NC, Zelikovsky A, Knight R, Crandall KA, Mangul S. 2022. [Unlocking capacities of genomics for the COVID-19 response and future pandemics](#). *Nat Methods* 19:374–380.

534. Aarestrup FM, Bonten M, Koopmans M. 2021. [Pandemics– One Health preparedness for the next](#). The Lancet Regional Health - Europe 9:100210.
535. Li J, Lai S, Gao GF, Shi W. 2021. [The emergence, genomic diversity and global spread of SARS-CoV-2](#). Nature 600:408–418.
536. Tessema SK, Inzaule SC, Christoffels A, Kebede Y, de Oliveira T, Ouma AEO, Happi CT, Nkengasong JN. 2020. [Accelerating genomics-based surveillance for COVID-19 response in Africa](#). The Lancet Microbe 1:e227–e228.
537. Hart OE, Halden RU. 2020. [Computational analysis of SARS-CoV-2/COVID-19 surveillance by wastewater-based epidemiology locally and globally: Feasibility, economy, opportunities and challenges](#). Science of The Total Environment 730:138875.
538. Medema G, Been F, Heijnen L, Petterson S. 2020. [Implementation of environmental surveillance for SARS-CoV-2 virus to support public health decisions: Opportunities and challenges](#). Current Opinion in Environmental Science & Health 17:49–71.
539. Ahmed W, Bivins A, Bertsch PM, Bibby K, Choi PM, Farkas K, Gyawali P, Hamilton KA, Haramoto E, Kitajima M, Simpson SL, Tandukar S, Thomas KV, Mueller JF. 2020. [Surveillance of SARS-CoV-2 RNA in wastewater: Methods optimization and quality control are crucial for generating reliable public health information](#). Current Opinion in Environmental Science & Health 17:82–93.
540. Wu Y, Guo C, Tang L, Hong Z, Zhou J, Dong X, Yin H, Xiao Q, Tang Y, Qu X, Kuang L, Fang X, Mishra N, Lu J, Shan H, Jiang G, Huang X. 2020. [Prolonged presence of SARS-CoV-2 viral RNA in faecal samples](#). The Lancet Gastroenterology & Hepatology 5:434–435.
541. Trujillo M, Cheung K, Gao A, Hoxie I, Kannoly S, Kubota N, San KM, Smyth DS, Dennehy JJ. 2021. [Protocol for safe, affordable, and reproducible isolation and quantitation of SARS-CoV-2 RNA from wastewater](#). PLoS ONE 16:e0257454.
542. Hoar C, Chauvin F, Clare A, McGibbon H, Castro E, Patinella S, Katehis D, Dennehy JJ, Trujillo M, Smyth DS, Silverman AI. 2022. [Monitoring SARS-CoV-2 in wastewater during New York City's second wave of COVID-19: sewershed-level trends and relationships to publicly available clinical testing data](#). Environ Sci: Water Res Technol 8:1021–1035.
543. Gerrity D, Papp K, Stoker M, Sims A, Frehner W. 2021. [Early-pandemic wastewater surveillance of SARS-CoV-2 in Southern Nevada: Methodology, occurrence, and incidence/prevalence considerations](#). Water Research X 10:100086.
544. Wu F, Zhang J, Xiao A, Gu X, Lee WL, Armas F, Kauffman K, Hanage W, Matus M, Ghaeli N, Endo N, Duvallet C, Poyet M, Moniz K, Washburne AD, Erickson TB, Chai PR, Thompson J, Alm EJ. 2020. [SARS-CoV-2 Titers in Wastewater Are Higher than Expected from Clinically Confirmed Cases](#). mSystems 5.

545. Schussman MK, Roguet A, Schmoldt A, Dinan B, McLellan SL. 2022. [Wastewater surveillance using ddPCR reveals highly accurate tracking of Omicron variant due to altered N1 probe binding efficiency](#). Cold Spring Harbor Laboratory.
546. Lee WL, Gu X, Armas F, Wu F, Chandra F, Chen H, Xiao A, Leifels M, Chua FJD, Kwok GW, Tay JY, Lim CY, Thompson J, Alm EJ. 2021. [Quantitative detection of SARS-CoV-2 Omicron BA.1 and BA.2 variants in wastewater through allele-specific RT-qPCR](#). Cold Spring Harbor Laboratory.
547. CDC. 2022. National Wastewater Surveillance System. Centers for Disease Control and Prevention. <https://www.cdc.gov/healthywater/surveillance/wastewater-surveillance/wastewater-surveillance.html>. Retrieved 5 December 2022.
548. Vavrek D, Speroni L, Curnow KJ, Oberholzer M, Moeder V, Febbo PG. 2021. [Genomic surveillance at scale is required to detect newly emerging strains at an early timepoint](#). Cold Spring Harbor Laboratory.
549. Dobson AP, Pimm SL, Hannah L, Kaufman L, Ahumada JA, Ando AW, Bernstein A, Busch J, Daszak P, Engelmann J, Kinnaird MF, Li BV, Loch-Temzelides T, Lovejoy T, Nowak K, Roehrdanz PR, Vale MM. 2020. [Ecology and economics for pandemic prevention](#). Science 369:379–381.
550. Carlson CJ, Albery GF, Merow C, Trisos CH, Zipfel CM, Eskew EA, Olival KJ, Ross N, Bansal S. 2022. [Climate change increases cross-species viral transmission risk](#). Nature 607:555–562.
551. Neiderud C-J. 2015. [How urbanization affects the epidemiology of emerging infectious diseases](#). Infection Ecology & Epidemiology 5:27060.
552. Matukas LM, Dhalla IA, Laupacis A. 2020. [Aggressively find, test, trace and isolate to beat COVID-19](#). CMAJ 192:E1164–E1165.
553. Jefferies S, French N, Gilkison C, Graham G, Hope V, Marshall J, McElroy C, McNeill A, Muellner P, Paine S, Prasad N, Scott J, Sherwood J, Yang L, Priest P. 2020. [COVID-19 in New Zealand and the impact of the national response: a descriptive epidemiological study](#). The Lancet Public Health 5:e612–e623.
554. Summers J, Cheng H-Y, Lin H-H, Barnard LT, Kvalsvig A, Wilson N, Baker MG. 2020. [Potential lessons from the Taiwan and New Zealand health responses to the COVID-19 pandemic](#). The Lancet Regional Health - Western Pacific 4:100044.
555. Novel Coronavirus – China. <https://www.who.int/emergencies/disease-outbreak-news/item/2020-DON233>. Retrieved 5 December 2022.
556. Corman VM, Landt O, Kaiser M, Molenkamp R, Meijer A, Chu DK, Bleicker T, Brünink S, Schneider J, Schmidt ML, Mulders DG, Haagmans BL, van der Veer B, van den Brink S, Wijsman L, Goderski G, Romette J-L, Ellis J, Zambon M, Peiris M, Goossens H, Reusken C, Koopmans MP,

- Drosten C. 2020. [Detection of 2019 novel coronavirus \(2019-nCoV\) by real-time RT-PCR](#). Eurosurveillance 25.
557. Garibyan L, Avashia N. 2013. [Polymerase Chain Reaction](#). Journal of Investigative Dermatology 133:1–4.
 558. Ngom B, Guo Y, Wang X, Bi D. 2010. [Development and application of lateral flow test strip technology for detection of infectious agents and chemical contaminants: a review](#). Anal Bioanal Chem 397:1113–1135.
 559. Alhajj M, Farhana A. 2022. Enzyme Linked Immunosorbent AssayStatPearls. StatPearls. <https://pubmed.ncbi.nlm.nih.gov/32310382>.
 560. Gong F, Wei H, Li Q, Liu L, Li B. 2021. [Evaluation and Comparison of Serological Methods for COVID-19 Diagnosis](#). Front Mol Biosci 8.
 561. Burbelo PD, Riedo FX, Morishima C, Rawlings S, Smith D, Das S, Strich JR, Chertow DS, Davey RT Jr, Cohen JL. 2020. [Sensitivity in Detection of Antibodies to Nucleocapsid and Spike Proteins of Severe Acute Respiratory Syndrome Coronavirus 2 in Patients With Coronavirus Disease 2019](#). The Journal of Infectious Diseases 222:206–213.
 562. To KK-W, Tsang OT-Y, Leung W-S, Tam AR, Wu T-C, Lung DC, Yip CC-Y, Cai J-P, Chan JM-C, Chik TS-H, Lau DP-L, Choi CY-C, Chen L-L, Chan W-M, Chan K-H, Ip JD, Ng AC-K, Poon RW-S, Luo C-T, Cheng VC-C, Chan JF-W, Hung IF-N, Chen Z, Chen H, Yuen K-Y. 2020. [Temporal profiles of viral load in posterior oropharyngeal saliva samples and serum antibody responses during infection by SARS-CoV-2: an observational cohort study](#). The Lancet Infectious Diseases 20:565–574.
 563. Zhang J, Wang S, Xue Y. 2020. [Fecal specimen diagnosis 2019 novel coronavirus-infected pneumonia](#). Journal of Medical Virology 92:680–682.
 564. Adams G. 2020. [A beginner's guide to RT-PCR, qPCR and RT-qPCR](#). The Biochemist 42:48–53.
 565. Bustin SA, Benes V, Garson JA, Hellemans J, Huggett J, Kubista M, Mueller R, Nolan T, Pfaffl MW, Shipley GL, Vandesompele J, Wittwer CT. 2009. [The MIQE Guidelines: Minimum Information for Publication of Quantitative Real-Time PCR Experiments](#). Clinical Chemistry 55:611–622.
 566. Chu DKW, Pan Y, Cheng SMS, Hui KPY, Krishnan P, Liu Y, Ng DYM, Wan CKC, Yang P, Wang Q, Peiris M, Poon LLM. 2020. [Molecular Diagnosis of a Novel Coronavirus \(2019-nCoV\) Causing an Outbreak of Pneumonia](#). Clinical Chemistry 66:549–555.
 567. La Scola B, Le Bideau M, Andreani J, Hoang VT, Grimaldier C, Colson P, Gautret P, Raoult D. 2020. [Viral RNA load as determined by cell culture as a management tool for discharge of SARS-CoV-2 patients from infectious disease wards](#). Eur J Clin Microbiol Infect Dis 39:1059–1061.
 568. van Kampen JJA, van de Vijver DAMC, Fraaij PLA, Haagmans BL, Lamers MM, Okba N, van den Akker JPC, Endeman H, Gommers DAMPJ,

Cornelissen JJ, Hoek RAS, van der Eerden MM, Hesselink DA, Metselaar HJ, Verbon A, de Steenwinkel JEM, Aron GI, van Gorp ECM, van Boheemen S, Voermans JC, Boucher CAB, Molenkamp R, Koopmans MPG, Geurtsvankessel C, van der Eijk AA. 2021. [Duration and key determinants of infectious virus shedding in hospitalized patients with coronavirus disease-2019 \(COVID-19\)](#). Nat Commun 12.

569. Dahdouh E, Lázaro-Perona F, Romero-Gómez MP, Mingorance J, García-Rodríguez J. 2021. [Ct values from SARS-CoV-2 diagnostic PCR assays should not be used as direct estimates of viral load](#). Journal of Infection 82:414–451.
570. Wikramaratna PS, Paton RS, Ghafari M, Lourenço J. 2020. [Estimating the false-negative test probability of SARS-CoV-2 by RT-PCR](#). Eurosurveillance 25.
571. Cohen AN, Kessel B, Milgroom MG. 2020. [Diagnosing SARS-CoV-2 infection: the danger of over-reliance on positive test results](#). Cold Spring Harbor Laboratory.
572. Tsang NNY, So HC, Ng KY, Cowling BJ, Leung GM, Ip DKM. 2021. [Diagnostic performance of different sampling approaches for SARS-CoV-2 RT-PCR testing: a systematic review and meta-analysis](#). The Lancet Infectious Diseases 21:1233–1245.
573. Chan JF-W, Yip CC-Y, To KK-W, Tang TH-C, Wong SC-Y, Leung K-H, Fung AY-F, Ng AC-K, Zou Z, Tsoi H-W, Choi GK-Y, Tam AR, Cheng VC-C, Chan K-H, Tsang OT-Y, Yuen K-Y. 2020. [Improved Molecular Diagnosis of COVID-19 by the Novel, Highly Sensitive and Specific COVID-19-RdRp/HeI Real-Time Reverse Transcription-PCR Assay Validated In Vitro and with Clinical Specimens](#). J Clin Microbiol 58.
574. Mercer TR, Salit M. 2021. [Testing at scale during the COVID-19 pandemic](#). Nat Rev Genet 22:415–426.
575. Ziegler K, Steininger P, Ziegler R, Steinmann J, Korn K, Ensser A. 2020. [SARS-CoV-2 samples may escape detection because of a single point mutation in the N gene](#). Eurosurveillance 25.
576. Peñarrubia L, Ruiz M, Porco R, Rao SN, Juanola-Falgarona M, Manissero D, López-Fontanals M, Pareja J. 2020. [Multiple assays in a real-time RT-PCR SARS-CoV-2 panel can mitigate the risk of loss of sensitivity by new genomic variants during the COVID-19 outbreak](#). International Journal of Infectious Diseases 97:225–229.
577. Taylor SC, Laperriere G, Germain H. 2017. [Droplet Digital PCR versus qPCR for gene expression analysis with low abundant targets: from variable nonsense to publication quality data](#). Sci Rep 7.
578. Quan P-L, Sauzade M, Brouzes E. 2018. [dPCR: A Technology Review](#). Sensors 18:1271.
579. Suo T, Liu X, Feng J, Guo M, Hu W, Guo D, Ullah H, Yang Y, Zhang Q, Wang X, Sajid M, Huang Z, Deng L, Chen T, Liu F, Xu K, Liu Y, Zhang Q, Liu Y, Xiong Y, Chen G, Lan K, Chen Y. 2020. [ddPCR: a more accurate](#)

[tool for SARS-CoV-2 detection in low viral load specimens](#). Emerging Microbes & Infections 9:1259–1268.

580. Dong L, Zhou J, Niu C, Wang Q, Pan Y, Sheng S, Wang X, Zhang Y, Yang J, Liu M, Zhao Y, Zhang X, Zhu T, Peng T, Xie J, Gao Y, Wang D, Dai X, Fang X. 2021. [Highly accurate and sensitive diagnostic detection of SARS-CoV-2 by digital PCR](#). Talanta 224:121726.
581. Hess JF, Kohl TA, Kotrová M, Rönsch K, Paprotka T, Mohr V, Hutzenlaub T, Brüggemann M, Zengerle R, Niemann S, Paust N. 2020. [Library preparation for next generation sequencing: A review of automation strategies](#). Biotechnology Advances 41:107537.
582. Udugama B, Kadhiresan P, Kozłowski HN, Malekjahani A, Osborne M, Li VYC, Chen H, Mubareka S, Gubbay JB, Chan WCW. 2020. [Diagnosing COVID-19: The Disease and Tools for Detection](#). ACS Nano 14:3822–3835.
583. Robishaw JD, Alter SM, Solano JJ, Shih RD, DeMets DL, Maki DG, Hennekens CH. 2021. [Genomic surveillance to combat COVID-19: challenges and opportunities](#). The Lancet Microbe 2:e481–e484.
584. Cov-Lineages. <https://cov-lineages.org/>. Retrieved 5 December 2022.
585. Brito AF, Semenova E, Dudas G, Hassler GW, Kalinich CC, Kraemer MUG, Ho J, Tegally H, Githinji G, Agoti CN, Matkin LE, Whittaker C, Howden BP, Sintchenko V, Zuckerman NS, Mor O, Blankenship HM, Oliveira T de, Lin RTP, Siqueira MM, Resende PC, Vasconcelos ATR, Spilki FR, Aguiar RS, Alexiev I, Ivanov IN, Philipova I, Carrington CVF, Sahadeo NSD, Gurry C, Maurer-Stroh S, Naidoo D, von Eije KJ, Perkins MD, Kerkhove M van, Hill SC, Sabino EC, Pybus OG, Dye C, Bhatt S, Flaxman S, Suchard MA, Grubaugh ND, Baele G, Faria NR. 2021. [Global disparities in SARS-CoV-2 genomic surveillance](#). Cold Spring Harbor Laboratory.
586. Yelin I, Aharony N, Tamar ES, Argoetti A, Messer E, Berenbaum D, Shafran E, Kuzli A, Gandali N, Shkedi O, Hashimshony T, Mandel-Gutfreund Y, Halberthal M, Geffen Y, Szwarcwort-Cohen M, Kishony R. 2020. [Evaluation of COVID-19 RT-qPCR Test in Multi sample Pools](#). Clinical Infectious Diseases 71:2073–2078.
587. Nelson AC, Auch B, Schomaker M, Gohl DM, Grady P, Johnson D, Kincaid R, Karnuth KE, Daniel J, Fiege JK, Fay EJ, Bold T, Langlois RA, Beckman KB, Yohe S. 2020. Analytical Validation of a COVID-19 qRT-PCR Detection Assay Using a 384-well Format and Three Extraction Methods. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.04.02.022186>.
588. Heidarzadeh A, Narayanan K. 2021. [Two-Stage Adaptive Pooling with RT-QPCR for Covid-19 Screening](#) ICASSP 2021 - 2021 IEEE International Conference on Acoustics, Speech and Signal Processing (ICASSP). IEEE.
589. Joachim A, Dewald F, Suárez I, Zemlin M, Lang I, Stutz R, Marthaler A, Bosse HM, Lübke N, Münch J, Bernard M-A, Jeltsch K, Tönshoff B, Weidner N, Kräusslich H-G, Birzele L, Hübner J, Schmied P, Meyer-Bühn M, Horemheb-Rubio G, Cornely OA, Haverkamp H, Wiesmüller G,

Fätkenheuer G, Hero B, Kaiser R, Dötsch J, Rybníček J, Cosgun ZC, Hünseler C, Schönenkorb J, Wurm J, Klein F, Heger E, Knops E, Sierra-Aragón S, Kretschmer AC, Sprute R, Kossow A, Hellmich M, Shah-Hosseini K, Weiss M, Goedicke-Fritz S, Kaiser E, Meyer S, Seiwert N, Smola S, Pfuhl T, Lohse S, Schupp A-K, Timm J, Gröne N, Lesmann H, Bredahl R, Schneble L, Turinsky M, Patry C, Hoffmann GF, Müller B, Börner K, Schnitzler P, Heuser A-M, Welker A, von Both U, Kern A. 2021. [Pooled RT-qPCR testing for SARS-CoV-2 surveillance in schools - a cluster randomised trial](#). *EClinicalMedicine* 39:101082.

590. Hirotsu Y, Maejima M, Shibusawa M, Nagakubo Y, Hosaka K, Amemiya K, Sueki H, Hayakawa M, Mochizuki H, Tsutsui T, Kakizaki Y, Miyashita Y, Omata M. 2020. [Pooling RT-qPCR testing for SARS-CoV-2 in 1000 individuals of healthy and infection-suspected patients](#). *Sci Rep* 10.
591. Notomi T. 2000. [Loop-mediated isothermal amplification of DNA](#). *Nucleic Acids Research* 28:63e–63.
592. Amaral C, Antunes W, Moe E, Duarte AG, Lima LMP, Santos C, Gomes IL, Afonso GS, Vieira R, Teles HSS, Reis MS, da Silva MAR, Henriques AM, Fevereiro M, Ventura MR, Serrano M, Pimentel C. 2021. [A molecular test based on RT-LAMP for rapid, sensitive and inexpensive colorimetric detection of SARS-CoV-2 in clinical samples](#). *Sci Rep* 11.
593. Al Bayat S, Mundodan J, Hasnain S, Sallam M, Khogali H, Ali D, Alateeg S, Osama M, Elberdiny A, Al-Romaihi H, Al-Thani MHJ. 2021. [Can the cycle threshold \(Ct\) value of RT-PCR test for SARS CoV2 predict infectivity among close contacts?](#) *Journal of Infection and Public Health* 14:1201–1205.
594. Mowrer CT, Creager H, Cawcutt K, Birge J, Lyden E, Van Schooneveld TC, Rupp ME, Hewlett A. 2021. [Evaluation of cycle threshold values at deisolation](#). *Infect Control Hosp Epidemiol* 43:794–796.
595. Tom MR, Mina MJ. 2020. [To Interpret the SARS-CoV-2 Test, Consider the Cycle Threshold Value](#). *Clinical Infectious Diseases* 71:2252–2254.
596. Zhang Y, Odiwuor N, Xiong J, Sun L, Nyaruaba RO, Wei H, Tanner NA. 2020. [Rapid Molecular Detection of SARS-CoV-2 \(COVID-19\) Virus RNA Using Colorimetric LAMP](#). Cold Spring Harbor Laboratory.
597. Subali AD, Wiyono L. 2021. [Reverse Transcriptase Loop Mediated Isothermal Amplification \(RT-LAMP\) for COVID-19 diagnosis: a systematic review and meta-analysis](#). *Pathog Glob Health* 115:281–291.
598. Adli M. 2018. [The CRISPR tool kit for genome editing and beyond](#). *Nat Commun* 9.
599. Gootenberg JS, Abudayyeh OO, Lee JW, Essletzbichler P, Dy AJ, Joung J, Verdine V, Donghia N, Daringer NM, Freije CA, Myhrvold C, Bhattacharyya RP, Livny J, Regev A, Koonin EV, Hung DT, Sabeti PC, Collins JJ, Zhang F. 2017. [Nucleic acid detection with CRISPR-Cas13a/C2c2](#). *Science* 356:438–442.
600. Hou T, Zeng W, Yang M, Chen W, Ren L, Ai J, Wu J, Liao Y, Gou X, Li Y, Wang X, Su H, Gu B, Wang J, Xu T. 2020. [Development and evaluation](#)

[of a rapid CRISPR-based diagnostic for COVID-19](#). PLoS Pathog 16:e1008705.

601. Metsky HC, Freije CA, Kosoko-Thoroddsen T-SF, Sabeti PC, Myhrvold C. 2020. CRISPR-based surveillance for COVID-19 using genomically-comprehensive machine learning design. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.26.967026>.
602. Rauch JN, Valois E, Solley SC, Braig F, Lach RS, Audouard M, Ponce-Rojas JC, Costello MS, Baxter NJ, Kosik KS, Arias C, Acosta-Alvear D, Wilson MZ. 2021. [A Scalable, Easy-to-Deploy Protocol for Cas13-Based Detection of SARS-CoV-2 Genetic Material](#). J Clin Microbiol 59.
603. Broughton JP, Deng X, Yu G, Fasching CL, Servellita V, Singh J, Miao X, Streithorst JA, Granados A, Sotomayor-Gonzalez A, Zorn K, Gopez A, Hsu E, Gu W, Miller S, Pan C-Y, Guevara H, Wadford DA, Chen JS, Chiu CY. 2020. [CRISPR-Cas12-based detection of SARS-CoV-2](#). Nature Biotechnology 38:870–874.
604. Lucia C, Federico P-B, Alejandra GC. 2020. An ultrasensitive, rapid, and portable coronavirus SARS-CoV-2 sequence detection method based on CRISPR-Cas12. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.29.971127>.
605. Xiong D, Dai W, Gong J, Li G, Liu N, Wu W, Pan J, Chen C, Jiao Y, Deng H, Ye J, Zhang X, Huang H, Li Q, Xue L, Zhang X, Tang G. 2020. [Rapid detection of SARS-CoV-2 with CRISPR-Cas12a](#). PLoS Biol 18:e3000978.
606. Ding X, Yin K, Li Z, Lalla RV, Ballesteros E, Sfeir MM, Liu C. 2020. [Ultrasensitive and visual detection of SARS-CoV-2 using all-in-one dual CRISPR-Cas12a assay](#). Nat Commun 11.
607. Guo L, Sun X, Wang X, Liang C, Jiang H, Gao Q, Dai M, Qu B, Fang S, Mao Y, Chen Y, Feng G, Gu Q, Wang RR, Zhou Q, Li W. 2020. [SARS-CoV-2 detection with CRISPR diagnostics](#). Cell Discov 6.
608. Ramachandran A, Huyke DA, Sharma E, Sahoo MK, Huang C, Banaei N, Pinsky BA, Santiago JG. 2020. [Electric field-driven microfluidics for rapid CRISPR-based diagnostics and its application to detection of SARS-CoV-2](#). Proc Natl Acad Sci USA 117:29518–29525.
609. Makarova KS, Koonin EV. 2015. [Annotation and Classification of CRISPR-Cas Systems](#), p. 47–75. *In* Methods in Molecular Biology. Springer New York.
610. Pyenson NC, Marraffini LA. 2017. [Type III CRISPR-Cas systems: when DNA cleavage just isn't enough](#). Current Opinion in Microbiology 37:150–154.
611. Tamulaitis G, Venclovas Č, Siksnys V. 2017. [Type III CRISPR-Cas Immunity: Major Differences Brushed Aside](#). Trends in Microbiology 25:49–61.
612. Santiago-Frangos A, Hall LN, Nemudraia A, Nemudryi A, Krishna P, Wiegand T, Wilkinson RA, Snyder DT, Hedges JF, Cicha C, Lee HH, Graham A, Jutila MA, Taylor MP, Wiedenheft B. 2021. [Intrinsic signal](#)

[amplification by type III CRISPR-Cas systems provides a sequence-specific SARS-CoV-2 diagnostic](#). Cell Reports Medicine 2:100319.

613. Fozouni P, Son S, Díaz de León Derby M, Knott GJ, Gray CN, D'Ambrosio MV, Zhao C, Switz NA, Kumar GR, Stephens SI, Boehm D, Tsou C-L, Shu J, Bhuiya A, Armstrong M, Harris AR, Chen P-Y, Osterloh JM, Meyer-Franke A, Joehnk B, Walcott K, Sil A, Langelier C, Pollard KS, Crawford ED, Puschnik AS, Phelps M, Kistler A, DeRisi JL, Doudna JA, Fletcher DA, Ott M. 2021. [Amplification-free detection of SARS-CoV-2 with CRISPR-Cas13a and mobile phone microscopy](#). Cell 184:323–333.e9.
614. Brandsma E, Verhagen HJMP, van de Laar TJW, Claas ECJ, Cornelissen M, van den Akker E. 2020. [Rapid, Sensitive, and Specific Severe Acute Respiratory Syndrome Coronavirus 2 Detection: A Multicenter Comparison Between Standard Quantitative Reverse-Transcriptase Polymerase Chain Reaction and CRISPR-Based DETECTR](#). The Journal of Infectious Diseases 223:206–213.
615. Azhar Mohd, Phutela R, Kumar M, Ansari AH, Rauthan R, Gulati S, Sharma N, Sinha D, Sharma S, Singh S, Acharya S, Paul D, Kathpalia P, Aich M, Sehgal P, Ranjan G, Bhoyar RC, Singhal K, Lad H, Patra PK, Makharia G, Chandak GR, Pesala B, Chakraborty D, Maiti S. 2020. [Rapid, accurate, nucleobase detection using FnCas9](#). Cold Spring Harbor Laboratory.
616. Drain PK. 2022. [Rapid Diagnostic Testing for SARS-CoV-2](#). N Engl J Med 386:264–272.
617. Alghounaim M, Bastaki H, Bin Essa F, Motlagh H, Al-Sabah S. 2021. [The Performance of Two Rapid Antigen Tests During Population-Level Screening for SARS-CoV-2 Infection](#). Front Med 8.
618. Posthuma-Trumpie GA, Korf J, van Amerongen A. 2008. [Lateral flow \(immuno\)assay: its strengths, weaknesses, opportunities and threats. A literature survey](#). Anal Bioanal Chem 393:569–582.
619. Mistry DA, Wang JY, Moeser M-E, Starkey T, Lee LYW. 2021. [A systematic review of the sensitivity and specificity of lateral flow devices in the detection of SARS-CoV-2](#). BMC Infect Dis 21:828.
620. Government sets out next steps for living with COVID. GOV.UK. <https://www.gov.uk/government/news/government-sets-out-next-steps-for-living-with-covid>. Retrieved 5 December 2022.
621. House TW. 2022. Fact Sheet: The Biden Administration to Begin Distributing At-Home, Rapid COVID-19 Tests to Americans for Free. The White House. <https://www.whitehouse.gov/briefing-room/statements-releases/2022/01/14/fact-sheet-the-biden-administration-to-begin-distributing-at-home-rapid-covid-19-tests-to-americans-for-free/>. Retrieved 5 December 2022.
622. Peeling RW, Olliaro PL, Boeras DI, Fongwen N. 2021. [Scaling up COVID-19 rapid antigen tests: promises and challenges](#). The Lancet Infectious Diseases 21:e290–e295.

623. Rai P, Kumar BK, Deekshit VK, Karunasagar I, Karunasagar I. 2021. [Detection technologies and recent developments in the diagnosis of COVID-19 infection](#). Appl Microbiol Biotechnol 105:441–455.
624. Lequin RM. 2005. [Enzyme Immunoassay \(EIA\)/Enzyme-Linked Immunosorbent Assay \(ELISA\)](#). Clinical Chemistry 51:2415–2418.
625. Schuurs AHW, Van Weemen BK. 1980. [Enzyme-Immunoassay: A Powerful Analytical Tool](#). Journal of Immunoassay 1:229–249.
626. Adams ER, Ainsworth M, Anand R, Andersson MI, Auckland K, Baillie JK, Barnes E, Beer S, Bell JI, Berry T, Bibi S, Carroll M, Chinnakannan SK, Clutterbuck E, Cornall RJ, Crook DW, de Silva T, Dejnirattisai W, Dingle KE, Dold C, Espinosa A, Eyre DW, Farmer H, Fernandez Mendoza M, Georgiou D, Hoosdally SJ, Hunter A, Jefferey K, Kelly DF, Klenerman P, Knight J, Knowles C, Kwok AJ, Leuschner U, Levin R, Liu C, López-Camacho C, Martinez J, Matthews PC, McGivern H, Mentzer AJ, Milton J, Mongkolsapaya J, Moore SC, Oliveira MS, Pereira F, Perez E, Peto T, Ploeg RJ, Pollard A, Prince T, Roberts DJ, Rudkin JK, Sanchez V, Screaton GR, Semple MG, Slon-Campos J, Skelly DT, Smith EN, Sobrinodiaz A, Staves J, Stuart DI, Supasa P, Surik T, Thraves H, Tsang P, Turtle L, Walker AS, Wang B, Washington C, Watkins N, Whitehouse J. 2020. [Antibody testing for COVID-19: A report from the National COVID Scientific Advisory Panel](#). Wellcome Open Res 5:139.
627. Adnan N, Khandker SS, Haq A, Chaity MA, Khalek A, Nazim AQ, Kaitsuka T, Tomizawa K, Mie M, Kobatake E, Ahmed S, Ali nor AA, Khondoker MU, Haque M, Jamiruddin MohdR. 2021. [Detection of SARS-CoV-2 by antigen ELISA test is highly swayed by viral load and sample storage condition](#). Expert Review of Anti-infective Therapy 20:473–481.
628. Winichakoon P, Chaiwarith R, Liwsrisakun C, Salee P, Goonna A, Limsukon A, Kaewpoowat Q. 2020. [Negative Nasopharyngeal and Oropharyngeal Swabs Do Not Rule Out COVID-19](#). Journal of Clinical Microbiology 58:e00297–20.
629. Sheridan C. 2020. [Coronavirus and the race to distribute reliable diagnostics](#). Nature Biotechnology 38:382–384.
630. CDC, CDC. 2022. Isolation. Centers for Disease Control and Prevention. <https://www.cdc.gov/coronavirus/2019-ncov/your-health/isolation.html>. Retrieved 5 December 2022.
631. Kumar P, Malik YS, Ganesh B, Rahangdale S, Saurabh S, Natesan S, Srivastava A, Sharun K, Yattoo MohdI, Tiwari R, Singh RK, Dhama K. 2020. [CRISPR-Cas System: An Approach With Potentials for COVID-19 Diagnosis and Therapeutics](#). Front Cell Infect Microbiol 10.
632. Service R. 2020. The standard coronavirus test, if available, works well—but can new diagnostics help in this pandemic? Science <https://doi.org/10.1126/science.abb8400>.
633. Tang Y-W, Schmitz JE, Persing DH, Stratton CW. 2020. [Laboratory Diagnosis of COVID-19: Current Issues and Challenges](#). Journal of Clinical Microbiology 58:e00512–20.

634. Sloss L. 2021. [The Plane Is Boarding, Where Are Your Test Results?](#) The New York Times.
635. Kresge N. 2022. A Stark Contrast Between the U.S. and Europe on Tests. Bloomberg. <https://www.bloomberg.com/news/newsletters/2022-01-07/a-stark-contrast-between-the-u-s-and-europe-on-tests>.
636. Vidarsson G, Dekkers G, Rispen T. 2014. [IgG Subclasses and Allotypes: From Structure to Effector Functions](#). Front Immunol 5.
637. 2001. The distribution and functions of immunoglobulin isotypes Immunobiology: The Immune System in Health and Disease. Garland Science. <https://www.ncbi.nlm.nih.gov/books/NBK27162>.
638. MO H, ZENG G, REN X, LI H, KE C, TAN Y, CAI C, LAI K, CHEN R, CHAN-YEUNG M, ZHONG N. 2006. [Longitudinal profile of antibodies against SARS-coronavirus in SARS patients and their clinical significance](#). Respirology 11:49–53.
639. Du Z, Zhu F, Guo F, Yang B, Wang T. 2020. [Detection of antibodies against SARS-CoV-2 in patients with COVID-19](#). Journal of Medical Virology 92:1735–1738.
640. Espejo AP, Akgun Y, Al Mana AF, Tjendra Y, Millan NC, Gomez-Fernandez C, Cray C. 2020. [Review of Current Advances in Serologic Testing for COVID-19](#). American Journal of Clinical Pathology 154:293–304.
641. Amanat F, Stadlbauer D, Strohmeier S, Nguyen THO, Chromikova V, McMahon M, Jiang K, Arunkumar GA, Jurczynski D, Polanco J, Bermudez-Gonzalez M, Kleiner G, Aydiello T, Miorin L, Fierer DS, Lugo LA, Kojic EM, Stoeve J, Liu STH, Cunningham-Rundles C, Felgner PL, Moran T, García-Sastre A, Caplivski D, Cheng AC, Kedzierska K, Vapalahti O, Hepojoki JM, Simon V, Krammer F. 2020. [A serological assay to detect SARS-CoV-2 seroconversion in humans](#). Nat Med 26:1033–1036.
642. Mehdi F, Chattopadhyay S, Thiruvengadam R, Yadav S, Kumar M, Sinha SK, Goswami S, Kshetrapal P, Wadhwa N, Chandramouli Natchu U, Sopory S, Koundinya Desiraju B, Pandey AK, Das A, Verma N, Sharma N, Sharma P, Bhartiya V, Gosain M, Lodha R, Lamminmäki U, Shrivastava T, Bhatnagar S, Batra G. 2021. [Development of a Fast SARS-CoV-2 IgG ELISA, Based on Receptor-Binding Domain, and Its Comparative Evaluation Using Temporally Segregated Samples From RT-PCR Positive Individuals](#). Front Microbiol 11.
643. Villafañe L, Vaulet LG, Viere FM, Klepp LI, Forrellad MA, Bigi MM, Romano MI, Magistrelli G, Fermepin MR, Bigi F. 2022. [Development and evaluation of a low cost IgG ELISA test based in RBD protein for COVID-19](#). Journal of Immunological Methods 500:113182.
644. Wang Y, Dzakah EE, Kang Y, Cai Y, Wu P, Tang B, Li R, He X. 2019. [A sensitive and rapid chemiluminescence immunoassay for point-of-care testing \(POCT\) of copeptin in serum based on high-affinity monoclonal](#)

[antibodies via cytokine-assisted immunization](#). IJN Volume 14:4293–4307.

645. Cinquanta L, Fontana DE, Bizzaro N. 2017. [Chemiluminescent immunoassay technology: what does it change in autoantibody detection?](#) Autoimmun Highlights 8.
646. Cai X, Chen J, Li H, Long Q, Deng H, Liu P, Fan K, Liao P, Liu B, Wu G, Chen Y, Li Z, Wang K, Zhang X, Tian W, Xiang J, Du H, Wang J, Hu Y, Tang N, Lin Y, Ren J, Huang L, Wei J, Gan C, Chen Y, Gao Q, Chen A, He C, Wang D-X, Hu P, Zhou F-C, Huang A, Wang D. 2020. [A Peptide-Based Magnetic Chemiluminescence Enzyme Immunoassay for Serological Diagnosis of Coronavirus Disease 2019](#). The Journal of Infectious Diseases 222:189–193.
647. Coste AT, Jaton K, Papadimitriou-Olivgeris M, Greub G, Croxatto A. 2021. [Comparison of SARS-CoV-2 serological tests with different antigen targets](#). Journal of Clinical Virology 134:104690.
648. Nicol T, Lefeuvre C, Serri O, Pivert A, Joubaud F, Dubée V, Kouatchet A, Ducancelle A, Lunel-Fabiani F, Le Guillou-Guillemette H. 2020. [Assessment of SARS-CoV-2 serological tests for the diagnosis of COVID-19 through the evaluation of three immunoassays: Two automated immunoassays \(Euroimmun and Abbott\) and one rapid lateral flow immunoassay \(NG Biotech\)](#). Journal of Clinical Virology 129:104511.
649. Charlton CL, Kanji JN, Johal K, Bailey A, Plitt SS, MacDonald C, Kunst A, Buss E, Burnes LE, Fonseca K, Berenger BM, Schnabl K, Hu J, Stokes W, Zelyas N, Tipples G. 2020. [Evaluation of Six Commercial Mid- to High-Volume Antibody and Six Point-of-Care Lateral Flow Assays for Detection of SARS-CoV-2 Antibodies](#). J Clin Microbiol 58.
650. Infantino M, Grossi V, Lari B, Bambi R, Perri A, Manneschi M, Terenzi G, Liotti I, Ciotta G, Taddei C, Benucci M, Casprini P, Veneziani F, Fabbri S, Pompetti A, Manfredi M. 2020. [Diagnostic accuracy of an automated chemiluminescent immunoassay for anti-SARS-CoV-2 IgM and IgG antibodies: an Italian experience](#). J Med Virol 92:1671–1675.
651. Sekirov I, Barakauskas VE, Simons J, Cook D, Bates B, Burns L, Masud S, Charles M, McLennan M, Mak A, Chahil N, Vih R, Hayden A, Goldfarb D, Levett PN, Krajden M, Morshed M. 2021. [SARS-CoV-2 serology: Validation of high-throughput chemiluminescent immunoassay \(CLIA\) platforms and a field study in British Columbia](#). Journal of Clinical Virology 142:104914.
652. Cellex. 2020. Cellex qSARS-CoV-2 IgG/IgM Rapid Test. <https://www.fda.gov/media/136625/download>.
653. Nam M, Seo JD, Moon H-W, Kim H, Hur M, Yun Y-M. 2021. [Evaluation of Humoral Immune Response after SARS-CoV-2 Vaccination Using Two Binding Antibody Assays and a Neutralizing Antibody Assay](#). Microbiol Spectr 9.
654. Muruato AE, Fontes-Garfias CR, Ren P, Garcia-Blanco MA, Menachery VD, Xie X, Shi P-Y. 2020. [A high-throughput neutralizing antibody assay for COVID-19 diagnosis and vaccine evaluation](#). Nat Commun 11.

655. Khoury DS, Cromer D, Reynaldi A, Schlub TE, Wheatley AK, Juno JA, Subbarao K, Kent SJ, Triccas JA, Davenport MP. 2021. [Neutralizing antibody levels are highly predictive of immune protection from symptomatic SARS-CoV-2 infection](#). Nat Med 27:1205–1211.
656. Addetia A, Crawford KHD, Dingens A, Zhu H, Roychoudhury P, Huang M-L, Jerome KR, Bloom JD, Greninger AL. 2020. [Neutralizing Antibodies Correlate with Protection from SARS-CoV-2 in Humans during a Fishery Vessel Outbreak with a High Attack Rate](#). J Clin Microbiol 58.
657. Cromer D, Steain M, Reynaldi A, Schlub TE, Wheatley AK, Juno JA, Kent SJ, Triccas JA, Khoury DS, Davenport MP. 2022. [Neutralising antibody titres as predictors of protection against SARS-CoV-2 variants and the impact of boosting: a meta-analysis](#). The Lancet Microbe 3:e52–e61.
658. Mravinacova S, Jönsson M, Christ W, Klingström J, Yousef J, Hellström C, Hedhammar M, Havervall S, Thålin C, Pin E, Tegel H, Nilsson P, Månberg A, Hober S. 2022. [A cell-free high throughput assay for assessment of SARS-CoV-2 neutralizing antibodies](#). New Biotechnology 66:46–52.
659. Liu W, Fontanet A, Zhang P, Zhan L, Xin Z, Baril L, Tang F, Lv H, Cao W. 2006. [Two-Year Prospective Study of the Humoral Immune Response of Patients with Severe Acute Respiratory Syndrome](#). The Journal of Infectious Diseases 193:792–795.
660. Callow KA, Parry HF, Sergeant M, Tyrrell DAJ. 2009. [The time course of the immune response to experimental coronavirus infection of man](#). Epidemiology and Infection 105:435–446.
661. Choe PG, Kim K-H, Kang CK, Suh HJ, Kang E, Lee SY, Kim NJ, Yi J, Park WB, Oh M. 2021. [Antibody Responses 8 Months after Asymptomatic or Mild SARS-CoV-2 Infection](#). Emerging Infectious Diseases 27.
662. Dan JM, Mateus J, Kato Y, Hastie KM, Yu ED, Faliti CE, Grifoni A, Ramirez SI, Haupt S, Frazier A, Nakao C, Rayaprolu V, Rawlings SA, Peters B, Krammer F, Simon V, Saphire EO, Smith DM, Weiskopf D, Sette A, Crotty S. 2021. [Immunological memory to SARS-CoV-2 assessed for up to 8 months after infection](#). Science eabf4063.
663. Hartley GE, Edwards ESJ, Aui PM, Varese N, Stojanovic S, McMahon J, Peleg AY, Boo I, Drummer HE, Hogarth PM, O'Hehir RE, van Zelm MC. 2020. [Rapid generation of durable B cell memory to SARS-CoV-2 spike and nucleocapsid proteins in COVID-19 and convalescence](#). Sci Immunol 5.
664. Sherina N, Piralla A, Du L, Wan H, Kumagai-Braesch M, Andréll J, Braesch-Andersen S, Cassaniti I, Percivalle E, Sarasini A, Bergami F, Di Martino R, Colaneri M, Vecchia M, Sambo M, Zuccaro V, Bruno R, Sachs M, Oggionni T, Meloni F, Abolhassani H, Bertoglio F, Schubert M, Byrne-Steele M, Han J, Hust M, Xue Y, Hammarström L, Baldanti F, Marcotte H, Pan-Hammarström Q. 2021. [Persistence of SARS-CoV-2-specific B and T cell responses in convalescent COVID-19 patients 6–8 months after the infection](#). Med 2:281–295.e4.

665. Gaebler C, Wang Z, Lorenzi JCC, Muecksch F, Finkin S, Tokuyama M, Cho A, Jankovic M, Schaefer-Babajew D, Oliveira TY, Cipolla M, Viant C, Barnes CO, Bram Y, Breton G, Hägglöf T, Mendoza P, Hurley A, Turroja M, Gordon K, Millard KG, Ramos V, Schmidt F, Weisblum Y, Jha D, Tankelevich M, Martinez-Delgado G, Yee J, Patel R, Dizon J, Unson-O'Brien C, Shimeliovich I, Robbiani DF, Zhao Z, Gazumyan A, Schwartz RE, Hatzioannou T, Bjorkman PJ, Mehandru S, Bieniasz PD, Caskey M, Nussenzweig MC. 2021. [Evolution of antibody immunity to SARS-CoV-2](#). *Nature* 591:639–644.
666. Wajnberg A, Amanat F, Firpo A, Altman DR, Bailey MJ, Mansour M, McMahon M, Meade P, Mendu DR, Muellers K, Stadlbauer D, Stone K, Strohmeier S, Simon V, Aberg J, Reich DL, Krammer F, Cordon-Cardo C. 2020. [Robust neutralizing antibodies to SARS-CoV-2 infection persist for months](#). *Science* 370:1227–1230.
667. Seow J, Graham C, Merrick B, Acors S, Pickering S, Steel KJA, Hemmings O, O'Byrne A, Kouphou N, Galao RP, Betancor G, Wilson HD, Signell AW, Winstone H, Kerridge C, Huettner I, Jimenez-Guardeño JM, Lista MJ, Temperton N, Snell LB, Bisnauthsing K, Moore A, Green A, Martinez L, Stokes B, Honey J, Izquierdo-Barras A, Arbane G, Patel A, Tan MKI, O'Connell L, O'Hara G, MacMahon E, Douthwaite S, Nebbia G, Batra R, Martinez-Nunez R, Shankar-Hari M, Edgeworth JD, Neil SJD, Malim MH, Doores KJ. 2020. [Longitudinal observation and decline of neutralizing antibody responses in the three months following SARS-CoV-2 infection in humans](#). *Nat Microbiol* 5:1598–1607.
668. Wheatley AK, Juno JA, Wang JJ, Selva KJ, Reynaldi A, Tan H-X, Lee WS, Wragg KM, Kelly HG, Esterbauer R, Davis SK, Kent HE, Mordant FL, Schlub TE, Gordon DL, Khoury DS, Subbarao K, Cromer D, Gordon TP, Chung AW, Davenport MP, Kent SJ. 2021. [Evolution of immune responses to SARS-CoV-2 in mild-moderate COVID-19](#). *Nat Commun* 12.
669. Garcia-Beltran WF, Lam EC, Astudillo MG, Yang D, Miller TE, Feldman J, Hauser BM, Caradonna TM, Clayton KL, Nitido AD, Murali MR, Alter G, Charles RC, Dighe A, Branda JA, Lennerz JK, Lingwood D, Schmidt AG, lafrate AJ, Balazs AB. 2021. [COVID-19-neutralizing antibodies predict disease severity and survival](#). *Cell* 184:476–488.e11.
670. Dowell AC, Butler MS, Jinks E, Tut G, Lancaster T, Sylla P, Begum J, Bruton R, Pearce H, Verma K, Logan N, Tyson G, Spalkova E, Margielewska-Davies S, Taylor GS, Syrimi E, Baawuah F, Beckmann J, Okike IO, Ahmad S, Garstang J, Brent AJ, Brent B, Ireland G, Aiano F, Amin-Chowdhury Z, Jones S, Borrow R, Linley E, Wright J, Azad R, Waiblinger D, Davis C, Thomson EC, Palmarini M, Willett BJ, Barclay WS, Poh J, Amirthalingam G, Brown KE, Ramsay ME, Zuo J, Moss P, Ladhani S. 2021. [Children develop robust and sustained cross-reactive spike-specific immune responses to SARS-CoV-2 infection](#). *Nat Immunol* 23:40–49.
671. Gil-Manso S, Miguens Blanco I, Motyka B, Halpin A, López-Esteban R, Pérez-Fernández VA, Carbonell D, López-Fernández LA, West L, Correa-Rocha R, Pion M. 2021. [ABO blood group is involved in the quality of the specific immune response anti-SARS-CoV-2](#). *Virulence* 13:30–45.

672. Kaneko N, Kuo H-H, Boucau J, Farmer JR, Allard-Chamard H, Mahajan VS, Piechocka-Trocha A, Lefteri K, Osborn M, Bals J, Bartsch YC, Bonheur N, Caradonna TM, Chevalier J, Chowdhury F, Diefenbach TJ, Einkauf K, Fallon J, Feldman J, Finn KK, Garcia-Broncano P, Hartana CA, Hauser BM, Jiang C, Kaplonek P, Karpell M, Koscher EC, Lian X, Liu H, Liu J, Ly NL, Michell AR, Rassadkina Y, Seiger K, Sessa L, Shin S, Singh N, Sun W, Sun X, Ticheli HJ, Waring MT, Zhu AL, Alter G, Li JZ, Lingwood D, Schmidt AG, Lichterfeld M, Walker BD, Yu XG, Padera RF, Pillai S. 2020. [Loss of Bcl-6-Expressing T Follicular Helper Cells and Germinal Centers in COVID-19](#). *Cell* 183:143–157.e13.
673. Zuo J, Dowell A, Pearce H, Verma K, Long H, Begum J, Aiano F, Amin-Chowdhury Z, Hallis B, Stapley L, Borrow R, Linley E, Ahmad S, Parker B, Horsley A, Amirthalingam G, Brown K, Ramsay M, Ladhani S, Moss P. 2020. Robust SARS-CoV-2-specific T-cell immunity is maintained at 6 months following primary infection. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.11.01.362319>.
674. Breton G, Mendoza P, Hagglof T, Oliveira TY, Schaefer-Babajew D, Gaebler C, Turroja M, Hurley A, Caskey M, Nussenzweig MC. 2020. Persistent Cellular Immunity to SARS-CoV-2 Infection. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.12.08.416636>.
675. Grifoni A, Weiskopf D, Ramirez SI, Mateus J, Dan JM, Moderbacher CR, Rawlings SA, Sutherland A, Premkumar L, Jadi RS, Marrama D, de Silva AM, Frazier A, Carlin AF, Greenbaum JA, Peters B, Krammer F, Smith DM, Crotty S, Sette A. 2020. [Targets of T Cell Responses to SARS-CoV-2 Coronavirus in Humans with COVID-19 Disease and Unexposed Individuals](#). *Cell* 181:1489–1501.e15.
676. Sheridan C. 2021. [COVID-19 testing turns to T cells](#). *Nat Biotechnol* 39:533–534.
677. Commissioner O of the. 2021. Coronavirus (COVID-19) Update: FDA Authorizes Adaptive Biotechnologies T-Detect COVID Test. FDA. <https://www.fda.gov/news-events/press-announcements/coronavirus-covid-19-update-fda-authorizes-adaptive-biotechnologies-t-detect-covid-test>. Retrieved 5 December 2022.
678. Mina MJ, Parker R, Larremore DB. 2020. [Rethinking Covid-19 Test Sensitivity — A Strategy for Containment](#). *N Engl J Med* 383:e120.
679. Peeling RW, Olliaro PL. 2020. [The time to do serosurveys for COVID-19 is now](#). *The Lancet Respiratory Medicine* 8:836–838.
680. Schuler CF, Gherasim C, O'Shea K, Manthei DM, Chen J, Giacherio D, Troost JP, Baldwin JL, Baker JR. 2021. [Accurate point-of-care serology tests for COVID-19](#). *PLoS ONE* 16:e0248729.
681. Chen X, Chen Z, Azman AS, Sun R, Lu W, Zheng N, Zhou J, Wu Q, Deng X, Zhao Z, Chen X, Ge S, Yang J, Leung DT, Yu H. 2021. [Neutralizing Antibodies Against Severe Acute Respiratory Syndrome Coronavirus 2 \(SARS-CoV-2\) Variants Induced by Natural Infection or Vaccination: A Systematic Review and Pooled Analysis](#). *Clinical Infectious Diseases* 74:734–742.

682. Murhekar MV, Clapham H. 2021. [COVID-19 serosurveys for public health decision making](#). The Lancet Global Health 9:e559–e560.
683. Ghaffari A, Meurant R, Ardakani A. 2020. [COVID-19 Serological Tests: How Well Do They Actually Perform?](#) Diagnostics 10:453.
684. Bond K, Williams E, Howden BP, Williamson DA. 2020. [Serological tests for COVID-19](#). Medical Journal of Australia 213:397.
685. Rodriguez-Moncayo R, Cedillo-Alcantar DF, Guevara-Pantoja PE, Chavez-Pineda OG, Hernandez-Ortiz JA, Amador-Hernandez JU, Rojas-Velasco G, Sanchez-Muñoz F, Manzur-Sandoval D, Patino-Lopez LD, May-Arrioja DA, Posadas-Sanchez R, Vargas-Alarcon G, Garcia-Cordero JL. 2021. [A high-throughput multiplexed microfluidic device for COVID-19 serology assays](#). Lab Chip 21:93–104.
686. Huang AT, Garcia-Carreras B, Hitchings MDT, Yang B, Katzelnick L, Rattigan SM, Borgert B, Moreno C, Solomon BD, Rodriguez-Barraquer I, Lessler J, Salje H, Burke DS, Wesolowski A, Cummings DAT. 2020. A systematic review of antibody mediated immunity to coronaviruses: antibody kinetics, correlates of protection, and association of antibody responses with severity of disease. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.04.14.20065771>.
687. Spellberg B, Nielsen TB, Casadevall A. 2021. [Antibodies, Immunity, and COVID-19](#). JAMA Intern Med 181:460.
688. Goldberg Y, Mandel M, Bar-On YM, Bodenheimer O, Freedman L, Ash N, Alroy-Preis S, Huppert A, Milo R. 2021. [Protection and waning of natural and hybrid COVID-19 immunity](#). Cold Spring Harbor Laboratory.
689. Tostmann A, Bradley J, Bousema T, Yiek W-K, Holwerda M, Bleeker-Rovers C, ten Oever J, Meijer C, Rahamat-Langendoen J, Hopman J, van der Geest-Blankert N, Wertheim H. 2020. [Strong associations and moderate predictive value of early symptoms for SARS-CoV-2 test positivity among healthcare workers, the Netherlands, March 2020](#). Eurosurveillance 25.
690. Ren X, Liu Y, Chen H, Liu W, Guo Z, Zhang Y, Chen C, Zhou J, Xiao Q, Jiang G, Shan H. 2020. [Application and optimization of RT-PCR in diagnosis of SARS-CoV-2 infection](#). Cold Spring Harbor Laboratory.
691. Ai T, Yang Z, Hou H, Zhan C, Chen C, Lv W, Tao Q, Sun Z, Xia L. 2020. [Correlation of Chest CT and RT-PCR Testing for Coronavirus Disease 2019 \(COVID-19\) in China: A Report of 1014 Cases](#). Radiology 296:E32–E40.
692. Bai HX, Hsieh B, Xiong Z, Halsey K, Choi JW, Tran TML, Pan I, Shi L-B, Wang D-C, Mei J, Jiang X-L, Zeng Q-H, Egglin TK, Hu P-F, Agarwal S, Xie F-F, Li S, Healey T, Atalay MK, Liao W-H. 2020. [Performance of Radiologists in Differentiating COVID-19 from Non-COVID-19 Viral Pneumonia at Chest CT](#). Radiology 296:E46–E54.
693. Apostolopoulos ID, Mpesiana TA. 2020. [Covid-19: automatic detection from X-ray images utilizing transfer learning with convolutional neural](#)

[networks](#). Physical and Engineering Sciences in Medicine 43:635–640.

694. Roberts M, Driggs D, Thorpe M, Gilbey J, Yeung M, Ursprung S, Aviles-Rivero AI, Etmann C, McCague C, Beer L, Weir-McCall JR, Teng Z, Gkrania-Klotsas E, Ruggiero A, Korhonen A, Jefferson E, Ako E, Langs G, Gozaliasl G, Yang G, Prosch H, Preller J, Stanczuk J, Tang J, Hofmanninger J, Babar J, Sánchez LE, Thillai M, Gonzalez PM, Teare P, Zhu X, Patel M, Cafolla C, Azadbakht H, Jacob J, Lowe J, Zhang K, Bradley K, Wassin M, Holzer M, Ji K, Ortet MD, Ai T, Walton N, Lio P, Stranks S, Shadbahr T, Lin W, Zha Y, Niu Z, Rudd JHF, Sala E, Schönlieb C-B. 2021. [Common pitfalls and recommendations for using machine learning to detect and prognosticate for COVID-19 using chest radiographs and CT scans](#). Nat Mach Intell 3:199–217.
695. Ibrahim NK. 2020. [Epidemiologic surveillance for controlling Covid-19 pandemic: types, challenges and implications](#). Journal of Infection and Public Health 13:1630–1638.
696. Farkas K, Hillary LS, Malham SK, McDonald JE, Jones DL. 2020. [Wastewater and public health: the potential of wastewater surveillance for monitoring COVID-19](#). Current Opinion in Environmental Science & Health 17:14–20.
697. CDC. 2020. Healthcare Workers. Centers for Disease Control and Prevention. <https://www.cdc.gov/coronavirus/2019-ncov/hcp/testing-overview.html>. Retrieved 8 February 2021.
698. 2020. NY Forward: a guide to reopening New York & building back better. <https://www.governor.ny.gov/sites/governor.ny.gov/files/atoms/files/NYForwardReopeningGuide.pdf>.
699. Kyosei Y, Yamura S, Namba M, Yoshimura T, Watabe S, Ito E. 2021. [Antigen tests for COVID-19](#). BIOPHYSICS 18:28–39.
700. Waters A. 2022. [Covid-19: Show us evidence for lifting restrictions, doctors tell Johnson](#). BMJ o383.
701. Misra M, Joshi H, Sarwal R, Rao KD. 2022. [Exit strategies from lockdowns due to COVID-19: a scoping review](#). BMC Public Health 22.
702. Jefferson T, Spencer EA, Brassey J, Heneghan C. 2020. [Viral Cultures for Coronavirus Disease 2019 Infectivity Assessment: A Systematic Review](#). Clinical Infectious Diseases 73:e3884–e3899.
703. Cevik M, Tate M, Lloyd O, Maraolo AE, Schafers J, Ho A. 2021. [SARS-CoV-2, SARS-CoV, and MERS-CoV viral load dynamics, duration of viral shedding, and infectiousness: a systematic review and meta-analysis](#). The Lancet Microbe 2:e13–e22.
704. Agarwal V, Venkatakrishnan AJ, Puranik A, Kirkup C, Lopez-Marquez A, Challener DW, Theel ES, O'Horo JC, Binnicker MJ, Kremers WK, Faubion WA Jr., Badley AD, Williams AW, Gores GJ, Halamka JD, Morice WG II, Soundararajan V. 2020. [Long-term SARS-CoV-2 RNA shedding and its temporal association to IgG seropositivity](#). Cell Death Discov 6.

705. Stankiewicz Karita HC, Dong TQ, Johnston C, Neuzil KM, Paasche-Orlow MK, Kissinger PJ, Bershteyn A, Thorpe LE, Deming M, Kottkamp A, Laufer M, Landovitz RJ, Luk A, Hoffman R, Roychoudhury P, Magaret CA, Greninger AL, Huang M-L, Jerome KR, Wener M, Celum C, Chu HY, Baeten JM, Wald A, Barnabas RV, Brown ER. 2022. [Trajectory of Viral RNA Load Among Persons With Incident SARS-CoV-2 G614 Infection \(Wuhan Strain\) in Association With COVID-19 Symptom Onset and Severity](#). JAMA Netw Open 5:e2142796.
706. Armstrong S. 2020. [Covid-19: Tests on students are highly inaccurate, early findings show](#). BMJ m4941.
707. Kmietowicz Z. 2021. [Covid-19: Controversial rapid test policy divides doctors and scientists](#). BMJ n81.
708. Deerain J, Druce J, Tran T, Batty M, Yoga Y, Fennell M, Dwyer DE, Kok J, Williamson DA. 2022. [Assessment of the Analytical Sensitivity of 10 Lateral Flow Devices against the SARS-CoV-2 Omicron Variant](#). J Clin Microbiol 60.
709. Mina MJ, Peto TE, García-Fiñana M, Semple MG, Buchan IE. 2021. [Clarifying the evidence on SARS-CoV-2 antigen rapid tests in public health responses to COVID-19](#). The Lancet 397:1425–1427.
710. Larremore DB, Wilder B, Lester E, Shehata S, Burke JM, Hay JA, Tambe M, Mina MJ, Parker R. 2021. [Test sensitivity is secondary to frequency and turnaround time for COVID-19 screening](#). Sci Adv 7.
711. Sun LH, Achenbach J. 2020. [CDC chief says coronavirus cases may be 10 times higher than reported](#). Washington Post.
712. Liverpool coronavirus (COVID-19) community testing pilot: full evaluation report summary. GOVUK. <https://www.gov.uk/government/publications/liverpool-coronavirus-covid-19-community-testing-pilot-full-evaluation-report-summary/liverpool-coronavirus-covid-19-community-testing-pilot-full-evaluation-report-summary>. Retrieved 5 December 2022.
713. Iacobucci G. 2021. [Covid-19: Government rolls out twice weekly rapid testing to all in England](#). BMJ n902.
714. Jae Moon M, Suzuki K, Park TI, Sakuwa K. 2021. [A comparative study of COVID-19 responses in South Korea and Japan: political nexus triad and policy responses](#). International Review of Administrative Sciences 87:651–671.
715. Bennett M. 2021. [All things equal? Heterogeneity in policy effectiveness against COVID-19 spread in chile](#). World Development 137:105208.
716. 2022. UK ending Covid testing 'very worrying' as WHO chief warns pandemic 'isn't over'. The Independent. <https://www.independent.co.uk/news/health/who-covid-testing-anil-soni-b2032884.html>. Retrieved 5 December 2022.

717. Clarke J, Beaney T, Majeed A. 2022. [UK scales back routine covid-19 surveillance](#). BMJ o562.
718. 2022. Local groups continue push for COVID testing and vaccinations as larger state-run sites plan to close. KUSAcom. <https://www.9news.com/article/news/health/coronavirus/local-groups-continue-push-for-covid-testing-and-vaccinations/73-bbcd8384-d96a-425e-aaeb-16ac9f36e581>. Retrieved 5 December 2022.
719. Utah will stop daily COVID case counts, close test sites in wind-down, Gov. Cox announces. The Salt Lake Tribune. <https://www.sltrib.com/news/2022/02/18/utah-will-stop-daily/>. Retrieved 5 December 2022.
720. Killingley B, Mann AJ, Kalinova M, Boyers A, Goonawardane N, Zhou J, Lindsell K, Hare SS, Brown J, Frise R, Smith E, Hopkins C, Noulon N, Löndt B, Wilkinson T, Harden S, McShane H, Baillet M, Gilbert A, Jacobs M, Charman C, Mande P, Nguyen-Van-Tam JS, Semple MG, Read RC, Ferguson NM, Openshaw PJ, Rapeport G, Barclay WS, Catchpole AP, Chiu C. 2022. [Safety, tolerability and viral kinetics during SARS-CoV-2 human challenge in young adults](#). Nat Med 28:1031–1041.
721. Lordan R, Prior S, Hennessy E, Naik A, Ghosh S, Paschos GK, Skarke C, Barekat K, Hollingsworth T, Juska S, Mazaleuskaya LL, Teegarden S, Glascock AL, Anderson S, Meng H, Tang S-Y, Weljie A, Bottalico L, Ricciotti E, Cherfane P, Mrcela A, Grant G, Poole K, Mayer N, Waring M, Adang L, Becker J, Fries S, FitzGerald GA, Grosser T. 2021. [Considerations for the Safe Operation of Schools During the Coronavirus Pandemic](#). Front Public Health 9.
722. Theuring S, Thielecke M, van Loon W, Hommes F, Hülso C, von der Haar A, Körner J, Schmidt M, Böhringer F, Mall MA, Rosen A, von Kalle C, Kirchberger V, Kurth T, Seybold J, Mockenhaupt FP. 2021. [SARS-CoV-2 infection and transmission in school settings during the second COVID-19 wave: a cross-sectional study, Berlin, Germany, November 2020](#). Eurosurveillance 26.
723. Miller E. 2022. End to quarantines, contact tracing among changes in new Oregon guidance for schools, starting March 12. OPB. <https://www.opb.org/article/2022/03/02/oregon-schools-guidance-mask-mandate-end/>.
724. 2022. Palm Beach County public schools to stop COVID-19 contact tracing. WPTV News Channel 5 West Palm. <https://www.wptv.com/news/education/palm-beach-county-public-schools-to-stop-covid-19-contact-tracing>. Retrieved 5 December 2022.
725. Hassan A. 2022. [Coronavirus Pandemic: Covid News: C.D.C. Drops Contact Tracing Recommendation](#). The New York Times.
726. Children and COVID-19: State-Level Data Report. American Academy of Pediatrics. <http://www.aap.org/en/pages/2019-novel-coronavirus-covid-19-infections/children-and-covid-19-state-level-data-report/>. Retrieved 11 April 2022.

727. Schreiber M. 2022. [A fifth of all US child Covid deaths occurred during Omicron surge](#). The Guardian.
728. Unwin HJT, Hillis S, Cluver L, Flaxman S, Goldman PS, Butchart A, Bachman G, Rawlings L, Donnelly CA, Ratmann O, Green P, Nelson CA, Blenkinsop A, Bhatt S, Desmond C, Villaveces A, Sherr L. 2022. [Global, regional, and national minimum estimates of children affected by COVID-19-associated orphanhood and caregiver death, by age and family circumstance up to Oct 31, 2021: an updated modelling study](#). The Lancet Child & Adolescent Health 6:249–259.
729. Reed T, Waites W, Manheim D, de Walque D, Vallini C, Gatti R, Hallett TB. 2021. [Five Ways that COVID-19 Diagnostics Can Save Lives: Prioritizing Uses of Tests to Maximize Cost-Effectiveness](#). Brief. World Bank, Washington, DC.
730. Hua J, Wang G, Huang M, Hua S, Yang S. 2020. [A Visual Approach for the SARS \(Severe Acute Respiratory Syndrome\) Outbreak Data Analysis](#). IJERPH 17:3973.
731. Center for Systems Science and Engineering at Johns Hopkins University. COVID-19 Data Repository. GitHub. https://github.com/CSSEGISandData/COVID-19/tree/master/csse_covid_19_data/csse_covid_19_time_series. Retrieved 19 June 2020.
732. Dong E, Du H, Gardner L. 2020. [An interactive web-based dashboard to track COVID-19 in real time](#). The Lancet Infectious Diseases 20:533–534.
733. Severe Acute Respiratory Syndrome (SARS). <https://www.who.int/health-topics/severe-acute-respiratory-syndrome>. Retrieved 5 December 2022.
734. kp D. 2021. sars-2003-outbreak-data-with-web-scrapping-munging-and-cleaning-code. <https://github.com/imdevskp/sars-2003-outbreak-data-webscraping-code>. Retrieved 5 December 2022.
735. Tang JW, Marr LC, Li Y, Dancer SJ. 2021. [Covid-19 has redefined airborne transmission](#). BMJ n913.
736. Zaki AM, van Boheemen S, Bestebroer TM, Osterhaus ADME, Fouchier RAM. 2012. [Isolation of a Novel Coronavirus from a Man with Pneumonia in Saudi Arabia](#). New England Journal of Medicine 367:1814–1820.
737. Pushpakom S, Iorio F, Eyers PA, Escott KJ, Hopper S, Wells A, Doig A, Guilleams T, Latimer J, McNamee C, Norris A, Sanseau P, Cavalla D, Pirmohamed M. 2018. [Drug repurposing: progress, challenges and recommendations](#). Nat Rev Drug Discov 18:41–58.
738. Mohs RC, Greig NH. 2017. [Drug discovery and development: Role of basic biological research](#). Alzheimer's & Dementia: Translational Research & Clinical Interventions 3:651–657.

739. DeVito N, Inglesby P. 2020. Evidence-Based Medicine Data Lab COVID-19 TrialsTracker. GitHub.
https://github.com/ebmdatalab/covid_trials_tracker-covid.
740. Siemieniuk RA, Bartoszko JJ, Zeraatkar D, Kum E, Qasim A, Díaz Martínez JP, Izcovich A, Rochwerg B, Lamontagne F, Han MA, Agarwal A, Agoritsas T, Azab M, Bravo G, Chu DK, Couban R, Cusano E, Devji T, Escamilla Z, Foroutan F, Gao Y, Ge L, Ghadimi M, Heels-Ansdell D, Honarmand K, Hou L, Ibrahim S, Khamis A, Lam B, Mansilla C, Loeb M, Miroshnychenko A, Marcucci M, McLeod SL, Motaghi S, Murthy S, Mustafa RA, Pardo-Hernandez H, Rada G, Rizwan Y, Saadat P, Switzer C, Thabane L, Tomlinson G, Vandvik PO, Vernooij RW, Viteri-García A, Wang Y, Yao L, Zhao Y, Guyatt GH, Brignardello-Petersen R. 2020. [Drug treatments for covid-19: living systematic review and network meta-analysis](#). BMJ m2980.
741. Elezkurtaj S, Greuel S, Ihlow J, Michaelis E, Bischoff P, Kunze CA, Sinn BV, Gerhold M, Hauptmann K, Ingold-Heppner B, Miller F, Herbst H, Corman VM, Martin H, Radbruch H, Heppner FL, Horst D. 2020. Causes of Death and Comorbidities in Patients with COVID-19. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.06.15.20131540>.
742. Zhang B, Zhou X, Qiu Y, Song Y, Feng F, Feng J, Song Q, Jia Q, Wang J. 2020. [Clinical characteristics of 82 cases of death from COVID-19](#). PLOS ONE 15:e0235458.
743. Shi Y, Wang Y, Shao C, Huang J, Gan J, Huang X, Bucci E, Piacentini M, Ippolito G, Melino G. 2020. [COVID-19 infection: the perspectives on immune responses](#). Cell Death Differ 27:1451–1454.
744. Fajgenbaum DC, June CH. 2020. [Cytokine Storm](#). New England Journal of Medicine 383:2255–2273.
745. Nicholls JM, Poon LL, Lee KC, Ng WF, Lai ST, Leung CY, Chu CM, Hui PK, Mak KL, Lim W, Yan KW, Chan KH, Tsang NC, Guan Y, Yuen KY, Malik Peiris J. 2003. [Lung pathology of fatal severe acute respiratory syndrome](#). The Lancet 361:1773–1778.
746. Gomersall CD, Kargel MJ, Lapinsky SE. 2004. [Pro/con clinical debate: Steroids are a key component in the treatment of SARS](#). Critical Care 8:105.
747. Prager R, Pratte MT, Unni RR, Bala S, Ng Fat Hing N, Wu K, McGrath TA, Thomas A, Thoma B, Kyeremanteng K. 2021. Content Analysis and Characterization of Medical Tweets During the Early Covid-19 Pandemic. Cureus <https://doi.org/10.7759/cureus.13594>.
748. Parkway NPSM, Durham S200. 2020. Small Molecules vs Biologics | Drug Development Differences. PK / PD and Clinical Pharmacology Consultants. <https://www.nuventra.com/resources/blog/small-molecules-versus-biologics/>. Retrieved 8 February 2021.
749. Drews J. 2000. [Drug Discovery: A Historical Perspective](#). Science 287:1960–1964.

750. Thompson AE, Ranard BL, Wei Y, Jelic S. 2020. [Prone Positioning in Awake, Nonintubated Patients With COVID-19 Hypoxemic Respiratory Failure](#). JAMA Intern Med 180:1537.
751. Stockman LJ, Bellamy R, Garner P. 2006. [SARS: Systematic Review of Treatment Effects](#). PLoS Med 3:e343.
752. Fujii T, Iwamoto A, Nakamura T, Iwamoto A. 2004. [Current concepts in SARS treatment](#). Journal of Infection and Chemotherapy 10:1–7.
753. Stern A, Skalsky K, Avni T, Carrara E, Leibovici L, Paul M. 2017. [Corticosteroids for pneumonia](#). Cochrane Database of Systematic Reviews 2017.
754. Chen Y, Li K, Pu H, Wu T. 2011. Corticosteroids for pneumonia. Cochrane Database of Systematic Reviews <https://doi.org/10.1002/14651858.cd007720.pub2>.
755. Sibila O, Agusti C, Torres A. 2008. [Corticosteroids in severe pneumonia](#). European Respiratory Journal 32:259–264.
756. Mikami K, Suzuki M, Kitagawa H, Kawakami M, Hirota N, Yamaguchi H, Narumoto O, Kichikawa Y, Kawai M, Tashimo H, Arai H, Horiuchi T, Sakamoto Y. 2007. [Efficacy of Corticosteroids in the Treatment of Community-Acquired Pneumonia Requiring Hospitalization](#). Lung 185:249–255.
757. Nie W, Zhang Y, Cheng J, Xiu Q. 2012. [Corticosteroids in the Treatment of Community-Acquired Pneumonia in Adults: A Meta-Analysis](#). PLoS ONE 7:e47926.
758. Fernández-Serrano S, Dorca J, Garcia-Vidal C, Fernández-Sabé N, Carratalà J, Fernández-Agüera A, Corominas M, Padrones S, Gudiol F, Manresa F. 2011. [Effect of corticosteroids on the clinical course of community-acquired pneumonia: a randomized controlled trial](#). Critical Care 15:R96.
759. Villar J, Ferrando C, Martínez D, Ambrós A, Muñoz T, Soler JA, Aguilar G, Alba F, González-Higueras E, Conesa LA, Martín-Rodríguez C, Díaz-Domínguez FJ, Serna-Grande P, Rivas R, Ferreres J, Belda J, Capilla L, Tallet A, Añón JM, Fernández RL, González-Martín JM, Aguilar G, Alba F, Álvarez J, Ambrós A, Añón JM, Asensio MJ, Belda J, Blanco J, Blasco M, Cachafeiro L, del Campo R, Capilla L, Carbonell JA, Carbonell N, Cariñena A, Carriedo D, Chico M, Conesa LA, Corpas R, Cuervo J, Díaz-Domínguez FJ, Domínguez-Antelo C, Fernández L, Fernández RL, Ferrando C, Ferreres J, Gamboa E, González-Higueras E, González-Luengo RI, González-Martín JM, Martínez D, Martín-Rodríguez C, Muñoz T, Ortiz Díaz-Miguel R, Pérez-González R, Prieto AM, Prieto I, Rivas R, Rojas-Viguera L, Romera MA, Sánchez-Ballesteros J, Segura JM, Serna-Grande P, Serrano A, Solano R, Soler JA, Soro M, Tallet A, Villar J. 2020. [Dexamethasone treatment for the acute respiratory distress syndrome: a multicentre, randomised controlled trial](#). The Lancet Respiratory Medicine 8:267–276.
760. Reddy K, O'Kane C, McAuley D. 2020. [Corticosteroids in acute respiratory distress syndrome: a step forward, but more evidence is](#)

[needed](#). The Lancet Respiratory Medicine 8:220–222.

761. Calfee CS, Matthay MA. 2007. [Nonventilatory Treatments for Acute Lung Injury and ARDS](#). Chest 131:913–920.
762. Meduri GU, Marik PE, Pastores SM, Annane D. 2007. [Corticosteroids in ARDS](#). Chest 132:1093–1094.
763. 2006. [Efficacy and Safety of Corticosteroids for Persistent Acute Respiratory Distress Syndrome](#). N Engl J Med 354:1671–1684.
764. Peter JV, John P, Graham PL, Moran JL, George IA, Bersten A. 2008. [Corticosteroids in the prevention and treatment of acute respiratory distress syndrome \(ARDS\) in adults: meta-analysis](#). BMJ 336:1006–1009.
765. Yu WC. 2004. [Antiviral agents and corticosteroids in the treatment of severe acute respiratory syndrome \(SARS\)](#). Thorax 59:643–645.
766. Yin-Chun Yam L, Chun-Wing Lau A, Yuk-Lin Lai F, Shung E, Chan J, Wong V. 2007. [Corticosteroid treatment of severe acute respiratory syndrome in Hong Kong](#). Journal of Infection 54:28–39.
767. Li H, Chen C, Hu F, Wang J, Zhao Q, Gale RP, Liang Y. 2020. [Impact of corticosteroid therapy on outcomes of persons with SARS-CoV-2, SARS-CoV, or MERS-CoV infection: a systematic review and meta-analysis](#). Leukemia 34:1503–1511.
768. Wenzel RP, Edmond MB. 2003. [Managing SARS amidst Uncertainty](#). N Engl J Med 348:1947–1948.
769. Khan MOF, Lee HJ. 2008. [Synthesis and Pharmacology of Anti-Inflammatory Steroidal Antedrugs](#). Chemical Reviews 108:5131–5145.
770. Drug vignettes: Dexamethasone. The Centre for Evidence-Based Medicine. <https://www.cebm.net/covid-19/dexamethasone/>. Retrieved 8 February 2021.
771. Zabirowicz ES, Gan TJ. 2019. [Pharmacology of Postoperative Nausea and Vomiting](#), p. 671–692. *In*. Elsevier BV.
772. Zhou W, Liu Y, Tian D, Wang C, Wang S, Cheng J, Hu M, Fang M, Gao Y. 2020. [Potential benefits of precise corticosteroids therapy for severe 2019-nCoV pneumonia](#). Signal Transduction and Targeted Therapy 5:18.
773. Arth GE, Johnston DBR, Fried J, Spooncer WW, Hoff DR, Sarett LH. 2002. [16-METHYLATED STEROIDS. I. 16 \$\alpha\$ -METHYLATED ANALOGS OF CORTISONE, A NEW GROUP OF ANTI-INFLAMMATORY STEROIDS](#). Journal of the American Chemical Society 80:3160–3161.
774. Cohen A. 1960. [Treatment of Rheumatoid Arthritis with Dexamethasone](#). JAMA 174:831.
775. 2007. Dexamethasone. DailyMed. <https://dailymed.nlm.nih.gov/dailymed/drugInfo.cfm?setid=537b424a-3e07-4c81-978c-1ad99014032a>.

776. Orlicka K, Barnes E, Culver EL. 2013. [Prevention of infection caused by immunosuppressive drugs in gastroenterology](#). Therapeutic Advances in Chronic Disease 4:167–185.
777. Russell CD, Millar JE, Baillie JK. 2020. [Clinical evidence does not support corticosteroid treatment for 2019-nCoV lung injury](#). The Lancet 395:473–475.
778. Shang L, Zhao J, Hu Y, Du R, Cao B. 2020. [On the use of corticosteroids for 2019-nCoV pneumonia](#). The Lancet 395:683–684.
779. Ritchie AI, Singanayagam A. 2020. [Immunosuppression for hyperinflammation in COVID-19: a double-edged sword?](#) The Lancet 395:11111.
780. Horby P, Lim WS, Emberson J, Mafham M, Bell J, Linsell L, Staplin N, Brightling C, Ustianowski A, Elmahi E, Prudon B, Green C, Felton T, Chadwick D, Rege K, Fegan C, Chappell LC, Faust SN, Jaki T, Jeffery K, Montgomery A, Rowan K, Juszczak E, Baillie JK, Haynes R, Landray MJ, RECOVERY Collaborative Group. 2020. Effect of Dexamethasone in Hospitalized Patients with COVID-19 – Preliminary Report. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.06.22.20137273>.
781. The RECOVERY Collaborative Group. 2020. [Dexamethasone in Hospitalized Patients with Covid-19 — Preliminary Report](#). New England Journal of Medicine NEJMoa2021436.
782. Pasin L, Navalesi P, Zangrillo A, Kuzovlev A, Likhvantsev V, Hajjar LA, Fresilli S, Lacerda MVG, Landoni G. 2021. [Corticosteroids for Patients With Coronavirus Disease 2019 \(COVID-19\) With Different Disease Severity: A Meta-Analysis of Randomized Clinical Trials](#). Journal of Cardiothoracic and Vascular Anesthesia 35:578–584.
783. Lee DW, Gardner R, Porter DL, Louis CU, Ahmed N, Jensen M, Grupp SA, Mackall CL. 2014. [Current concepts in the diagnosis and management of cytokine release syndrome](#). Blood 124:188–195.
784. Prescott HC, Rice TW. 2020. [Corticosteroids in COVID-19 ARDS](#). JAMA 324:1292.
785. Noreen S, Maqbool I, Madni A. 2021. [Dexamethasone: Therapeutic potential, risks, and future projection during COVID-19 pandemic](#). European Journal of Pharmacology 894:173854.
786. Mahase E. 2020. [Covid-19: Demand for dexamethasone surges as RECOVERY trial publishes preprint](#). BMJ m2512.
787. Dimmock NJ, Easton AJ, Leppard KN. 2007. Introduction to modern virology 6th ed. Blackwell Pub, Malden, MA.
788. CORONA Data Viewer. Castleman Disease Collaborative Network. <https://cdcn.org/corona-data-viewer/>. Retrieved 12 August 2021.
789. 2015. Coronaviruses Methods in Molecular Biology. Springer New York. <https://doi.org/ggqfqx>.

790. Lung J, Lin Y, Yang Y, Chou Y, Shu L, Cheng Y, Liu HT, Wu C. 2020. [The potential chemical structure of anti-SARS-CoV-2 RNA-dependent RNA polymerase](#). Journal of Medical Virology 92:693–697.
791. Totura AL, Bavari S. 2019. [Broad-spectrum coronavirus antiviral drug discovery](#). Expert Opinion on Drug Discovery 14:397–412.
792. Tong S, Su Y, Yu Y, Wu C, Chen J, Wang S, Jiang J. 2020. [Ribavirin therapy for severe COVID-19: a retrospective cohort study](#). International Journal of Antimicrobial Agents 56:106114.
793. Seley-Radtke KL, Yates MK. 2018. [The evolution of nucleoside analogue antivirals: A review for chemists and non-chemists. Part 1: Early structural modifications to the nucleoside scaffold](#). Antiviral Research 154:66–86.
794. Jin Z, Smith LK, Rajwanshi VK, Kim B, Deval J. 2013. [The Ambiguous Base-Pairing and High Substrate Efficiency of T-705 \(Favipiravir\) Ribofuranosyl 5'-Triphosphate towards Influenza A Virus Polymerase](#). PLoS ONE 8:e68347.
795. 2020. Favipiravir. DrugBank. <https://www.drugbank.ca/drugs/DB12466>.
796. Furuta Y, Takahashi K, Fukuda Y, Kuno M, Kamiyama T, Kozaki K, Nomura N, Egawa H, Minami S, Watanabe Y, Narita H, Shiraki K. 2002. [In Vitro and In Vivo Activities of Anti-Influenza Virus Compound T-705](#). Antimicrobial Agents and Chemotherapy 46:977–981.
797. Sidwell RW, Barnard DL, Day CW, Smee DF, Bailey KW, Wong M-H, Morrey JD, Furuta Y. 2007. [Efficacy of Orally Administered T-705 on Lethal Avian Influenza A \(H5N1\) Virus Infections in Mice](#). Antimicrobial Agents and Chemotherapy 51:845–851.
798. Furuta Y, Takahashi K, Kuno-Maekawa M, Sangawa H, Uehara S, Kozaki K, Nomura N, Egawa H, Shiraki K. 2005. [Mechanism of Action of T-705 against Influenza Virus](#). Antimicrobial Agents and Chemotherapy 49:981–986.
799. Julander JG, Shafer K, Smee DF, Morrey JD, Furuta Y. 2009. [Activity of T-705 in a Hamster Model of Yellow Fever Virus Infection in Comparison with That of a Chemically Related Compound, T-1106](#). Antimicrobial Agents and Chemotherapy 53:202–209.
800. Gowen BB, Wong M-H, Jung K-H, Sanders AB, Mendenhall M, Bailey KW, Furuta Y, Sidwell RW. 2007. [In Vitro and In Vivo Activities of T-705 against Arenavirus and Bunyavirus Infections](#). Antimicrobial Agents and Chemotherapy 51:3168–3176.
801. Rocha-Pereira J, Jochmans D, Dallmeier K, Leyssen P, Nascimento MSJ, Neyts J. 2012. [Favipiravir \(T-705\) inhibits in vitro norovirus replication](#). Biochemical and Biophysical Research Communications 424:777–780.
802. Mendenhall M, Russell A, Juelich T, Messina EL, Smee DF, Freiberg AN, Holbrook MR, Furuta Y, de la Torre J-C, Nunberg JH, Gowen BB. 2011. [T-705 \(Favipiravir\) Inhibition of Arenavirus Replication in Cell Culture](#). Antimicrobial Agents and Chemotherapy 55:782–787.

803. FURUTA Y, KOMENO T, NAKAMURA T. 2017. [Favipiravir \(T-705\), a broad spectrum inhibitor of viral RNA polymerase](#). Proceedings of the Japan Academy, Series B 93:449–463.
804. Gordon CJ, Tchesnokov EP, Feng JY, Porter DP, Götte M. 2020. [The antiviral compound remdesivir potently inhibits RNA-dependent RNA polymerase from Middle East respiratory syndrome coronavirus](#). Journal of Biological Chemistry 295:4773–4779.
805. Agostini ML, Andres EL, Sims AC, Graham RL, Sheahan TP, Lu X, Smith EC, Case JB, Feng JY, Jordan R, Ray AS, Cihlar T, Siegel D, Mackman RL, Clarke MO, Baric RS, Denison MR. 2018. [Coronavirus Susceptibility to the Antiviral Remdesivir \(GS-5734\) Is Mediated by the Viral Polymerase and the Proofreading Exoribonuclease](#). mBio 9:e00221–18.
806. Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M, Shi Z, Hu Z, Zhong W, Xiao G. 2020. [Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus \(2019-nCoV\) in vitro](#). Cell Research 30:269–271.
807. Beigel JH, Tomashek KM, Dodd LE, Mehta AK, Zingman BS, Kalil AC, Hohmann E, Chu HY, Luetkemeyer A, Kline S, Lopez de Castilla D, Finberg RW, Dierberg K, Tapson V, Hsieh L, Patterson TF, Paredes R, Sweeney DA, Short WR, Touloumi G, Lye DC, Ohmagari N, Oh M, Ruiz-Palacios GM, Benfield T, Fätkenheuer G, Kortepeter MG, Atmar RL, Creech CB, Lundgren J, Babiker AG, Pett S, Neaton JD, Burgess TH, Bonnett T, Green M, Makowski M, Osinusi A, Nayak S, Lane HC. 2020. [Remdesivir for the Treatment of Covid-19 — Final Report](#). New England Journal of Medicine 383:1813–1826.
808. National Institute of Allergy and Infectious Diseases (NIAID). 2020. [A Multicenter, Adaptive, Randomized Blinded Controlled Trial of the Safety and Efficacy of Investigational Therapeutics for the Treatment of COVID-19 in Hospitalized Adults](#). NCT04280705. Clinical trial registration. clinicaltrials.gov.
809. Gilead Sciences. 2020. [A Phase 3 Randomized Study to Evaluate the Safety and Antiviral Activity of Remdesivir \(GS-5734™\) in Participants With Severe COVID-19](#). NCT04292899. Clinical trial registration. clinicaltrials.gov.
810. Grein J, Ohmagari N, Shin D, Diaz G, Asperges E, Castagna A, Feldt T, Green G, Green ML, Lescure F-X, Nicastrì E, Oda R, Yo K, Quiros-Roldan E, Studemeister A, Redinski J, Ahmed S, Bennett J, Chelliah D, Chen D, Chihara S, Cohen SH, Cunningham J, D'Arminio Monforte A, Ismail S, Kato H, Lapadula G, L'Her E, Maeno T, Majumder S, Massari M, Mora-Rillo M, Mutoh Y, Nguyen D, Verweij E, Zoufaly A, Osinusi AO, DeZure A, Zhao Y, Zhong L, Chokkalingam A, Elboudwarej E, Telep L, Timbs L, Henne I, Sellers S, Cao H, Tan SK, Winterbourne L, Desai P, Mera R, Gaggari A, Myers RP, Brainard DM, Childs R, Flanigan T. 2020. [Compassionate Use of Remdesivir for Patients with Severe Covid-19](#). New England Journal of Medicine 382:2327–2336.
811. WHO Solidarity Trial Consortium. 2020. [Repurposed Antiviral Drugs for Covid-19 — Interim WHO Solidarity Trial Results](#). New England Journal

of Medicine NEJMoa2023184.

812. 2020. PLAQUENIL - hydroxychloroquine sulfate tablet. DailyMed. <https://dailymed.nlm.nih.gov/dailymed/drugInfo.cfm?setid=34496b43-05a2-45fb-a769-52b12e099341>.
813. Kalia S, Dutz JP. 2007. [New concepts in antimalarial use and mode of action in dermatology](#). Dermatologic Therapy 20:160–174.
814. Vincent MJ, Bergeron E, Benjannet S, Erickson BR, Rollin PE, Ksiazek TG, Seidah NG, Nichol ST. 2005. [Chloroquine is a potent inhibitor of SARS coronavirus infection and spread](#). Virology Journal 2:69.
815. Yao X, Ye F, Zhang M, Cui C, Huang B, Niu P, Liu X, Zhao L, Dong E, Song C, Zhan S, Lu R, Li H, Tan W, Liu D. 2020. [In Vitro Antiviral Activity and Projection of Optimized Dosing Design of Hydroxychloroquine for the Treatment of Severe Acute Respiratory Syndrome Coronavirus 2 \(SARS-CoV-2\)](#). Clinical Infectious Diseases 71:732–739.
816. Gautret P, Lagier J-C, Parola P, Hoang VT, Meddeb L, Mailhe M, Doudier B, Courjon J, Giordanengo V, Vieira VE, Tissot Dupont H, Honoré S, Colson P, Chabrière E, La Scola B, Rolain J-M, Brouqui P, Raoult D. 2020. [Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial](#). International Journal of Antimicrobial Agents 56:105949.
817. Voss A. 2020. Official Statement from International Society of Antimicrobial Chemotherapy. <https://www.isac.world/news-and-publications/official-isac-statement>.
818. The RECOVERY Collaborative Group. 2020. [Effect of Hydroxychloroquine in Hospitalized Patients with Covid-19](#). New England Journal of Medicine 383:2030–2040.
819. No clinical benefit from use of hydroxychloroquine in hospitalised patients with COVID-19 — RECOVERY Trial. <https://www.recoverytrial.net/news/statement-from-the-chief-investigators-of-the-randomised-evaluation-of-covid-19-therapy-recovery-trial-on-hydroxychloroquine-5-june-2020-no-clinical-benefit-from-use-of-hydroxychloroquine-in-hospitalised-patients-with-covid-19>. Retrieved 5 December 2022.
820. Ōmura S, Crump A. 2004. [The life and times of ivermectin — a success story](#). Nat Rev Microbiol 2:984–989.
821. Burg RW, Miller BM, Baker EE, Birnbaum J, Currie SA, Hartman R, Kong Y-L, Monaghan RL, Olson G, Putter I, Tunac JB, Wallick H, Stapley EO, Oiwa R, Ōmura S. 1979. [Avermectins, New Family of Potent Anthelmintic Agents: Producing Organism and Fermentation](#). Antimicrob Agents Chemother 15:361–367.
822. CRUMP A, OMURA S. 2011. [Ivermectin, 'Wonder drug' from Japan: the human use perspective](#). Proceedings of the Japan Academy Ser B: Physical and Biological Sciences 87:13–28.

823. Crump A. 2017. [Ivermectin: enigmatic multifaceted 'wonder' drug continues to surprise and exceed expectations](#). J Antibiot 70:495–505.
824. Egerton JR, Ostlind DA, Blair LS, Eary CH, Suhayda D, Cifelli S, Riek RF, Campbell WC. 1979. [Avermectins, New Family of Potent Anthelmintic Agents: Efficacy of the B1a Component](#). Antimicrobial Agents and Chemotherapy 15:372–378.
825. Heidary F, Gharebaghi R. 2020. [Ivermectin: a systematic review from antiviral effects to COVID-19 complementary regimen](#). J Antibiot 73:593–602.
826. Sharun K, Dhama K, Patel SK, Pathak M, Tiwari R, Singh BR, Sah R, Bonilla-Aldana DK, Rodriguez-Morales AJ, Leblebicioglu H. 2020. [Ivermectin, a new candidate therapeutic against SARS-CoV-2/COVID-19](#). Ann Clin Microbiol Antimicrob 19.
827. Caly L, Druce JD, Catton MG, Jans DA, Wagstaff KM. 2020. [The FDA-approved drug ivermectin inhibits the replication of SARS-CoV-2 in vitro](#). Antiviral Research 178:104787.
828. Chaccour C, Hammann F, Ramón-García S, Rabinovich NR. 2020. [Ivermectin and COVID-19: Keeping Rigor in Times of Urgency](#). The American Journal of Tropical Medicine and Hygiene 102:1156–1157.
829. Schmith VD, Zhou J(Jessie), Lohmer LRL. 2020. [The Approved Dose of Ivermectin Alone is not the Ideal Dose for the Treatment of COVID-19](#). Clin Pharmacol Ther 108:762–765.
830. Momekov G, Momekova D. 2020. [Ivermectin as a potential COVID-19 treatment from the pharmacokinetic point of view: antiviral levels are not likely attainable with known dosing regimens](#). Biotechnology & Biotechnological Equipment 34:469–474.
831. Ménez C, Sutra J-F, Prichard R, Lespine A. 2012. [Relative Neurotoxicity of Ivermectin and Moxidectin in Mdr1ab \(-/-\) Mice and Effects on Mammalian GABA\(A\) Channel Activity](#). PLoS Negl Trop Dis 6:e1883.
832. Rajter JC, Sherman MS, Fatteh N, Vogel F, Sacks J, Rajter J-J. 2021. [Use of Ivermectin Is Associated With Lower Mortality in Hospitalized Patients With Coronavirus Disease 2019](#). Chest 159:85–92.
833. Camprubí D, Almuedo-Riera A, Martí-Soler H, Soriano A, Hurtado JC, Subirà C, Grau-Pujol B, Krolewiecki A, Muñoz J. 2020. [Lack of efficacy of standard doses of ivermectin in severe COVID-19 patients](#). PLoS ONE 15:e0242184.
834. Gheibi N, Shakhsi Niaee M, Namdar P, Allami A, Zolghadr L, Javadi A, Karampour A, Varnaseri M, Bijani B, Cheraghi F, Naderi Y, Amini F, Karamyan M, YadYad M, Jamshidian R. 2021. [Ivermectin as an adjunct treatment for hospitalized adult COVID-19 patients: A randomized multi-center clinical trial](#). Asian Pac J Trop Med 14:266.
835. Babalola OE, Bode CO, Ajayi AA, Alakaloko FM, Akase IE, Otrofanowei E, Salu OB, Adeyemo WL, Ademuyiwa AO, Omilabu S. 2021. [Ivermectin shows clinical benefits in mild to moderate COVID19: a randomized](#)

[controlled double-blind, dose-response study in Lagos](#). QJM: An International Journal of Medicine 114:780–788.

836. Pott-Junior H, Paoliello MMB, Miguel A de QC, da Cunha AF, de Melo Freire CC, Neves FF, da Silva de Avó LR, Roscani MG, dos Santos SDS, Chachá SGF. 2021. [RETRACTED: Use of ivermectin in the treatment of Covid-19: A pilot trial](#). Toxicology Reports 8:505–510.
837. Ahmed S, Karim MM, Ross AG, Hossain MS, Clemens JD, Sumiya MK, Phru CS, Rahman M, Zaman K, Somani J, Yasmin R, Hasnat MA, Kabir A, Aziz AB, Khan WA. 2021. [A five-day course of ivermectin for the treatment of COVID-19 may reduce the duration of illness](#). International Journal of Infectious Diseases 103:214–216.
838. Shahbaznejad L, Davoudi A, Eslami G, Markowitz JS, Navaeifar MR, Hosseinzadeh F, Movahedi FS, Rezai MS. 2021. [Effects of Ivermectin in Patients With COVID-19: A Multicenter, Double-blind, Randomized, Controlled Clinical Trial](#). Clinical Therapeutics 43:1007–1019.
839. Chaccour C, Casellas A, Blanco-Di Matteo A, Pineda I, Fernandez-Montero A, Ruiz-Castillo P, Richardson M-A, Rodríguez-Mateos M, Jordán-Iborra C, Brew J, Carmona-Torre F, Giráldez M, Laso E, Gabaldón-Figueira JC, Dobaño C, Moncunill G, Yuste JR, Del Pozo JL, Rabinovich NR, Schöning V, Hammann F, Reina G, Sadaba B, Fernández-Alonso M. 2021. [The effect of early treatment with ivermectin on viral load, symptoms and humoral response in patients with non-severe COVID-19: A pilot, double-blind, placebo-controlled, randomized clinical trial](#). EClinicalMedicine 32:100720.
840. Mohan A, Tiwari P, Suri T, Mittal S, Patel A, Jain A, T. V, Das UK, Bopanna TK, Pandey R, Shelke S, Singh AR, Bhatnagar S, Masih S, Mahajan S, Dwivedi T, Sahoo B, Pandit A, Bhopale S, Vig S, Gupta R, Madan K, Hadda V, Gupta N, Garg R, Meena VP, Guleria R. 2021. [Ivermectin in mild and moderate COVID-19 \(RIVET-COV\): a randomized, placebo-controlled trial](#). Research Square Platform LLC.
841. Ravikirti, Roy R, Pattadar C, Raj R, Agarwal N, Biswas B, Manjhi PK, Rai DK, Shyama, Kumar A, Sarfaraz A. 2021. [Evaluation of Ivermectin as a Potential Treatment for Mild to Moderate COVID-19: A Double-Blind Randomized Placebo Controlled Trial in Eastern India](#). J Pharm Pharm Sci 24:343–350.
842. Elgazzar A, Eltaweel A, Youssef SA, Hany B, Hafez M, Moussa H. 2020. [Efficacy and Safety of Ivermectin for Treatment and prophylaxis of COVID-19 Pandemic](#). Research Square Platform LLC.
843. Lawrence J. 2021. Why Was a Major Study on Ivermectin for COVID-19 Just Retracted? <https://grftr.news/why-was-a-major-study-on-ivermectin-for-covid-19-just-retracted>.
844. Brown N. 2021. Nick Brown's blog: Some problems in the dataset of a large study of Ivermectin for the treatment of Covid-19. Nick Brown's blog. <https://steamtraen.blogspot.com/2021/07/Some-problems-with-the-data-from-a-Covid-study.html>. Retrieved 5 December 2022.

845. 2021. [Efficacy and Safety of Ivermectin for Treatment and prophylaxis of COVID-19 Pandemic](#). Research Square Platform LLC.
846. López-Medina E, López P, Hurtado IC, Dávalos DM, Ramirez O, Martínez E, Díazgranados JA, Oñate JM, Chavarriaga H, Herrera S, Parra B, Libreros G, Jaramillo R, Avendaño AC, Toro DF, Torres M, Lesmes MC, Rios CA, Caicedo I. 2021. [Effect of Ivermectin on Time to Resolution of Symptoms Among Adults With Mild COVID-19](#). JAMA 325:1426.
847. Roman YM, Burela PA, Pasupuleti V, Piscoya A, Vidal JE, Hernandez AV. 2021. [Ivermectin for the treatment of COVID-19: A systematic review and meta-analysis of randomized controlled trials](#). Cold Spring Harbor Laboratory.
848. Castañeda-Sabogal A, Chamberg-Michilot D, Toro-Huamanchumo CJ, Silva-Rengifo C, Gonzales-Zamora J, Barboza JJ. 2021. [Outcomes of Ivermectin in the treatment of COVID-19: a systematic review and meta-analysis](#). Cold Spring Harbor Laboratory.
849. Hill A, Garratt A, Levi J, Falconer J, Ellis L, McCann K, Pilkington V, Qavi A, Wang J, Wentzel H. 2021. [RETRACTED: Expression of Concern: "Meta-analysis of Randomized Trials of Ivermectin to Treat SARS-CoV-2 Infection"](#). Open Forum Infectious Diseases 8.
850. Zein AFMZ, Sulistiyana CS, Raffaello WM, Pranata R. 2021. [Ivermectin and mortality in patients with COVID-19: A systematic review, meta-analysis, and meta-regression of randomized controlled trials](#). Diabetes & Metabolic Syndrome: Clinical Research & Reviews 15:102186.
851. Bryant A, Lawrie TA, Dowswell T, Fordham EJ, Mitchell S, Hill SR, Tham TC. 2021. [Ivermectin for Prevention and Treatment of COVID-19 Infection](#). American Journal of Therapeutics Publish Ahead of Print.
852. Hariyanto TI, Halim DA, Rosalind J, Gunawan C, Kurniawan A. 2021. [Ivermectin and outcomes from Covid-19 pneumonia: A systematic review and meta-analysis of randomized clinical trial studies](#). Reviews in Medical Virology 32.
853. Bryant A, Lawrie TA, Dowswell T, Fordham E, Scott M MD, Hill SR, Tham TC. 2021. [Ivermectin for prevention and treatment of COVID-19 infection: a systematic review and meta-analysis](#). Center for Open Science.
854. Kory P, Meduri GU, Varon J, Iglesias J, Marik PE. 2021. [Review of the Emerging Evidence Demonstrating the Efficacy of Ivermectin in the Prophylaxis and Treatment of COVID-19](#). American Journal of Therapeutics 28:e299–e318.
855. Kow CS, Merchant HA, Mustafa ZU, Hasan SS. 2021. [The association between the use of ivermectin and mortality in patients with COVID-19: a meta-analysis](#). Pharmacol Rep 73:1473–1479.
856. Karale S, Bansal V, Makadia J, Tayyeb M, Khan H, Ghanta SS, Singh R, Tekin A, Bhurwal A, Mutneja H, Mehra I, Kashyap R. 2021. [An Updated](#)

[Systematic Review and Meta-Analysis of Mortality, Need for ICU admission, Use of Mechanical Ventilation, Adverse effects and other Clinical Outcomes of Ivermectin Treatment in COVID-19 Patients](#). Cold Spring Harbor Laboratory.

857. M-K G. 2021. Does Ivermectin Work for Covid-19? <https://gidmk.medium.com/does-ivermectin-work-for-covid-19-1166126c364a>.
858. Hill A, Garratt A, Levi J, Falconer J, Ellis L, McCann K, Pilkington V, Qavi A, Wang J, Wentzel H. 2021. [Retracted: Meta-analysis of Randomized Trials of Ivermectin to Treat SARS-CoV-2 Infection](#). Open Forum Infectious Diseases 8.
859. Medicine C for V. 2022. [FAQ: COVID-19 and Ivermectin Intended for Animals](#). FDA.
860. Cardresearch. 2022. [A Multicenter, Prospective, Adaptive, Double-blind, Randomized, Placebo-controlled Study to Evaluate the Effect of Interferon Lambda 1A, Fluvoxamina + Budesonida, Fluoxetina + Budesonida in Mild COVID-19 and High Risk of Complications](#). NCT04727424. Clinical trial registration. clinicaltrials.gov.
861. [Ivermectin to be investigated in adults aged 18+ as a possible treatment for COVID-19 in the PRINCIPLE trial — PRINCIPLE Trial](#).
862. Reis G, Moreira Silva EA dos S, Medeiros Silva DC, Thabane L, Singh G, Park JJH, Forrest JI, Harari O, Quirino dos Santos CV, Guimarães de Almeida APF, Figueiredo Neto AD de, Savassi LCM, Milagres AC, Teixeira MM, Simplicio MIC, Ribeiro LB, Oliveira R, Mills EJ. 2021. [Effect of Early Treatment With Hydroxychloroquine or Lopinavir and Ritonavir on Risk of Hospitalization Among Patients With COVID-19](#). JAMA Netw Open 4:e216468.
863. terren.green@duke.edu. 2021. August 6, 2021: Early Treatment of COVID-19 with Repurposed Therapies: The TOGETHER Adaptive Platform Trial (Edward Mills, PhD, FRCP). Rethinking Clinical Trials. <https://rethinkingclinicaltrials.org/news/august-6-2021-early-treatment-of-covid-19-with-repurposed-therapies-the-together-adaptive-platform-trial-edward-mills-phd-frcp/>. Retrieved 5 December 2022.
864. Lisinopril - Drug Usage Statistics. ClinCalc DrugStats Database. <https://clincalc.com/DrugStats/Drugs/Lisinopril>. Retrieved 30 July 2020.
865. Byrd JB, Chertow GM, Bhalla V. 2019. [Hypertension Hot Potato — Anatomy of the Angiotensin-Receptor Blocker Recalls](#). New England Journal of Medicine 380:1589–1591.
866. Bian B, Kelton CML, Guo JJ, Wigle PR. 2010. [ACE Inhibitor and ARB Utilization and Expenditures in the Medicaid Fee-For-Service Program from 1991 to 2008](#). JMCP 16:671–679.
867. Turner AJ, Hiscox JA, Hooper NM. 2004. [ACE2: from vasopeptidase to SARS virus receptor](#). Trends in Pharmacological Sciences 25:291–294.

868. Huentelman MJ, Zubcevic J, Hernández Prada JA, Xiao X, Dimitrov DS, Raizada MK, Ostrov DA. 2004. [Structure-Based Discovery of a Novel Angiotensin-Converting Enzyme 2 Inhibitor](#). *Hypertension* 44:903–906.
869. Dimitrov DS. 2003. [The Secret Life of ACE2 as a Receptor for the SARS Virus](#). *Cell* 115:652–653.
870. Kreutz R, Algharably EAE-H, Azizi M, Dobrowolski P, Guzik T, Januszewicz A, Persu A, Prejbisz A, Riemer TG, Wang J-G, Burnier M. 2020. [Hypertension, the renin-angiotensin system, and the risk of lower respiratory tract infections and lung injury: implications for COVID-19](#). *Cardiovascular Research* 116:1688–1699.
871. Walters TE, Kalman JM, Patel SK, Mearns M, Velkoska E, Burrell LM. 2016. [Angiotensin converting enzyme 2 activity and human atrial fibrillation: increased plasma angiotensin converting enzyme 2 activity is associated with atrial fibrillation and more advanced left atrial structural remodelling](#). *Europace* euw246.
872. Mehra MR, Desai SS, Kuy S, Henry TD, Patel AN. 2020. [Cardiovascular Disease, Drug Therapy, and Mortality in Covid-19](#). *New England Journal of Medicine* 382:e102.
873. Mehra MR, Desai SS, Kuy S, Henry TD, Patel AN. 2020. [Retraction: Cardiovascular Disease, Drug Therapy, and Mortality in Covid-19. N Engl J Med. DOI: 10.1056/NEJMoa2007621](#). *N Engl J Med* 382:2582–2582.
874. Cohen JB, Hanff TC, William P, Sweitzer N, Rosado-Santander NR, Medina C, Rodriguez-Mori JE, Renna N, Chang TI, Corrales-Medina V, Andrade-Villanueva JF, Barbagelata A, Cristodulo-Cortez R, Díaz-Cucho OA, Spaak J, Alfonso CE, Valdivia-Vega R, Villavicencio-Carranza M, Ayala-García RJ, Castro-Callirgos CA, González-Hernández LA, Bernales-Salas EF, Coacalla-Guerra JC, Salinas-Herrera CD, Nicolosi L, Basconcel M, Byrd JB, Sharkoski T, Bendeزú-Huwasquiche LE, Chittams J, Edmonston DL, Vasquez CR, Chirinos JA. 2021. [Continuation versus discontinuation of renin-angiotensin system inhibitors in patients admitted to hospital with COVID-19: a prospective, randomised, open-label trial](#). *The Lancet Respiratory Medicine* 9:275–284.
875. Lopes RD, Macedo AVS, de Barros E Silva PGM, Moll-Bernardes RJ, dos Santos TM, Mazza L, Feldman A, D'Andréa Saba Arruda G, de Albuquerque DC, Camiletti AS, de Sousa AS, de Paula TC, Giusti KGD, Domiciano RAM, Noya-Rabelo MM, Hamilton AM, Loures VA, Dionísio RM, Furquim TAB, De Luca FA, dos Santos Sousa ÍB, Bandeira BS, Zukowski CN, de Oliveira RGG, Ribeiro NB, de Moraes JL, Petriz JLF, Pimentel AM, Miranda JS, de Jesus Abufaiad BE, Gibson CM, Granger CB, Alexander JH, de Souza OF. 2021. [Effect of Discontinuing vs Continuing Angiotensin-Converting Enzyme Inhibitors and Angiotensin II Receptor Blockers on Days Alive and Out of the Hospital in Patients Admitted With COVID-19](#). *JAMA* 325:254.
876. 2020. Frequently Asked Questions on the Revocation of the Emergency Use Authorization for Hydroxychloroquine Sulfate and Chloroquine

Phosphate. US Food and Drug Administration.
<https://www.fda.gov/media/138946/download>.

877. 2020. COVID-19: chloroquine and hydroxychloroquine only to be used in clinical trials or emergency use programmes.
<https://www.ema.europa.eu/en/news/covid-19-chloroquine-hydroxychloroquine-only-be-used-clinical-trials-emergency-use-programmes>.
878. Shire SJ. 2009. [Formulation and manufacturability of biologics](#). Current Opinion in Biotechnology 20:708–714.
879. Baumann A. 2006. [Early Development of Therapeutic Biologics - Pharmacokinetics](#). Current Drug Metabolism 7:15–21.
880. Lu R-M, Hwang Y-C, Liu I-J, Lee C-C, Tsai H-Z, Li H-J, Wu H-C. 2020. [Development of therapeutic antibodies for the treatment of diseases](#). Journal of Biomedical Science 27:1.
881. Corti D, Lanzavecchia A. 2013. [Broadly Neutralizing Antiviral Antibodies](#). Annual Review of Immunology 31:705–742.
882. Iacob SA, Iacob DG. 2017. [Ibalizumab Targeting CD4 Receptors, An Emerging Molecule in HIV Therapy](#). Frontiers in Microbiology 8:2323.
883. Resch B. 2017. [Product review on the monoclonal antibody palivizumab for prevention of respiratory syncytial virus infection](#). Human Vaccines & Immunotherapeutics 13:2138–2149.
884. Houser KV, Gretebeck L, Ying T, Wang Y, Vogel L, Lamirande EW, Bock KW, Moore IN, Dimitrov DS, Subbarao K. 2016. [Prophylaxis With a Middle East Respiratory Syndrome Coronavirus \(MERS-CoV\)-Specific Human Monoclonal Antibody Protects Rabbits From MERS-CoV Infection](#). J Infect Dis 213:1557–1561.
885. van Doremalen N, Falzarano D, Ying T, de Wit E, Bushmaker T, Feldmann F, Okumura A, Wang Y, Scott DP, Hanley PW, Feldmann H, Dimitrov DS, Munster VJ. 2017. [Efficacy of antibody-based therapies against Middle East respiratory syndrome coronavirus \(MERS-CoV\) in common marmosets](#). Antiviral Research 143:30–37.
886. Tanaka T, Narazaki M, Kishimoto T. 2014. [IL-6 in Inflammation, Immunity, and Disease](#). Cold Spring Harbor Perspectives in Biology 6:a016295–a016295.
887. 2020. ACTEMRA - tocilizumab. DailyMed.
<https://dailymed.nlm.nih.gov/dailymed/drugInfo.cfm?setid=2e5365ff-cb2a-4b16-b2c7-e35c6bf2de13>.
888. Guaraldi G, Meschiari M, Cozzi-Lepri A, Milic J, Tonelli R, Menozzi M, Franceschini E, Cuomo G, Orlando G, Borghi V, Santoro A, Di Gaetano M, Puzzolante C, Carli F, Bedini A, Corradi L, Fantini R, Castaniere I, Tabbì L, Girardis M, Tedeschi S, Giannella M, Bartoletti M, Pascale R, Dolci G, Brugioni L, Pietrangelo A, Cossarizza A, Pea F, Clini E, Salvarani C, Massari M, Viale PL, Mussini C. 2020. [Tocilizumab in patients with](#)

[severe COVID-19: a retrospective cohort study](#). The Lancet Rheumatology 2:e474–e484.

889. Price CC, Altice FL, Shyr Y, Koff A, Pischel L, Goshua G, Azar MM, Mcmanus D, Chen S-C, Gleeson SE, Britto CJ, Azmy V, Kaman K, Gaston DC, Davis M, Burrello T, Harris Z, Villanueva MS, Aoun-Barakat L, Kang I, Seropian S, Chupp G, Bucala R, Kaminski N, Lee AI, LoRusso PM, Topal JE, Dela Cruz C, Malinis M. 2020. [Tocilizumab Treatment for Cytokine Release Syndrome in Hospitalized Patients With Coronavirus Disease 2019](#). Chest 158:1397–1408.
890. Capra R, De Rossi N, Mattioli F, Romanelli G, Scarpazza C, Sormani MP, Cossi S. 2020. [Impact of low dose tocilizumab on mortality rate in patients with COVID-19 related pneumonia](#). European Journal of Internal Medicine 76:31–35.
891. Klopfenstein T, Zayet S, Lohse A, Balblanc J-C, Badie J, Royer P-Y, Toko L, Mezher C, Kadiane-Oussou NJ, Bossert M, Bozgan A-M, Charpentier A, Roux M-F, Contreras R, Mazurier I, Dussert P, Gendrin V, Conrozier T. 2020. [Tocilizumab therapy reduced intensive care unit admissions and/or mortality in COVID-19 patients](#). Médecine et Maladies Infectieuses 50:397–400.
892. Rojas-Marte G, Khalid M, Mukhtar O, Hashmi AT, Waheed MA, Ehrlich S, Aslam A, Siddiqui S, Agarwal C, Malyshev Y, Henriquez-Felipe C, Sharma D, Sharma S, Chukwuka N, Rodriguez DC, Alliu S, Le J, Shani J. 2020. [Outcomes in patients with severe COVID-19 disease treated with tocilizumab: a case-controlled study](#). QJM: An International Journal of Medicine 113:546–550.
893. Xu X, Han M, Li T, Sun W, Wang D, Fu B, Zhou Y, Zheng X, Yang Y, Li X, Zhang X, Pan A, Wei H. 2020. [Effective treatment of severe COVID-19 patients with tocilizumab](#). Proceedings of the National Academy of Sciences 117:10970–10975.
894. , Horby PW, Pessoa-Amorim G, Peto L, Brightling CE, Sarkar R, Thomas K, Jeebun V, Ashish A, Tully R, Chadwick D, Sharafat M, Stewart R, Rudran B, Baillie JK, Buch MH, Chappell LC, Day JN, Furst SN, Jaki T, Jeffery K, Juszczak E, Lim WS, Montgomery A, Mumford A, Rowan K, Thwaites G, Mafham M, Haynes R, Landray MJ. 2021. [Tocilizumab in patients admitted to hospital with COVID-19 \(RECOVERY\): preliminary results of a randomised, controlled, open-label, platform trial](#). Cold Spring Harbor Laboratory.
895. Rosas IO, Bräu N, Waters M, Go RC, Hunter BD, Bhagani S, Skiest D, Aziz MS, Cooper N, Douglas IS, Savic S, Youngstein T, Del Sorbo L, Cubillo Gracian A, De La Zerda DJ, Ustianowski A, Bao M, Dimonaco S, Graham E, Matharu B, Spotswood H, Tsai L, Malhotra A. 2021. [Tocilizumab in Hospitalized Patients with Severe Covid-19 Pneumonia](#). N Engl J Med 384:1503–1516.
896. Salama C, Han J, Yau L, Reiss WG, Kramer B, Neidhart JD, Criner GJ, Kaplan-Lewis E, Baden R, Pandit L, Cameron ML, Garcia-Diaz J, Chávez V, Mekebeb-Reuter M, Lima de Menezes F, Shah R, González-Lara MF,

- Assman B, Freedman J, Mohan SV. 2021. [Tocilizumab in Patients Hospitalized with Covid-19 Pneumonia](#). N Engl J Med 384:20–30.
897. Hoffmann-La Roche. 2022. [A Phase III, Randomized, Double-Blind, Multicenter Study to Evaluate the Efficacy and Safety of Remdesivir Plus Tocilizumab Compared With Remdesivir Plus Placebo in Hospitalized Patients With Severe COVID-19 Pneumonia](#). NCT04409262. Clinical trial registration. clinicaltrials.gov.
898. Hoffmann-La Roche. 2021. [A Randomized, Double-Blind, Placebo-Controlled, Multicenter Study to Evaluate the Safety and Efficacy of Tocilizumab in Patients With Severe COVID-19 Pneumonia](#). NCT04320615. Clinical trial registration. clinicaltrials.gov.
899. Genentech, Inc. 2021. [A Randomized, Double-Blind, Placebo-Controlled, Multicenter Study to Evaluate the Efficacy and Safety of Tocilizumab in Hospitalized Patients With COVID-19 Pneumonia](#). NCT04372186. Clinical trial registration. clinicaltrials.gov.
900. 2021. Genentech tocilizumab Letter of Authority. US Food and Drug Administration. <https://www.fda.gov/media/150319/download>.
901. Synairgen Research Ltd. 2021. [A Randomised Double-blind Placebo-controlled Trial to Determine the Safety and Efficacy of Inhaled SNG001 \(IFN- \$\beta\$ 1a for Nebulisation\) for the Treatment of Patients With Confirmed SARS-CoV-2 Infection](#). NCT04385095. Clinical trial registration. clinicaltrials.gov.
902. 2020. Synairgen announces positive results from trial of SNG001 in hospitalised COVID-19 patients. Synairgen plc press release. <http://synairgen.web01.hosting.bdc.co.uk/umbraco/Surface/Download/GetFile?cid=1130026e-0983-4338-b648-4ac7928b9a37>.
903. Monk PD, Marsden RJ, Tear VJ, Brookes J, Batten TN, Mankowski M, Gabbay FJ, Davies DE, Holgate ST, Ho L-P, Clark T, Djukanovic R, Wilkinson TMA, Crooks MG, Dosanjh DP, Siddiqui S, Rahman NM, Smith JA, Horsley A, Harrison TW, Saralaya D, McGarvey L, Watson A, Foster E, Fleet A, Singh D, Hemmings S, Aitken S, Dudley S, Beegan R, Thompson A, Rodrigues PM. 2021. [Safety and efficacy of inhaled nebulised interferon beta-1a \(SNG001\) for treatment of SARS-CoV-2 infection: a randomised, double-blind, placebo-controlled, phase 2 trial](#). The Lancet Respiratory Medicine 9:196–206.
904. Chen L, Xiong J, Bao L, Shi Y. 2020. [Convalescent plasma as a potential therapy for COVID-19](#). The Lancet Infectious Diseases 20:398–400.
905. Roback JD, Guarner J. 2020. [Convalescent Plasma to Treat COVID-19](#). JAMA 323:1561.
906. Rajendran K, Krishnasamy N, Rangarajan J, Rathinam J, Natarajan M, Ramachandran A. 2020. [Convalescent plasma transfusion for the treatment of COVID-19: Systematic review](#). J Med Virol 92:1475–1483.
907. Janiaud P, Axfors C, Schmitt AM, Gloy V, Ebrahimi F, Hepprich M, Smith ER, Haber NA, Khanna N, Moher D, Goodman SN, Ioannidis JPA,

- Hemkens LG. 2021. [Association of Convalescent Plasma Treatment With Clinical Outcomes in Patients With COVID-19](#). JAMA 325:1185.
908. Joyner MJ, Carter RE, Senefeld JW, Klassen SA, Mills JR, Johnson PW, Theel ES, Wiggins CC, Bruno KA, Klompas AM, Lesser ER, Kunze KL, Sexton MA, Diaz Soto JC, Baker SE, Shepherd JRA, van Helmond N, Verdun NC, Marks P, van Buskirk CM, Winters JL, Stubbs JR, Rea RF, Hodge DO, Herasevich V, Whelan ER, Clayburn AJ, Larson KF, Ripoll JG, Andersen KJ, Buras MR, Vogt MNP, Dennis JJ, Regimbal RJ, Bauer PR, Blair JE, Paneth NS, Fairweather D, Wright RS, Casadevall A. 2021. [Convalescent Plasma Antibody Levels and the Risk of Death from Covid-19](#). New England Journal of Medicine NEJMoa2031893.
909. , Horby PW, Estcourt L, Peto L, Emberson JR, Staplin N, Spata E, Pessoa-Amorim G, Campbell M, Roddick A, Brunskill NE, George T, Zehnder D, Tiberi S, Aung NN, Uriel A, Widdrington J, Koshy G, Brown T, Scott S, Baillie JK, Buch MH, Chappell LC, Day JN, Faust SN, Jaki T, Jeffery K, Juszczak E, Lim WS, Montgomery A, Mumford A, Rowan K, Thwaites G, Mafham M, Roberts D, Haynes R, Landray MJ. 2021. [Convalescent plasma in patients admitted to hospital with COVID-19 \(RECOVERY\): a randomised, controlled, open-label, platform trial](#). Cold Spring Harbor Laboratory.
910. Hsueh P-R, Huang L-M, Chen P-J, Kao C-L, Yang P-C. 2004. [Chronological evolution of IgM, IgA, IgG and neutralisation antibodies after infection with SARS-associated coronavirus](#). Clinical Microbiology and Infection 10:1062–1066.
911. Yuchun N, Guangwen W, Xuanling S, Hong Z, Yan Q, Zhongping H, Wei W, Gewei L, Xiaolei Y, Liying D, Lili R, Jianwei W, Xiong H, Taisheng L, Hongkui D, Mingxiao D. 2004. [Neutralizing Antibodies in Patients with Severe Acute Respiratory Syndrome-Associated Coronavirus Infection](#). The Journal of Infectious Diseases 190:1119–1126.
912. Prabakaran P, Zhu Z, Xiao X, Biragyn A, Dimitrov AS, Broder CC, Dimitrov DS. 2009. [Potent human monoclonal antibodies against SARS CoV, Nipah and Hendra viruses](#). Expert Opinion on Biological Therapy 9:355–368.
913. Sun C, Chen L, Yang J, Luo C, Zhang Y, Li J, Yang J, Zhang J, Xie L. 2020. SARS-CoV-2 and SARS-CoV Spike-RBD Structure and Receptor Binding Comparison and Potential Implications on Neutralizing Antibody and Vaccine Development. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.16.951723>.
914. Renn A, Fu Y, Hu X, Hall MD, Simeonov A. 2020. [Fruitful Neutralizing Antibody Pipeline Brings Hope To Defeat SARS-Cov-2](#). Trends in Pharmacological Sciences 41:815–829.
915. Pinto D, Park Y-J, Beltramello M, Walls AC, Tortorici MA, Bianchi S, Jaconi S, Culap K, Zatta F, De Marco A, Peter A, Guarino B, Spreafico R, Cameroni E, Case JB, Chen RE, Havenar-Daughton C, Snell G, Telenti A, Virgin HW, Lanzavecchia A, Diamond MS, Fink K, Veasler D, Corti D. 2020. [Cross-neutralization of SARS-CoV-2 by a human monoclonal SARS-CoV antibody](#). Nature 583:290–295.

916. Wang C, Li W, Drabek D, Okba NMA, van Haperen R, Osterhaus ADME, van Kuppeveld FJM, Haagmans BL, Grosveld F, Bosch B-J. 2020. A human monoclonal antibody blocking SARS-CoV-2 infection. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.11.987958>.
917. Deb P, Molla MdMA, Saif-Ur-Rahman KM. 2021. [An update to monoclonal antibody as therapeutic option against COVID-19](#). Biosafety and Health 3:87–91.
918. Jones BE, Brown-Augsburger PL, Corbett KS, Westendorf K, Davies J, Cujec TP, Wiethoff CM, Blackbourne JL, Heinz BA, Foster D, Higgs RE, Balasubramaniam D, Wang L, Bidshahri R, Kraft L, Hwang Y, Žentelis S, Jepson KR, Goya R, Smith MA, Collins DW, Hinshaw SJ, Tycho SA, Pellacani D, Xiang P, Muthuraman K, Sobhanifar S, Piper MH, Triana FJ, Hendle J, Pustilnik A, Adams AC, Berens SJ, Baric RS, Martinez DR, Cross RW, Geisbert TW, Borisevich V, Abiona O, Belli HM, de Vries M, Mohamed A, Dittmann M, Samanovic M, Mulligan MJ, Goldsmith JA, Hsieh C-L, Johnson NV, Wrapp D, McLellan JS, Barnhart BC, Graham BS, Mascola JR, Hansen CL, Falconer E. 2020. [LY-CoV555, a rapidly isolated potent neutralizing antibody, provides protection in a non-human primate model of SARS-CoV-2 infection](#). Cold Spring Harbor Laboratory.
919. Eli Lilly and Company. 2021. [A Randomized, Placebo-Controlled, Double-Blind, Sponsor Unblinded, Single Ascending Dose, Phase 1 First in Human Study to Evaluate the Safety, Tolerability, Pharmacokinetics and Pharmacodynamics of Intravenous LY3819253 in Participants Hospitalized for COVID-19](#). NCT04411628. Clinical trial registration. clinicaltrials.gov.
920. Eli Lilly and Company. 2021. [A Phase 1, Randomized, Placebo-Controlled Study to Evaluate the Tolerability, Safety, Pharmacokinetics, and Immunogenicity of LY3832479 Given as a Single Intravenous Dose in Healthy Participants](#). NCT04441931. Clinical trial registration. clinicaltrials.gov.
921. Gottlieb RL, Nirula A, Chen P, Boscia J, Heller B, Morris J, Huhn G, Cardona J, Mocherla B, Stosor V, Shawa I, Kumar P, Adams AC, Van Naarden J, Custer KL, Durante M, Oakley G, Schade AE, Holzer TR, Ebert PJ, Higgs RE, Kallewaard NL, Sabo J, Patel DR, Klekotka P, Shen L, Skovronsky DM. 2021. [Effect of Bamlanivimab as Monotherapy or in Combination With Etesevimab on Viral Load in Patients With Mild to Moderate COVID-19](#). JAMA 325:632.
922. Eli Lilly and Company. 2022. [A Randomized, Double-blind, Placebo-Controlled, Phase 2/3 Study to Evaluate the Efficacy and Safety of LY3819253 and LY3832479 in Participants With Mild to Moderate COVID-19 Illness](#). NCT04427501. Clinical trial registration. clinicaltrials.gov.
923. Chen P, Nirula A, Heller B, Gottlieb RL, Boscia J, Morris J, Huhn G, Cardona J, Mocherla B, Stosor V, Shawa I, Adams AC, Van Naarden J, Custer KL, Shen L, Durante M, Oakley G, Schade AE, Sabo J, Patel DR, Klekotka P, Skovronsky DM. 2021. [SARS-CoV-2 Neutralizing Antibody LY-CoV555 in Outpatients with Covid-19](#). N Engl J Med 384:229–237.

924. 2021. Bamlanivimab and Etesevimab EUA Letter of Authorization. US Food and Drug Administration.
<https://www.fda.gov/media/145801/download>.
925. Hansen J, Baum A, Pascal KE, Russo V, Giordano S, Wloga E, Fulton BO, Yan Y, Koon K, Patel K, Chung KM, Hermann A, Ullman E, Cruz J, Rafique A, Huang T, Fairhurst J, Libertiny C, Malbec M, Lee W, Welsh R, Farr G, Pennington S, Deshpande D, Cheng J, Watty A, Bouffard P, Babb R, Levenkova N, Chen C, Zhang B, Romero Hernandez A, Saotome K, Zhou Y, Franklin M, Sivapalasingam S, Lye DC, Weston S, Logue J, Haupt R, Frieman M, Chen G, Olson W, Murphy AJ, Stahl N, Yancopoulos GD, Kyratsous CA. 2020. [Studies in humanized mice and convalescent humans yield a SARS-CoV-2 antibody cocktail](#). Science 369:1010–1014.
926. Regeneron Pharmaceuticals. 2022. [A Master Protocol Assessing the Safety, Tolerability, and Efficacy of Anti-Spike \(S\) SARS-CoV-2 Monoclonal Antibodies for the Treatment of Hospitalized Patients With COVID-19](#). NCT04426695. Clinical trial registration. clinicaltrials.gov.
927. Regeneron Pharmaceuticals. 2022. [A Master Protocol Assessing the Safety, Tolerability, and Efficacy of Anti-Spike \(S\) SARS-CoV-2 Monoclonal Antibodies for the Treatment of Ambulatory Patients With COVID-19](#). NCT04425629. Clinical trial registration. clinicaltrials.gov.
928. Weinreich DM, Sivapalasingam S, Norton T, Ali S, Gao H, Bhore R, Musser BJ, Soo Y, Rofail D, Im J, Perry C, Pan C, Hosain R, Mahmood A, Davis JD, Turner KC, Hooper AT, Hamilton JD, Baum A, Kyratsous CA, Kim Y, Cook A, Kampman W, Kohli A, Sachdeva Y, Graber X, Kowal B, DiCioccio T, Stahl N, Lipsich L, Braunstein N, Herman G, Yancopoulos GD. 2021. [REGN-COV2, a Neutralizing Antibody Cocktail, in Outpatients with Covid-19](#). N Engl J Med 384:238–251.
929. Commissioner O of the. 2020. Coronavirus (COVID-19) Update: FDA Authorizes Monoclonal Antibodies for Treatment of COVID-19. FDA.
<https://www.fda.gov/news-events/press-announcements/coronavirus-covid-19-update-fda-authorizes-monoclonal-antibodies-treatment-covid-19>. Retrieved 5 December 2022.
930. Traggiai E, Becker S, Subbarao K, Kolesnikova L, Uematsu Y, Gismondo MR, Murphy BR, Rappuoli R, Lanzavecchia A. 2004. [An efficient method to make human monoclonal antibodies from memory B cells: potent neutralization of SARS coronavirus](#). Nat Med 10:871–875.
931. Dolgin E. 2021. [‘Super-antibodies’ could curb COVID-19 and help avert future pandemics](#). Nat Biotechnol 39:783–785.
932. Walker LM, Burton DR. 2018. [Passive immunotherapy of viral infections: ‘super-antibodies’ enter the fray](#). Nat Rev Immunol 18:297–308.
933. Vir Biotechnology, Inc. 2022. [A Phase II/III Randomized, Multi-center, Double-blind, Placebo-controlled Study to Assess the Safety and Efficacy of Monoclonal Antibody VIR-7831 for the Early Treatment of Coronavirus Disease 2019 \(COVID-19\) in Non-hospitalized Patients](#). NCT04545060. Clinical trial registration. clinicaltrials.gov.

934. Gupta A, Gonzalez-Rojas Y, Juarez E, Casal MC, Moya J, Falci DR, Sarkis E, Solis J, Zheng H, Scott N, Cathcart AL, Hebner CM, Sager J, Mogalian E, Tipple C, Peppercorn A, Alexander E, Pang PS, Free A, Brinson C, Aldinger M, Shapiro AE. 2021. [Early Covid-19 Treatment With SARS-CoV-2 Neutralizing Antibody Sotrovimab](#). Cold Spring Harbor Laboratory.
935. Liu Z, VanBlargan LA, Bloyet L-M, Rothlauf PW, Chen RE, Stumpf S, Zhao H, Errico JM, Theel ES, Liebeskind MJ, Alford B, Buchser WJ, Ellebedy AH, Fremont DH, Diamond MS, Whelan SPJ. 2021. [Identification of SARS-CoV-2 spike mutations that attenuate monoclonal and serum antibody neutralization](#). Cell Host & Microbe 29:477–488.e4.
936. Diamond M, Chen R, Xie X, Case J, Zhang X, VanBlargan L, Liu Y, Liu J, Errico J, Winkler E, Suryadevara N, Tahan S, Turner J, Kim W, Schmitz A, Thapa M, Wang D, Boon A, Pinto D, Presti R, O'Halloran J, Kim A, Deepak P, Fremont D, Corti D, Virgin H, Crowe J, Droit L, Ellebedy A, Shi P-Y, Gilchuk P. 2021. [SARS-CoV-2 variants show resistance to neutralization by many monoclonal and serum-derived polyclonal antibodies](#). Research Square Platform LLC.
937. Graham C, Seow J, Huettner I, Khan H, Kouphou N, Acors S, Winstone H, Pickering S, Pedro Galao R, Jose Lista M, Jimenez-Guardeno JM, Laing AG, Wu Y, Joseph M, Muir L, Ng WM, Duyvesteyn HME, Zhao Y, Bowden TA, Shankar-Hari M, Rosa A, Cherepanov P, McCoy LE, Hayday AC, Neil SJD, Malim MH, Doores KJ. 2021. [Impact of the B.1.1.7 variant on neutralizing monoclonal antibodies recognizing diverse epitopes on SARS-CoV-2 Spike](#). Cold Spring Harbor Laboratory.
938. Wang P, Nair MS, Liu L, Iketani S, Luo Y, Guo Y, Wang M, Yu J, Zhang B, Kwong PD, Graham BS, Mascola JR, Chang JY, Yin MT, Sobieszczyk M, Kyratsous CA, Shapiro L, Sheng Z, Huang Y, Ho DD. 2021. [Antibody Resistance of SARS-CoV-2 Variants B.1.351 and B.1.1.7](#). Cold Spring Harbor Laboratory.
939. 2021. Pause in the Distribution of bamlanivimab/etesevimab. US Department of Health and Human Services. <https://www.phe.gov/emergency/events/COVID19/investigation-MCM/Bamlanivimab-etesevimab/Pages/bamlanivimab-etesevimab-distribution-pause.aspx>.
940. Sun Z-YJ, Oh KJ, Kim M, Yu J, Brusic V, Song L, Qiao Z, Wang J, Wagner G, Reinherz EL. 2008. [HIV-1 Broadly Neutralizing Antibody Extracts Its Epitope from a Kinked gp41 Ectodomain Region on the Viral Membrane](#). Immunity 28:52–63.
941. Ekiert DC, Bhabha G, Elsliger M-A, Friesen RHE, Jongeneelen M, Throsby M, Goudsmit J, Wilson IA. 2009. [Antibody Recognition of a Highly Conserved Influenza Virus Epitope](#). Science 324:246–251.
942. He Y, Li J, Du L, Yan X, Hu G, Zhou Y, Jiang S. 2006. [Identification and characterization of novel neutralizing epitopes in the receptor-binding domain of SARS-CoV spike protein: Revealing the critical antigenic determinants in inactivated SARS-CoV vaccine](#). Vaccine 24:5498–5508.

943. Rockx B, Donaldson E, Frieman M, Sheahan T, Corti D, Lanzavecchia A, Baric Ralph S. 2010. [Escape from Human Monoclonal Antibody Neutralization Affects In Vitro and In Vivo Fitness of Severe Acute Respiratory Syndrome Coronavirus](#). J INFECT DIS 201:946–955.
944. Stanford Coronavirus Antiviral & Resistance Database (CoVDB). <https://covdb.stanford.edu/page/susceptibility-data>. Retrieved 5 December 2022.
945. Rappazzo CG, Tse LV, Kaku CI, Wrapp D, Sakharkar M, Huang D, Deveau LM, Yockachonis TJ, Herbert AS, Battles MB, O'Brien CM, Brown ME, Geoghegan JC, Belk J, Peng L, Yang L, Hou Y, Scobey TD, Burton DR, Nemazee D, Dye JM, Voss JE, Gunn BM, McLellan JS, Baric RS, Gralinski LE, Walker LM. 2021. [Broad and potent activity against SARS-like viruses by an engineered human monoclonal antibody](#). Science 371:823–829.
946. Parvathaneni V, Kulkarni NS, Muth A, Gupta V. 2019. [Drug repurposing: a promising tool to accelerate the drug discovery process](#). Drug Discovery Today 24:2076–2085.
947. Zheng W, Sun W, Simeonov A. 2017. [Drug repurposing screens and synergistic drug-combinations for infectious diseases](#). British Journal of Pharmacology 175:181–191.
948. Muratov EN, Amaro R, Andrade CH, Brown N, Ekins S, Fourches D, Isayev O, Kozakov D, Medina-Franco JL, Merz KM, Oprea TI, Poroikov V, Schneider G, Todd MH, Varnek A, Winkler DA, Zakharov AV, Cherkasov A, Tropsha A. 2021. [A critical overview of computational approaches employed for COVID-19 drug discovery](#). Chem Soc Rev 50:9121–9151.
949. Law GL, Tisoncik-Go J, Korth MJ, Katze MG. 2013. [Drug repurposing: a better approach for infectious disease drug discovery?](#) Current Opinion in Immunology 25:588–592.
950. Pereira DA, Williams JA. 2007. [Origin and evolution of high throughput screening](#). British Journal of Pharmacology 152:53–61.
951. Vincent F, Loria P, Pregel M, Stanton R, Kitching L, Nocka K, Doyonnas R, Steppan C, Gilbert A, Schroeter T, Peakman M-C. 2015. [Developing predictive assays: The phenotypic screening “rule of 3”](#). Sci Transl Med 7.
952. Swinney DC. 2013. [Phenotypic vs. Target-Based Drug Discovery for First-in-Class Medicines](#). Clin Pharmacol Ther 93:299–301.
953. Moffat JG, Rudolph J, Bailey D. 2014. [Phenotypic screening in cancer drug discovery — past, present and future](#). Nat Rev Drug Discov 13:588–602.
954. Diversity Libraries. ChemBridge. <https://chembridge.com/diversity-and-pre-plated-libraries/diversity-libraries/>. Retrieved 5 December 2022.
955. Wagner Bridget K, Schreiber Stuart L. 2016. [The Power of Sophisticated Phenotypic Screening and Modern Mechanism-of-Action Methods](#). Cell

956. Mercorelli B, Palù G, Loregian A. 2018. [Drug Repurposing for Viral Infectious Diseases: How Far Are We?](#) Trends in Microbiology 26:865–876.
957. Challa AP, Lavieri RR, Lewis JT, Zaleski NM, Shirey-Rice JK, Harris PA, Aronoff DM, Pulley JM. 2019. [Systematically Prioritizing Candidates in Genome-Based Drug Repurposing](#). ASSAY and Drug Development Technologies 17:352–363.
958. Edwards A. 2020. [What Are the Odds of Finding a COVID-19 Drug from a Lab Repurposing Screen?](#) J Chem Inf Model 60:5727–5729.
959. Kido H, Okumura Y, Yamada H, Quang Le T, Yano M. 2007. [Proteases Essential for Human Influenza Virus Entry into Cells and Their Inhibitors as Potential Therapeutic Agents](#). Current Pharmaceutical Design 13:405–414.
960. Zhou Y, Vedantham P, Lu K, Agudelo J, Carrion R, Nunneley JW, Barnard D, Pöhlmann S, McKerrow JH, Renslo AR, Simmons G. 2015. [Protease inhibitors targeting coronavirus and filovirus entry](#). Antiviral Research 116:76–84.
961. Jin Z, Du X, Xu Y, Deng Y, Liu M, Zhao Y, Zhang B, Li X, Zhang L, Peng C, Duan Y, Yu J, Wang L, Yang K, Liu F, Jiang R, Yang X, You T, Liu X, Yang X, Bai F, Liu H, Liu X, Guddat LW, Xu W, Xiao G, Qin C, Shi Z, Jiang H, Rao Z, Yang H. 2020. [Structure of Mpro from SARS-CoV-2 and discovery of its inhibitors](#). Nature 582:289–293.
962. Yang H, Xie W, Xue X, Yang K, Ma J, Liang W, Zhao Q, Zhou Z, Pei D, Ziebuhr J, Hilgenfeld R, Yuen KY, Wong L, Gao G, Chen S, Chen Z, Ma D, Bartlam M, Rao Z. 2005. [Design of Wide-Spectrum Inhibitors Targeting Coronavirus Main Proteases](#). PLoS Biology 3:e324.
963. Ren Z, Yan L, Zhang N, Guo Y, Yang C, Lou Z, Rao Z. 2013. [The newly emerged SARS-Like coronavirus HCoV-EMC also has an “Achilles’ heel”: current effective inhibitor targeting a 3C-like protease](#). Protein & Cell 4:248–250.
964. Wang F, Chen C, Tan W, Yang K, Yang H. 2016. [Structure of Main Protease from Human Coronavirus NL63: Insights for Wide Spectrum Anti-Coronavirus Drug Design](#). Sci Rep 6.
965. Xue X, Yu H, Yang H, Xue F, Wu Z, Shen W, Li J, Zhou Z, Ding Y, Zhao Q, Zhang XC, Liao M, Bartlam M, Rao Z. 2008. [Structures of Two Coronavirus Main Proteases: Implications for Substrate Binding and Antiviral Drug Design](#). J Virol 82:2515–2527.
966. Azad GK, Tomar RS. 2014. [Ebselen, a promising antioxidant drug: mechanisms of action and targets of biological pathways](#). Molecular Biology Reports 41:4865–4879.
967. Menéndez CA, Byléhn F, Perez-Lemus GR, Alvarado W, de Pablo JJ. 2020. [Molecular characterization of ebselen binding activity to SARS-CoV-2 main protease](#). Sci Adv 6.

968. Chen Z, Jiang Z, Chen N, Shi Q, Tong L, Kong F, Cheng X, Chen H, Wang C, Tang B. 2018. [Target discovery of ebselen with a biotinylated probe](#). Chemical Communications 54:9506–9509.
969. FDA Clears SPI's Ebselen for Phase II COVID-19 Trials. Contract Pharma. https://www.contractpharma.com/contents/view_breaking-news/2020-08-31/fda-clears-spis-ebselen-for-phase-ii-covid-19-trials/. Retrieved 5 December 2022.
970. Sound Pharmaceuticals, Incorporated. 2022. [A Phase 2, Randomized, Double-Blind, Placebo-Controlled, Dose Escalation Study to Evaluate the Safety and Efficacy of SPI-1005 in Moderate COVID-19 Patients](#). NCT04484025. Clinical trial registration. clinicaltrials.gov.
971. Sound Pharmaceuticals, Incorporated. 2022. [A Phase 2, Randomized, Double-Blind, Placebo-Controlled, Dose Escalation Study to Evaluate the Safety and Efficacy of SPI-1005 in Severe COVID-19 Patients](#). NCT04483973. Clinical trial registration. clinicaltrials.gov.
972. Pfizer. 2021. [A PHASE 1B, 2-PART, DOUBLE-BLIND, PLACEBO-CONTROLLED, SPONSOR-OPEN STUDY, TO EVALUATE THE SAFETY, TOLERABILITY AND PHARMACOKINETICS OF SINGLE ASCENDING \(24-HOUR, PART 1\) AND MULTIPLE ASCENDING \(120-HOUR, PART 2\) INTRAVENOUS INFUSIONS OF PF-07304814 IN HOSPITALIZED PARTICIPANTS WITH COVID-19](#). NCT04535167. Clinical trial registration. clinicaltrials.gov.
973. Pfizer. 2022. [AN INTERVENTIONAL EFFICACY AND SAFETY, PHASE 2/3, DOUBLE-BLIND, 2-ARM STUDY TO INVESTIGATE ORALLY ADMINISTERED PF-07321332/RITONAVIR COMPARED WITH PLACEBO IN NONHOSPITALIZED SYMPTOMATIC ADULT PARTICIPANTS WITH COVID-19 WHO ARE AT INCREASED RISK OF PROGRESSING TO SEVERE ILLNESS](#). NCT04960202. Clinical trial registration. clinicaltrials.gov.
974. Qian T, Zhu S, Hoshida Y. 2019. [Use of big data in drug development for precision medicine: an update](#). Expert Review of Precision Medicine and Drug Development 4:189–200.
975. Kuleshov MV, Stein DJ, Clarke DJB, Kropiwnicki E, Jagodnik KM, Bartal A, Evangelista JE, Hom J, Cheng M, Bailey A, Zhou A, Ferguson LB, Lachmann A, Ma'ayan A. 2020. [The COVID-19 Drug and Gene Set Library](#). Patterns 1:100090.
976. Sadegh S, Matschinske J, Blumenthal DB, Galindez G, Kacprowski T, List M, Nasirigerdeh R, Oubounyt M, Pichlmair A, Rose TD, Salgado-Albarrán M, Späth J, Stukalov A, Wenke NK, Yuan K, Pauling JK, Baumbach J. 2020. [Exploring the SARS-CoV-2 virus-host-drug interactome for drug repurposing](#). Nat Commun 11.
977. Zhou Y, Wang F, Tang J, Nussinov R, Cheng F. 2020. [Artificial intelligence in COVID-19 drug repurposing](#). The Lancet Digital Health 2:e667–e676.
978. Maurice T, Su T-P. 2009. [The pharmacology of sigma-1 receptors](#). Pharmacology & Therapeutics 124:195–206.

979. Gordon DE, Hiatt J, Bouhaddou M, Rezelj VV, Ulferts S, Braberg H, Jureka AS, Obernier K, Guo JZ, Batra J, others. 2020. [Comparative host-coronavirus protein interaction networks reveal pan-viral disease mechanisms](#). *Science* 370:eabe9403.
980. Vela JM. 2020. [Repurposing Sigma-1 Receptor Ligands for COVID-19 Therapy?](#) *Front Pharmacol* 11.
981. Hayashi T, Su T-. 2005. [The Sigma Receptor: Evolution of the Concept in Neuropsychopharmacology](#). *CN* 3:267–280.
982. Tummino TA, Rezelj VV, Fischer B, Fischer A, O'Meara MJ, Monel B, Vallet T, White KM, Zhang Z, Alon A, Schadt H, O'Donnell HR, Lyu J, Rosales R, McGovern BL, Rathnasinghe R, Jangra S, Schotsaert M, Galarneau J-R, Krogan NJ, Urban L, Shokat KM, Kruse AC, García-Sastre A, Schwartz O, Moretti F, Vignuzzi M, Pognan F, Shoichet BK. 2021. [Drug-induced phospholipidosis confounds drug repurposing for SARS-CoV-2](#). *Science* 373:541–547.
983. Breiden B, Sandhoff K. 2019. [Emerging mechanisms of drug-induced phospholipidosis](#). *Biological Chemistry* 401:31–46.
984. Obeidat M, Isaacson AL, Chen SJ, Ivanovic M, Holanda D. 2020. [Zebra-like bodies in COVID-19: is phospholipidosis evidence of hydroxychloroquine induced acute kidney injury?](#) *Ultrastructural Pathology* 44:519–523.
985. Dittmar M, Lee JS, Whig K, Segrist E, Li M, Kamalia B, Castellana L, Ayyanathan K, Cardenas-Diaz FL, Morrissey EE, Truitt R, Yang W, Jurado K, Samby K, Ramage H, Schultz DC, Cherry S. 2021. [Drug repurposing screens reveal cell-type-specific entry pathways and FDA-approved drugs active against SARS-Cov-2](#). *Cell Reports* 35:108959.
986. Edwards A, Hartung IV. 2021. [No shortcuts to SARS-CoV-2 antivirals](#). *Science* 373:488–489.
987. Hashimoto K. 2021. [Repurposing of CNS drugs to treat COVID-19 infection: targeting the sigma-1 receptor](#). *Eur Arch Psychiatry Clin Neurosci* 271:249–258.
988. Lenze EJ, Mattar C, Zorumski CF, Stevens A, Schweiger J, Nicol GE, Miller JP, Yang L, Yingling M, Avidan MS, Reiersen AM. 2020. [Fluvoxamine vs Placebo and Clinical Deterioration in Outpatients With Symptomatic COVID-19](#). *JAMA* 324:2292.
989. Seftel D, Boulware DR. 2021. [Prospective Cohort of Fluvoxamine for Early Treatment of Coronavirus Disease 19](#). *Open Forum Infectious Diseases* 8.
990. Rosen DA, Seki SM, Fernández-Castañeda A, Beiter RM, Eccles JD, Woodfolk JA, Gaultier A. 2019. [Modulation of the sigma-1 receptor–IRE1 pathway is beneficial in preclinical models of inflammation and sepsis](#). *Sci Transl Med* 11.
991. Sukhatme VP, Reiersen AM, Vayttaden SJ, Sukhatme VV. 2021. [Fluvoxamine: A Review of Its Mechanism of Action and Its Role in](#)

992. Lowe D. 2021. Too Many Papers. In the Pipeline. <https://blogs.sciencemag.org/pipeline/archives/2021/07/19/too-many-papers>.
993. Prinz F, Schlange T, Asadullah K. 2011. [Believe it or not: how much can we rely on published data on potential drug targets?](#) Nat Rev Drug Discov 10:712–712.
994. Swinney DC, Anthony J. 2011. [How were new medicines discovered?](#) Nat Rev Drug Discov 10:507–519.
995. Kupferschmidt K. 2020. [Big studies dim hopes for hydroxychloroquine](#). Science 368:1166–1167.
996. Bugin K, Woodcock J. 2021. [Trends in COVID-19 therapeutic clinical trials](#). Nat Rev Drug Discov 20:254–255.
997. Dron L, Dillman A, Zoratti MJ, Haggstrom J, Mills EJ, Park JJH. 2021. [Clinical Trial Data Sharing for COVID-19–Related Research](#). J Med Internet Res 23:e26718.
998. Lee Z, Rayner CR, Forrest JI, Nachega JB, Senchaudhuri E, Mills EJ. 2021. [The Rise and Fall of Hydroxychloroquine for the Treatment and Prevention of COVID-19](#). The American Journal of Tropical Medicine and Hygiene 104:35–38.
999. Park JJH, Dron L, Mills EJ. 2021. [Moving forward in clinical research with master protocols](#). Contemporary Clinical Trials 106:106438.
1000. 2020. Main protease structure and XChem fragment screen. Diamond. <https://www.diamond.ac.uk/covid-19/for-scientists/Main-protease-structure-and-XChem.html>.
1001. Barnes MA, Carson MJ, Nair MG. 2015. [Non-traditional cytokines: How catecholamines and adipokines influence macrophages in immunity, metabolism and the central nervous system](#). Cytokine 72:210–219.
1002. ELENKOV IJ, CHROUSOS GP. 2002. [Stress Hormones, Proinflammatory and Antiinflammatory Cytokines, and Autoimmunity](#). Annals of the New York Academy of Sciences 966:290–303.
1003. García A, Martí O, Vallès A, Dal-Zotto S, Armario A. 2000. [Recovery of the Hypothalamic-Pituitary-Adrenal Response to Stress](#). Neuroendocrinology 72:114–125.
1004. Elenkov IJ, Papanicolaou DA, Wilder RL, Chrousos GP. 1996. [Modulatory effects of glucocorticoids and catecholamines on human interleukin-12 and interleukin-10 production: clinical implications](#). Proc Assoc Am Physicians 108:374–81.
1005. Theoharides TC. 2020. [Dexamethasone for COVID-19? Not so fast](#). JOURNAL OF BIOLOGICAL REGULATORS AND HOMEOSTATIC AGENTS 34.

1006. Matthay MA, Thompson BT. 2020. [Dexamethasone in hospitalised patients with COVID-19: addressing uncertainties](#). The Lancet Respiratory Medicine 8:1170–1172.
1007. Brotherton H, Usuf E, Nadjm B, Forrest K, Bojang K, Samateh AL, Bittaye M, Roberts CA, d'Alessandro U, Roca A. 2020. [Dexamethasone for COVID-19: data needed from randomised clinical trials in Africa](#). The Lancet Global Health 8:e1125–e1126.
1008. Cai Q, Yang M, Liu D, Chen J, Shu D, Xia J, Liao X, Gu Y, Cai Q, Yang Y, Shen C, Li X, Peng L, Huang D, Zhang J, Zhang S, Wang F, Liu J, Chen L, Chen S, Wang Z, Zhang Z, Cao R, Zhong W, Liu Y, Liu L. 2020. [Experimental Treatment with Favipiravir for COVID-19: An Open-Label Control Study](#). Engineering 6:1192–1198.
1009. Horby PW, Mafham M, Bell JL, Linsell L, Staplin N, Emberson J, Palfreeman A, Raw J, Elmahi E, Prudon B, Green C, Carley S, Chadwick D, Davies M, Wise MP, Baillie JK, Chappell LC, Faust SN, Jaki T, Jefferey K, Lim WS, Montgomery A, Rowan K, Juszczak E, Haynes R, Landray MJ. 2020. [Lopinavir–ritonavir in patients admitted to hospital with COVID-19 \(RECOVERY\): a randomised, controlled, open-label, platform trial](#). The Lancet 396:1345–1352.
1010. Harrington DP, Baden LR, Hogan JW. 2020. [A Large, Simple Trial Leading to Complex Questions](#). New England Journal of Medicine NEJMe2034294.
1011. 2020. Retracted coronavirus (COVID-19) papers. Retraction Watch. <https://retractionwatch.com/retracted-coronavirus-covid-19-papers/>. Retrieved 8 February 2021.
1012. Lou Y, Liu L, Yao H, Hu X, Su J, Xu K, Luo R, Yang X, He L, Lu X, Zhao Q, Liang T, Qiu Y. 2021. [Clinical Outcomes and Plasma Concentrations of Baloxavir Marboxil and Favipiravir in COVID-19 Patients: An Exploratory Randomized, Controlled Trial](#). European Journal of Pharmaceutical Sciences 157:105631.
1013. Dabbous HM, Abd-Elsalam S, El-Sayed MH, Sherief AF, Ebeid FFS, El Ghafar MSA, Soliman S, Elbahnasawy M, Badawi R, Tageldin MA. 2021. Efficacy of favipiravir in COVID-19 treatment: a multi-center randomized study. Archives of Virology <https://doi.org/10.1007/s00705-021-04956-9>.
1014. Ivashchenko AA, Dmitriev KA, Vostokova NV, Azarova VN, Blinow AA, Egorova AN, Gordeev IG, Ilin AP, Karapetian RN, Kravchenko DV, Lomakin NV, Merkulova EA, Papazova NA, Pavlikova EP, Savchuk NP, Simakina EN, Sitdekov TA, Smolyarchuk EA, Tikhomolova EG, Yakubova EV, Ivachtchenko AV. 2020. [AVIFAVIR for Treatment of Patients With Moderate Coronavirus Disease 2019 \(COVID-19\): Interim Results of a Phase II/III Multicenter Randomized Clinical Trial](#). Clinical Infectious Diseases ciaa1176.
1015. Pilkington V, Pepperrell T, Hill A. 2020. [A review of the safety of favipiravir – a potential treatment in the COVID-19 pandemic?](#) Journal of Virus Eradication 6:45–51.

1016. Mulangu S, Dodd LE, Davey RT, Tshiani Mbaya O, Proschan M, Mukadi D, Lusakibanza Manzo M, Nzolo D, Tshomba Oloma A, Ibanda A, Ali R, Coulibaly S, Levine AC, Grais R, Diaz J, Lane HC, Muyembe-Tamfum J-J, the PALM Writing Group. 2019. [A Randomized, Controlled Trial of Ebola Virus Disease Therapeutics](#). New England Journal of Medicine 381:2293–2303.
1017. Sheahan TP, Sims AC, Graham RL, Menachery VD, Gralinski LE, Case JB, Leist SR, Pyrc K, Feng JY, Trantcheva I, Bannister R, Park Y, Babusis D, Clarke MO, Mackman RL, Spahn JE, Palmiotti CA, Siegel D, Ray AS, Cihlar T, Jordan R, Denison MR, Baric RS. 2017. [Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses](#). Science Translational Medicine 9:eaal3653.
1018. Cohen J. 2020. Did an experimental drug help a U.S. coronavirus patient? Science <https://doi.org/10.1126/science.abb7243>.
1019. Kujawski SA, Wong KK, Collins JP, Epstein L, Killerby ME, Midgley CM, Abedi GR, Ahmed NS, Almendares O, Alvarez FN, Anderson KN, Balter S, Barry V, Bartlett K, Beer K, Ben-Aderet MA, Benowitz I, Biggs H, Binder AM, Black SR, Bonin B, Brown CM, Bruce H, Bryant-Genevier J, Budd A, Buell D, Bystritsky R, Cates J, Charles EM, Chatham-Stephens K, Chea N, Chiou H, Christiansen D, Chu V, Cody S, Cohen M, Conners E, Curns A, Dasari V, Dawson P, DeSalvo T, Diaz G, Donahue M, Donovan S, Duca LM, Erickson K, Esona MD, Evans S, Falk J, Feldstein LR, Fenstersheib M, Fischer M, Fisher R, Foo C, Fricchione MJ, Friedman O, Fry AM, Galang RR, Garcia MM, Gerber SI, Gerrard G, Ghinai I, Gounder P, Grein J, Grigg C, Gunzenhauser JD, Gutkin GI, Haddix M, Hall AJ, Han G, Harcourt J, Harriman K, Haupt T, Haynes A, Holshue M, Hoover C, Hunter JC, Jacobs MW, Jarashow C, Jhung MA, Joshi K, Kamali T, Kamili S, Kim L, Kim M, King J, Kirking HL, Kita-Yarbro A, Klos R, Kobayashi M, Kocharian A, Komatsu KK, Koppaka R, Layden JE, Li Y, Lindquist S, Lindstrom S, Link-Gelles R, Lively J, Livingston M, Lo K, Lo J, Lu X, Lynch B, Madoff L, Malapati L, Marks G, Marlow M, Mathisen GE, McClung N, McGovern O, McPherson TD, Mehta M, Meier A, Mello L, Moon S, Morgan M, Moro RN, Murray J, Murthy R, Novosad S, Oliver SE, O'Shea J, Pacilli M, Paden CR, Pallansch MA, Patel M, Patel S, Pedraza I, Pillai SK, Pindyck T, Pray I, Queen K, Quick N, Reese H, Rha B, Rhodes H, Robinson S, Robinson P, Rolfes M, Routh J, Rubin R, Rudman SL, Sakthivel SK, Scott S, Shepherd C, Shetty V, Smith EA, Smith S, Stierman B, Stoecker W, Sunenshine R, Sy-Santos R, Tamin A, Tao Y, Terashita D, Thornburg NJ, Tong S, Traub E, Tural A, Uehara A, Uyeki TM, Vahey G, Verani JR, Villarino E, Wallace M, Wang L, Watson JT, Westercamp M, Whitaker B, Wilkerson S, Woodruff RC, Wortham JM, Wu T, Xie A, Yousaf A, Zahn M, Zhang J, The COVID-19 Investigation Team. 2020. First 12 patients with coronavirus disease 2019 (COVID-19) in the United States. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.09.20032896>.
1020. Holshue ML, DeBolt C, Lindquist S, Lofy KH, Wiesman J, Bruce H, Spitters C, Ericson K, Wilkerson S, Tural A, Diaz G, Cohn A, Fox L, Patel A, Gerber SI, Kim L, Tong S, Lu X, Lindstrom S, Pallansch MA, Weldon WC, Biggs HM, Uyeki TM, Pillai SK. 2020. [First Case of 2019 Novel](#)

[Coronavirus in the United States](#). New England Journal of Medicine 382:929–936.

1021. Goldman JD, Lye DCB, Hui DS, Marks KM, Bruno R, Montejano R, Spinner CD, Galli M, Ahn M-Y, Nahass RG, Chen Y-S, SenGupta D, Hyland RH, Osinusi AO, Cao H, Blair C, Wei X, Gaggar A, Brainard DM, Towner WJ, Muñoz J, Mullane KM, Marty FM, Tashima KT, Diaz G, Subramanian A. 2020. [Remdesivir for 5 or 10 Days in Patients with Severe Covid-19](#). New England Journal of Medicine 383:1827–1837.
1022. Hinton DM. 2020. Remdesivir EUA Letter of Authorization. <https://www.fda.gov/media/137564/download>.
1023. Gilead Sciences. 2021. [A Phase 3 Randomized Study to Evaluate the Safety and Antiviral Activity of Remdesivir \(GS-5734™\) in Participants With Moderate COVID-19 Compared to Standard of Care Treatment](#). NCT04292730. Clinical trial registration. clinicaltrials.gov.
1024. 2020. Multi-centre, adaptive, randomized trial of the safety and efficacy of treatments of COVID-19 in hospitalized adults. EU Clinical Trials Register. <https://www.clinicaltrialsregister.eu/ctr-search/trial/2020-000936-23/FR>.
1025. A Trial of Remdesivir in Adults With Mild and Moderate COVID-19 - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04252664>. Retrieved 8 February 2021.
1026. Cao B. 2020. [A Phase 3 Randomized, Double-blind, Placebo-controlled, Multicenter Study to Evaluate the Efficacy and Safety of Remdesivir in Hospitalized Adult Patients With Severe COVID-19](#). NCT04257656. Clinical trial registration. clinicaltrials.gov.
1027. Commissioner O of the. 2020. FDA Approves First Treatment for COVID-19. FDA. <https://www.fda.gov/news-events/press-announcements/fda-approves-first-treatment-covid-19>. Retrieved 8 February 2021.
1028. Gilead Sciences Statement on the Solidarity Trial. <https://www.gilead.com/news-and-press/company-statements/gilead-sciences-statement-on-the-solidarity-trial>. Retrieved 8 February 2021.
1029. Galiuto L, Patrono C. 2020. [Conflicting results on the efficacy of remdesivir in hospitalized Covid-19 patients: comment on the Adaptive Covid-19 Treatment Trial](#). European Heart Journal 41:4387–4388.
1030. Cohen J, Kupferschmidt K. 2020. The ‘very, very bad look’ of remdesivir, the first FDA-approved COVID-19 drug. Science | AAAS. <https://www.sciencemag.org/news/2020/10/very-very-bad-look-remdesivir-first-fda-approved-covid-19-drug>. Retrieved 8 February 2021.
1031. Spinner CD, Gottlieb RL, Criner GJ, Arribas López JR, Cattelan AM, Soriano Viladomiu A, Ogbuagu O, Malhotra P, Mullane KM, Castagna A, Chai LYA, Roestenberg M, Tsang OTY, Bernasconi E, Le Turnier P, Chang S-C, SenGupta D, Hyland RH, Osinusi AO, Cao H, Blair C, Wang

- H, Gaggar A, Brainard DM, McPhail MJ, Bhagani S, Ahn MY, Sanyal AJ, Huhn G, Marty FM, for the GS-US-540-5774 Investigators. 2020. [Effect of Remdesivir vs Standard Care on Clinical Status at 11 Days in Patients With Moderate COVID-19](#). JAMA 324:1048.
1032. Kalil AC, Patterson TF, Mehta AK, Tomashek KM, Wolfe CR, Ghazaryan V, Marconi VC, Ruiz-Palacios GM, Hsieh L, Kline S, Tapson V, Iovine NM, Jain MK, Sweeney DA, El Sahly HM, Branche AR, Regalado Pineda J, Lye DC, Sandkovsky U, Luetkemeyer AF, Cohen SH, Finberg RW, Jackson PEH, Taiwo B, Paules CI, Arguinchona H, Erdmann N, Ahuja N, Frank M, Oh M, Kim E-S, Tan SY, Mularski RA, Nielsen H, Ponce PO, Taylor BS, Larson L, Roupahel NG, Saklawi Y, Cantos VD, Ko ER, Engemann JJ, Amin AN, Watanabe M, Billings J, Elie M-C, Davey RT, Burgess TH, Ferreira J, Green M, Makowski M, Cardoso A, de Bono S, Bonnett T, Proschan M, Deye GA, Dempsey W, Nayak SU, Dodd LE, Beigel JH. 2020. [Baricitinib plus Remdesivir for Hospitalized Adults with Covid-19](#). New England Journal of Medicine NEJMoa2031994.
1033. Hinton DM. 2020. Letter of Authorization: EUA for baricitinib (Olumiant), in combination with remdesivir (Veklury), for the treatment of suspected or laboratory confirmed coronavirus disease 2019 (COVID-19). Food and Drug Administration. <https://www.fda.gov/media/143822/download>.
1034. Ashfaq UA, Javed T, Rehman S, Nawaz Z, Riazuddin S. 2011. [Lysosomotropic agents as HCV entry inhibitors](#). Virology Journal 8:163.
1035. Müller-Calleja N, Manukyan D, Ruf W, Lackner K. 2016. [Mechanism of Action of Hydroxychloroquine in the Antiphospholipid Syndrome](#). Blood 128:5023–5023.
1036. Erkan D, Aguiar CL, Andrade D, Cohen H, Cuadrado MJ, Danowski A, Levy RA, Ortel TL, Rahman A, Salmon JE, Tektonidou MG, Willis R, Lockshin MD. 2014. [14th International Congress on Antiphospholipid Antibodies Task Force Report on Antiphospholipid Syndrome Treatment Trends](#). Autoimmunity Reviews 13:685–696.
1037. Wang T-F, Lim W. 2016. [What is the role of hydroxychloroquine in reducing thrombotic risk in patients with antiphospholipid antibodies?](#) Hematology 2016:714–716.
1038. Zhou D, Dai S-M, Tong Q. 2020. [COVID-19: a recommendation to examine the effect of hydroxychloroquine in preventing infection and progression](#). Journal of Antimicrobial Chemotherapy 75:1667–1670.
1039. Sperber K, Louie M, Kraus T, Proner J, Sapira E, Lin S, Stecher V, Mayer L. 1995. [Hydroxychloroquine treatment of patients with human immunodeficiency virus type 1](#). Clinical Therapeutics 17:622–636.
1040. Helal GK, Gad MA, Abd-Eliah MF, Eid MS. 2016. [Hydroxychloroquine augments early virological response to pegylated interferon plus ribavirin in genotype-4 chronic hepatitis C patients](#). Journal of Medical Virology 88:2170–2178.
1041. Coster WJ. 2013. [Making the Best Match: Selecting Outcome Measures for Clinical Trials and Outcome Studies](#). The American Journal of

1042. Molina JM, Delaugerre C, Le Goff J, Mela-Lima B, Ponscarne D, Goldwirt L, de Castro N. 2020. [No evidence of rapid antiviral clearance or clinical benefit with the combination of hydroxychloroquine and azithromycin in patients with severe COVID-19 infection](#). Médecine et Maladies Infectieuses 50:384.
1043. Chen Z, Hu J, Zhang Z, Jiang S, Han S, Yan D, Zhuang R, Hu B, Zhang Z. 2020. Efficacy of hydroxychloroquine in patients with COVID-19: results of a randomized clinical trial. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.22.20040758>.
1044. 2020. Therapeutic effect of hydroxychloroquine on novel coronavirus pneumonia (COVID-19). Chinese Clinical Trial Registry. <http://www.chictr.org.cn/showprojen.aspx?proj=48880>.
1045. CHEN Jun, LIU Danping, LIU Li, LIU Ping, XU Qingnian, XIA Lu, LING Yun, HUANG Dan, SONG Shuli, ZHANG Dandan, QIAN Zhiping, LI Tao, SHEN Yinzong, LU Hongzhou. 2020. [A pilot study of hydroxychloroquine in treatment of patients with common coronavirus disease-19 \(COVID-19\)](#). Journal of Zhejiang University (Medical Sciences) 49.
1046. Gao J, Tian Z, Yang X. 2020. [Breakthrough: Chloroquine phosphate has shown apparent efficacy in treatment of COVID-19 associated pneumonia in clinical studies](#). BioScience Trends 14:72–73.
1047. Yang N, Shen H-M. 2020. [Targeting the Endocytic Pathway and Autophagy Process as a Novel Therapeutic Strategy in COVID-19](#). International Journal of Biological Sciences 16:1724–1731.
1048. Zheng J. 2020. [SARS-CoV-2: an Emerging Coronavirus that Causes a Global Threat](#). International Journal of Biological Sciences 16:1678–1685.
1049. Mehra MR, Desai SS, Ruschitzka F, Patel AN. 2020. [RETRACTED: Hydroxychloroquine or chloroquine with or without a macrolide for treatment of COVID-19: a multinational registry analysis](#). The Lancet S0140673620311806.
1050. Mehra MR, Ruschitzka F, Patel AN. 2020. [Retraction—Hydroxychloroquine or chloroquine with or without a macrolide for treatment of COVID-19: a multinational registry analysis](#). The Lancet 395:1820.
1051. O’Laughlin JP, Mehta PH, Wong BC. 2016. [Life Threatening Severe QTc Prolongation in Patient with Systemic Lupus Erythematosus due to Hydroxychloroquine](#). Case Reports in Cardiology 2016:1–4.
1052. Roden DM. 2008. [Keep the QT interval: It is a reliable predictor of ventricular arrhythmias](#). Heart Rhythm 5:1213–1215.
1053. C.E.Lane J, Weaver J, Kostka K, Duarte-Salles T, Abrahao MTF, Alghoul H, Alser O, Alshammari TM, Biedermann P, Burn E, Casajust P, Conover M, Culhane AC, Davydov A, DuVall SL, Dymshyts D, Fernandez-Bertolin S, Fišter K, Hardin J, Hester L, Hripcsak G, Kent S, Khosla S, Kolovos S,

Lambert CG, van der Lei J, Londhe AA, Lynch KE, Makadia R, Margulis AV, Matheny ME, Mehta P, Morales DR, Morgan-Stewart H, Mosseveld M, Newby D, Nyberg F, Ostropolets A, Park RW, Prats-Urbe A, Rao GA, Reich C, Reps J, Rijnbeek P, Kumaran Sathappan SM, Schuemie M, Seager S, Sena A, Shoaibi A, Spotnitz M, Suchard MA, Swerdel J, Torre CO, Vizcaya D, Wen H, de Wilde M, You SC, Zhang L, Zhuk O, Ryan P, Prieto-Alhambra D. 2020. Safety of hydroxychloroquine, alone and in combination with azithromycin, in light of rapid wide-spread use for COVID-19: a multinational, network cohort and self-controlled case series study. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.04.08.20054551>.

1054. Silva Borba MG, Almeida Val FF, Sampaio VS, Araújo Alexandre MA, Melo GC, Brito M, Gomes Mourão MP, Brito-Sousa JD, Baía-da-Silva D, Farias Guerra MV, Abrahão Hajjar L, Pinto RC, Silva Balieiro AA, Naveca FG, Simão Xavier M, Salomão A, Siqueira AM, Schwarzbolt A, Rosa Croda JH, Nogueira ML, Sierra Romero GA, Bassat Q, Fontes CJ, Cláudio Albuquerque B, Daniel-Ribeiro CT, Monteiro WM, Guimarães Lacerda MV, CloroCovid-19 Team. 2020. Chloroquine diphosphate in two different dosages as adjunctive therapy of hospitalized patients with severe respiratory syndrome in the context of coronavirus (SARS-CoV-2) infection: Preliminary safety results of a randomized, double-blinded, phase IIb clinical trial (CloroCovid-19 Study). Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.04.07.20056424>.
1055. Howard J, Cohen E, Kounang N, Nyberg P. 2020. Heart risk concerns mount around use of chloroquine and hydroxychloroquine for Covid-19 treatment. CNN. <https://www.cnn.com/2020/04/13/health/chloroquine-risks-coronavirus-treatment-trials-study/index.html>.
1056. 2020. WHO Director-General's opening remarks at the media briefing on COVID-19. World Health Organization. <https://www.who.int/dg/speeches/detail/who-director-general-s-opening-remarks-at-the-media-briefing-on-covid-19---25-may-2020>.
1057. Tang W, Cao Z, Han M, Wang Z, Chen J, Sun W, Wu Y, Xiao W, Liu S, Chen E, Chen W, Wang X, Yang J, Lin J, Zhao Q, Yan Y, Xie Z, Li D, Yang Y, Liu L, Qu J, Ning G, Shi G, Xie Q. 2020. Hydroxychloroquine in patients mainly with mild to moderate COVID-19: an open-label, randomized, controlled trial. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.04.10.20060558>.
1058. Magagnoli J, Narendran S, Pereira F, Cummings T, Hardin JW, Sutton SS, Ambati J. 2020. Outcomes of hydroxychloroquine usage in United States veterans hospitalized with Covid-19. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.04.16.20065920>.
1059. Mitjà O, Corbacho-Monné M, Ubals M, Tebé C, Peñafiel J, Tobias A, Ballana E, Alemany A, Riera-Martí N, Pérez CA, Suñer C, Laporte P, Admella P, Mitjà J, Clua M, Bertran L, Sarquella M, Gavilán S, Ara J, Argimon JM, Casabona J, Cuatrecasas G, Cañadas P, Elizalde-Torrent A, Fabregat R, Farré M, Forcada A, Flores-Mateo G, Muntada E, Nadal N, Narejos S, Nieto A, Prat N, Puig J, Quiñones C, Reyes-Ureña J, Ramírez-Viaplana F, Ruiz L, Riveira-Muñoz E, Sierra A, Velasco C, Vivanco-

Hidalgo RM, Sentís A, G-Beiras C, Clotet B, Vall-Mayans M. 2020. [Hydroxychloroquine for Early Treatment of Adults With Mild Coronavirus Disease 2019: A Randomized, Controlled Trial](#). Clinical Infectious Diseases ciaa1009.

1060. Boulware DR, Pullen MF, Bangdiwala AS, Pastick KA, Lofgren SM, Okafor EC, Skipper CP, Nascene AA, Nicol MR, Abassi M, Engen NW, Cheng MP, LaBar D, Lothar SA, MacKenzie LJ, Drobot G, Marten N, Zarychanski R, Kelly LE, Schwartz IS, McDonald EG, Rajasingham R, Lee TC, Hullsiek KH. 2020. [A Randomized Trial of Hydroxychloroquine as Postexposure Prophylaxis for Covid-19](#). New England Journal of Medicine 383:517–525.
1061. Abella BS, Jolkovsky EL, Biney BT, Uspal JE, Hyman MC, Frank I, Hensley SE, Gill S, Vogl DT, Maillard I, Babushok DV, Huang AC, Nasta SD, Walsh JC, Wiletyo EP, Gimotty PA, Milone MC, Amaravadi RK, Prevention and Treatment of COVID-19 With Hydroxychloroquine (PATCH) Investigators. 2021. [Efficacy and Safety of Hydroxychloroquine vs Placebo for Pre-exposure SARS-CoV-2 Prophylaxis Among Health Care Workers](#). JAMA Internal Medicine 181:195.
1062. Schrezenmeier E, Dörner T. 2020. [Mechanisms of action of hydroxychloroquine and chloroquine: implications for rheumatology](#). Nature Reviews Rheumatology 16:155–166.
1063. Elimination or Prolongation of ACE Inhibitors and ARB in Coronavirus Disease 2019 - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04338009>. Retrieved 8 February 2021.
1064. Stopping ACE-inhibitors in COVID-19 - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04353596>. Retrieved 8 February 2021.
1065. Losartan for Patients With COVID-19 Not Requiring Hospitalization - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04311177>. Retrieved 8 February 2021.
1066. Losartan for Patients With COVID-19 Requiring Hospitalization - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04312009>. Retrieved 8 February 2021.
1067. McEvoy PJW. 2020. [The CORONAvirus Disease 2019 Angiotensin Converting Enzyme Inhibitor/Angiotensin Receptor Blocker InvestigatiON \(CORONACION\) Randomized Clinical Trial](#). NCT04330300. Clinical trial registration. clinicaltrials.gov.
1068. Ramipril for the Treatment of COVID-19 - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04366050>. Retrieved 8 February 2021.
1069. D'Or Institute for Research and Education. 2020. [Suspension of Angiotensin Receptor Blockers and Angiotensin-converting Enzyme Inhibitors and Adverse Outcomes in Hospitalized Patients With](#)

[Coronavirus Infection \(COVID-19\). A Randomized Trial](#). NCT04364893.
Clinical trial registration. clinicaltrials.gov.

1070. Cohen JB, Hanff TC, South AM, Sparks MA, Hiremath S, Bress AP, Byrd JB, Chirinos JA. 2020. [Response by Cohen et al to Letter Regarding Article, "Association of Inpatient Use of Angiotensin-Converting Enzyme Inhibitors and Angiotensin II Receptor Blockers With Mortality Among Patients With Hypertension Hospitalized With COVID-19"](#). *Circulation Research* 126.
1071. Sparks MA, South A, Welling P, Luther JM, Cohen J, Byrd JB, Burrell LM, Batlle D, Tomlinson L, Bhalla V, Rheault MN, Soler MJ, Swaminathan S, Hiremath S. 2020. [Sound Science before Quick Judgement Regarding RAS Blockade in COVID-19](#). *Clinical Journal of the American Society of Nephrology* 15:714–716.
1072. Sparks M, Hiremath S. The Coronavirus Conundrum: ACE2 and Hypertension Edition. *NephJC*.
<http://www.nephjc.com/news/covidace2>. Retrieved 30 July 2020.
1073. Luo Y, Zheng SG. 2016. [Hall of Fame among Pro-inflammatory Cytokines: Interleukin-6 Gene and Its Transcriptional Regulation Mechanisms](#). *Frontiers in Immunology* 7.
1074. Rose-John S. 2012. [IL-6 Trans-Signaling via the Soluble IL-6 Receptor: Importance for the Pro-Inflammatory Activities of IL-6](#). *International Journal of Biological Sciences* 8:1237–1247.
1075. Scheller J, Rose-John S. 2006. [Interleukin-6 and its receptor: from bench to bedside](#). *Medical Microbiology and Immunology* 195:173–183.
1076. Garbers C, Hermanns HM, Schaper F, Müller-Newen G, Grötzinger J, Rose-John S, Scheller J. 2012. [Plasticity and cross-talk of Interleukin 6-type cytokines](#). *Cytokine & Growth Factor Reviews* 23:85–97.
1077. Rose-John S, Heinrich PC. 1994. [Soluble receptors for cytokines and growth factors: generation and biological function](#). *Biochemical Journal* 300:281–290.
1078. Tanaka T, Narazaki M, Masuda K, Kishimoto T. 2013. [Interleukin-6; pathogenesis and treatment of autoimmune inflammatory diseases](#). *Inflammation and Regeneration* 33:054–065.
1079. Watad A, Bragazzi NL, Bridgewood C, Mansour M, Mahroum N, Riccò M, Nasr A, Hussein A, Gendelman O, Shoenfeld Y, Lindar M, Amital H, Wu J, McGonagle D. 2020. Systematic Review and Meta-Analysis of Case-Control Studies from 7,000 COVID-19 Pneumonia Patients Suggests a Beneficial Impact of Tocilizumab with Benefit Most Evident in Non-Corticosteroid Exposed Subjects. *SSRN Electronic Journal* <https://doi.org/10.2139/ssrn.3642653>.
1080. Kaye AG, Siegel R. 2020. [The efficacy of IL-6 inhibitor Tocilizumab in reducing severe COVID-19 mortality: a systematic review](#). *PeerJ* 8:e10322.

1081. Cortegiani A, Ippolito M, Greco M, Granone V, Protti A, Gregoret C, Giarratano A, Einav S, Cecconi M. 2021. [Rationale and evidence on the use of tocilizumab in COVID-19: a systematic review](#). Pulmonology 27:52–66.
1082. Jones G, Panova E. 2018. [New insights and long-term safety of tocilizumab in rheumatoid arthritis](#). Therapeutic Advances in Musculoskeletal Disease 10:195–199.
1083. Saito J, Yakuwa N, Kaneko K, Takai C, Goto M, Nakajima K, Yamatani A, Murashima A. 2019. [Tocilizumab during pregnancy and lactation: drug levels in maternal serum, cord blood, breast milk and infant serum](#). Rheumatology 58:1505–1507.
1084. Chen L-F, Mo Y-Q, Jing J, Ma J-D, Zheng D-H, Dai L. 2017. [Short-course tocilizumab increases risk of hepatitis B virus reactivation in patients with rheumatoid arthritis: a prospective clinical observation](#). International Journal of Rheumatic Diseases 20:859–869.
1085. Fu B, Xu X, Wei H. 2020. [Why tocilizumab could be an effective treatment for severe COVID-19?](#) Journal of Translational Medicine 18:164.
1086. Campbell L, Chen C, Bhagat SS, Parker RA, Ostor AJK. 2010. [Risk of adverse events including serious infections in rheumatoid arthritis patients treated with tocilizumab: a systematic literature review and meta-analysis of randomized controlled trials](#). Rheumatology 50:552–562.
1087. Pawar A, Desai RJ, Solomon DH, Santiago Ortiz AJ, Gale S, Bao M, Sarsour K, Schneeweiss S, Kim SC. 2019. [Risk of serious infections in tocilizumab versus other biologic drugs in patients with rheumatoid arthritis: a multidatabase cohort study](#). Annals of the Rheumatic Diseases 78:456–464.
1088. Lang VR, Englbrecht M, Rech J, Nüsslein H, Manger K, Schuch F, Tony H-P, Fleck M, Manger B, Schett G, Zwerina J. 2012. [Risk of infections in rheumatoid arthritis patients treated with tocilizumab](#). Rheumatology 51:852–857.
1089. Radbel J, Narayanan N, Bhatt PJ. 2020. [Use of Tocilizumab for COVID-19-Induced Cytokine Release Syndrome](#). Chest 158:e15–e19.
1090. Tzotzos SJ, Fischer B, Fischer H, Zeitlinger M. 2020. [Incidence of ARDS and outcomes in hospitalized patients with COVID-19: a global literature survey](#). Crit Care 24.
1091. Kaye A, Siegel R. 2020. The Efficacy of IL-6 Inhibitor Tocilizumab in Reducing Severe COVID-19 Mortality: A Systematic Review. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.07.10.20150938>.
1092. Hassoun A, Thottacherry ED, Muklewicz J, Aziz Q, Edwards J. 2020. [Utilizing tocilizumab for the treatment of cytokine release syndrome in COVID-19](#). Journal of Clinical Virology 128:104443.

1093. Spiegel M, Pichlmair A, Mühlberger E, Haller O, Weber F. 2004. [The antiviral effect of interferon-beta against SARS-Coronavirus is not mediated by MxA protein](#). Journal of Clinical Virology 30:211–213.
1094. DeDiego ML, Nieto-Torres JL, Jimenez-Guardeño JM, Regla-Nava JA, Castaño-Rodríguez C, Fernandez-Delgado R, Usera F, Enjuanes L. 2014. [Coronavirus virulence genes with main focus on SARS-CoV envelope gene](#). Virus Research 194:124–137.
1095. 2020. Synairgen to start trial of SNG001 in COVID-19 imminently. Synairgen plc press release. <http://synairgen.web01.hosting.bdc1.co.uk/umbraco/Surface/Download/GetFile?cid=23c9b12c-508b-48c3-9081-36605c5a9ccd>.
1096. Peiffer-Smadja N, Yazdanpanah Y. 2021. [Nebulised interferon beta-1a for patients with COVID-19](#). The Lancet Respiratory Medicine 9:122–123.
1097. Ranieri VM, Pettilä V, Karvonen MK, Jalkanen J, Nightingale P, Brealey D, Mancebo J, Ferrer R, Mercat A, Patroniti N, Quintel M, Vincent J-L, Okkonen M, Meziani F, Bellani G, MacCallum N, Creteur J, Kluge S, Artigas-Raventos A, Maksimow M, Piippo I, Elima K, Jalkanen S, Jalkanen M, Bellingan G. 2020. [Effect of Intravenous Interferon \$\beta\$ -1a on Death and Days Free From Mechanical Ventilation Among Patients With Moderate to Severe Acute Respiratory Distress Syndrome](#). JAMA 323:725.
1098. Davoudi-Monfared E, Rahmani H, Khalili H, Hajiabdolbaghi M, Salehi M, Abbasian L, Kazemzadeh H, Yekaninejad MS. 2020. [A Randomized Clinical Trial of the Efficacy and Safety of Interferon \$\beta\$ -1a in Treatment of Severe COVID-19](#). Antimicrob Agents Chemother 64.
1099. National Institute of Allergy and Infectious Diseases (NIAID). 2022. [A Multicenter, Adaptive, Randomized Blinded Controlled Trial of the Safety and Efficacy of Investigational Therapeutics for the Treatment of COVID-19 in Hospitalized Adults \(ACTT-3\)](#). NCT04492475. Clinical trial registration. clinicaltrials.gov.
1100. 2015. Tocilizumab (Actemra): Adult Patients with Moderately to Severely Active Rheumatoid Arthritis. Canadian Agency for Drugs and Technologies in Health, Ottawa (ON). <https://www.ncbi.nlm.nih.gov/books/NBK349513/table/T43/>.
1101. Hankin C, Feldman S, Szczotka A, Stinger R, Fish L, Hankin D. 2005. [A Cost Comparison of Treatments of Moderate to Severe Psoriasis](#). Drug Benefit Trends 17:200–214.
1102. Tobinick E. 2008. [TNF- \$\alpha\$ inhibition for potential therapeutic modulation of SARS coronavirus infection](#). Current Medical Research and Opinion 20:39–40.
1103. 2020. Sanofi and Regeneron begin global Kevzara® (sarilumab) clinical trial program in patients with severe COVID-19. Sanofi. <http://www.news.sanofi.us/2020-03-16-Sanofi-and-Regeneron-begin-global-Kevzara-R-sarilumab-clinical-trial-program-in-patients-with-severe-COVID-19>.

1104. Sarilumab COVID-19 - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04327388>. Retrieved 8 February 2021.
1105. Stebbing J, Phelan A, Griffin I, Tucker C, Oechsle O, Smith D, Richardson P. 2020. [COVID-19: combining antiviral and anti-inflammatory treatments](#). The Lancet Infectious Diseases 20:400–402.
1106. Richardson P, Griffin I, Tucker C, Smith D, Oechsle O, Phelan A, Rawling M, Savory E, Stebbing J. 2020. [Baricitinib as potential treatment for 2019-nCoV acute respiratory disease](#). The Lancet 395:e30–e31.
1107. Lilly Begins Clinical Testing of Therapies for COVID-19 | Eli Lilly and Company. <https://investor.lilly.com/news-releases/news-release-details/lilly-begins-clinical-testing-therapies-covid-19>. Retrieved 8 February 2021.
1108. Cantini F. 2020. [Baricitinib Combined With Antiviral Therapy in Symptomatic Patients Infected by COVID-19: an Open-label, Pilot Study](#). NCT04320277. Clinical trial registration. clinicaltrials.gov.
1109. Biot C, Daher W, Chavain N, Fandeur T, Khalife J, Dive D, De Clercq E. 2006. [Design and Synthesis of Hydroxyferroquine Derivatives with Antimalarial and Antiviral Activities](#). Journal of Medicinal Chemistry 49:2845–2849.
1110. Sheahan TP, Sims AC, Zhou S, Graham RL, Pruijssers AJ, Agostini ML, Leist SR, Schäfer A, Dinnon KH, Stevens LJ, Chappell JD, Lu X, Hughes TM, George AS, Hill CS, Montgomery SA, Brown AJ, Bluemling GR, Natchus MG, Saindane M, Kolykhalov AA, Painter G, Harcourt J, Tamin A, Thornburg NJ, Swanstrom R, Denison MR, Baric RS. 2020. [An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 in human airway epithelial cell cultures and multiple coronaviruses in mice](#). Science Translational Medicine 12:eabb5883.
1111. Pelegrin M, Naranjo-Gomez M, Piechaczyk M. 2015. [Antiviral Monoclonal Antibodies: Can They Be More Than Simple Neutralizing Agents?](#) Trends in Microbiology 23:653–665.
1112. Zhao J, Wohlford-Lenane C, Zhao J, Fleming E, Lane TE, McCray PB, Perlman S. 2012. [Intranasal Treatment with Poly\(I:C\) Protects Aged Mice from Lethal Respiratory Virus Infections](#). Journal of Virology 86:11416–11424.
1113. Plotkin S. 2014. [History of vaccination](#). Proc Natl Acad Sci USA 111:12283–12287.
1114. Krammer F. 2020. [SARS-CoV-2 vaccines in development](#). Nature 586:516–527.
1115. COVID19 Vaccine Tracker. <https://covid19.trackvaccines.org/>. Retrieved 5 December 2022.
1116. Mathieu E, Ritchie H, Ortiz-Ospina E, Roser M, Hasell J, Appel C, Giattino C, Rodés-Guirao L. 2021. [A global database of COVID-19 vaccinations](#). Nat Hum Behav 5:947–953.

1117. Corey L, Mascola JR, Fauci AS, Collins FS. 2020. [A strategic approach to COVID-19 vaccine R&D](#). Science 368:948–950.
1118. Vaccine efficacy, effectiveness and protection. [https://www.who.int/news-room/feature-stories/detail/vaccine-
efficacy-effectiveness-and-protection](https://www.who.int/news-room/feature-stories/detail/vaccine-efficacy-effectiveness-and-protection). Retrieved 5 December 2022.
1119. Jackson LA, Anderson EJ, Roupael NG, Roberts PC, Makhene M, Coler RN, McCullough MP, Chappell JD, Denison MR, Stevens LJ, Pruijssers AJ, McDermott A, Flach B, Doria-Rose NA, Corbett KS, Morabito KM, O'Dell S, Schmidt SD, Swanson PA, Padilla M, Mascola JR, Neuzil KM, Bennett H, Sun W, Peters E, Makowski M, Albert J, Cross K, Buchanan W, Pikaart-Tautges R, Ledgerwood JE, Graham BS, Beigel JH. 2020. [An mRNA Vaccine against SARS-CoV-2 — Preliminary Report](#). New England Journal of Medicine 383:1920–1931.
1120. Davidson MH. 2006. [Differences between clinical trial efficacy and real-world effectiveness](#). Am J Manag Care 12:S405–11.
1121. Monti S, Grosso V, Todoerti M, Caporali R. 2018. [Randomized controlled trials and real-world data: differences and similarities to untangle literature data](#). Rheumatology 57:vii54–vii58.
1122. Stewart AJ, Devlin PM. 2006. [The history of the smallpox vaccine](#). Journal of Infection 52:329–334.
1123. Leung AKC. 2011. [“Variolation” and Vaccination in Late Imperial China, Ca 1570–1911](#), p. 5–12. *In* History of Vaccine Development. Springer New York.
1124. Stern AM, Markel H. 2005. [The History Of Vaccines And Immunization: Familiar Patterns, New Challenges](#). Health Affairs 24:611–621.
1125. Esparza J, Schrick L, Damaso CR, Nitsche A. 2017. [Equination \(inoculation of horsepox\): An early alternative to vaccination \(inoculation of cowpox\) and the potential role of horsepox virus in the origin of the smallpox vaccine](#). Vaccine 35:7222–7230.
1126. Minor PD. 2015. [Live attenuated vaccines: Historical successes and current challenges](#). Virology 479-480:379–392.
1127. Zhang C, Maruggi G, Shan H, Li J. 2019. [Advances in mRNA Vaccines for Infectious Diseases](#). Frontiers in Immunology 10:594.
1128. Cui Z. 2005. [DNA Vaccine](#), p. 257–289. *In* Non-Viral Vectors for Gene Therapy, Second Edition: Part 2. Elsevier.
1129. Barranco C. 2020. The first live attenuated vaccines. Nature Research <https://doi.org/10.1038/d42859-020-00008-5>.
1130. Flint J, Skalka AM, Rall GF, Racaniello VR. 2015. Principles of Virology, Volume I: Molecular Biology. American Society of Microbiology. <https://doi.org/gmqjck>.
1131. Sanchez-Felipe L, Vercruysse T, Sharma S, Ma J, Lemmens V, Van Looveren D, Arkalagud Javarappa MP, Boudewijns R, Malengier-Devlies

B, Liesenborghs L, Kaptein SJF, De Keyzer C, Bervoets L, Debaveye S, Rasulova M, Seldeslachts L, Li L-H, Jansen S, Yakass MB, Verstrepen BE, Böszörményi KP, Kiemenyi-Kayere G, van Driel N, Quaye O, Zhang X, ter Horst S, Mishra N, Deboutte W, Matthijnsens J, Coelmont L, Vandermeulen C, Heylen E, Vergote V, Schols D, Wang Z, Bogers W, Kuiken T, Verschoor E, Cawthorne C, Van Laere K, Opdenakker G, Vande Velde G, Weynand B, Teuwen DE, Matthys P, Neyts J, Jan Thibaut H, Dallmeier K. 2020. [A single-dose live-attenuated YF17D-vectored SARS-CoV-2 vaccine candidate](#). *Nature* 590:320–325.

1132. Siegrist C-A. 2018. [Vaccine Immunology](#), p. 16–34.e7. *In* Plotkin's Vaccines. Elsevier.
1133. Zhao J, Zhao S, Ou J, Zhang J, Lan W, Guan W, Wu X, Yan Y, Zhao W, Wu J, Chodosh J, Zhang Q. 2020. [COVID-19: Coronavirus Vaccine Development Updates](#). *Front Immunol* 11.
1134. 2021. Vaccine Types. US Department of Health & Human Services. <https://www.hhs.gov/immunization/basics/types/index.html>. Retrieved 5 December 2022.
1135. Lee J, Khang D. 2023. [Mucosal delivery of nanovaccine strategy against COVID-19 and its variants](#). *Acta Pharmaceutica Sinica B* 13:2897–2925.
1136. Alu A, Chen L, Lei H, Wei Y, Tian X, Wei X. 2022. [Intranasal COVID-19 vaccines: From bench to bed](#). *eBioMedicine* 76:103841.
1137. Sharma S, Vercruysse T, Sanchez-Felipe L, Kerstens W, Rasulova M, Abdelnabi R, Foo CS, Lemmens V, Van Looveren D, Maes P, Baele G, Weynand B, Lemey P, Neyts J, Thibaut HJ, Dallmeier K. 2021. [Updated vaccine protects from infection with SARS-CoV-2 variants, prevents transmission and is immunogenic against Omicron in hamsters](#). Cold Spring Harbor Laboratory.
1138. Marshall D. Griffith University researchers on the road to COVID-19 vaccine. <https://news.griffith.edu.au/2020/04/23/griffith-university-researchers-on-the-road-to-covid-19-vaccine/>. Retrieved 5 December 2022.
1139. COVID-19 Treatment and Vaccine Tracker. Milken Institute. <https://covid-19tracker.milkeninstitute.org/>. Retrieved 5 August 2022.
1140. Wang Y, Yang C, Song Y, Coleman JR, Stawowczyk M, Tafrova J, Tasker S, Boltz D, Baker R, Garcia L, Seale O, Kushnir A, Wimmer E, Mueller S. 2021. [Scalable live-attenuated SARS-CoV-2 vaccine candidate demonstrates preclinical safety and efficacy](#). *Proc Natl Acad Sci USA* 118.
1141. Coleman JR, Papamichail D, Skiena S, Fitcher B, Wimmer E, Mueller S. 2008. [Virus Attenuation by Genome-Scale Changes in Codon Pair Bias](#). *Science* 320:1784–1787.
1142. Codagenix, Inc. 2022. [First-in-human, Randomised, Double-blind, Placebo-controlled, Dose-escalation Study in Healthy Young Adults Evaluating the Safety and Immunogenicity of COVI-VAC, a Live](#)

[Attenuated Vaccine Candidate for Prevention of COVID-19.](#)

NCT04619628. Clinical trial registration. clinicaltrials.gov.

1143. Kushnir A, Mueller S, Tasker S, Robert Coleman J. 2021. [577. COVI-VAC™, a Live Attenuated COVID-19 Vaccine, Provides Single Dose Protection Against Heterologous Challenge with SARS-CoV-2 Beta \(B.1.351\) in the Syrian Golden Hamster Model](#). Open Forum Infect Dis 8:S390.
1144. He DC, He C-Q. 2022. [Discovery of vaccine-like recombinant SARS-CoV-2 circulating in human](#). Virol J 19.
1145. Tioni MF, Jordan R, Pena AS, Garg A, Wu D, Phan SI, Weiss CM, Cheng X, Greenhouse J, Orekov T, Valentin D, Kar S, Pessaint L, Andersen H, Stobart CC, Bloodworth MH, Stokes Peebles R, Liu Y, Xie X, Shi P-Y, Moore ML, Tang RS. 2022. [Mucosal administration of a live attenuated recombinant COVID-19 vaccine protects nonhuman primates from SARS-CoV-2](#). npj Vaccines 7.
1146. Gao S, Song S, Zhang L. 2019. [Recent Progress in Vaccine Development Against Chikungunya Virus](#). Front Microbiol 10.
1147. Meissa Vaccines, Inc. 2022. [Phase 1, Open-Label, Dose-Escalation Study to Evaluate Tolerability, Safety, and Immunogenicity of an Intranasal Live Attenuated Respiratory Syncytial Virus Vaccine Expressing Spike Protein of SARS-CoV-2 in Healthy Adults Ages 18 - 69 Years](#). NCT04798001. Clinical trial registration. clinicaltrials.gov.
1148. Rando HM, Greene CS, Robson MP, Boca SM, Wellhausen N, Lordan R, Brueffer C, Ray S, McGowan LD, Gitter A, Dattoli AA, Velazquez R, Barton JP, Field JM, Ramsundar B, MacLean AL, Lee AJ, Immunology Institute of the Icahn School of Medicine, Hu F, Jadavji NM, Sell E, Wang J, Rafizadeh DN, Skelly AN, Guebila MB, Kolla L, Manheim D, Ghosh S, Byrd JB, Park Y, Bansal V, Capone S, Dziak JJ, Sun Y, Qi Y, Shinholster L, Lukan T, Knyazev S, Perrin D, Mangul S, Das S, Szeto GL, Lubiana T, Mai D, COVID-19 Review Consortium, Goel RR, Boerckel JD, Naik A, Sun Y, Himmelstein DS, Kamil JP, Meyer JG, Mundo AI. 2023. [SARS-CoV-2 and COVID-19: An Evolving Review of Diagnostics and Therapeutics](#).
1149. O'Neill LAJ, Netea MG. 2020. [BCG-induced trained immunity: can it offer protection against COVID-19?](#) Nat Rev Immunol 20:335–337.
1150. Moorlag SJCFM, Taks E, ten Doesschate T, van der Vaart TW, Janssen AB, Müller L, Ostermann P, Dijkstra H, Lemmers H, Simonetti E, Mazur M, Schaal H, ter Heine R, van de Veerdonk FL, Bleeker-Rovers CP, van Crevel R, ten Oever J, de Jonge MI, Bonten MJ, van Werkhoven CH, Netea MG. 2022. [Efficacy of BCG Vaccination Against Respiratory Tract Infections in Older Adults During the Coronavirus Disease 2019 Pandemic](#). Clinical Infectious Diseases 75:e938–e946.
1151. BCG Vaccination to Protect Healthcare Workers Against COVID-19 - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04327206>. Retrieved 8 February 2021.

1152. BCG Vaccine for Health Care Workers as Defense Against COVID 19 - Full Text View - ClinicalTrials.gov.
<https://clinicaltrials.gov/ct2/show/NCT04348370>. Retrieved 8 February 2021.
1153. Codagenix Inc: COVI-VAC – COVID19 Vaccine Tracker.
<https://covid19.trackvaccines.org/vaccines/59/>. Retrieved 30 July 2023.
1154. Robert-Guroff M. 2007. [Replicating and non-replicating viral vectors for vaccine development](#). Current Opinion in Biotechnology 18:546–556.
1155. Barrett PN, Terpening SJ, Snow D, Cobb RR, Kistner O. 2017. [Vero cell technology for rapid development of inactivated whole virus vaccines for emerging viral diseases](#). Expert Review of Vaccines 16:883–894.
1156. Bachmann MF, Jennings GT. 2010. [Vaccine delivery: a matter of size, geometry, kinetics and molecular patterns](#). Nat Rev Immunol 10:787–796.
1157. Roberts A, Lamirande EW, Vogel L, Jackson JP, Paddock CD, Guarner J, Zaki SR, Sheahan T, Baric R, Subbarao K. 2008. [Animal models and vaccines for SARS-CoV infection](#). Virus Research 133:20–32.
1158. Scherle PA, Gerhard W. 1986. [Functional analysis of influenza-specific helper T cell clones in vivo. T cells specific for internal viral proteins provide cognate help for B cell responses to hemagglutinin](#). Journal of Experimental Medicine 164:1114–1128.
1159. Brüssow H. 2021. [COVID-19: vaccine's progress](#). Microb Biotechnol 14:1246–1257.
1160. Reuters. 2020. [China gives its first COVID-19 vaccine approval to Sinopharm](#). Reuters.
1161. Wee S-L, Simões M. 2020. [In Coronavirus Vaccine Race, China Strays From the Official Paths](#). The New York Times.
1162. Ball P. 2020. [The lightning-fast quest for COVID vaccines — and what it means for other diseases](#). Nature 589:16–18.
1163. Tsunetsugu-Yokota Y, Ohnishi K, Takemori T. 2006. [Severe acute respiratory syndrome \(SARS\) coronavirus: application of monoclonal antibodies and development of an effective vaccine](#). Rev Med Virol 16:117–131.
1164. Takasuka N. 2004. [A subcutaneously injected UV-inactivated SARS coronavirus vaccine elicits systemic humoral immunity in mice](#). International Immunology 16:1423–1430.
1165. Su S, Du L, Jiang S. 2020. [Learning from the past: development of safe and effective COVID-19 vaccines](#). Nat Rev Microbiol 19:211–219.
1166. Wang Q, Zhang L, Kuwahara K, Li L, Liu Z, Li T, Zhu H, Liu J, Xu Y, Xie J, Morioka H, Sakaguchi N, Qin C, Liu G. 2016. [Immunodominant SARS Coronavirus Epitopes in Humans Elicited both Enhancing and](#)

[Neutralizing Effects on Infection in Non-human Primates](#). ACS Infect Dis 2:361–376.

1167. Agrawal AS, Tao X, Algaissi A, Garron T, Narayanan K, Peng B-H, Couch RB, Tseng C-TK. 2016. [Immunization with inactivated Middle East Respiratory Syndrome coronavirus vaccine leads to lung immunopathology on challenge with live virus](#). Human Vaccines & Immunotherapeutics 12:2351–2356.
1168. Li K, Li Z, Wohlford-Lenane C, Meyerholz DK, Channappanavar R, An D, Perlman S, McCray PB Jr., He B. 2020. [Single-Dose, Intranasal Immunization with Recombinant Parainfluenza Virus 5 Expressing Middle East Respiratory Syndrome Coronavirus \(MERS-CoV\) Spike Protein Protects Mice from Fatal MERS-CoV Infection](#). mBio 11.
1169. Types of Vaccines – COVID19 Vaccine Tracker. <https://covid19.trackvaccines.org/types-of-vaccines/>. Retrieved 5 December 2022.
1170. Jordahl K, Van Den Bossche J, Fleischmann M, McBride J, Wasserman J, Badaracco AG, Gerard J, Snow AD, Tratner J, Perry M, Farmer C, Hjelle GA, Cochran M, Gillies S, Culbertson L, Bartos M, Ward B, Caria G, Taves M, Eubank N, Sangarshanan, Flavin J, Richards M, Rey S, Maxalbert, Bilogur A, Ren C, Arribas-Bel D, Mesejo-León D, Wasser L. 2021. geopandas/geopandas: v0.10.2 (v0.10.2). Zenodo. <https://doi.org/gqkzpv>.
1171. Background document on the inactivated vaccine Sinovac-CoronaVac against COVID-19. https://www.who.int/publications-detail-redirect/WHO-2019-nCoV-vaccines-SAGE_recommendation-Sinovac-CoronaVac-background-2021.1. Retrieved 5 December 2022.
1172. Wu Z, Hu Y, Xu M, Chen Z, Yang W, Jiang Z, Li M, Jin H, Cui G, Chen P, Wang L, Zhao G, Ding Y, Zhao Y, Yin W. 2021. [Safety, tolerability, and immunogenicity of an inactivated SARS-CoV-2 vaccine \(CoronaVac\) in healthy adults aged 60 years and older: a randomised, double-blind, placebo-controlled, phase 1/2 clinical trial](#). The Lancet Infectious Diseases 21:803–812.
1173. Gao Q, Bao L, Mao H, Wang L, Xu K, Yang M, Li Y, Zhu L, Wang N, Lv Z, Gao H, Ge X, Kan B, Hu Y, Liu J, Cai F, Jiang D, Yin Y, Qin C, Li J, Gong X, Lou X, Shi W, Wu D, Zhang H, Zhu L, Deng W, Li Y, Lu J, Li C, Wang X, Yin W, Zhang Y, Qin C. 2020. [Development of an inactivated vaccine candidate for SARS-CoV-2](#). Science 369:77–81.
1174. Zhang Y, Zeng G, Pan H, Li C, Hu Y, Chu K, Han W, Chen Z, Tang R, Yin W, Chen X, Hu Y, Liu X, Jiang C, Li J, Yang M, Song Y, Wang X, Gao Q, Zhu F. 2021. [Safety, tolerability, and immunogenicity of an inactivated SARS-CoV-2 vaccine in healthy adults aged 18–59 years: a randomised, double-blind, placebo-controlled, phase 1/2 clinical trial](#). The Lancet Infectious Diseases 21:181–192.
1175. Sinovac: CoronaVac – COVID19 Vaccine Tracker. <https://covid19.trackvaccines.org/vaccines/7/>. Retrieved 5 December 2022.

1176. Bueno SM, Abarca K, González PA, Gálvez NMS, Soto JA, Duarte LF, Schultz BM, Pacheco GA, González LA, Vázquez Y, Ríos M, Melo-González F, Rivera-Pérez D, Iturriaga C, Urzúa M, Domínguez A, Andrade CA, Berríos-Rojas RV, Canedo-Marroquín G, Covián C, Moreno-Tapia D, Saavedra F, Vallejos OP, Donato P, Espinoza P, Fuentes D, González M, Guzmán P, Muñoz Venturelli P, Pérez CM, Potin M, Rojas Á, Fasce RA, Fernández J, Mora J, Ramírez E, Gaete-Argel A, Oyarzún-Arrau A, Valiente-Echeverría F, Soto-Rifo R, Weiskopf D, Sette A, Zeng G, Meng W, González-Aramundiz JV, Kalergis AM. 2021. [Safety and Immunogenicity of an Inactivated Severe Acute Respiratory Syndrome Coronavirus 2 Vaccine in a Subgroup of Healthy Adults in Chile](#). *Clinical Infectious Diseases* 75:e792–e804.
1177. Tanrıover MD, Doğanay HL, Akova M, Güner HR, Azap A, Akhan S, Köse Ş, Erdinç FŞ, Akalın EH, Tabak ÖF, Pullukçu H, Batum Ö, Şimşek Yavuz S, Turhan Ö, Yıldırım MT, Köksal İ, Taşova Y, Korten V, Yılmaz G, Çelen MK, Altın S, Çelik İ, Bayındır Y, Karaoğlu İ, Yılmaz A, Özkul A, Gür H, Unal S, Kayaaslan B, Hasanoğlu İ, Dalkıran A, Aydos Ö, Çınar G, Akdemir-Kalkan İ, İnkaya AÇ, Aydın M, Çakır H, Yıldız J, Kocabıyık Ö, Arslan S, Nallı B, Demir Ö, Singil S, Ataman-Hatipoğlu Ç, Tuncer-Ertem G, Kınıklı S, Önal U, Mete B, Dalgan G, Taşbakan M, Yamazhan T, Kömürcüoğlu B, Yalnız E, Benli A, Keskin-Sarıtaş Ç, Ertosun MG, Özkan Ö, Emre S, Arica S, Kuşçu F, Candevir A, Ertürk-Şengel B, Ayvaz F, Aksoy F, Mermutluoğlu Ç, Demir Y, Günlüoğlu G, Tural-Önür S, Kılıç-Toker A, Eren E, Otlı B, Mete AÖ, Koçak K, Ateş H, Koca-Kalkan İ, Aksu K. 2021. [Efficacy and safety of an inactivated whole-virion SARS-CoV-2 vaccine \(CoronaVac\): interim results of a double-blind, randomised, placebo-controlled, phase 3 trial in Turkey](#). *The Lancet* 398:213–222.
1178. Corum J, Zimmer C. 2020. [How the Sinovac Vaccine Works](#). *The New York Times*.
1179. Palacios R, Patiño EG, de Oliveira Pirelli R, Conde MTRP, Batista AP, Zeng G, Xin Q, Kallas EG, Flores J, Ockenhouse CF, Gast C. 2020. [Double-Blind, Randomized, Placebo-Controlled Phase III Clinical Trial to Evaluate the Efficacy and Safety of treating Healthcare Professionals with the Adsorbed COVID-19 \(Inactivated\) Vaccine Manufactured by Sinovac – PROFISCOV: A structured summary of a study protocol for a randomised controlled trial](#). *Trials* 21.
1180. Palacios R, Batista AP, Albuquerque CSN, Patiño EG, Santos J do P, Tilli Reis Pessoa Conde M, Pirelli R de O, Pereira Júnior LC, Raboni SM, Ramos F, Sierra Romero GA, Leal FE, Camargo LFA, Aoki FH, Coelho EB, Oliveira DS, Fontes CJF, Pileggi GCS, Oliveira ALL de, Siqueira AM de, Oliveira DBL de, Botosso VF, Zeng G, Xin Q, Teixeira MM, Nogueira ML, Kallas EG. 2021. Efficacy and Safety of a COVID-19 Inactivated Vaccine in Healthcare Professionals in Brazil: The PROFISCOV Study. SSRN Journal <https://doi.org/10.2139/ssrn.3822780>.
1181. Medeiros-Ribeiro AC, Aikawa NE, Saad CGS, Yuki EFN, Pedrosa T, Fusco SRG, Rojo PT, Pereira RMR, Shinjo SK, Andrade DCO, Sampaio-Barros PD, Ribeiro CT, Deveza GBH, Martins VAO, Silva CA, Lopes MH, Duarte AJS, Antonangelo L, Sabino EC, Kallas EG, Pasoto SG, Bonfa E. 2021. [Immunogenicity and safety of the CoronaVac inactivated vaccine in](#)

[patients with autoimmune rheumatic diseases: a phase 4 trial](#). Nat Med 27:1744–1751.

1182. Han B, Song Y, Li C, Yang W, Ma Q, Jiang Z, Li M, Lian X, Jiao W, Wang L, Shu Q, Wu Z, Zhao Y, Li Q, Gao Q. 2021. [Safety, tolerability, and immunogenicity of an inactivated SARS-CoV-2 vaccine \(CoronaVac\) in healthy children and adolescents: a double-blind, randomised, controlled, phase 1/2 clinical trial](#). The Lancet Infectious Diseases 21:1645–1653.
1183. Fonseca P. 2020. [Brazil institute says CoronaVac efficacy above 50%, but delays full results](#). Reuters.
1184. Zimmer C, Londoño E. 2020. [Turkey and Brazil Say Chinese Vaccine Effective, With Sparse Supporting Data](#). The New York Times.
1185. Jara A, Undurraga EA, González C, Paredes F, Fontecilla T, Jara G, Pizarro A, Acevedo J, Leo K, Leon F, Sans C, Leighton P, Suárez P, García-Escorza H, Araos R. 2021. [Effectiveness of an Inactivated SARS-CoV-2 Vaccine in Chile](#). N Engl J Med 385:875–884.
1186. Zimmer C, Corum J, Wee S-L, Kristoffersen M. 2020. [Coronavirus Vaccine Tracker](#). The New York Times.
1187. Tunjungputri RN, Tetraswi EN, Veronica M, Pandelaki J, Ibrahim F, Nelwan EJ. 2021. [Vaccine-Associated Disease Enhancement \(VADE\): Considerations in Postvaccination COVID-19](#). Case Reports in Medicine 2021:1–5.
1188. Wee S-L. 2021. [They Relied on Chinese Vaccines. Now They're Battling Outbreaks](#). The New York Times.
1189. Vacharathit V, Aiewsakun P, Manopwisedjaroen S, Srisaowakarn C, Laopanupong T, Ludowyke N, Phuphuakrat A, Setthaudom C, Ekronarongchai S, Srichatrapimuk S, Wongsirisin P, Sangrajrang S, Imsuwansri T, Kirdlarp S, Nualkaew S, Sensorn I, Sawaengdee W, Wichukchinda N, Sungkanuparph S, Chantratita W, Kunakorn M, Rojanamatin J, Hongeng S, Thitithanyanont A. 2021. [CoronaVac induces lower neutralising activity against variants of concern than natural infection](#). The Lancet Infectious Diseases 21:1352–1354.
1190. Lu L, Mok BWY, Chen LL, Chan JMC, Tsang OTY, Lam BHS, Chuang VWM, Chu AWH, Chan WM, Ip JD, Chan BPC, Zhang R, Yip CCY, Cheng VCC, Chan KH, Jin DY, Hung IFN, Yuen KY, Chen H, To KKW. 2021. [Neutralization of Severe Acute Respiratory Syndrome Coronavirus 2 Omicron Variant by Sera From BNT162b2 or CoronaVac Vaccine Recipients](#). Clinical Infectious Diseases 75:e822–e826.
1191. Wang G-L, Wang Z-Y, Duan L-J, Meng Q-C, Jiang M-D, Cao J, Yao L, Zhu K-L, Cao W-C, Ma M-J. 2021. [Susceptibility of Circulating SARS-CoV-2 Variants to Neutralization](#). N Engl J Med 384:2354–2356.
1192. Angkasekwinai N, Sewatanon J, Niyomnaitham S, Phumiamorn S, Sukapirom K, Sapsutthipas S, Sirijatuphat R, Wittawatmongkol O, Senawong S, Mahasirimongkol S, Trisiriwanich S, Chokephaibulkit K. 2022. [Comparison of safety and immunogenicity of CoronaVac and](#)

[ChAdOx1 against the SARS-CoV-2 circulating variants of concern \(Alpha, Delta, Beta\) in Thai healthcare workers](#). Vaccine: X 10:100153.

1193. Hua Q, Zhang H, Yao P, Xu N, Sun Y, Lu H, Xu F, Liao Y, Yang J, Mao H, Zhang Y, Zhu H, Hu X, Lv H, Jiang J. 2022. [Immunogenicity and immune-persistence of the CoronaVac or Covilo inactivated COVID-19 Vaccine: a 6-month population-based cohort study](#). Front Immunol 13.
1194. Li M, Yang J, Wang L, Wu Q, Wu Z, Zheng W, Wang L, Lu W, Deng X, Peng C, Han B, Zhao Y, Yu H, Yin W. 2021. [A booster dose is immunogenic and will be needed for older adults who have completed two doses vaccination with CoronaVac: a randomised, double-blind, placebo-controlled, phase 1/2 clinical trial](#). Cold Spring Harbor Laboratory.
1195. Zeng G, Wu Q, Pan H, Li M, Yang J, Wang L, Wu Z, Jiang D, Deng X, Chu K, Zheng W, Wang L, Lu W, Han B, Zhao Y, Zhu F, Yu H, Yin W. 2022. [Immunogenicity and safety of a third dose of CoronaVac, and immune persistence of a two-dose schedule, in healthy adults: interim results from two single-centre, double-blind, randomised, placebo-controlled phase 2 clinical trials](#). The Lancet Infectious Diseases 22:483–495.
1196. 2021. China approves first mixed-vaccine trial as Delta spreads. <https://medicalxpress.com/news/2021-08-china-mixed-vaccine-trial-delta.html>. Retrieved 5 December 2022.
1197. Reactogenicity, Safety, and Immunogenicity of Covid-19 Vaccine Booster - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04992182>. Retrieved 30 July 2023.
1198. Li J, Hou L, Guo X, Jin P, Wu S, Zhu J, Pan H, Wang X, Song Z, Wan J, Cui L, Li J, Chen Y, Wang X, Jin L, Liu J, Shi F, Xu X, Zhu T, Chen W, Zhu F. 2022. [Heterologous AD5-nCOV plus CoronaVac versus homologous CoronaVac vaccination: a randomized phase 4 trial](#). Nat Med 28:401–409.
1199. Pérez-Then E, Lucas C, Monteiro VS, Miric M, Brache V, Cochon L, Vogels CBF, Malik AA, De la Cruz E, Jorge A, De los Santos M, Leon P, Breban MI, Billig K, Yildirim I, Pearson C, Downing R, Gagnon E, Muyombwe A, Razeq J, Campbell M, Ko AI, Omer SB, Grubaugh ND, Vermund SH, Iwasaki A. 2022. [Neutralizing antibodies against the SARS-CoV-2 Delta and Omicron variants following heterologous CoronaVac plus BNT162b2 booster vaccination](#). Nat Med 28:481–485.
1200. 2022. The Sinovac-CoronaVac COVID-19 vaccine: What you need to know. <https://www.who.int/news-room/feature-stories/detail/the-sinovac-covid-19-vaccine-what-you-need-to-know>. Retrieved 5 December 2022.
1201. Kyriakidis NC, López-Cortés A, González EV, Grimaldos AB, Prado EO. 2021. [SARS-CoV-2 vaccines strategies: a comprehensive review of phase 3 candidates](#). npj Vaccines 6.
1202. Wang H, Zhang Y, Huang B, Deng W, Quan Y, Wang W, Xu W, Zhao Y, Li N, Zhang J, Liang H, Bao L, Xu Y, Ding L, Zhou W, Gao H, Liu J, Niu P,

- Zhao L, Zhen W, Fu H, Yu S, Zhang Z, Xu G, Li C, Lou Z, Xu M, Qin C, Wu G, Gao GF, Tan W, Yang X. 2020. [Development of an Inactivated Vaccine Candidate, BBIBP-CorV, with Potent Protection against SARS-CoV-2](#). Cell 182:713–721.e9.
1203. Xia S, Zhang Y, Wang Y, Wang H, Yang Y, Gao GF, Tan W, Wu G, Xu M, Lou Z, Huang W, Xu W, Huang B, Wang H, Wang W, Zhang W, Li N, Xie Z, Ding L, You W, Zhao Y, Yang X, Liu Y, Wang Q, Huang L, Yang Y, Xu G, Luo B, Wang W, Liu P, Guo W, Yang X. 2021. [Safety and immunogenicity of an inactivated SARS-CoV-2 vaccine, BBIBP-CorV: a randomised, double-blind, placebo-controlled, phase 1/2 trial](#). The Lancet Infectious Diseases 21:39–51.
1204. Xia S, Duan K, Zhang Y, Zhao D, Zhang H, Xie Z, Li X, Peng C, Zhang Y, Zhang W, Yang Y, Chen W, Gao X, You W, Wang X, Wang Z, Shi Z, Wang Y, Yang X, Zhang L, Huang L, Wang Q, Lu J, Yang Y, Guo J, Zhou W, Wan X, Wu C, Wang W, Huang S, Du J, Meng Z, Pan A, Yuan Z, Shen S, Guo W, Yang X. 2020. [Effect of an Inactivated Vaccine Against SARS-CoV-2 on Safety and Immunogenicity Outcomes](#). JAMA 324:951.
1205. Al Kaabi N, Zhang Y, Xia S, Yang Y, Al Qahtani MM, Abdulrazzaq N, Al Nusair M, Hassany M, Jawad JS, Abdalla J, Hussein SE, Al Mazrouei SK, Al Karam M, Li X, Yang X, Wang W, Lai B, Chen W, Huang S, Wang Q, Yang T, Liu Y, Ma R, Hussain ZM, Khan T, Saifuddin Fasihuddin M, You W, Xie Z, Zhao Y, Jiang Z, Zhao G, Zhang Y, Mahmoud S, ElTantawy I, Xiao P, Koshy A, Zaher WA, Wang H, Duan K, Pan A, Yang X. 2021. Effect of 2 Inactivated SARS-CoV-2 Vaccines on Symptomatic COVID-19 Infection in Adults. JAMA <https://doi.org/10.1001/jama.2021.8565>.
1206. 2020. [Sinopharm's COVID-19 vaccine 79% effective, seeks approval in China](#). Reuters.
1207. Jeewandara C, Aberathna IS, Pushpakumara PD, Kamaladasa A, Guruge D, Jayathilaka D, Gunasekara B, Tanussiya S, Kuruppu H, Ranasinghe T, Dayarathne S, Dissanayake O, Gamalath N, Ekanayake D, Jayamali M, Wijesinghe A, Dissanayake M, Madusanka D, Jayadas TT, Mudunkotuwa A, Somathilake G, Harvie M, Nimasha T, Danasekara S, Wijayamuni R, Schimanski L, Tan TK, Dong T, Townsend A, Ogg GS, Malavige GN. 2021. [Antibody and T cell responses to Sinopharm/BBIBP-CorV in naïve and previously infected individuals in Sri Lanka](#). Cold Spring Harbor Laboratory.
1208. Huang B, Dai L, Wang H, Hu Z, Yang X, Tan W, Gao GF. 2021. [Neutralization of SARS-CoV-2 VOC 501Y.V2 by human antisera elicited by both inactivated BBIBP-CorV and recombinant dimeric RBD ZF2001 vaccines](#). Cold Spring Harbor Laboratory.
1209. Liu Y, Zeng Q, Deng C, Li M, Li L, Liu D, Liu M, Ruan X, Mei J, Mo R, Zhou Q, Liu M, Peng S, Wang J, Zhang H, Xiao H. 2022. [Robust induction of B cell and T cell responses by a third dose of inactivated SARS-CoV-2 vaccine](#). Cell Discov 8.
1210. Mohandas S, Yadav PD, Shete A, Abraham P, Mohan K, Sapkal G, Mote C, Nyayanit D, Gupta N, Srini VK, Kadam M, Kumar A, Jain R, Majumdar T, Deshpande G, Patil S, Sarkale P, Patil D, Ella R, Prasad SD, Sharma S,

Ella KM, Panda S, Bhargava B. 2020. [Immunogenicity and protective efficacy of BBV152: a whole virion inactivated SARS CoV-2 vaccine in the Syrian hamster model](#). Research Square Platform LLC.

1211. Yadav PD, Ella R, Kumar S, Patil DR, Mohandas S, Shete AM, Vadrevu KM, Bhati G, Sapkal G, Kaushal H, Patil S, Jain R, Deshpande G, Gupta N, Agarwal K, Gokhale M, Mathapati B, Metkari S, Mote C, Nyayanit D, Patil DY, Sai Prasad BS, Suryawanshi A, Kadam M, Kumar A, Daigude S, Gopale S, Majumdar T, Mali D, Sarkale P, Baradkar S, Gawande P, Joshi Y, Fulari S, Dighe H, Sharma S, Gunjekar R, Kumar A, Kalele K, Srinivas VK, Gangakhedkar RR, Ella KM, Abraham P, Panda S, Bhargava B. 2021. [Immunogenicity and protective efficacy of inactivated SARS-CoV-2 vaccine candidate, BBV152 in rhesus macaques](#). Nat Commun 12.
1212. Ella R, Vadrevu KM, Jogdand H, Prasad S, Reddy S, Sarangi V, Ganneru B, Sapkal G, Yadav P, Abraham P, Panda S, Gupta N, Reddy P, Verma S, Kumar Rai S, Singh C, Redkar SV, Gillurkar CS, Kushwaha JS, Mohapatra S, Rao V, Guleria R, Ella K, Bhargava B. 2021. [Safety and immunogenicity of an inactivated SARS-CoV-2 vaccine, BBV152: a double-blind, randomised, phase 1 trial](#). The Lancet Infectious Diseases 21:637–646.
1213. Ella R, Reddy S, Jogdand H, Sarangi V, Ganneru B, Prasad S, Das D, Raju D, Praturi U, Sapkal G, Yadav P, Reddy P, Verma S, Singh C, Redkar SV, Gillurkar CS, Kushwaha JS, Mohapatra S, Bhate A, Rai S, Panda S, Abraham P, Gupta N, Ella K, Bhargava B, Vadrevu KM. 2021. [Safety and immunogenicity of an inactivated SARS-CoV-2 vaccine, BBV152: interim results from a double-blind, randomised, multicentre, phase 2 trial, and 3-month follow-up of a double-blind, randomised phase 1 trial](#). The Lancet Infectious Diseases 21:950–961.
1214. 2021. Bharat Biotech Announces Phase 3 Results of COVAXIN. Bharat Biotech. <https://www.bharatbiotech.com/images/press/covaxin-phase3-efficacy-results.pdf>.
1215. 2021. Ocugen's COVID-19 Vaccine Co-Development Partner, Bharat Biotech shares Phase 3 Interim Results of COVAXIN, Demonstrates Efficacy of 81%. Biogenetech. <https://www.biogenetech.co.th/wp-content/uploads/2021/03/5-Ocugen.pdf>.
1216. 2021. Zimbabwe authorizes use of India's first indigenous COVID-19 vaccine - Xinhua | English.news.cn. http://www.xinhuanet.com/english/2021-03/04/c_139783893.htm. Retrieved 5 December 2022.
1217. Ella R, Reddy S, Blackwelder W, Potdar V, Yadav P, Sarangi V, Aileni VK, Kanungo S, Rai S, Reddy P, Verma S, Singh C, Redkar S, Mohapatra S, Pandey A, Ranganadin P, Gumashta R, Multani M, Mohammad S, Bhatt P, Kumari L, Sapkal G, Gupta N, Abraham P, Panda S, Prasad S, Bhargava B, Ella K, Vadrevu KM. 2021. [Efficacy, safety, and lot to lot immunogenicity of an inactivated SARS-CoV-2 vaccine \(BBV152\): a double-blind, randomised, controlled phase 3 trial](#). Cold Spring Harbor Laboratory.

1218. Ella R, Reddy S, Blackwelder W, Potdar V, Yadav P, Sarangi V, Aileni VK, Kanungo S, Rai S, Reddy P, Verma S, Singh C, Redkar S, Mohapatra S, Pandey A, Ranganadin P, Gumashta R, Multani M, Mohammad S, Bhatt P, Kumari L, Sapkal G, Gupta N, Abraham P, Panda S, Prasad S, Bhargava B, Ella K, Vadrevu KM, Aggarwal P, Aglawe V, Ali A, Anand N, Awad N, Bafna V, Balasubramaniam G, Bandkar A, Basha P, Bharge V, Bhate A, Bhate S, Bhavani V, Bhosale R, Chalapathy D, Chaubal C, Chaudhary D, Chavan A, Desai P, Dhodi D, Dutta S, Garg R, Garg K, George M, Goyal P, Guleria R, Gupta S, Jain M, Jain MK, Jindal S, Kalra M, Kant S, Khosla P, Kulkarni P, Kumar P, Kumar Y, Majumdar A, Meshram P, Mishra V, Mohanty S, Nair J, Pandey S, Panigrahi SK, Patil B, Patil V, Rahate P, Raj V, Ramanand S, Rami K, Ramraj B, Rane S, Rao EV, Rao N, Raphael R, Reddy G, Redkar V, Redkar S, Sachdeva A, Saha J, Sahoo J, Sampath P, Savith A, Shah M, Shanmugam L, Sharma R, Sharma P, Sharma D, Singh A, Singh J, Singh P, Sivaprakasam S, Subramaniam S, Sudheer D, Tandon S, Tariq M, Tripathi V, Vable M, Verma R, Waghmare S. 2021. [Efficacy, safety, and lot-to-lot immunogenicity of an inactivated SARS-CoV-2 vaccine \(BBV152\): interim results of a randomised, double-blind, controlled, phase 3 trial](#). The Lancet 398:2173–2184.
1219. Bharat Biotech: Covaxin – COVID19 Vaccine Tracker. <https://covid19.trackvaccines.org/vaccines/9/>. Retrieved 5 December 2022.
1220. Yadav PD, Sahay RR, Sapkal G, Nyayanit D, Shete AM, Deshpande G, Patil DY, Gupta N, Kumar S, Abraham P, Panda S, Bhargava B. 2021. [Comparable neutralization of SARS-CoV-2 Delta AY.1 and Delta in individuals sera vaccinated with BBV152](#). Cold Spring Harbor Laboratory.
1221. Yadav PD, Sapkal GN, Ella R, Sahay RR, Nyayanit DA, Patil DY, Deshpande G, Shete AM, Gupta N, Mohan VK, Abraham P, Panda S, Bhargava B. 2021. [Neutralization of Beta and Delta variant with sera of COVID-19 recovered cases and vaccinees of inactivated COVID-19 vaccine BBV152/Covaxin](#). Journal of Travel Medicine 28.
1222. Sapkal GN, Yadav PD, Ella R, Deshpande GR, Sahay RR, Gupta N, Vadrevu KM, Abraham P, Panda S, Bhargava B. 2021. [Inactivated COVID-19 vaccine BBV152/COVAXIN effectively neutralizes recently emerged B.1.1.7 variant of SARS-CoV-2](#). Journal of Travel Medicine 28.
1223. Yadav PD, Sahay RR, Sapkal G, Nyayanit D, Shete AM, Deshpande G, Patil DY, Gupta N, Kumar S, Abraham P, Panda S, Bhargava B. 2021. [Comparable neutralization of SARS-CoV-2 Delta AY.1 and Delta with individuals sera vaccinated with BBV152](#). Journal of Travel Medicine 28.
1224. Edara V-V, Patel M, Suthar MS. 2022. [Covaxin \(BBV152\) Vaccine Neutralizes SARS-CoV-2 Delta and Omicron variants](#). Cold Spring Harbor Laboratory.
1225. Yadav PD, Sapkal GN, Sahay RR, Patil DY, Deshpande GR, Jain R, Nyayanit DA, Shete AM, Suryawanshi A, Nandapurkar A, Gurav YK, Abraham P. 2022. [Elevated neutralization of Omicron with sera of](#)

[COVID-19 recovered and breakthrough cases vaccinated with Covaxin than two dose naïve vaccinees](#). Journal of Infection 84:834–872.

1226. Rajpurohit P, Suva M, Rajpurohit H, Singh Y, Boda P. 2021. [A Retrospective observational survey of adverse events following immunization comparing tolerability of covishield and covaxin vaccines in the real world](#). jpadr 2:21–26.
1227. Parida SP, Sahu DP, Singh AK, Alekhya G, Subba SH, Mishra A, Padhy BM, Patro BK. 2022. [Adverse events following immunization of COVID-19 \(Covaxin\) vaccine at a tertiary care center of India](#). Journal of Medical Virology 94:2453–2459.
1228. Kaur U, K.L A, Chauhan M, Joshi A, Das A, Kansal S, Jaisawal V, Patwardhan K, Chakrabarti SS. 2022. [A Prospective Observational Study on BBV152 Coronavirus Vaccine Use in Adolescents and Comparison with Adults: Interim Results of the First Real-World Safety Analysis](#). Drug Saf 45:1099–1109.
1229. Behera P, Singh AK, Subba SH, Mc A, Sahu DP, Chandanshive PD, Pradhan SK, Parida SP, Mishra A, Patro BK, Batmanabane G. 2022. [Effectiveness of COVID-19 vaccine \(Covaxin\) against breakthrough SARS-CoV-2 infection in India](#). Human Vaccines & Immunotherapeutics 18.
1230. Sagiraju HKR, Elavarasi A, Gupta N, Garg RK, Paul SS, Vig S, Sirohiya P, Ratre B, Garg R, Pandit A, Nalwa R, Kumar B, Meena VP, Wig N, Mittal S, Pahuja S, Madan K, Das N, Dwivedi T, Gupta R, Wundawalli L, Singh AR, Singh S, Mishra A, Pandey M, Matharoo KS, Kumar S, Mohan A, Guleria R, Bhatnagar S. 2021. [The effectiveness of SARS-CoV-2 vaccination in preventing severe illness and death – real-world data from a cohort of patients hospitalized with COVID-19](#). Cold Spring Harbor Laboratory.
1231. 2021. Covaxin booster dose: What is it? What does govt say about this? Hindustan Times. <https://www.hindustantimes.com/india-news/covaxin-booster-dose-what-is-it-what-does-govt-say-about-this-101625644184446.html>. Retrieved 5 December 2022.
1232. Das S. 2021. Booster dose: Bharat Biotech's nasal vaccine may be used with Covaxin. https://www.business-standard.com/article/current-affairs/booster-dose-bharat-biotech-s-nasal-vaccine-may-be-used-with-covaxin-121092500034_1.html. Retrieved 5 December 2022.
1233. 2021. [Covaxin kids trial over, Bharat Biotech to submit data to DCGI next week](#). The Times of India.
1234. WHO emergency approval for Covaxin delayed till October 5. India Today. <https://www.indiatoday.in/coronavirus-covid-19-outbreak/video/who-emergency-approval-for-covaxin-delayed-till-october-1856178-2021-09-23>. Retrieved 5 December 2022.
1235. 2022. Suspension of supply of COVID-19 vaccine (COVAXIN®). WHO - Prequalification of Medical Products (IVDs, Medicines, Vaccines and Immunization Devices, Vector Control). <https://extranet.who.int/pqweb/vaccines/suspension-supply-covid-19-vaccine-covaxin>. Retrieved 5 December 2022.

1236. Thiagarajan K. 2022. [Covid-19: WHO suspends supplies of India's Covaxin through UN agencies](#). BMJ o902.
1237. 2022. FDA lifts hold on Bharat Biotech's Covaxin clinical trials in US. Business Today. <https://www.businesstoday.in/coronavirus/story/fda-lifts-hold-on-bharat-biotechs-covaxin-clinical-trials-in-us-334915-2022-05-24>. Retrieved 30 July 2023.
1238. Moir NL. 2020. [To Boldly Remember Where We Have Already Been](#). J Appl Hist 2:17–35.
1239. McMenamin ME, Cowling BJ. 2021. [CoronaVac efficacy data from Turkey](#). The Lancet 398:1873–1874.
1240. Tanriover MD, Doğanay HL, Unal S, Akova M. 2021. [CoronaVac efficacy data from Turkey – Authors' reply](#). The Lancet 398:1874.
1241. 2022. The Valneva VLA2001 COVID-19 vaccine: What you need to know. <https://www.who.int/news-room/feature-stories/detail/the-valneva-vla2001-covid-19-vaccine--what-you-need-to-know>. Retrieved 1 December 2022.
1242. 2022. The Bharat Biotech BBV152 COVAXIN vaccine against COVID-19: What you need to know. <https://www.who.int/news-room/feature-stories/detail/the-bharat-biotech-bbv152-covaxin-vaccine-against-covid-19-what-you-need-to-know>. Retrieved 5 December 2022.
1243. Is the Sinopharm BIBP (Covilo) COVID-19 vaccine safe and effective? | COVID-19 Info Vaccines. <https://www.covid19infovaccines.com/en-posts/is-the-sinopharm-bibp-covilo-covid-19-vaccine-safe-and-effective>. Retrieved 5 December 2022.
1244. 2022. COVID-19 Vaccine (Vero Cell), Inactivated (Sinopharm). World Health Organization COVID-19 Vaccine Explainer. <https://cdn.who.int/media/docs/default-source/immunization/covid-19/16-june-22080-sinopharm-vaccine-explainer-update.pdf>. Retrieved 5 December 2022.
1245. Xiao C, Su J, Zhang C, Huang B, Mao L, Ren Z, Bai W, Li H, Lei G, Zheng J, Chen G, Liang X, Qiu C. 2022. [Effectiveness of Booster Doses of the SARS-CoV-2 Inactivated Vaccine KCONVAC against the Mutant Strains](#). Viruses 14:2016.
1246. Sattwika PD. 2022. [A Phase II Non-Randomized Open Labelled Clinical Trial to Evaluate the Safety & Immunogenicity of SARS-COV-2 Vaccine \(Vero Cell\) Inactivated as A Booster Dose](#). NCT05172193. Clinical trial registration. clinicaltrials.gov.
1247. China National Biotec Group Company Limited. 2022. [Multicenter, Randomized, Double Blind, Parallel Placebo Controlled, Phase III Clinical Trial to Evaluate the Protective Efficacy, Safety and Immunogenicity of Inactivated SARS-CoV-2 Vaccines \(Vero Cell\) in Healthy Population Aged 18 Years Old and Above](#). NCT04510207. Clinical trial registration. clinicaltrials.gov.

1248. Omma A, Batirel A, Aydin M, Yilmaz Karadag F, Erden A, Kucuksahin O, Armagan B, Güven SC, Karakas O, Gokdemir S, Altunal LN, Buber AA, Gemcioglu E, Zengin O, Inan O, Sahiner ES, Korukluoglu G, Sezer Z, Ozdarendeli A, Kara A, Ates I. 2022. Safety and immunogenicity of inactive vaccines as booster doses for COVID-19 in Türkiye: A randomized trial. *Human Vaccines & Immunotherapeutics* <https://doi.org/10.1080/21645515.2022.2122503>.
1249. Ahi M, Hamidi Farahani R, Basiri P, Karimi Rahjerdi A, Sheidaei A, Gohari K, Rahimi Z, Gholami F, Moradi M, Ghafoori Naeeni F, Saffar KN, Ghasemi S, Barati B, Moradi S, Monazah A, Pouranvari F, Forooghizadeh M. 2022. [Comparison of the Safety and Immunogenicity of FAKHRAVAC and BBIBP-CorV Vaccines when Administrated as Booster Dose: A Parallel Two Arms, Randomized, Double Blind Clinical Trial](#). *Vaccines* 10:1800.
1250. Silva VO, Yamashiro R, Ahagon CM, de Campos IB, de Oliveira IP, de Oliveira EL, López-Lopes GIS, Matsuda EM, Castejon MJ, de Macedo Brígido LF. 2021. [Inhibition of receptor-binding domain—ACE2 interaction after two doses of Sinovac's CoronaVac or AstraZeneca/Oxford's AZD1222 SARS-CoV-2 vaccines](#). *Journal of Medical Virology* 94:1217–1223.
1251. Rappuoli R, Pizza M, Del Giudice G, De Gregorio E. 2014. [Vaccines, new opportunities for a new society](#). *Proc Natl Acad Sci USA* 111:12288–12293.
1252. Donaldson B, Al-Barwani F, Young V, Scullion S, Ward V, Young S. 2014. [Virus-Like Particles, a Versatile Subunit Vaccine Platform](#), p. 159–180. *In* *Advances in Delivery Science and Technology*. Springer New York.
1253. Noad R, Roy P. 2003. [Virus-like particles as immunogens](#). *Trends in Microbiology* 11:438–444.
1254. Pollet J, Chen W-H, Strych U. 2021. [Recombinant protein vaccines, a proven approach against coronavirus pandemics](#). *Advanced Drug Delivery Reviews* 170:71–82.
1255. Nooraei S, Bahrulolum H, Hoseini ZS, Katalani C, Hajizade A, Easton AJ, Ahmadian G. 2021. [Virus-like particles: preparation, immunogenicity and their roles as nanovaccines and drug nanocarriers](#). *J Nanobiotechnol* 19.
1256. 1995. *Vaccine Design* Pharmaceutical Biotechnology. Springer US. <https://doi.org/gh3zp9>.
1257. Li W, Joshi M, Singhania S, Ramsey K, Murthy A. 2014. [Peptide Vaccine: Progress and Challenges](#). *Vaccines* 2:515–536.
1258. Garçon N, Wettendorff M, Van Mechelen M. 2011. [Role of AS04 in human papillomavirus vaccine: mode of action and clinical profile](#). *Expert Opinion on Biological Therapy* 11:667–677.
1259. Shi S, Zhu H, Xia X, Liang Z, Ma X, Sun B. 2019. [Vaccine adjuvants: Understanding the structure and mechanism of adjuvanticity](#). *Vaccine* 37:3167–3178.

1260. Stils HF. 2005. [Adjuvants and Antibody Production: Dispelling the Myths Associated with Freund's Complete and Other Adjuvants](#). ILAR Journal 46:280–293.
1261. He Y, Li J, Heck S, Lustigman S, Jiang S. 2006. [Antigenic and Immunogenic Characterization of Recombinant Baculovirus-Expressed Severe Acute Respiratory Syndrome Coronavirus Spike Protein: Implication for Vaccine Design](#). J Virol 80:5757–5767.
1262. Li J, Ulitzky L, Silberstein E, Taylor DR, Viscidi R. 2013. [Immunogenicity and Protection Efficacy of Monomeric and Trimeric Recombinant SARS Coronavirus Spike Protein Subunit Vaccine Candidates](#). Viral Immunology 26:126–132.
1263. ZHOU Z, POST P, CHUBET R, HOLTZ K, MCPHERSON C, PETRIC M, COX M. 2006. [A recombinant baculovirus-expressed S glycoprotein vaccine elicits high titers of SARS-associated coronavirus \(SARS-CoV\) neutralizing antibodies in mice](#). Vaccine 24:3624–3631.
1264. Du L, Zhao G, He Y, Guo Y, Zheng B-J, Jiang S, Zhou Y. 2007. [Receptor-binding domain of SARS-CoV spike protein induces long-term protective immunity in an animal model](#). Vaccine 25:2832–2838.
1265. Du L, Zhao G, Li L, He Y, Zhou Y, Zheng B-J, Jiang S. 2009. [Antigenicity and immunogenicity of SARS-CoV S protein receptor-binding domain stably expressed in CHO cells](#). Biochemical and Biophysical Research Communications 384:486–490.
1266. Du L, Zhao G, Chan CC, Li L, He Y, Zhou Y, Zheng B-J, Jiang S. 2010. [A 219-mer CHO-Expressing Receptor-Binding Domain of SARS-CoV S Protein Induces Potent Immune Responses and Protective Immunity](#). Viral Immunology 23:211–219.
1267. Zakhartchouk AN, Sharon C, Satkunarajah M, Auperin T, Viswanathan S, Mutwiri G, Petric M, See RH, Brunham RC, Finlay BB, Cameron C, Kelvin DJ, Cochrane A, Rini JM, Babiuk LA. 2007. [Immunogenicity of a receptor-binding domain of SARS coronavirus spike protein in mice: Implications for a subunit vaccine](#). Vaccine 25:136–143.
1268. Guo Y, Sun S, Wang K, Zhang S, Zhu W, Chen Z. 2005. [Elicitation of Immunity in Mice After Immunization with the S2 Subunit of the Severe Acute Respiratory Syndrome Coronavirus](#). DNA and Cell Biology 24:510–515.
1269. He Y, Zhou Y, Siddiqui P, Niu J, Jiang S. 2005. [Identification of Immunodominant Epitopes on the Membrane Protein of the Severe Acute Respiratory Syndrome-Associated Coronavirus](#). J Clin Microbiol 43:3718–3726.
1270. Zheng N, Xia R, Yang C, Yin B, Li Y, Duan C, Liang L, Guo H, Xie Q. 2009. [Boosted expression of the SARS-CoV nucleocapsid protein in tobacco and its immunogenicity in mice](#). Vaccine 27:5001–5007.
1271. LIU S, LENG C, LIEN S, CHI H, HUANG C, LIN C, LIAN W, CHEN C, HSIEH S, CHONG P. 2006. [Immunological characterizations of the](#)

[nucleocapsid protein based SARS vaccine candidates](#). Vaccine 24:3100–3108.

1272. Wang N, Shang J, Jiang S, Du L. 2020. [Subunit Vaccines Against Emerging Pathogenic Human Coronaviruses](#). Front Microbiol 11.
1273. Chen W-H, Du L, Chag SM, Ma C, Tricoche N, Tao X, Seid CA, Hudspeth EM, Lustigman S, Tseng C-TK, Bottazzi ME, Hotez PJ, Zhan B, Jiang S. 2013. [Yeast-expressed recombinant protein of the receptor-binding domain in SARS-CoV spike protein with deglycosylated forms as a SARS vaccine candidate](#). Human Vaccines & Immunotherapeutics 10:648–658.
1274. Lan J, Yao Y, Deng Y, Chen H, Lu G, Wang W, Bao L, Deng W, Wei Q, Gao GF, Qin C, Tan W. 2015. [Recombinant Receptor Binding Domain Protein Induces Partial Protective Immunity in Rhesus Macaques Against Middle East Respiratory Syndrome Coronavirus Challenge](#). EBioMedicine 2:1438–1446.
1275. Lan J, Deng Y, Chen H, Lu G, Wang W, Guo X, Lu Z, Gao GF, Tan W. 2014. [Tailoring Subunit Vaccine Immunity with Adjuvant Combinations and Delivery Routes Using the Middle East Respiratory Coronavirus \(MERS-CoV\) Receptor-Binding Domain as an Antigen](#). PLoS ONE 9:e112602.
1276. Nyon MP, Du L, Tseng C-TK, Seid CA, Pollet J, Naceanceno KS, Agrawal A, Algaissi A, Peng B-H, Tai W, Jiang S, Bottazzi ME, Strych U, Hotez PJ. 2018. [Engineering a stable CHO cell line for the expression of a MERS-coronavirus vaccine antigen](#). Vaccine 36:1853–1862.
1277. Tai W, Zhao G, Sun S, Guo Y, Wang Y, Tao X, Tseng C-TK, Li F, Jiang S, Du L, Zhou Y. 2016. [A recombinant receptor-binding domain of MERS-CoV in trimeric form protects human dipeptidyl peptidase 4 \(hDPP4\) transgenic mice from MERS-CoV infection](#). Virology 499:375–382.
1278. Pallesen J, Wang N, Corbett KS, Wrapp D, Kirchdoerfer RN, Turner HL, Cottrell CA, Becker MM, Wang L, Shi W, Kong W-P, Andres EL, Kettenbach AN, Denison MR, Chappell JD, Graham BS, Ward AB, McLellan JS. 2017. [Immunogenicity and structures of a rationally designed prefusion MERS-CoV spike antigen](#). Proc Natl Acad Sci USA 114.
1279. Yang Y, Deng Y, Wen B, Wang H, Meng X, Lan J, Gao GF, Tan W. 2014. [The Amino Acids 736–761 of the MERS-CoV Spike Protein Induce Neutralizing Antibodies: Implications for the Development of Vaccines and Antiviral Agents](#). Viral Immunology 27:543–550.
1280. Jiaming L, Yanfeng Y, Yao D, Yawei H, Linlin B, Baoying H, Jinghua Y, Gao GF, Chuan Q, Wenjie T. 2017. [The recombinant N-terminal domain of spike proteins is a potential vaccine against Middle East respiratory syndrome coronavirus \(MERS-CoV\) infection](#). Vaccine 35:10–18.
1281. Wang L, Shi W, Joyce MG, Modjarrad K, Zhang Y, Leung K, Lees CR, Zhou T, Yassine HM, Kanekiyo M, Yang Z, Chen X, Becker MM, Freeman M, Vogel L, Johnson JC, Olinger G, Todd JP, Bagci U, Solomon J, Mollura DJ, Hensley L, Jahrling P, Denison MR, Rao SS, Subbarao K, Kwong PD,

- Mascola JR, Kong W-P, Graham BS. 2015. [Evaluation of candidate vaccine approaches for MERS-CoV](#). Nat Commun 6.
1282. Wang Q, Wong G, Lu G, Yan J, Gao GF. 2016. [MERS-CoV spike protein: Targets for vaccines and therapeutics](#). Antiviral Research 133:165–177.
1283. Hashemzadeh A, Avan A, Ferns GA, Khazaei M. 2020. [Vaccines based on virus-like nano-particles for use against Middle East Respiratory Syndrome \(MERS\) coronavirus](#). Vaccine 38:5742–5746.
1284. Coleman CM, Liu YV, Mu H, Taylor JK, Massare M, Flyer DC, Glenn GM, Smith GE, Frieman MB. 2014. [Purified coronavirus spike protein nanoparticles induce coronavirus neutralizing antibodies in mice](#). Vaccine 32:3169–3174.
1285. Wang C, Zheng X, Gai W, Zhao Y, Wang H, Wang H, Feng N, Chi H, Qiu B, Li N, Wang T, Gao Y, Yang S, Xia X. 2016. [MERS-CoV virus-like particles produced in insect cells induce specific humoral and cellular immunity in rhesus macaques](#). Oncotarget 8:12686–12694.
1286. Lan J, Deng Y, Song J, Huang B, Wang W, Tan W. 2018. [Significant Spike-Specific IgG and Neutralizing Antibodies in Mice Induced by a Novel Chimeric Virus-Like Particle Vaccine Candidate for Middle East Respiratory Syndrome Coronavirus](#). Virol Sin 33:453–455.
1287. Li Y-D, Chi W-Y, Su J-H, Ferrall L, Hung C-F, Wu T-C. 2020. [Coronavirus vaccine development: from SARS and MERS to COVID-19](#). J Biomed Sci 27.
1288. Liljeqvist S, Ståhl S. 1999. [Production of recombinant subunit vaccines: protein immunogens, live delivery systems and nucleic acid vaccines](#). Journal of Biotechnology 73:1–33.
1289. Ausiello CM, Cassone A. 2014. [Acellular Pertussis Vaccines and Pertussis Resurgence: Revise or Replace?](#) mBio 5.
1290. Cherry JD. 2013. [Pertussis: Challenges Today and for the Future](#). PLoS Pathog 9:e1003418.
1291. McLemore MR. 2006. [Gardasil®: Introducing the New Human Papillomavirus Vaccine](#). Clinical Journal of Oncology Nursing 10:559–560.
1292. Zhang N, Zheng B-J, Lu L, Zhou Y, Jiang S, Du L. 2015. [Advancements in the development of subunit influenza vaccines](#). Microbes and Infection 17:123–134.
1293. Vaccines – COVID19 Vaccine Tracker.
<https://covid19.trackvaccines.org/vaccines/>. Retrieved 5 December 2022.
1294. Wang M, Jiang S, Wang Y. 2016. [Recent advances in the production of recombinant subunit vaccines in Pichia pastoris](#). Bioengineered 7:155–165.

1295. The Novavax vaccine against COVID-19: What you need to know. <https://www.who.int/news-room/feature-stories/detail/the-novavax-vaccine-against-covid-19-what-you-need-to-know>. Retrieved 30 July 2023.
1296. Keech C, Albert G, Cho I, Robertson A, Reed P, Neal S, Plested JS, Zhu M, Cloney-Clark S, Zhou H, Smith G, Patel N, Frieman MB, Haupt RE, Logue J, McGrath M, Weston S, Piedra PA, Desai C, Callahan K, Lewis M, Price-Abbott P, Formica N, Shinde V, Fries L, Lickliter JD, Griffin P, Wilkinson B, Glenn GM. 2020. [Phase 1-2 Trial of a SARS-CoV-2 Recombinant Spike Protein Nanoparticle Vaccine](#). N Engl J Med 383:2320–2332.
1297. Tian J-H, Patel N, Haupt R, Zhou H, Weston S, Hammond H, Logue J, Portnoff AD, Norton J, Guebre-Xabier M, Zhou B, Jacobson K, Maciejewski S, Khatoon R, Wisniewska M, Moffitt W, Kluepfel-Stahl S, Ekechukwu B, Papin J, Boddapati S, Jason Wong C, Piedra PA, Frieman MB, Massare MJ, Fries L, Bengtsson KL, Stertman L, Ellingsworth L, Glenn G, Smith G. 2021. [SARS-CoV-2 spike glycoprotein vaccine candidate NVX-CoV2373 immunogenicity in baboons and protection in mice](#). Nat Commun 12.
1298. Yee CM, Zak AJ, Hill BD, Wen F. 2018. [The Coming Age of Insect Cells for Manufacturing and Development of Protein Therapeutics](#). Ind Eng Chem Res 57:10061–10070.
1299. Riley TP, Chou H-T, Hu R, Bzymek KP, Correia AR, Partin AC, Li D, Gong D, Wang Z, Yu X, Manzanillo P, Garces F. 2021. [Enhancing the Prefusion Conformational Stability of SARS-CoV-2 Spike Protein Through Structure-Guided Design](#). Front Immunol 12.
1300. Heath PT, Galiza EP, Baxter DN, Boffito M, Browne D, Burns F, Chadwick DR, Clark R, Cosgrove C, Galloway J, Goodman AL, Heer A, Higham A, Iyengar S, Jamal A, Jeanes C, Kalra PA, Kyriakidou C, McAuley DF, Meyrick A, Minassian AM, Minton J, Moore P, Munsoor I, Nicholls H, Osanlou O, Packham J, Pretswell CH, San Francisco Ramos A, Saralaya D, Sheridan RP, Smith R, Soiza RL, Swift PA, Thomson EC, Turner J, Viljoen ME, Albert G, Cho I, Dubovsky F, Glenn G, Rivers J, Robertson A, Smith K, Toback S. 2021. [Safety and Efficacy of NVX-CoV2373 Covid-19 Vaccine](#). N Engl J Med 385:1172–1183.
1301. Dunkle LM, Kotloff KL, Gay CL, Áñez G, Adelglass JM, Barrat Hernández AQ, Harper WL, Duncanson DM, McArthur MA, Florescu DF, McClelland RS, Garcia-Fragoso V, Riesenberger RA, Musante DB, Fried DL, Safirstein BE, McKenzie M, Jeanfreau RJ, Kingsley JK, Henderson JA, Lane DC, Ruíz-Palacios GM, Corey L, Neuzil KM, Coombs RW, Greninger AL, Hutter J, Ake JA, Smith K, Woo W, Cho I, Glenn GM, Dubovsky F. 2022. [Efficacy and Safety of NVX-CoV2373 in Adults in the United States and Mexico](#). N Engl J Med 386:531–543.
1302. Dunkle LM, Kotloff KL, Gay CL, Áñez G, Adelglass JM, Barrat Hernández AQ, Harper WL, Duncanson DM, McArthur MA, Florescu DF, McClelland RS, Garcia-Fragoso V, Riesenberger RA, Musante DB, Fried DL, Safirstein BE, McKenzie M, Jeanfreau RJ, Kingsley JK, Henderson JA, Lane DC, Ruíz-Palacios GM, Corey L, Neuzil KM, Coombs RW, Greninger AL, Hutter J,

Ake JA, Smith K, Woo W, Cho I, Glenn GM, Dubovsky F. 2021. [Efficacy and Safety of NVX-CoV2373 in Adults in the United States and Mexico](#). Cold Spring Harbor Laboratory.

1303. Novavax missed its global moonshot but is angling to win over mRNA defectors. <https://www.asbmb.org/asbmb-today/industry/060422/novavax-missed-its-global-moonshot-but-is-angling>. Retrieved 30 July 2023.
1304. Tinari S, Riva C. 2021. [Covid-19: Whatever happened to the Novavax vaccine?](#) BMJ n2965.
1305. Price WN II, Rai AK, Minssen T. 2020. [Knowledge transfer for large-scale vaccine manufacturing](#). Science 369:912–914.
1306. 2021. 'They rushed the process': Vaccine maker's woes hamper global inoculation campaign. POLITICO. <https://www.politico.com/news/2021/10/19/novavax-vaccine-rush-process-global-campaign-516298>. Retrieved 30 July 2023.
1307. Kimball S. 2022. Novavax confident Covid vaccine will receive FDA authorization in June after delays. CNBC. <https://www.cnbc.com/2022/05/13/novavax-confident-covid-vaccine-will-receive-fda-authorization-in-june-after-delays.html>. Retrieved 30 July 2023.
1308. WHO lists 9th COVID-19 vaccine for emergency use with aim to increase access to vaccination in lower-income countries. <https://www.who.int/news/item/17-12-2021-who-lists-9th-covid-19-vaccine-for-emergency-use-with-aim-to-increase-access-to-vaccination-in-lower-income-countries>. Retrieved 30 July 2023.
1309. Novavax COVID-19 vaccine Nuvaxovid approved by MHRA. GOV.UK. <https://www.gov.uk/government/news/novavax-covid-19-vaccine-nuvaxovid-approved-by-mhra>. Retrieved 5 December 2022.
1310. 2021. EMA recommends Nuvaxovid for authorisation in the EU. <https://www.ema.europa.eu/en/news/ema-recommends-nuvaxovid-authorisation-eu>.
1311. Commissioner O of the. 2022. Coronavirus (COVID-19) Update: FDA Authorizes Emergency Use of Novavax COVID-19 Vaccine, Adjuvanted. FDA. <https://www.fda.gov/news-events/press-announcements/coronavirus-covid-19-update-fda-authorizes-emergency-use-novavax-covid-19-vaccine-adjuvanted>. Retrieved 5 December 2022.
1312. Reuters. 2022. [Gavi rejects Novavax's claim on COVID vaccine deal breach](#). Reuters.
1313. Shinde V, Bhikha S, Hoosain Z, Archary M, Bhorat Q, Fairlie L, Lalloo U, Masilela MSL, Moodley D, Hanley S, Fouche L, Louw C, Tameris M, Singh N, Goga A, Dheda K, Grobbelaar C, Kruger G, Carrim-Ganey N, Baillie V, de Oliveira T, Lombard Koen A, Lombaard JJ, Mngqibisa R, Bhorat AE, Benadé G, Lalloo N, Pitsi A, Vollgraaff P-L, Luabeya A, Esmail A, Petrick FG, Oommen-Jose A, Foulkes S, Ahmed K, Thombrayil A, Fries

- L, Cloney-Clark S, Zhu M, Bennett C, Albert G, Faust E, Plested JS, Robertson A, Neal S, Cho I, Glenn GM, Dubovsky F, Madhi SA. 2021. [Efficacy of NVX-CoV2373 Covid-19 Vaccine against the B.1.351 Variant](#). N Engl J Med 384:1899–1909.
1314. Mallory R, Formica N, Pfeiffer S, Wilkinson B, Marcheschi A, Albert G, McFall H, Robinson M, Plested JS, Zhu M, Cloney-Clark S, Zhou B, Chau G, Robertson A, Maciejewski S, Smith G, Patel N, Glenn GM, Dubovsky F. 2021. [Immunogenicity and Safety Following a Homologous Booster Dose of a SARS-CoV-2 recombinant spike protein vaccine \(NVX-CoV2373\): A Phase 2 Randomized Placebo-Controlled Trial](#). Cold Spring Harbor Laboratory.
1315. Novavax. 2022. [A 2-Part, Phase 1/2, Randomized, Observer-Blinded Study To Evaluate The Safety And Immunogenicity Of A SARS-CoV-2 Recombinant Spike Protein Nanoparticle Vaccine \(SARS-CoV-2 rS\) With Or Without MATRIX-M™ Adjuvant In Healthy Subjects](#). NCT04368988. Clinical trial registration. clinicaltrials.gov.
1316. Bhiman JN, Richardson SI, Lambson BE, Kgagudi P, Mzindle N, Kaldine H, Crowther C, Gray G, Bekker L-G, Shinde V, Bennett C, Glenn GM, Madhi S, Moore PL. 2022. [Novavax NVX-COV2373 triggers potent neutralization of Omicron sub-lineages](#). Cold Spring Harbor Laboratory.
1317. Toback S, Galiza E, Cosgrove C, Galloway J, Goodman AL, Swift PA, Rajaram S, Graves-Jones A, Edelman J, Burns F, Minassian AM, Cho I, Kumar L, Plested JS, Rivers EJ, Robertson A, Dubovsky F, Glenn G, Heath PT. 2021. [Safety, Immunogenicity, and Efficacy of a COVID-19 Vaccine \(NVX-CoV2373\) Co-administered With Seasonal Influenza Vaccines](#). Cold Spring Harbor Laboratory.
1318. Novavax Initiates Phase 1/2 Clinical Trial of Combination Vaccine for COVID-19 and Seasonal Influenza. Novavax Investor Relations. <https://ir.novavax.com/2021-09-08-Novavax-Initiates-Phase-1-2-Clinical-Trial-of-Combination-Vaccine-for-COVID-19-and-Seasonal-Influenza>. Retrieved 5 December 2022.
1319. Massare MJ, Patel N, Zhou B, Maciejewski S, Flores R, Guebre-Xabier M, Tian J-H, Portnoff AD, Fries L, Shinde V, Ellingsworth L, Glenn G, Smith G. 2021. [Combination Respiratory Vaccine Containing Recombinant SARS-CoV-2 Spike and Quadrivalent Seasonal Influenza Hemagglutinin Nanoparticles with Matrix-M Adjuvant](#). Cold Spring Harbor Laboratory.
1320. Reardon S. 2021. [Cuba's bet on home-grown COVID vaccines is paying off](#). Nature 600:15–16.
1321. Osterholm M, Rabadán-Diehl C, Anzinger J, Bottazzi M, Christie-Samuels C, Erondú N, Marrazzo J, Milan S, Quashie P, Schwaab T, Williams D. 2022. [EXECUTIVE SUMMARY Insights from Cuba's COVID-19 Vaccine Enterprise: Report from a High Level Fact-Finding Delegation to Cuba](#). MEDICC Review 24.
1322. Toledo-Romaní ME, García-Carmenate M, Valenzuela-Silva C, Baldoquín-Rodríguez W, Martínez-Pérez M, Rodríguez-González M,

Paredes-Moreno B, Mendoza-Hernández I, González-Mujica Romero R, Samón-Tabio O, Velazco-Villares P, Bacallao-Castillo JP, Licea-Martín E, Rodríguez-Ortega M, Herrera-Marrero N, Caballero-González E, Egües-Torres L, Duarte-González R, García-Blanco S, Pérez-Cabrera S, Huete-Ferreira S, Idalmis-Cisnero K, Fonte-Galindo O, Meliá-Pérez D, Rojas-Remedios I, Doroud D, Gouya MM, Biglari A, Fernández-Castillo S, Climent-Ruiz Y, Valdes-Balbín Y, García-Rivera D, Van der Stuyft P, Verev-Bencomo V. 2023. [Safety and efficacy of the two doses conjugated protein-based SOBERANA-02 COVID-19 vaccine and of a heterologous three-dose combination with SOBERANA-Plus: a double-blind, randomised, placebo-controlled phase 3 clinical trial](#). The Lancet Regional Health - Americas 18:100423.

1323. Toledo-Romani ME, García-Carmenate M, Verdecia-Sánchez L, Pérez-Rodríguez S, Rodríguez-González M, Valenzuela-Silva C, Paredes-Moreno B, Sanchez-Ramirez B, González-Mugica R, Hernández-Garcia T, Orosa-Vázquez I, Díaz-Hernández M, Pérez-Guevara MT, Enriquez-Puertas J, Noa-Romero E, Palenzuela-Diaz A, Baro-Roman G, Mendoza-Hernández I, Muñoz Y, Gómez-Maceo Y, Santos-Vega BL, Fernandez-Castillo S, Climent-Ruiz Y, Rodríguez-Noda L, Santana-Mederos D, García-Vega Y, Chen G-W, Doroud D, Biglari A, Boggiano-Ayo T, Valdés-Balbín Y, Rivera DG, García-Rivera D, Vérez-Bencomo V, Cubas-Curbelo M, Rodríguez-Castillo PG, Acevedo-Martínez Y, Estoque-Cabrera S, Ávila-Cabreja JA, Alfaro-Guzmán A, Zulueta-Pérez L, Espino-Rojas NT, Medinas-Santos GM, Sarda-Rodríguez IL, Acosta-Martinez MA, Reyes-Matienzo R, Coviella-Arttime JM, Morffi-Cinta I, Martínez-Pérez M, Valera-Fernández R, Garcés-Hechavarría A, Martínez-Bedoya D, Garrido-Arteaga R, Cardoso-SanJorge F, Ramírez-Gonzalez U, Quintero-Moreno L, Ontivero-Pino I, Martínez-Rivera R, Guillén-Obregón B, Lora-García J, Medina-Nápoles M, Espi-Ávila J, Fontanies-Fernández M, Domínguez-Pentón YR, Bergado-Baez G, Pi-Estopián F, Ojito-Magaz E, Rodríguez M, Cruz-Sui O, García-Montero M, Dubed-Echevarría M, García-López E, Galano-Frutos E, Perez-Perez A, Morales-Ruano S, Brito-Pascual I, Amoroto M, Arteaga-García A. 2022. [Safety and immunogenicity of anti-SARS-CoV-2 heterologous scheme with SOBERANA 02 and SOBERANA Plus vaccines: Phase IIb clinical trial in adults](#). Med 3:760–773.e5.
1324. Limonta-Fernández M, Chinea-Santiago G, Martín-Dunn AM, Gonzalez-Roche D, Bequet-Romero M, Marquez-Perera G, González-Moya I, Canaan-Haden-Ayala C, Cabrales-Rico A, Espinosa-Rodríguez LA, Ramos-Gómez Y, Andujar-Martínez I, González-López LJ, de la Iglesia MP, Zamora-Sanchez J, Cruz-Sui O, Lemos-Pérez G, Cabrera-Herrera G, Valdes-Hernández J, Martinez-Diaz E, Pimentel-Vazquez E, Ayala-Avila M, Guillén-Nieto G. 2022. [An engineered SARS-CoV-2 receptor-binding domain produced in Pichia pastoris as a candidate vaccine antigen](#). New Biotechnology 72:11–21.
1325. Hernández-Bernal F, Ricardo-Cobas MC, Martín-Bauta Y, Navarro-Rodríguez Z, Piñera-Martínez M, Quintana-Guerra J, Urrutia-Pérez K, Urrutia-Pérez K, Chávez-Chong CO, Azor-Hernández JL, Rodríguez-Reinoso JL, Lobaina-Lambert L, Colina-Ávila E, Bizet-Almeida J, Rodríguez-Nuviola J, del Valle-Piñera S, Ramírez-Domínguez M, Tablada-Ferreiro E, Alonso-Valdés M, Lemos-Pérez G, Guillén-Nieto GE,

Palenzuela-Díaz A, Noa-Romero E, Limonta-Fernández M, Fernández-Ávila JM, Ali-Mros NA, del Toro-Lahera L, Remedios-Reyes R, Ayala-Ávila M, Muzio-González VL. 2022. [Safety, tolerability, and immunogenicity of a SARS-CoV-2 recombinant spike RBD protein vaccine: A randomised, double-blind, placebo-controlled, phase 1-2 clinical trial \(ABDALA Study\)](#). eClinicalMedicine 46:101383.

1326. Cuba's Abdala COVID-19 vaccine enters phase 3 clinical trial - Xinhua | English.news.cn.
http://www.xinhuanet.com/english/northamerica/2021-03/20/c_139823225.htm. Retrieved 30 July 2023.
1327. Reuters. 2021. [Cuba says Abdala vaccine 92.28% effective against coronavirus](#). Reuters.
1328. 2021. [Abdala, con tres dosis, tiene una eficacia de 92,28 %](#).
1329. Staff O. 2021. How was the efficacy of the Cuban COVID-19 vaccine candidates calculated? OnCubaNews English.
<https://oncubanews.com/en/cuba/how-was-the-efficacy-of-the-cuban-covid-19-vaccine-candidates-calculated/>. Retrieved 30 July 2023.
1330. 2021. Aprueba el CECMED el Autorizo de Uso de Emergencia del candidato vacunal cubano ABDALA.
<https://www.cecmed.cu/noticias/aprueba-cecmed-autorizo-uso-emergencia-candidato-vacunal-cubano-abdala>. Retrieved 22 December 2022.
1331. Taylor L. 2021. [Cuba's home-grown vaccines have massively cut covid-19 cases](#).
1332. Hernández-Bernal F, Ricardo-Cobas MC, Martín-Bauta Y, Rodríguez-Martínez E, Urrutia-Pérez K, Urrutia-Pérez K, Quintana-Guerra J, Navarro-Rodríguez Z, Piñera-Martínez M, Rodríguez-Reinoso JL, Chávez-Chong CO, Baladrón-Castrillo I, Melo-Suárez G, Batista-Izquierdo A, Pupo-Micó A, Mora-Betancourt R, Soler-Cano D, Bizet-Almeida J, Martínez-Rodríguez MC, Lobaina-Lambert L, Velázquez-Pérez VM, Soler-Díaz J, Blanco-Garrido Y, Laurencio-Vallina S, Meriño-Hechavarría T, Carmenaty-Campos N, Rodríguez-Montero E, Limonta-Fernández M, Alonso-Valdés M, Hernández-Rodríguez R, Pimentel-Vázquez E, Catasús-Álvarez KM, Cabrera-Núñez MV, Ayala-Ávila M, Muzio-González VL. 2022. [A phase 3, randomised, double-blind, placebo-controlled clinical trial for adult evaluation of the efficacy and safety of a SARS-CoV-2 recombinant spike RBD protein vaccine \(ABDALA-3 Study\)](#). Cold Spring Harbor Laboratory.
1333. Más-Bermejo PI, Dickinson-Meneses FO, Almenares-Rodríguez K, Sánchez-Valdés L, Guinovart-Díaz R, Vidal-Ledo M, Galbán-García E, Olivera-Nodarse Y, Morgado-Vega I, Dueñas-Carrera S, Pujol M, Hernández-Bernal F, Limonta-Fernández M, Guillén-Nieto G, Muzio-González VL, Ayala-Ávila M. 2022. [Cuban Abdala vaccine: Effectiveness in preventing severe disease and death from COVID-19 in Havana, Cuba; A cohort study](#). The Lancet Regional Health - Americas 16:100366.

1334. 2023. [Actualización de la estrategia para el desarrollo de las vacunas cubanas](#) Ministerio de Salud Pública de Cuba.
1335. Gorry C. 2022. [Vaccines and Public Trust: Containing COVID-19 in Cuba](#) Verena Muzio-González PhD DSc Director of Clinical Research, Genetic Engineering and Biotechnology Center. MEDICC Review 24:9.
1336. 2021. [Status of COVID-19 Vaccines within WHO EUL/PQ evaluation process](#) Assessment. Status of COVID-19 Vaccines within WHO EUL/PQ Evaluation Process.
1337. Taylor L. 2022. [Covid-19: Cuba will request WHO approval for homegrown vaccine](#). BMJ o230.
1338. Serrano-Barrera OR, Bello-Rodríguez MM, Pupo-Rodríguez OL, Robinson-Agramonte M de los A, Pérez O. 2022. [Variantes ómicron y delta de SARS-CoV-2 conservan epítopes presentes en vacunas cubanas anti-covid-19 Abdala y Soberana](#). Revista Electrónica Dr Zoilo E Marinello Vidaurreta 47.
1339. EFE. 2021. Cuba to apply booster doses of its COVID-19 vaccines. OnCubaNews English. <https://oncubanews.com/en/coronavirus/cuba-to-apply-booster-doses-of-its-covid-19-vaccines/>. Retrieved 30 July 2023.
1340. Meredith S. 2022. Why Cuba's extraordinary Covid vaccine success could provide the best hope for low-income countries. CNBC. <https://www.cnbc.com/2022/01/13/why-cubas-extraordinary-covid-vaccine-success-could-provide-the-best-hope-for-the-global-south.html>. Retrieved 30 July 2023.
1341. Frank M, Sherwood D. 2021. [Cuba to fast-track boosters as Omicron looms](#). Reuters.
1342. Cuba COVID - Coronavirus Statistics - Worldometer. <https://www.worldometers.info/coronavirus/country/cuba>. Retrieved 30 July 2023.
1343. Taylor L. 2021. [Why Cuba developed its own covid vaccine—and what happened next](#). BMJ n1912.
1344. Duong D, Vogel L. 2022. [Why is WHO pushing back on a Health Canada-approved Medicago SARS-CoV-2 vaccine?](#) CMAJ 194:E504–E505.
1345. Covifenz®. Medicago. <https://medicago.com/en/our-products/our-vaccines/covifenz-covid-19-vlp-vaccine/>. Retrieved 5 December 2022.
1346. Ward BJ, Makarkov A, Séguin A, Pillet S, Trépanier S, Dhaliwall J, Libman MD, Vesikari T, Landry N. 2020. [Efficacy, immunogenicity, and safety of a plant-derived, quadrivalent, virus-like particle influenza vaccine in adults \(18–64 years\) and older adults \(≥65 years\): two multicentre, randomised phase 3 trials](#). The Lancet 396:1491–1503.
1347. Ward BJ, Gobeil P, Séguin A, Atkins J, Boulay I, Charbonneau P-Y, Couture M, D'Aoust M-A, Dhaliwall J, Finkle C, Hager K, Mahmood A,

- Makarkov A, Cheng MP, Pillet S, Schimke P, St-Martin S, Trépanier S, Landry N. 2021. [Phase 1 randomized trial of a plant-derived virus-like particle vaccine for COVID-19](#). *Nat Med* 27:1071–1078.
1348. Garçon N, Vaughn DW, Didierlaurent AM. 2012. [Development and evaluation of AS03, an Adjuvant System containing \$\alpha\$ -tocopherol and squalene in an oil-in-water emulsion](#). *Expert Review of Vaccines* 11:349–366.
1349. Hager KJ, Pérez Marc G, Gobeil P, Diaz RS, Heizer G, Llapur C, Makarkov AI, Vasconcellos E, Pillet S, Riera F, Saxena P, Geller Wolff P, Bhutada K, Wallace G, Aazami H, Jones CE, Polack FP, Ferrara L, Atkins J, Boulay I, Dhaliwall J, Charland N, Couture MMJ, Jiang-Wright J, Landry N, Lapointe S, Lorin A, Mahmood A, Moulton LH, Pahmer E, Parent J, Séguin A, Tran L, Breuer T, Ceregido M-A, Koutsoukos M, Roman F, Namba J, D'Aoust M-A, Trepanier S, Kimura Y, Ward BJ. 2022. [Efficacy and Safety of a Recombinant Plant-Based Adjuvanted Covid-19 Vaccine](#). *N Engl J Med* 386:2084–2096.
1350. Medicago and GSK announce the approval by Health Canada of COVIFENZ®, an adjuvanted plant-based COVID-19 vaccine | GSK. <https://www.gsk.com/en-gb/media/press-releases/medicago-and-gsk-announce-the-approval-by-health-canada-of-covifenz/>. Retrieved 5 December 2022.
1351. D'Aoust M-A, Lavoie P-O, Couture MM-J, Trépanier S, Guay J-M, Dargis M, Mongrand S, Landry N, Ward BJ, Vézina L-P. 2008. [Influenza virus-like particles produced by transient expression in *Nicotiana benthamiana* induce a protective immune response against a lethal viral challenge in mice](#). *Plant Biotechnology Journal* 6:930–940.
1352. Dyer O. 2022. [Covid-19: WHO set to reject Canadian plant based vaccine because of links with tobacco industry](#). *BMJ* o811.
1353. Chen Q, Lai H. 2013. [Plant-derived virus-like particles as vaccines](#). *Human Vaccines & Immunotherapeutics* 9:26–49.
1354. News · PL-C. 2022. Big Tobacco's divestment from Quebec's Medicago 'a step in the right direction' for its COVID vaccine | CBC News. CBC. <https://www.cbc.ca/news/health/philips-morris-divestment-medicago-1.6700103>. Retrieved 30 July 2023.
1355. 2022. COVID-19 vaccine: Canadian Immunization Guide. <https://www.canada.ca/en/public-health/services/publications/healthy-living/canadian-immunization-guide-part-4-active-vaccines/page-26-covid-19-vaccine.html>. Retrieved 22 December 2022.
1356. . <https://medicago.com/en/news-releases>.
1357. Mohammadi M, Shayestehpour M, Mirzaei H. 2021. [The impact of spike mutated variants of SARS-CoV2 \[Alpha, Beta, Gamma, Delta, and Lambda\] on the efficacy of subunit recombinant vaccines](#). *The Brazilian Journal of Infectious Diseases* 25:101606.
1358. Triccas JA, Kint J, Wurm FM. 2022. [Affordable SARS-CoV-2 protein vaccines for the pandemic endgame](#). *npj Vaccines* 7.

1359. Rodríguez Mega E. 2020. [Latin American scientists join the coronavirus vaccine race: 'No one's coming to rescue us'](#). Nature 582:470–471.
1360. Rapaka RR, Hammershaimb EA, Neuzil KM. 2021. [Are Some COVID-19 Vaccines Better Than Others? Interpreting and Comparing Estimates of Efficacy in Vaccine Trials](#). Clinical Infectious Diseases 74:352–358.
1361. Mills MC, Salisbury D. 2021. [The challenges of distributing COVID-19 vaccinations](#). EClinicalMedicine 31:100674.
1362. Emanuel EJ, Persad G, Kern A, Buchanan A, Fabre C, Halliday D, Heath J, Herzog L, Leland RJ, Lemango ET, Luna F, McCoy MS, Norheim OF, Ottersen T, Schaefer GO, Tan K-C, Wellman CH, Wolff J, Richardson HS. 2020. [An ethical framework for global vaccine allocation](#). Science 369:1309–1312.
1363. Matrajt L, Eaton J, Leung T, Brown ER. 2021. [Vaccine optimization for COVID-19: Who to vaccinate first?](#) Sci Adv 7.
1364. Holder J. 2021. [Tracking Coronavirus Vaccinations Around the World](#). The New York Times.
1365. Mathieu E, Ritchie H, Rodés-Guirao L, Appel C, Giattino C, Hasell J, Macdonald B, Dattani S, Beltekian D, Ortiz-Ospina E, Roser M. 2020. [Coronavirus Pandemic \(COVID-19\)](#). Our World in Data.
1366. Goodman PS. 2020. [One Vaccine Side Effect: Global Economic Inequality](#). The New York Times.
1367. ECCLESTON-TURNER M, UPTON H. 2021. [International Collaboration to Ensure Equitable Access to Vaccines for COVID-19: The ACT-Accelerator and the COVAX Facility](#). The Milbank Quarterly 99:426–449.
1368. Fair allocation mechanism for COVID-19 vaccines through the COVAX Facility. <https://www.who.int/publications/m/item/fair-allocation-mechanism-for-covid-19-vaccines-through-the-covax-facility>. Retrieved 5 December 2022.
1369. Kupferschmidt K. 2020. [Global plan seeks to promote vaccine equity, spread risks](#). Science 369:489–490.
1370. Usher AD. 2021. [A beautiful idea: how COVAX has fallen short](#). The Lancet 397:2322–2325.
1371. Kibirige H. 2022. plotnine. <https://github.com/has2k1/plotnine>. Retrieved 5 December 2022.
1372. Smoothed conditional means — geom_smooth. https://ggplot2.tidyverse.org/reference/geom_smooth.html. Retrieved 5 December 2022.
1373. 2022. COVID-19 Dataset by Our World in Data. Our World in Data. <https://github.com/owid/covid-19-data/blob/9e0b8314da28f155a88bbaf7617390c01c1c26da/public/data/vaccinations/locations.csv>. Retrieved 5 December 2022.

1374. Twohey M, Collins K, Thomas K. 2020. [With First Dibs on Vaccines, Rich Countries Have 'Cleared the Shelves'](#). The New York Times.
1375. Obinna DN. 2021. [Solidarity across borders: A pragmatic need for global COVID-19 vaccine equity](#). Health Planning & Management 37:21–29.
1376. 2021. [Covid-19 vaccinations: African nations miss WHO target](#). BBC News.
1377. Herzog LM, Norheim OF, Emanuel EJ, McCoy MS. 2021. [Covax must go beyond proportional allocation of covid vaccines to ensure fair and equitable access](#). BMJ m4853.
1378. COVAX. <https://www.who.int/initiatives/act-accelerator/covax>. Retrieved 5 December 2022.
1379. COVAX vaccine roll-out. <https://www.gavi.org/covax-vaccine-roll-out>. Retrieved 5 December 2022.
1380. 2021. [Countries now scrambling for COVID-19 vaccines may soon have surpluses to donate](#). AAAS Articles DO Group. American Association for the Advancement of Science (AAAS).
1381. Bubar KM, Reinholt K, Kissler SM, Lipsitch M, Cobey S, Grad YH, Larremore DB. 2021. [Model-informed COVID-19 vaccine prioritization strategies by age and serostatus](#). Science 371:916–921.
1382. Grauer J, Löwen H, Liebchen B. 2020. [Strategic spatiotemporal vaccine distribution increases the survival rate in an infectious disease like Covid-19](#). Sci Rep 10.
1383. Williams J, Degeling C, McVernon J, Dawson A. 2021. [How should we conduct pandemic vaccination?](#) Vaccine 39:994–999.
1384. Jecker NS, Wightman AG, Diekema DS. 2021. [Vaccine ethics: an ethical framework for global distribution of COVID-19 vaccines](#). J Med Ethics medethics-2020-107036.
1385. Tran TN-A, Wikle N, Albert J, Inam H, Strong E, Brinda K, Leighow SM, Yang F, Hossain S, Pritchard JR, Chan P, Hanage WP, Hanks EM, Boni MF. 2021. [Optimal SARS-CoV-2 vaccine allocation using real-time seroprevalence estimates in Rhode Island and Massachusetts](#). Cold Spring Harbor Laboratory.
1386. Mathieu E, Ritchie H, Rodés-Guirao L, Appel C, Giattino C, Hasell J, Macdonald B, Dattani S, Beltekian D, Ortiz-Ospina E, Roser M. 2020. [Coronavirus Pandemic \(COVID-19\)](#). Our World in Data.
1387. Ferran M. 2022. CORBEVAX, a new patent-free COVID-19 vaccine, could be a pandemic game changer globally. The Conversation. <http://theconversation.com/corbevax-a-new-patent-free-covid-19-vaccine-could-be-a-pandemic-game-changer-globally-174672>. Retrieved 30 July 2023.

1388. Biological E Limited: Corbevax – COVID19 Vaccine Tracker. <https://covid19.trackvaccines.org/vaccines/54/>. Retrieved 30 July 2023.
1389. James ER. 2021. [Disrupting vaccine logistics](#). International Health 13:211–214.
1390. AboulFotouh K, Cui Z, Williams RO III. 2021. [Next-Generation COVID-19 Vaccines Should Take Efficiency of Distribution into Consideration](#). AAPS PharmSciTech 22.
1391. Sguazzin A. 2021. [Africa Left With Few Options for Vaccines, South Africa Says](#). Bloomberg.
1392. COVAX: ensuring global equitable access to COVID-19 vaccines | UNICEF Supply Division. <https://www.unicef.org/supply/covax-ensuring-global-equitable-access-covid-19-vaccines>. Retrieved 30 July 2023.
1393. Bajaj SS, Maki L, Stanford FC. 2022. [Vaccine apartheid: global cooperation and equity](#). The Lancet 399:1452–1453.
1394. Sparke M, Levy O. 2022. [Competing Responses to Global Inequalities in Access to COVID Vaccines: Vaccine Diplomacy and Vaccine Charity Versus Vaccine Liberty](#). Clinical Infectious Diseases 75:S86–S92.
1395. Grenfell RFQ, Oyeyemi OT. 2022. [Access to COVID-19 vaccines and testing in Africa: the importance of COVAX - Nigeria as a case study](#). Pathogens and Global Health 117:152–166.
1396. Taylor A. 2022. [Covax promised 2 billion vaccine doses to help the world's neediest in 2021. It won't deliver even half that](#). Washington Post.
1397. Ten health issues WHO will tackle this year. <https://www.who.int/news-room/spotlight/ten-threats-to-global-health-in-2019>. Retrieved 30 July 2023.
1398. Lopes L, 2021. 2021. KFF COVID-19 Vaccine Monitor: January 2021. KFF. <https://www.kff.org/coronavirus-covid-19/report/kff-covid-19-vaccine-monitor-january-2021/>. Retrieved 30 July 2023.
1399. Rosenbaum L. 2021. [Escaping Catch-22 — Overcoming Covid Vaccine Hesitancy](#). N Engl J Med 384:1367–1371.
1400. Unroe KT, Evans R, Weaver L, Rusyniak D, Blackburn J. 2021. [Willingness of Long-Term Care Staff to Receive a COVID-19 Vaccine: A Single State Survey](#). J Am Geriatr Soc 69:593–599.
1401. Ruiz JB, Bell RA. 2022. [Parental COVID-19 Vaccine Hesitancy in the United States](#). Public Health Rep 137:1162–1169.
1402. Cénat JM, Noorishad P, Moshirian Farahi SMM, Darius WP, Mesbahi EI, Aouame A, Onesi O, Broussard C, Furyk SE, Yaya S, Caulley L, Chomienne M, Etowa J, Labelle PR. 2022. [Prevalence and factors related to COVID-19 vaccine hesitancy and unwillingness in Canada: A systematic review and meta-analysis](#). Journal of Medical Virology 95.

1403. Steinert JI, Sternberg H, Prince H, Fasolo B, Galizzi MM, Bütthe T, Veltri GA. 2022. [COVID-19 vaccine hesitancy in eight European countries: Prevalence, determinants, and heterogeneity](#). Sci Adv 8.
1404. Ackah BBB, Woo M, Stallwood L, Fazal ZA, Okpani A, Ukah UV, Adu PA. 2022. [COVID-19 vaccine hesitancy in Africa: a scoping review](#). glob health res policy 7.
1405. Harapan H, Anwar S, Yufika A, Sharun K, Gachabayov M, Fahriani M, Husnah M, Raad R, Abdalla RY, Adam RY, Khiri NME, Ismaeil MIH, Ismail AY, Kacem W, Dahman NBH, Teyeb Z, Aloui K, Hafsi M, Ferjani M, Deeb DA, Emad D, Abbas KS, Monib FA, Sami FS, Subramaniam R, Panchawagh S, Anandu S, Haque MA, Ferreto LED, Briones MFC, Morales RBI, Díaz SAL, Aburto JTO, Rojas JET, Balogun EO, Enitan SS, Yomi AR, Durosini A, Ezigbo ED, Adejumo EN, Babadi E, Kakemam E, Malik NI, Ullah I, Rosiello DF, Emran TB, Wendt GW, Arab-Zozani M, Wagner AL, Mudatsir M. 2021. [Vaccine hesitancy among communities in ten countries in Asia, Africa, and South America during the COVID-19 pandemic](#). Pathogens and Global Health 116:236–243.
1406. Dereje N, Tesfaye A, Tamene B, Alemeshet D, Abe H, Tesfa N, Gedion S, Biruk T, Lakew Y. 2022. [COVID-19 vaccine hesitancy in Addis Ababa, Ethiopia: a mixed-method study](#). BMJ Open 12:e052432.
1407. Rathje S, He JK, Roozenbeek J, Van Bavel JJ, van der Linden S. 2022. [Social media behavior is associated with vaccine hesitancy](#). PNAS Nexus 1.
1408. Pertwee E, Simas C, Larson HJ. 2022. [An epidemic of uncertainty: rumors, conspiracy theories and vaccine hesitancy](#). Nat Med 28:456–459.
1409. Ng JWJ, Vaithilingam S, Nair M, Hwang L-A, Musa KI. 2022. [Key predictors of COVID-19 vaccine hesitancy in Malaysia: An integrated framework](#). PLoS ONE 17:e0268926.
1410. Schernhammer E, Weitzer J, Laubichler MD, Birmann BM, Bertau M, Zenk L, Caniglia G, Jäger CC, Steiner G. 2021. [Correlates of COVID-19 vaccine hesitancy in Austria: trust and the government](#). Journal of Public Health 44:e106–e116.
1411. Head KJ, Kasting ML, Sturm LA, Hartsock JA, Zimet GD. 2020. [A National Survey Assessing SARS-CoV-2 Vaccination Intentions: Implications for Future Public Health Communication Efforts](#). Science Communication 42:698–723.
1412. Salmon D, Opel DJ, Dudley MZ, Brewer J, Breiman R. 2021. [Reflections On Governance, Communication, And Equity: Challenges And Opportunities In COVID-19 Vaccination](#). Health Affairs 40:419–425.
1413. Anand P, Stahel VP. 2021. [The safety of Covid-19 mRNA vaccines: a review](#). Patient Saf Surg 15.
1414. Salmon DA, Dudley MZ, Glanz JM, Omer SB. 2015. [Vaccine hesitancy](#). Vaccine 33:D66–D71.

1415. Jacobson RM, St. Sauver JL, Finney Rutten LJ. 2015. [Vaccine Hesitancy](#). Mayo Clinic Proceedings 90:1562–1568.
1416. 2022. What sets apart Novavax option from other COVID-19 vaccines. American Medical Association. <https://www.ama-assn.org/delivering-care/public-health/what-sets-apart-novavax-option-other-covid-19-vaccines>. Retrieved 30 July 2023.
1417. RUPP DMBAR. 2022. Novavax COVID vaccine is for those hesitant to get vaccinated. The Daily News. https://www.galvnews.com/health/free/novavax-covid-vaccine-is-for-those-hesitant-to-get-vaccinated/article_8ddebef4-1db0-579a-b0d2-852f2031d881.html. Retrieved 30 July 2023.
1418. Rubin R. 2022. [Despite Its Fan Base, Newly Authorized “Traditional” Novavax COVID-19 Vaccine Is Having Trouble Gaining a Foothold in the US](#). JAMA 328:1026.
1419. Constantino AK. 2022. Novavax's new Covid vaccine is perfect for people scared of mRNA tech—but it won't win over the unvaccinated. CNBC. <https://www.cnbc.com/2022/07/15/why-new-novavax-covid-vaccine-wont-win-over-unvaccinated-americans.html>. Retrieved 30 July 2023.
1420. Patel MK, Bergeri I, Bresee JS, Cowling BJ, Crowcroft NS, Fahmy K, Hirve S, Kang G, Katz MA, Lanata CF, L'Aizou Jackson M, Joshi S, Lipsitch M, Mwenda JM, Nogareda F, Orenstein WA, Ortiz JR, Pebody R, Schrag SJ, Smith PG, Srikanthiah P, Subissi L, Valenciano M, Vaughn DW, Verani JR, Wilder-Smith A, Feikin DR. 2021. [Evaluation of post-introduction COVID-19 vaccine effectiveness: Summary of interim guidance of the World Health Organization](#). Vaccine 39:4013–4024.
1421. Jiang S, He Y, Liu S. 2005. [SARS Vaccine Development](#). Emerg Infect Dis 11:1016–1020.
1422. Graham RL, Donaldson EF, Baric RS. 2013. [A decade after SARS: strategies for controlling emerging coronaviruses](#). Nat Rev Microbiol 11:836–848.
1423. Lurie N, Saville M, Hatchett R, Halton J. 2020. [Developing Covid-19 Vaccines at Pandemic Speed](#). New England Journal of Medicine 382:1969–1973.
1424. Maslow JN. 2017. [Vaccines for emerging infectious diseases: Lessons from MERS coronavirus and Zika virus](#). Human Vaccines & Immunotherapeutics 13:2918–2930.
1425. Sharma O, Sultan AA, Ding H, Trigg CR. 2020. [A Review of the Progress and Challenges of Developing a Vaccine for COVID-19](#). Front Immunol 11.
1426. Li Y-P, Liang Z-L, Gao Q, Huang L-R, Mao Q-Y, Wen S-Q, Liu Y, Yin W-D, Li R-C, Wang J-Z. 2012. [Safety and immunogenicity of a novel human Enterovirus 71 \(EV71\) vaccine: A randomized, placebo-controlled, double-blind, Phase I clinical trial](#). Vaccine 30:3295–3303.

1427. Magnusson SE, Altenburg AF, Bengtsson KL, Bosman F, de Vries RD, Rimmelzwaan GF, Stertman L. 2018. [Matrix-M™ adjuvant enhances immunogenicity of both protein- and modified vaccinia virus Ankara-based influenza vaccines in mice](#). Immunol Res 66:224–233.
1428. Magnusson SE, Reimer JM, Karlsson KH, Lilja L, Bengtsson KL, Stertman L. 2013. [Immune enhancing properties of the novel Matrix-M™ adjuvant leads to potentiated immune responses to an influenza vaccine in mice](#). Vaccine 31:1725–1733.
1429. Madhun AS, Haaheim LR, Nilsen MV, Cox RJ. 2009. [Intramuscular Matrix-M-adjuvanted virosomal H5N1 vaccine induces high frequencies of multifunctional Th1 CD4+ cells and strong antibody responses in mice](#). Vaccine 27:7367–7376.
1430. Pedersen G, Major D, Roseby S, Wood J, Madhun AS, Cox RJ. 2011. [Matrix-M adjuvanted virosomal H5N1 vaccine confers protection against lethal viral challenge in a murine model](#). Influenza and Other Respiratory Viruses 5:426–437.
1431. Cox RJ, Pedersen G, Madhun AS, Svindland S, Sævik M, Breakwell L, Hoschler K, Willemsen M, Campitelli L, Nøstbakken JK, Weverling GJ, Klap J, McCullough KC, Zambon M, Kompier R, Sjursen H. 2011. [Evaluation of a virosomal H5N1 vaccine formulated with Matrix M™ adjuvant in a phase I clinical trial](#). Vaccine 29:8049–8059.
1432. Formica N, Mallory R, Albert G, Robinson M, Plested JS, Cho I, Robertson A, Dubovsky F, Glenn GM. 2021. [Different dose regimens of a SARS-CoV-2 recombinant spike protein vaccine \(NVX-CoV2373\) in younger and older adults: A phase 2 randomized placebo-controlled trial](#). PLoS Med 18:e1003769.
1433. Novavax COVID-19 Vaccine Demonstrates 89.3% Efficacy in UK Phase 3 Trial. Novavax Investor Relations. <https://ir.novavax.com/2021-01-28-Novavax-COVID-19-Vaccine-Demonstrates-89-3-Efficacy-in-UK-Phase-3-Trial>. Retrieved 5 December 2022.
1434. 2021. Novavax COVID-19 Vaccine Demonstrates 90% Overall Efficacy and 100% Protection Against Moderate and Severe Disease in PREVENT-19 Phase 3 Trial. Novavax Investor Relations. <https://ir.novavax.com/2021-06-14-Novavax-COVID-19-Vaccine-Demonstrates-90-Overall-Efficacy-and-100-Protection-Against-Moderate-and-Severe-Disease-in-PREVENT-19-Phase-3-Trial>. Retrieved 1 December 2022.
1435. Raj VS, Mou H, Smits SL, Dekkers DHW, Müller MA, Dijkman R, Muth D, Demmers JAA, Zaki A, Fouchier RAM, Thiel V, Drosten C, Rottier PJM, Osterhaus ADME, Bosch BJ, Haagmans BL. 2013. [Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC](#). Nature 495:251–254.
1436. Lu G, Hu Y, Wang Q, Qi J, Gao F, Li Y, Zhang Y, Zhang W, Yuan Y, Bao J, Zhang B, Shi Y, Yan J, Gao GF. 2013. [Molecular basis of binding between novel human coronavirus MERS-CoV and its receptor CD26](#). Nature 500:227–231.

1437. Mou H, Raj VS, van Kuppeveld FJM, Rottier PJM, Haagmans BL, Bosch BJ. 2013. [The Receptor Binding Domain of the New Middle East Respiratory Syndrome Coronavirus Maps to a 231-Residue Region in the Spike Protein That Efficiently Elicits Neutralizing Antibodies](#). J Virol 87:9379–9383.
1438. Du L, Kou Z, Ma C, Tao X, Wang L, Zhao G, Chen Y, Yu F, Tseng C-TK, Zhou Y, Jiang S. 2013. [A Truncated Receptor-Binding Domain of MERS-CoV Spike Protein Potently Inhibits MERS-CoV Infection and Induces Strong Neutralizing Antibody Responses: Implication for Developing Therapeutics and Vaccines](#). PLoS ONE 8:e81587.
1439. Zhang N, Channappanavar R, Ma C, Wang L, Tang J, Garron T, Tao X, Tasneem S, Lu L, Tseng C-TK, Zhou Y, Perlman S, Jiang S, Du L. 2015. [Identification of an ideal adjuvant for receptor-binding domain-based subunit vaccines against Middle East respiratory syndrome coronavirus](#). Cell Mol Immunol 13:180–190.
1440. Netea MG, Domínguez-Andrés J, Barreiro LB, Chavakis T, Divangahi M, Fuchs E, Joosten LAB, van der Meer JWM, Mhlanga MM, Mulder WJM, Riksen NP, Schlitzer A, Schultze JL, Stabell Benn C, Sun JC, Xavier RJ, Latz E. 2020. [Defining trained immunity and its role in health and disease](#). Nature Reviews Immunology 20:375–388.
1441. Netea MG, Giamarellos-Bourboulis EJ, Domínguez-Andrés J, Curtis N, van Crevel R, van de Veerdonk FL, Bonten M. 2020. [Trained Immunity: a Tool for Reducing Susceptibility to and the Severity of SARS-CoV-2 Infection](#). Cell 181:969–977.
1442. Reducing Health Care Workers Absenteeism in Covid-19 Pandemic Through BCG Vaccine - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04328441>. Retrieved 8 February 2021.
1443. Khattab A. 2020. [Application of BCG Vaccine for Immune-prophylaxis Among Egyptian Healthcare Workers During the Pandemic of COVID-19](#). NCT04350931. Clinical trial registration. clinicaltrials.gov.
1444. Universidad de Antioquia. 2020. [Performance Evaluation of BCG Vaccination in Healthcare Personnel to Reduce the Severity of SARS-COV-2 Infection in Medellín, Colombia, 2020](#). NCT04362124. Clinical trial registration. clinicaltrials.gov.
1445. COVID-19: BCG As Therapeutic Vaccine, Transmission Limitation, and Immunoglobulin Enhancement - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04369794>. Retrieved 8 February 2021.
1446. Using BCG Vaccine to Protect Health Care Workers in the COVID-19 Pandemic - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04373291>. Retrieved 8 February 2021.
1447. TASK Applied Science. 2020. [Reducing Morbidity and Mortality in Health Care Workers Exposed to SARS-CoV-2 by Enhancing Non-specific Immune Responses Through Bacillus Calmette-Guérin](#)

[Vaccination, a Randomized Controlled Trial](#). NCT04379336. Clinical trial registration. [clinicaltrials.gov](#).

1448. Assistance Publique - Hôpitaux de Paris. 2020. [Randomized Controlled Trial Evaluating the Efficacy of Vaccination With Bacillus Calmette and Guérin \(BCG\) in the Prevention of COVID-19 Via the Strengthening of Innate Immunity in Health Care Workers](#). NCT04384549. Clinical trial registration. [clinicaltrials.gov](#).
1449. Study to Assess VPM1002 in Reducing Healthcare Professionals' Absenteeism in COVID-19 Pandemic - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04387409>. Retrieved 8 February 2021.
1450. Hellenic Institute for the Study of Sepsis. 2020. [A Randomized Clinical Trial for Enhanced Trained Immune Responses Through Bacillus Calmette-Guérin Vaccination to Prevent Infections by COVID-19: The ACTIVATE II Trial](#). NCT04414267. Clinical trial registration. [clinicaltrials.gov](#).
1451. Radboud University. 2020. [Reducing Hospital Admission of Elderly in SARS-CoV-2 Pandemic Via the Induction of Trained Immunity by Bacillus Calmette-Guérin Vaccination, a Randomized Controlled Trial](#). NCT04417335. Clinical trial registration. [clinicaltrials.gov](#).
1452. Study to Assess VPM1002 in Reducing Hospital Admissions and/or Severe Respiratory Infectious Diseases in Elderly in COVID-19 Pandemic - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04435379>. Retrieved 8 February 2021.
1453. Efficacy and Safety of VPM1002 in Reducing SARS-CoV-2 (COVID-19) Infection Rate and Severity - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04439045>. Retrieved 8 February 2021.
1454. Giamarellos-Bourboulis EJ, Netea MG, Rovina N, Akinosoglou K, Antoniadou A, Antonakos N, Damoraki G, Gkavogianni T, Adami M-E, Katsaounou P, Ntaganou M, Kyriakopoulou M, Dimopoulos G, Koutsodimitropoulos I, Velissaris D, Koufargyris P, Karageorgos A, Katrini K, Lekakis V, Lupse M, Kotsaki A, Renieris G, Theodoulou D, Panou V, Koukaki E, Koulouris N, Gogos C, Koutsoukou A. 2020. [Complex Immune Dysregulation in COVID-19 Patients with Severe Respiratory Failure](#). *Cell Host & Microbe* 27:992–1000.e3.
1455. Park A, Iwasaki A. 2020. [Type I and Type III Interferons – Induction, Signaling, Evasion, and Application to Combat COVID-19](#). *Cell Host & Microbe* 27:870–878.
1456. 2021. [Covaxin: India approves two Covid vaccines for children under 12](#). BBC News.
1457. Service TN. DCGI grants EUA to Corbevax for those aged 5-12, Covaxin for 6-12 age group. Tribuneindia News Service. <https://www.tribuneindia.com/news/nation/covaxin-cleared-for-6-12-age-group-by-drugs-regulator-389581>. Retrieved 5 December 2022.

1458. 2020. [Novavax aims for 2 billion COVID-19 vaccine doses with expanded India deal](#). Reuters.
1459. iNCOVACC - Intranasal Vaccine For Covid-19 | Bharat Biotech. <https://www.bharatbiotech.com/intranasal-vaccine.html>. Retrieved 5 December 2022.
1460. Wee S-L. 2021. [China Wanted to Show Off Its Vaccines. It's Backfiring](#). The New York Times.
1461. Sharfstein JM, Goodman JL, Borio L. 2021. [The US Regulatory System and COVID-19 Vaccines](#). JAMA 325:1153.
1462. Reuters. 2022. [Chinese Omicron-specific mRNA COVID vaccine candidate to be trialed in UAE](#). Reuters.
1463. 2021. Philippines receives COVID-19 vaccine after delays. AP NEWS. <https://apnews.com/article/cabinets-philippines-coronavirus-pandemic-asia-east-asia-e6f826359395951b2c805dbd90f34205>. Retrieved 5 December 2022.
1464. Dubai CD in T and JM in. China's Covid-19 Vaccine Makers Struggle to Meet Demand. WSJ. <https://www.wsj.com/articles/chinas-covid-19-vaccine-makers-struggle-to-meet-demand-11612958560>. Retrieved 5 December 2022.
1465. Cohen J. 2014. [Ebola vaccine: Little and late](#). Science 345:1441–1442.
1466. Coller B-AG, Blue J, Das R, Dubey S, Finelli L, Gupta S, Helmond F, Grant-Klein RJ, Liu K, Simon J, Troth S, VanRheenen S, Waterbury J, Wivel A, Wolf J, Heppner DG, Kemp T, Nichols R, Monath TP. 2017. [Clinical development of a recombinant Ebola vaccine in the midst of an unprecedented epidemic](#). Vaccine 35:4465–4469.
1467. Our Story. Moderna. <https://www.modernatx.com/en-US/about-us/our-story?slug=about-us%2Four-story>. Retrieved 5 December 2022.
1468. Thanh Le T, Andreadakis Z, Kumar A, Gómez Román R, Tollefsen S, Saville M, Mayhew S. 2020. [The COVID-19 vaccine development landscape](#). Nat Rev Drug Discov 19:305–306.
1469. Voysey M, Clemens SAC, Madhi SA, Weckx LY, Folegatti PM, Aley PK, Angus B, Baillie VL, Barnabas SL, Bhorat QE, Bibi S, Briner C, Cicconi P, Collins AM, Colin-Jones R, Cutland CL, Darton TC, Dheda K, Duncan CJA, Emary KRW, Ewer KJ, Fairlie L, Faust SN, Feng S, Ferreira DM, Finn A, Goodman AL, Green CM, Green CA, Heath PT, Hill C, Hill H, Hirsch I, Hodgson SHC, Izu A, Jackson S, Jenkin D, Joe CCD, Kerridge S, Koen A, Kwatra G, Lazarus R, Lawrie AM, Lelliott A, Libri V, Lillie PJ, Mallory R, Mendes AVA, Milan EP, Minassian AM, McGregor A, Morrison H, Mujadidi YF, Nana A, O'Reilly PJ, Padayachee SD, Pittella A, Plested E, Pollock KM, Ramasamy MN, Rhead S, Schwarzbald AV, Singh N, Smith A, Song R, Snape MD, Sprinz E, Sutherland RK, Tarrant R, Thomson EC, Török ME, Toshner M, Turner DPJ, Vekemans J, Villafana TL, Watson MEE, Williams CJ, Douglas AD, Hill AVS, Lambe T, Gilbert SC, Pollard AJ, Aban M, Abayomi F, Abeyskera K, Aboagye J, Adam M, Adams K, Adamson J, Adelaja YA, Adewetan G, Adlou S, Ahmed K, Akhalwaya Y,

Akhalwaya S, Alcock A, Ali A, Allen ER, Allen L, Almeida TCDSC, Alves MPS, Amorim F, Andritsou F, Anslow R, Appleby M, Arbe-Barnes EH, Ariaans MP, Arns B, Arruda L, Azi P, Azi L, Babbage G, Bailey C, Baker KF, Baker M, Baker N, Baker P, Baldwin L, Baleanu I, Bandeira D, Bara A, Barbosa MAS, Barker D, Barlow GD, Barnes E, Barr AS, Barrett JR, Barrett J, Bates L, Batten A, Beadon K, Beales E, Beckley R, Belij-Rammerstorfer S, Bell J, Bellamy D, Bellei N, Belton S, Berg A, Bermejo L, Berrie E, Berry L, Berzenyi D, Beveridge A, Bewley KR, Bexhell H, Bhikha S, Bhorat AE, Bhorat ZE, Bijker E, Birch G, Birch S, Bird A, Bird O, Bisnauthsing K, Bittaye M, Blackstone K, Blackwell L, Bletchly H, Blundell CL, Blundell SR, Bodalia P, Boettger BC, Bolam E, Boland E, Bormans D, Borthwick N, Bowring F, Boyd A, Bradley P, Brenner T, Brown P, Brown C, Brown-O'Sullivan C, Bruce S, Brunt E, Buchan R, Budd W, Bulbulia YA, Bull M, Burbage J, Burhan H, Burn A, Buttigieg KR, Byard N, Cabera Puig I, Calderon G, Calvert A, Camara S, Cao M, Cappuccini F, Cardoso JR, Carr M, Carroll MW, Carson-Stevens A, Carvalho Y de M, Carvalho JAM, Casey HR, Cashen P, Castro T, Castro LC, Cathie K, Cavey A, Cerbino-Neto J, Chadwick J, Chapman D, Charlton S, Chelysheva I, Chester O, Chita S, Cho J-S, Cifuentes L, Clark E, Clark M, Clarke A, Clutterbuck EA, Collins SLK, Conlon CP, Connarty S, Coombes N, Cooper C, Cooper R, Cornelissen L, Corrah T, Cosgrove C, Cox T, Crocker WEM, Crosbie S, Cullen L, Cullen D, Cunha DRMF, Cunningham C, Cuthbertson FC, Da Guarda SNF, da Silva LP, Damratoski BE, Danos Z, Dantas MTDC, Darroch P, Datto MS, Datta C, Davids M, Davies SL, Davies H, Davis E, Davis J, Davis J, De Nobrega MMD, De Oliveira Kalid LM, Dearlove D, Demissie T, Desai A, Di Marco S, Di Maso C, Dinelli MIS, Dinesh T, Docksey C, Dold C, Dong T, Donnellan FR, Dos Santos T, dos Santos TG, Dos Santos EP, Douglas N, Downing C, Drake J, Drake-Brockman R, Driver K, Drury R, Dunachie SJ, Durham BS, Dutra L, Easom NJW, van Eck S, Edwards M, Edwards NJ, El Muhanna OM, Elias SC, Elmore M, English M, Esmail A, Essack YM, Farmer E, Farooq M, Farrar M, Farrugia L, Faulkner B, Fedosyuk S, Felle S, Feng S, Ferreira Da Silva C, Field S, Fisher R, Flaxman A, Fletcher J, Fofie H, Fok H, Ford KJ, Fowler J, Fraiman PHA, Francis E, Franco MM, Frater J, Freire MSM, Fry SH, Fudge S, Furze J, Fuskova M, Galian-Rubio P, Galiza E, Garland H, Gavrilu M, Geddes A, Gibbons KA, Gilbride C, Gill H, Glynn S, Godwin K, Gokani K, Goldoni UC, Goncalves M, Gonzalez IGS, Goodwin J, Goondiwala A, Gordon-Quayle K, Gorini G, Grab J, Gracie L, Greenland M, Greenwood N, Greffrath J, Groenewald MM, Grossi L, Gupta G, Hackett M, Hallis B, Hamaluba M, Hamilton E, Hamlyn J, Hammersley D, Hanrath AT, Hanumunthadu B, Harris SA, Harris C, Harris T, Harrison TD, Harrison D, Hart TC, Hartnell B, Hassan S, Haughney J, Hawkins S, Hay J, Head I, Henry J, Hermosin Herrera M, Hettle DB, Hill J, Hodges G, Horne E, Hou MM, Houlihan C, Howe E, Howell N, Humphreys J, Humphries HE, Hurley K, Huson C, Hyder-Wright A, Hyams C, Ikram S, Ishwarbhai A, Ivan M, Iveson P, Iyer V, Jackson F, De Jager J, Jaumdally S, Jeffers H, Jesudason N, Jones B, Jones K, Jones E, Jones C, Jorge MR, Jose A, Joshi A, Júnior EAMS, Kadziola J, Kailath R, Kana F, Karampatsas K, Kasanyinga M, Keen J, Kelly EJ, Kelly DM, Kelly D, Kelly S, Kerr D, Kfoury R de Á, Khan L, Khozoe B, Kidd S, Killen A, Kinch J, Kinch P, King LDW, King TB, Kingham L, Klenerman P, Knapper F, Knight JC, Knott D, Koleva S, Lang M, Lang G, Larkworthy CW, Larwood JPJ, Law R, Lazarus EM, Leach A, Lees EA, Lemm N-M, Lessa A, Leung S, Li Y, Lias AM, Liatsikos K, Linder A, Lipworth S, Liu S,

Liu X, Lloyd A, Lloyd S, Loew L, Lopez Ramon R, Lora L, Lowthorpe V, Luz K, MacDonald JC, MacGregor G, Madhavan M, Mainwaring DO, Makambwa E, Makinson R, Malahleha M, Malamatsho R, Mallett G, Mansatta K, Maoko T, Mapetla K, Marchevsky NG, Marinou S, Marlow E, Marques GN, Marriott P, Marshall RP, Marshall JL, Martins FJ, Masenya M, Masilela M, Masters SK, Mathew M, Matlebjane H, Matshidiso K, Mazur O, Mazzella A, McCaughan H, McEwan J, McGlashan J, McInroy L, McIntyre Z, McLenaghan D, McRobert N, McSwiggan S, Megson C, Mehdipour S, Meijs W, Mendonça RNÁ, Mentzer AJ, Mirtorabi N, Mitton C, Mnyakeni S, Moghaddas F, Molapo K, Moloi M, Moore M, Moraes-Pinto MI, Moran M, Morey E, Morgans R, Morris S, Morris S, Morris HC, Morselli F, Morshead G, Morter R, Mottal L, Moultrie A, Moya N, Mpelembue M, Msomi S, Mugodi Y, Mukhopadhyay E, Muller J, Munro A, Munro C, Murphy S, Mweu P, Myasaki CH, Naik G, Naker K, Nastouli E, Nazir A, Ndlovu B, Neffa F, Njenga C, Noal H, Noé A, Novaes G, Nugent FL, Nunes G, O'Brien K, O'Connor D, Odam M, Oelofse S, Oguti B, Olchawski V, Oldfield NJ, Oliveira MG, Oliveira C, Oosthuizen A, O'Reilly P, Osborne P, Owen DRJ, Owen L, Owens D, Owino N, Pacurar M, Paiva BVB, Palhares EMF, Palmer S, Parkinson S, Parracho HMRT, Parsons K, Patel D, Patel B, Patel F, Patel K, Patrick-Smith M, Payne RO, Peng Y, Penn EJ, Pennington A, Peralta Alvarez MP, Perring J, Perry N, Perumal R, Petkar S, Philip T, Phillips DJ, Phillips J, Phohu MK, Pickup L, Pieterse S, Piper J, Pipini D, Plank M, Du Plessis J, Pollard S, Pooley J, Pooran A, Poulton I, Powers C, Presa FB, Price DA, Price V, Primeira M, Proud PC, Provstgaard-Morys S, Pueschel S, Pulido D, Quaid S, Rabara R, Radford A, Radia K, Rajapaska D, Rajeswaran T, Ramos ASF, Ramos Lopez F, Rampling T, Rand J, Ratcliffe H, Rawlinson T, Rea D, Rees B, Reiné J, Resuello-Dauti M, Reyes Pabon E, Ribiero CM, Ricamara M, Richter A, Ritchie N, Ritchie AJ, Robbins AJ, Roberts H, Robinson RE, Robinson H, Rocchetti TT, Rocha BP, Roche S, Rollier C, Rose L, Ross Russell AL, Rossouw L, Royal S, Rudiansyah I, Ruiz S, Saich S, Sala C, Sale J, Salman AM, Salvador N, Salvador S, Sampaio M, Samson AD, Sanchez-Gonzalez A, Sanders H, Sanders K, Santos E, Santos Guerra MFS, Satti I, Saunders JE, Saunders C, Sayed A, Schim van der Loeff I, Schmid AB, Schofield E, Screation G, Seddiqi S, Segireddy RR, Senger R, Serrano S, Shah R, Shaik I, Sharpe HE, Sharrocks K, Shaw R, Shea A, Shepherd A, Shepherd JG, Shiham F, Sidhom E, Silk SE, da Silva Moraes AC, Silva-Junior G, Silva-Reyes L, Silveira AD, Silveira MBV, Sinha J, Skelly DT, Smith DC, Smith N, Smith HE, Smith DJ, Smith CC, Soares A, Soares T, Solórzano C, Sorio GL, Sorley K, Sosa-Rodriguez T, Souza CMCDL, Souza BSDF, Souza AR, Spencer AJ, Spina F, Spoors L, Stafford L, Stamford I, Starinskij I, Stein R, Steven J, Stockdale L, Stockwell LV, Strickland LH, Stuart AC, Sturdy A, Sutton N, Szigeti A, Tahiri-Alaoui A, Tanner R, Taoushanis C, Tarr AW, Taylor K, Taylor U, Taylor IJ, Taylor J, te Water Naude R, Themistocleous Y, Themistocleous A, Thomas M, Thomas K, Thomas TM, Thombrayil A, Thompson F, Thompson A, Thompson K, Thompson A, Thomson J, Thornton-Jones V, Tighe PJ, Tinoco LA, Tiongson G, Tladinyane B, Tomasicchio M, Tomic A, Tonks S, Towner J, Tran N, Tree J, Trillana G, Trinhnam C, Trivett R, Truby A, Tsheko BL, Turabi A, Turner R, Turner C, Ulaszewska M, Underwood BR, Varughese R, Verbart D, Verheul M, Vichos I, Vieira T, Waddington CS, Walker L, Wallis E, Wand M, Warbick D, Wardell T, Warimwe G, Warren SC, Watkins B, Watson E, Webb S, Webb-Bridges A, Webster A,

Welch J, Wells J, West A, White C, White R, Williams P, Williams RL, Winslow R, Woodyer M, Worth AT, Wright D, Wroblewska M, Yao A, Zimmer R, Zizi D, Zuidewind P. 2021. [Safety and efficacy of the ChAdOx1 nCoV-19 vaccine \(AZD1222\) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK](#). The Lancet 397:99–111.

1470. Duan L, Zheng Q, Zhang H, Niu Y, Lou Y, Wang H. 2020. [The SARS-CoV-2 Spike Glycoprotein Biosynthesis, Structure, Function, and Antigenicity: Implications for the Design of Spike-Based Vaccine Immunogens](#). Front Immunol 11.
1471. BioRender. <https://biorender.com/>. Retrieved 5 December 2022.
1472. Human Coronavirus Structure. <https://app.biorender.com/biorender-templates/figures/all/t-5f21e90283765600b08fbe9d-human-coronavirus-structure>. Retrieved 5 December 2022.
1473. New Images of Novel Coronavirus SARS-CoV-2 Now Available | NIH: National Institute of Allergy and Infectious Diseases. <https://www.niaid.nih.gov/news-events/novel-coronavirus-sarscov2-images>. Retrieved 5 December 2022.
1474. Rhee JH. 2014. [Towards Vaccine 3.0: new era opened in vaccine research and industry](#). Clin Exp Vaccine Res 3:1.
1475. Seib KL, Zhao X, Rappuoli R. 2012. [Developing vaccines in the era of genomics: a decade of reverse vaccinology](#). Clinical Microbiology and Infection 18:109–116.
1476. Liu. 2019. [A Comparison of Plasmid DNA and mRNA as Vaccine Technologies](#). Vaccines 7:37.
1477. Ellis RW. 1994. [New Vaccine Technologies](#). JAMA 271:929.
1478. Liu MA. 2003. [DNA vaccines: a review](#). J Intern Med 253:402–410.
1479. Kutzler MA, Weiner DB. 2008. [DNA vaccines: ready for prime time?](#) Nat Rev Genet 9:776–788.
1480. Sternberg A, Naujokat C. 2020. [Structural features of coronavirus SARS-CoV-2 spike protein: Targets for vaccination](#). Life Sciences 257:118056.
1481. Kirchdoerfer RN, Cottrell CA, Wang N, Pallesen J, Yassine HM, Turner HL, Corbett KS, Graham BS, McLellan JS, Ward AB. 2016. [Pre-fusion structure of a human coronavirus spike protein](#). Nature 531:118–121.
1482. Liu C, Mendonça L, Yang Y, Gao Y, Shen C, Liu J, Ni T, Ju B, Liu C, Tang X, Wei J, Ma X, Zhu Y, Liu W, Xu S, Liu Y, Yuan J, Wu J, Liu Z, Zhang Z, Liu L, Wang P, Zhang P. 2020. [The Architecture of Inactivated SARS-CoV-2 with Postfusion Spikes Revealed by Cryo-EM and Cryo-ET](#). Structure 28:1218–1224.e4.
1483. Ke Z, Oton J, Qu K, Cortese M, Zila V, McKeane L, Nakane T, Zivanov J, Neufeldt CJ, Cerikan B, Lu JM, Peukes J, Xiong X, Kräusslich H-G,

- Scheres SHW, Bartenschlager R, Briggs JAG. 2020. [Structures and distributions of SARS-CoV-2 spike proteins on intact virions](#). Nature 588:498–502.
1484. Hsieh C-L, Goldsmith JA, Schaub JM, DiVenere AM, Kuo H-C, Javanmardi K, Le KC, Wrapp D, Lee AG, Liu Y, Chou C-W, Byrne PO, Hjorth CK, Johnson NV, Ludes-Meyers J, Nguyen AW, Park J, Wang N, Amengor D, Lavinder JJ, Ippolito GC, Maynard JA, Finkelstein IJ, McLellan JS. 2020. [Structure-based design of prefusion-stabilized SARS-CoV-2 spikes](#). Science 369:1501–1505.
1485. Bos R, Rutten L, van der Lubbe JEM, Bakkers MJG, Hardenberg G, Wegmann F, Zuijdgeest D, de Wilde AH, Koornneef A, Verwilligen A, van Manen D, Kwaks T, Vogels R, Dalebout TJ, Myeni SK, Kikkert M, Snijder EJ, Li Z, Barouch DH, Vellinga J, Langedijk JPM, Zahn RC, Custers J, Schuitemaker H. 2020. [Ad26 vector-based COVID-19 vaccine encoding a prefusion-stabilized SARS-CoV-2 Spike immunogen induces potent humoral and cellular immune responses](#). npj Vaccines 5.
1486. Marrack P, McKee AS, Munks MW. 2009. [Towards an understanding of the adjuvant action of aluminium](#). Nature Reviews Immunology 9:287–293.
1487. Hayashi T, Momota M, Kuroda E, Kusakabe T, Kobari S, Makisaka K, Ohno Y, Suzuki Y, Nakagawa F, Lee MSJ, Coban C, Onodera R, Higashi T, Motoyama K, Ishii KJ, Arima H. 2018. [DAMP-Inducing Adjuvant and PAMP Adjuvants Parallely Enhance Protective Type-2 and Type-1 Immune Responses to Influenza Split Vaccination](#). Frontiers in Immunology 9:2619.
1488. Wang Z-B, Xu J. 2020. [Better Adjuvants for Better Vaccines: Progress in Adjuvant Delivery Systems, Modifications, and Adjuvant–Antigen Codelivery](#). Vaccines 8:128.
1489. Hobernik D, Bros M. 2018. [DNA Vaccines—How Far From Clinical Use?](#) IJMS 19:3605.
1490. Ghaffarifar F. 2018. [Plasmid DNA vaccines: where are we now?](#) Drugs Today 54:315.
1491. Glenting J, Wessels S. 2005. [Ensuring safety of DNA vaccines](#). Microb Cell Fact 4.
1492. Williams J. 2013. [Vector Design for Improved DNA Vaccine Efficacy, Safety and Production](#). Vaccines 1:225–249.
1493. Lim M, Badruddoza AZM, Firdous J, Azad M, Mannan A, Al-Hilal TA, Cho C-S, Islam MA. 2020. [Engineered Nanodelivery Systems to Improve DNA Vaccine Technologies](#). Pharmaceutics 12:30.
1494. DNA vaccines. World Health Organization. <https://www.who.int/biologicals/areas/vaccines/dna/en>. Retrieved 5 August 2022.
1495. Lapuente D, Stab V, Storcksdieck genannt Bonsmann M, Maaske A, Köster M, Xiao H, Ehrhardt C, Tenbusch M. 2020. [Innate signalling](#)

[molecules as genetic adjuvants do not alter the efficacy of a DNA-based influenza A vaccine](#). PLoS ONE 15:e0231138.

1496. Liu MA, Ulmer JB. 2005. [Human Clinical Trials of Plasmid DNA Vaccines](#), p. 25–40. *In* Advances in Genetics. Elsevier.
1497. Roy MJ, Wu MS, Barr LJ, Fuller JT, Tussey LG, Speller S, Culp J, Burkholder JK, Swain WF, Dixon RM, Widera G, Vessey R, King A, Ogg G, Gallimore A, Haynes JR, Heydenburg Fuller D. 2000. [Induction of antigen-specific CD8+ T cells, T helper cells, and protective levels of antibody in humans by particle-mediated administration of a hepatitis B virus DNA vaccine](#). Vaccine 19:764–778.
1498. Weiner DB, Nabel GJ. 2018. [Development of Gene-Based Vectors for Immunization](#), p. 1305–1319.e8. *In* Plotkin's Vaccines. Elsevier.
1499. A. Gómez L, A. Oñate A. 2019. [Plasmid-Based DNA Vaccines](#) Plasmid. IntechOpen.
1500. Eusébio D, Neves AR, Costa D, Biswas S, Alves G, Cui Z, Sousa Â. 2021. [Methods to improve the immunogenicity of plasmid DNA vaccines](#). Drug Discovery Today 26:2575–2592.
1501. Garver K, LaPatra S, Kurath G. 2005. [Efficacy of an infectious hematopoietic necrosis \(IHN\) virus DNA vaccine in Chinook Oncorhynchus tshawytscha and sockeye O. nerka salmon](#). Dis Aquat Org 64:13–22.
1502. Thacker EL, Holtkamp DJ, Khan AS, Brown PA, Draghia-Akli R. 2006. [Plasmid-mediated growth hormone-releasing hormone efficacy in reducing disease associated with Mycoplasma hyopneumoniae and porcine reproductive and respiratory syndrome virus infection1](#). Journal of Animal Science 84:733–742.
1503. Davidson AH, Traub-Dargatz JL, Rodeheaver RM, Ostlund EN, Pedersen DD, Moorhead RG, Stricklin JB, Dewell RD, Roach SD, Long RE, Albers SJ, Callan RJ, Salman MD. 2005. [Immunologic responses to West Nile virus in vaccinated and clinically affected horses](#). JAVMA 226:240–245.
1504. Langellotti CA, Gammella M, Soria I, Bellusci C, Quattrocchi V, Vermeulen M, Mongini C, Zamorano PI. 2021. [An Improved DNA Vaccine Against Bovine Herpesvirus-1 Using CD40L and a Chemical Adjuvant Induces Specific Cytotoxicity in Mice](#). Viral Immunology 34:68–78.
1505. Collins C, Lorenzen N, Collet B. 2019. [DNA vaccination for finfish aquaculture](#). Fish & Shellfish Immunology 85:106–125.
1506. Chakraborty C, Agoramoorthy G. 2020. [India's cost-effective COVID-19 vaccine development initiatives](#). Vaccine 38:7883–7884.
1507. Mallapaty S. 2021. [India's DNA COVID vaccine is a world first – more are coming](#). Nature 597:161–162.
1508. Abbasi J. 2021. [India's New COVID-19 DNA Vaccine for Adolescents and Adults Is a First](#). JAMA 326:1365.

1509. Safety, Tolerability and Immunogenicity of INO-4800 for COVID-19 in Healthy Volunteers - Full Text View - ClinicalTrials.gov.
<https://clinicaltrials.gov/ct2/show/NCT04336410>. Retrieved 8 February 2021.
1510. Diehl MC, Lee JC, Daniels SE, Tebas P, Khan AS, Giffear M, Sardesai NY, Bagarazzi ML. 2013. [Tolerability of intramuscular and intradermal delivery by CELLECTRA® adaptive constant current electroporation device in healthy volunteers](#). Human Vaccines & Immunotherapeutics 9:2246–2252.
1511. Sardesai NY, Weiner DB. 2011. [Electroporation delivery of DNA vaccines: prospects for success](#). Current Opinion in Immunology 23:421–429.
1512. Tebas P, Yang S, Boyer JD, Reuschel EL, Patel A, Christensen-Quick A, Andrade VM, Morrow MP, Kraynyak K, Agnes J, Purwar M, Sylvester A, Pawlicki J, Gillespie E, Maricic I, Zaidi FI, Kim KY, Dia Y, Frase D, Pezzoli P, Schultheis K, Smith TRF, Ramos SJ, McMullan T, Buttigieg K, Carroll MW, Ervin J, Diehl MC, Blackwood E, Mammen MP, Lee J, Dallas MJ, Brown AS, Shea JE, Kim JJ, Weiner DB, Broderick KE, Humeau LM. 2021. [Safety and immunogenicity of INO-4800 DNA vaccine against SARS-CoV-2: A preliminary report of an open-label, Phase 1 clinical trial](#). EClinicalMedicine 31:100689.
1513. Mammen MP Jr., Tebas P, Agnes J, Giffear M, Kraynyak KA, Blackwood E, Amante D, Reuschel EL, Purwar M, Christensen-Quick A, Liu N, Andrade VM, Carter J, Garufi G, Diehl MC, Sylvester A, Morrow MP, Pezzoli P, Kulkarni AJ, Zaidi FI, Frase D, Liaw K, Badie H, Simon KO, Smith TRF, Ramos S, Spitz R, Juba RJ, Lee J, Dallas M, Brown AS, Shea JE, Kim JJ, Weiner DB, Broderick KE, Boyer JD, Humeau LM. 2021. [Safety and immunogenicity of INO-4800 DNA vaccine against SARS-CoV-2: a preliminary report of a randomized, blinded, placebo-controlled, Phase 2 clinical trial in adults at high risk of viral exposure](#). Cold Spring Harbor Laboratory.
1514. INOVIO Receives U.S. FDA Authorization to Proceed with INNOVATE Phase 3 Segment for its COVID-19 Vaccine Candidate, INO-4800, in the U.S. <https://ir.inovio.com/news-releases/news-releases-details/2021/INOVIO-Receives-U.S.-FDA-Authorization-to-Proceed-with-INNOVATE-Phase-3-Segment-for-its-COVID-19-Vaccine-Candidate-INO-4800-in-the-U.S/default.aspx>. Retrieved 5 December 2022.
1515. INOVIO Further Expands INNOVATE Phase 3 Trial for COVID-19 DNA Vaccine Candidate INO-4800 With Regulatory Authorization from India. <https://ir.inovio.com/news-releases/news-releases-details/2021/INOVIO-Further-Expands-INNOVATE-Phase-3-Trial-for-COVID-19-DNA-Vaccine-Candidate-INO-4800-With-Regulatory-Authorization-from-India/default.aspx>. Retrieved 5 December 2022.
1516. INOVIO Expands INNOVATE Phase 3 for INO-4800, its DNA Vaccine Candidate for COVID-19, to include Colombia following Regulatory Authorization. <https://ir.inovio.com/news-releases/news-releases-details/2021/INOVIO-Expands-INNOVATE-Phase-3-for-INO-4800-its->

[DNA-Vaccine-Candidate-for-COVID-19-to-include-Colombia-following-Regulatory-Authorization/default.aspx](#). Retrieved 5 December 2022.

1517. INOVIO Receives Regulatory Authorization to Conduct Phase 3 Efficacy Trial of its COVID-19 DNA Vaccine Candidate, INO-4800, in Mexico. <https://ir.inovio.com/news-releases/news-releases-details/2021/INOVIO-Receives-Regulatory-Authorization-to-Conduct-Phase-3-Efficacy-Trial-of-its-COVID-19-DNA-Vaccine-Candidate-INO-4800-in-Mexico/default.aspx>. Retrieved 5 December 2022.
1518. Momin T, Kansagra K, Patel H, Sharma S, Sharma B, Patel J, Mittal R, Sanmukhani J, Maithal K, Dey A, Chandra H, Rajanathan CT, Pericherla HP, Kumar P, Narkhede A, Parmar D. 2021. [Safety and Immunogenicity of a DNA SARS-CoV-2 vaccine \(ZyCoV-D\): Results of an open-label, non-randomized phase I part of phase I/II clinical study by intradermal route in healthy subjects in India](#). EClinicalMedicine 38:101020.
1519. Khobragade A, Bhate S, Ramaiah V, Deshpande S, Giri K, Phophle H, Supe P, Godara I, Revanna R, Nagarkar R, Sanmukhani J, Dey A, Rajanathan TMC, Kansagra K, Koradia P. 2022. [Efficacy, safety, and immunogenicity of the DNA SARS-CoV-2 vaccine \(ZyCoV-D\): the interim efficacy results of a phase 3, randomised, double-blind, placebo-controlled study in India](#). The Lancet 399:1313–1321.
1520. Zydus Cadila: ZyCoV-D – COVID19 Vaccine Tracker. <https://covid19.trackvaccines.org/vaccines/29/>. Retrieved 5 December 2022.
1521. 2021. Covishield, Covaxin, ZyCoV-D Makers to Assess Efficacy of Vaccine as They Await Data on Omicron. News18. <https://www.news18.com/news/india/covishield-covaxin-zycov-d-makers-to-assess-efficacy-of-their-vaccine-as-they-await-data-on-omicron-4504295.html>. Retrieved 5 December 2022.
1522. Andrade VM, Christensen-Quick A, Agnes J, Tur J, Reed C, Kalia R, Marrero I, Elwood D, Schultheis K, Purwar M, Reuschel E, McMullan T, Pezzoli P, Kraynyak K, Sylvester A, Mammen MP, Tebas P, Joseph Kim J, Weiner DB, Smith TRF, Ramos SJ, Humeau LM, Boyer JD, Broderick KE. 2021. [INO-4800 DNA vaccine induces neutralizing antibodies and T cell activity against global SARS-CoV-2 variants](#). npj Vaccines 6.
1523. Kraynyak KA, Blackwood E, Agnes J, Tebas P, Giffear M, Amante D, Reuschel EL, Purwar M, Christensen-Quick A, Liu N, Andrade VM, Diehl MC, Wani S, Lupicka M, Sylvester A, Morrow MP, Pezzoli P, McMullan T, Kulkarni AJ, Zaidi FI, Frase D, Liaw K, Smith TRF, Ramos SJ, Ervin J, Adams M, Lee J, Dallas M, Shah Brown A, Shea JE, Kim JJ, Weiner DB, Broderick KE, Humeau LM, Boyer JD, Mammen MP Jr. 2022. [SARS-CoV-2 DNA Vaccine INO-4800 Induces Durable Immune Responses Capable of Being Boosted in a Phase 1 Open-Label Trial](#). The Journal of Infectious Diseases 225:1923–1932.
1524. Inc IP. INOVIO Announces Strategy to Address Omicron (B.1.1.529) and Future SARS-CoV-2 Variants. <https://www.prnewswire.com/news-releases/inovio-announces-strategy-to-address-omicron-b1-1-529-and->

[future-sars-cov-2-variants-301433776.html](https://doi.org/10.1016/j.vaccine.2022.115376). Retrieved 5 December 2022.

1525. Walters JN, Schouest B, Patel A, Reuschel EL, Schultheis K, Parzych E, Maricic I, Gary EN, Purwar M, Andrade VM, Doan A, Elwood D, Eblimit Z, Nguyen B, Frase D, Zaidi FI, Kulkarni A, Generotti A, Joseph Kim J, Humeau LM, Ramos SJ, Smith TRF, Weiner DB, Broderick KE. 2022. [Prime-boost vaccination regimens with INO-4800 and INO-4802 augment and broaden immune responses against SARS-CoV-2 in nonhuman primates](#). *Vaccine* 40:2960–2969.
1526. Reed CC, Schultheis K, Andrade VM, Kalia R, Tur J, Schouest B, Elwood D, Walters JN, Maricic I, Doan A, Vazquez M, Eblimit Z, Pezzoli P, Amante D, Porto M, Narvaez B, Lok M, Spence B, Bradette H, Horn H, Yang M, Fader J, Ferrer R, Weiner DB, Kar S, Kim JJ, Humeau LM, Ramos SJ, Smith TRF, Broderick KE. 2021. [Design, immunogenicity and efficacy of a Pan-SARS-CoV-2 synthetic DNA vaccine](#). Cold Spring Harbor Laboratory.
1527. Lauer KB, Borrow R, Blanchard TJ. 2017. [Multivalent and Multipathogen Viral Vector Vaccines](#). *Clin Vaccine Immunol* 24.
1528. Ewer KJ, Lambe T, Rollier CS, Spencer AJ, Hill AV, Dorrell L. 2016. [Viral vectors as vaccine platforms: from immunogenicity to impact](#). *Current Opinion in Immunology* 41:47–54.
1529. Antrobus RD, Coughlan L, Berthoud TK, Dicks MD, Hill AV, Lambe T, Gilbert SC. 2014. [Clinical Assessment of a Novel Recombinant Simian Adenovirus ChAdOx1 as a Vectored Vaccine Expressing Conserved Influenza A Antigens](#). *Molecular Therapy* 22:668–674.
1530. Al-Kassmy J, Pedersen J, Kobinger G. 2020. [Vaccine Candidates against Coronavirus Infections. Where Does COVID-19 Stand?](#) *Viruses* 12:861.
1531. Pastoret P-P, Vanderplasschen A. 2003. [Poxviruses as vaccine vectors](#). *Comparative Immunology, Microbiology and Infectious Diseases* 26:343–355.
1532. García-Arriaza J, Esteban M. 2014. [Enhancing poxvirus vectors vaccine immunogenicity](#). *Human Vaccines & Immunotherapeutics* 10:2235–2244.
1533. Lasaro MO, Ertl HC. 2009. [New Insights on Adenovirus as Vaccine Vectors](#). *Molecular Therapy* 17:1333–1339.
1534. Roberts A, Buonocore L, Price R, Forman J, Rose JK. 1999. [Attenuated vesicular stomatitis viruses as vaccine vectors](#). *J Virol* 73:3723–32.
1535. Lichty BD, Power AT, Stojdl DF, Bell JC. 2004. [Vesicular stomatitis virus: re-inventing the bullet](#). *Trends in Molecular Medicine* 10:210–216.
1536. Rollier CS, Reyes-Sandoval A, Cottingham MG, Ewer K, Hill AV. 2011. [Viral vectors as vaccine platforms: deployment in sight](#). *Current Opinion in Immunology* 23:377–382.

1537. Nayak S, Herzog RW. 2009. [Progress and prospects: immune responses to viral vectors](#). *Gene Ther* 17:295–304.
1538. Ura T, Okuda K, Shimada M. 2014. [Developments in Viral Vector-Based Vaccines](#). *Vaccines* 2:624–641.
1539. Moffatt S, Hays J, HogenEsch H, Mittal SK. 2000. [Circumvention of Vector-Specific Neutralizing Antibody Response by Alternating Use of Human and Non-Human Adenoviruses: Implications in Gene Therapy](#). *Virology* 272:159–167.
1540. Fausther-Bovendo H, Kobinger GP. 2014. [Pre-existing immunity against Ad vectors](#). *Human Vaccines & Immunotherapeutics* 10:2875–2884.
1541. Vrba SM, Kirk NM, Brisse ME, Liang Y, Ly H. 2020. [Development and Applications of Viral Vectored Vaccines to Combat Zoonotic and Emerging Public Health Threats](#). *Vaccines* 8:680.
1542. Ollmann Saphire E. 2020. [A Vaccine against Ebola Virus](#). *Cell* 181:6.
1543. Bliss CM, Drammeh A, Bowyer G, Sanou GS, Jagne YJ, Ouedraogo O, Edwards NJ, Tarama C, Ouedraogo N, Ouedraogo M, Njie-Jobe J, Diarra A, Afolabi MO, Tiono AB, Yaro JB, Adetifa UJ, Hodgson SH, Anagnostou NA, Roberts R, Duncan CJA, Cortese R, Viebig NK, Leroy O, Lawrie AM, Flanagan KL, Kampmann B, Imoukhuede EB, Sirima SB, Bojang K, Hill AVS, Nébié I, Ewer KJ. 2017. [Viral Vector Malaria Vaccines Induce High-Level T Cell and Antibody Responses in West African Children and Infants](#). *Molecular Therapy* 25:547–559.
1544. Li S, Locke E, Bruder J, Clarke D, Doolan DL, Havenga MJE, Hill AVS, Liljestrom P, Monath TP, Naim HY, Ockenhouse C, Tang DC, Van Kampen KR, Viret J-F, Zavala F, Dubovsky F. 2007. [Viral vectors for malaria vaccine development](#). *Vaccine* 25:2567–2574.
1545. Ledgerwood JE, DeZure AD, Stanley DA, Coates EE, Novik L, Enama ME, Berkowitz NM, Hu Z, Joshi G, Ploquin A, Sitar S, Gordon IJ, Plummer SA, Holman LA, Hendel CS, Yamshchikov G, Roman F, Nicosia A, Colloca S, Cortese R, Bailer RT, Schwartz RM, Roederer M, Mascola JR, Koup RA, Sullivan NJ, Graham BS. 2017. [Chimpanzee Adenovirus Vector Ebola Vaccine](#). *N Engl J Med* 376:928–938.
1546. Geisbert TW, Feldmann H. 2011. [Recombinant Vesicular Stomatitis Virus-Based Vaccines Against Ebola and Marburg Virus Infections](#). *The Journal of Infectious Diseases* 204:S1075–S1081.
1547. Marzi A, Feldmann H. 2014. [Ebola virus vaccines: an overview of current approaches](#). *Expert Review of Vaccines* 13:521–531.
1548. Parks CL, Picker LJ, King CR. 2013. [Development of replication-competent viral vectors for HIV vaccine delivery](#). *Current Opinion in HIV and AIDS* 8:402–411.
1549. Trivedi S, Jackson RJ, Ranasinghe C. 2014. [Different HIV pox viral vector-based vaccines and adjuvants can induce unique antigen presenting cells that modulate CD8 T cell avidity](#). *Virology* 468-470:479–489.

1550. See RH, Zakhartchouk AN, Petric M, Lawrence DJ, Mok CPY, Hogan RJ, Rowe T, Zitzow LA, Karunakaran KP, Hitt MM, Graham FL, Prevec L, Mahony JB, Sharon C, Auperin TC, Rini JM, Tingle AJ, Scheifele DW, Skowronski DM, Patrick DM, Voss TG, Babiuk LA, Gauldie J, Roper RL, Brunham RC, Finlay BB. 2006. [Comparative evaluation of two severe acute respiratory syndrome \(SARS\) vaccine candidates in mice challenged with SARS coronavirus](#). Journal of General Virology 87:641–650.
1551. See RH, Petric M, Lawrence DJ, Mok CPY, Rowe T, Zitzow LA, Karunakaran KP, Voss TG, Brunham RC, Gauldie J, Finlay BB, Roper RL. 2008. [Severe acute respiratory syndrome vaccine efficacy in ferrets: whole killed virus and adenovirus-vectored vaccines](#). Journal of General Virology 89:2136–2146.
1552. Yeung HT. 2018. Update with the development of Ebola vaccines and implications of emerging evidence to inform future policy recommendations. World Health Organization SAGE meeting background.
https://www.who.int/immunization/sage/meetings/2018/october/2_Ebola_SAGE2018Oct_BgDoc_20180919.pdf.
1553. Alharbi NK, Padron-Regalado E, Thompson CP, Kupke A, Wells D, Sloan MA, Grehan K, Temperton N, Lambe T, Warimwe G, Becker S, Hill AVS, Gilbert SC. 2017. [ChAdOx1 and MVA based vaccine candidates against MERS-CoV elicit neutralising antibodies and cellular immune responses in mice](#). Vaccine 35:3780–3788.
1554. van Doremalen N, Haddock E, Feldmann F, Meade-White K, Bushmaker T, Fischer RJ, Okumura A, Hanley PW, Saturday G, Edwards NJ, Clark MHA, Lambe T, Gilbert SC, Munster VJ. 2020. [A single dose of ChAdOx1 MERS provides protective immunity in rhesus macaques](#). Sci Adv 6.
1555. Folegatti PM, Bittaye M, Flaxman A, Lopez FR, Bellamy D, Kupke A, Mair C, Makinson R, Sheridan J, Rohde C, Halwe S, Jeong Y, Park Y-S, Kim J-O, Song M, Boyd A, Tran N, Silman D, Poulton I, Datoo M, Marshall J, Themistocleous Y, Lawrie A, Roberts R, Berrie E, Becker S, Lambe T, Hill A, Ewer K, Gilbert S. 2020. [Safety and immunogenicity of a candidate Middle East respiratory syndrome coronavirus viral-vectored vaccine: a dose-escalation, open-label, non-randomised, uncontrolled, phase 1 trial](#). The Lancet Infectious Diseases 20:816–826.
1556. van Doremalen N, Lambe T, Spencer A, Belij-Rammerstorfer S, Purushotham JN, Port JR, Avanzato VA, Bushmaker T, Flaxman A, Ulaszewska M, Feldmann F, Allen ER, Sharpe H, Schulz J, Holbrook M, Okumura A, Meade-White K, Pérez-Pérez L, Edwards NJ, Wright D, Bissett C, Gilbride C, Williamson BN, Rosenke R, Long D, Ishwarbhai A, Kailath R, Rose L, Morris S, Powers C, Lovaglio J, Hanley PW, Scott D, Saturday G, de Wit E, Gilbert SC, Munster VJ. 2020. [ChAdOx1 nCoV-19 vaccine prevents SARS-CoV-2 pneumonia in rhesus macaques](#). Nature 586:578–582.
1557. Folegatti PM, Ewer KJ, Aley PK, Angus B, Becker S, Belij-Rammerstorfer S, Bellamy D, Bibi S, Bittaye M, Clutterbuck EA, Dold C, Faust SN, Finn A, Flaxman AL, Hallis B, Heath P, Jenkin D, Lazarus R, Makinson R,

Minassian AM, Pollock KM, Ramasamy M, Robinson H, Snape M, Tarrant R, Voysey M, Green C, Douglas AD, Hill AVS, Lambe T, Gilbert SC, Pollard AJ, Aboagye J, Adams K, Ali A, Allen E, Allison JL, Anslow R, Arbe-Barnes EH, Babbage G, Baillie K, Baker M, Baker N, Baker P, Baleanu I, Ballaminut J, Barnes E, Barrett J, Bates L, Batten A, Beadon K, Beckley R, Berrie E, Berry L, Beveridge A, Bewley KR, Bijker EM, Bingham T, Blackwell L, Blundell CL, Bolam E, Boland E, Borthwick N, Bower T, Boyd A, Brenner T, Bright PD, Brown-O'Sullivan C, Brunt E, Burbage J, Burge S, Buttigieg KR, Byard N, Cabera Puig I, Calvert A, Camara S, Cao M, Cappuccini F, Carr M, Carroll MW, Carter V, Cathie K, Challis RJ, Charlton S, Chelysheva I, Cho J-S, Cicconi P, Cifuentes L, Clark H, Clark E, Cole T, Colin-Jones R, Conlon CP, Cook A, Coombes NS, Cooper R, Cosgrove CA, Coy K, Crocker WEM, Cunningham CJ, Damratowski BE, Dando L, Datoo MS, Davies H, De Graaf H, Demissie T, Di Maso C, Dietrich I, Dong T, Donnellan FR, Douglas N, Downing C, Drake J, Drake-Brockman R, Drury RE, Dunachie SJ, Edwards NJ, Edwards FDL, Edwards CJ, Elias SC, Elmore MJ, Emary KRW, English MR, Fagerbrink S, Felle S, Feng S, Field S, Fixmer C, Fletcher C, Ford KJ, Fowler J, Fox P, Francis E, Frater J, Furze J, Fuskova M, Galiza E, Gbesemete D, Gilbride C, Godwin K, Gorini G, Goulston L, Grabau C, Gracie L, Gray Z, Guthrie LB, Hackett M, Halwe S, Hamilton E, Hamlyn J, Hanumunthadu B, Harding I, Harris SA, Harris A, Harrison D, Harrison C, Hart TC, Haskell L, Hawkins S, Head I, Henry JA, Hill J, Hodgson SHC, Hou MM, Howe E, Howell N, Hutlin C, Ikram S, Isitt C, Iveson P, Jackson S, Jackson F, James SW, Jenkins M, Jones E, Jones K, Jones CE, Jones B, Kailath R, Karampatsas K, Keen J, Kelly S, Kelly D, Kerr D, Kerridge S, Khan L, Khan U, Killen A, Kinch J, King TB, King L, King J, Kingham-Page L, Klenerman P, Knapper F, Knight JC, Knott D, Koleva S, Kupke A, Larkworthy CW, Larwood JPJ, Laskey A, Lawrie AM, Lee A, Ngan Lee KY, Lees EA, Legge H, Lelliott A, Lemm N-M, Lias AM, Linder A, Lipworth S, Liu X, Liu S, Lopez Ramon R, Lwin M, Mabesa F, Madhavan M, Mallett G, Mansatta K, Marcal I, Marinou S, Marlow E, Marshall JL, Martin J, McEwan J, McInroy L, Meddaugh G, Mentzer AJ, Mirtorabi N, Moore M, Moran E, Morey E, Morgan V, Morris SJ, Morrison H, Morshead G, Morter R, Mujadidi YF, Muller J, Munera-Huertas T, Munro C, Munro A, Murphy S, Munster VJ, Mweu P, Noé A, Nugent FL, Nuthall E, O'Brien K, O'Connor D, Oguti B, Oliver JL, Oliveira C, O'Reilly PJ, Osborn M, Osborne P, Owen C, Owens D, Owino N, Pacurar M, Parker K, Parracho H, Patrick-Smith M, Payne V, Pearce J, Peng Y, Peralta Alvarez MP, Perring J, Pfafferott K, Pipini D, Plested E, Pluess-Hall H, Pollock K, Poulton I, Presland L, Provstgaard-Morys S, Pulido D, Radia K, Ramos Lopez F, Rand J, Ratcliffe H, Rawlinson T, Rhead S, Riddell A, Ritchie AJ, Roberts H, Robson J, Roche S, Rohde C, Rollier CS, Romani R, Rudiansyah I, Saich S, Sajjad S, Salvador S, Sanchez Riera L, Sanders H, Sanders K, Sapaun S, Sayce C, Schofield E, Screation G, Selby B, Semple C, Sharpe HR, Shaik I, Shea A, Shelton H, Silk S, Silva-Reyes L, Skelly DT, Smee H, Smith CC, Smith DJ, Song R, Spencer AJ, Stafford E, Steele A, Stefanova E, Stockdale L, Szigeti A, Tahiri-Alaoui A, Tait M, Talbot H, Tanner R, Taylor IJ, Taylor V, Te Water Naude R, Thakur N, Themistocleous Y, Themistocleous A, Thomas M, Thomas TM, Thompson A, Thomson-Hill S, Tomlins J, Tonks S, Towner J, Tran N, Tree JA, Truby A, Turkentine K, Turner C, Turner N, Turner S, Tuthill T, Ulaszewska M, Varughese R, Van Doremalen N, Veighey K, Verheul MK, Vichos I, Vitale E, Walker L, Watson MEE, Welham B, Wheat J, White C,

- White R, Worth AT, Wright D, Wright S, Yao XL, Yau Y. 2020. [Safety and immunogenicity of the ChAdOx1 nCoV-19 vaccine against SARS-CoV-2: a preliminary report of a phase 1/2, single-blind, randomised controlled trial](#). The Lancet 396:467–478.
1558. Jones I, Roy P. 2021. [Sputnik V COVID-19 vaccine candidate appears safe and effective](#). The Lancet 397:642–643.
1559. Barouch DH, Kik SV, Weverling GJ, Dilan R, King SL, Maxfield LF, Clark S, Ng'ang'a D, Brandariz KL, Abbink P, Sinangil F, de Bruyn G, Gray GE, Roux S, Bekker L-G, Dilraj A, Kibuuka H, Robb ML, Michael NL, Anzala O, Amornkul PN, Gilmour J, Hural J, Buchbinder SP, Seaman MS, Dolin R, Baden LR, Carville A, Mansfield KG, Pau MG, Goudsmit J. 2011. [International seroepidemiology of adenovirus serotypes 5, 26, 35, and 48 in pediatric and adult populations](#). Vaccine 29:5203–5209.
1560. Office USGA. Operation Warp Speed: Accelerated COVID-19 Vaccine Development Status and Efforts to Address Manufacturing Challenges. <https://www.gao.gov/products/gao-21-319>. Retrieved 5 December 2022.
1561. Johnson & Johnson Announces a Lead Vaccine Candidate for COVID-19; Landmark New Partnership with U.S. Department of Health & Human Services; and Commitment to Supply One Billion Vaccines Worldwide for Emergency Pandemic Use | Johnson & Johnson. Content Lab US. <https://www.jnj.com/johnson-johnson-announces-a-lead-vaccine-candidate-for-covid-19-landmark-new-partnership-with-u-s-department-of-health-human-services-and-commitment-to-supply-one-billion-vaccines-worldwide-for-emergency-pandemic-use>. Retrieved 5 December 2022.
1562. Sadoff J, Le Gars M, Shukarev G, Heerwegh D, Truyers C, de Groot AM, Stoop J, Tete S, Van Damme W, Leroux-Roels I, Berghmans P-J, Kimmel M, Van Damme P, de Hoon J, Smith W, Stephenson KE, De Rosa SC, Cohen KW, McElrath MJ, Cormier E, Scheper G, Barouch DH, Hendriks J, Struyf F, Douoguih M, Van Hoof J, Schuitemaker H. 2021. [Interim Results of a Phase 1–2a Trial of Ad26.COV2.S Covid-19 Vaccine](#). N Engl J Med 384:1824–1835.
1563. Mercado NB, Zahn R, Wegmann F, Loos C, Chandrashekar A, Yu J, Liu J, Peter L, McMahan K, Tostanoski LH, He X, Martinez DR, Rutten L, Bos R, van Manen D, Vellinga J, Custers J, Langedijk JP, Kwaks T, Bakkers MJG, Zuijdgeest D, Rosendahl Huber SK, Atyeo C, Fischinger S, Burke JS, Feldman J, Hauser BM, Caradonna TM, Bondzie EA, Dagotto G, Gebre MS, Hoffman E, Jacob-Dolan C, Kirilova M, Li Z, Lin Z, Mahrokhian SH, Maxfield LF, Nampanya F, Nityanandam R, Nkolola JP, Patel S, Ventura JD, Verrington K, Wan H, Pessaint L, Van Ry A, Blade K, Strasbaugh A, Cabus M, Brown R, Cook A, Zouantchangadou S, Teow E, Andersen H, Lewis MG, Cai Y, Chen B, Schmidt AG, Reeves RK, Baric RS, Lauffenburger DA, Alter G, Stoffels P, Mammen M, Van Hoof J, Schuitemaker H, Barouch DH. 2020. [Single-shot Ad26 vaccine protects against SARS-CoV-2 in rhesus macaques](#). Nature 586:583–588.
1564. Tostanoski LH, Wegmann F, Martinot AJ, Loos C, McMahan K, Mercado NB, Yu J, Chan CN, Bondoc S, Starke CE, Nekorchuk M, Busman-Sahay

- K, Piedra-Mora C, Wrijil LM, Ducat S, Custers J, Atyeo C, Fischinger S, Burke JS, Feldman J, Hauser BM, Caradonna TM, Bondzie EA, Dagotto G, Gebre MS, Jacob-Dolan C, Lin Z, Mahrokhian SH, Nampanya F, Nityanandam R, Pessaint L, Porto M, Ali V, Benetiene D, Tevi K, Andersen H, Lewis MG, Schmidt AG, Lauffenburger DA, Alter G, Estes JD, Schuitemaker H, Zahn R, Barouch DH. 2020. [Ad26 vaccine protects against SARS-CoV-2 severe clinical disease in hamsters](#). Nat Med 26:1694–1700.
1565. Solforosi L, Kuipers H, Huber SKR, van der Lubbe JEM, Dekking L, Czapska-Casey DN, Gil AI, Baert MRM, Drijver J, Vaneman J, van Huizen E, Choi Y, Vreugdenhil J, Kroos S, de Wilde AH, Kourkouta E, Custers J, Dalebout TJ, Myeni SK, Kikkert M, Snijder EJ, Barouch DH, Böszörményi KP, Stammes MA, Kondova I, Verschoor EJ, Verstrepen BE, Koopman G, Mooij P, Bogers WMJM, van Heerden M, Muchene L, Tolboom JTBM, Roozendaal R, Schuitemaker H, Wegmann F, Zahn RC. 2020. [Immunogenicity and protective efficacy of one- and two-dose regimens of the Ad26.COV2.S COVID-19 vaccine candidate in adult and aged rhesus macaques](#). Cold Spring Harbor Laboratory.
1566. Roozendaal R, Solforosi L, Stieh D, Serroyen J, Straetemans R, Wegmann F, Rosendahl Huber SK, van der Lubbe JEM, Hendriks J, le Gars M, Dekking L, Czapska-Casey DN, Guimera N, Janssen S, Tete S, Chandrashekar A, Mercado N, Yu J, Koudstaal W, Sadoff J, Barouch DH, Schuitemaker H, Zahn R. 2021. [SARS-CoV-2 binding and neutralizing antibody levels after vaccination with Ad26.COV2.S predict durable protection in rhesus macaques](#). Cold Spring Harbor Laboratory.
1567. Sadoff J, Gray G, Vandebosch A, Cárdenas V, Shukarev G, Grinsztejn B, Goepfert PA, Truysers C, Van Dromme I, Spiessens B, Vingerhoets J, Custers J, Scheper G, Robb ML, Treanor J, Ryser MF, Barouch DH, Swann E, Marovich MA, Neuzil KM, Corey L, Stoddard J, Hardt K, Ruiz-Guiñazú J, Le Gars M, Schuitemaker H, Van Hoof J, Struyf F, Douoguih M. 2022. [Final Analysis of Efficacy and Safety of Single-Dose Ad26.COV2.S](#). N Engl J Med 386:847–860.
1568. AstraZeneca's COVID-19 vaccine authorised for emergency supply in the UK. <https://www.astrazeneca.com/media-centre/press-releases/2020/astrazenecas-covid-19-vaccine-authorised-in-uk.html>. Retrieved 5 December 2022.
1569. The Brussels Times. <https://www.brusselstimes.com/news-contents/world/149039/1-5-million-people-have-received-sputnik-v-vaccine-russia-says-russian-direct-investment-fund-mikhail-murashko>. Retrieved 5 December 2022.
1570. Daventry M. 2021. Hungary becomes first EU country to deploy Russia's COVID-19 vaccine. euronews. <https://www.euronews.com/2021/02/12/hungary-to-begin-using-russia-s-sputnik-v-vaccine-today>. Retrieved 5 December 2022.
1571. 2021. San Marino buys Russia's Sputnik V after EU vaccine delivery delays. euronews. <https://www.euronews.com/2021/02/24/san-marino-buys-russia-s-sputnik-v-after-eu-vaccine-delivery-delays>. Retrieved 5 December 2022.

1572. AFP. 2020. Belarus Starts Coronavirus Vaccination With Sputnik V. The Moscow Times.
<https://www.themoscowtimes.com/2020/12/29/belarus-starts-coronavirus-vaccination-with-sputnik-v-a72512>. Retrieved 5 December 2022.
1573. Logunov DY, Dolzhikova IV, Shcheblyakov DV, Tukhvatulin AI, Zubkova OV, Dzharullaeva AS, Kovyshina AV, Lubenets NL, Grousova DM, Erokhova AS, Botikov AG, Izhaeva FM, Popova O, Ozharovskaya TA, Esmagambetov IB, Favorskaya IA, Zrelkin DI, Voronina DV, Shcherbinin DN, Semikhin AS, Simakova YV, Tokarskaya EA, Egorova DA, Shmarov MM, Nikitenko NA, Gushchin VA, Smolyarchuk EA, Zyryanov SK, Borisevich SV, Naroditsky BS, Gintsburg AL. 2021. [Safety and efficacy of an rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine: an interim analysis of a randomised controlled phase 3 trial in Russia](#). The Lancet 397:671–681.
1574. Sadoff J, Gray G, Vandebosch A, Cárdenas V, Shukarev G, Grinsztejn B, Goepfert PA, Truyers C, Fennema H, Spiessens B, Offergeld K, Scheper G, Taylor KL, Robb ML, Treanor J, Barouch DH, Stoddard J, Ryser MF, Marovich MA, Neuzil KM, Corey L, Cauwenberghs N, Tanner T, Hardt K, Ruiz-Guiñazú J, Le Gars M, Schuitemaker H, Van Hoof J, Struyf F, Douoguih M. 2021. [Safety and Efficacy of Single-Dose Ad26.COV2.S Vaccine against Covid-19](#). N Engl J Med 384:2187–2201.
1575. 2021. Janssen Investigational COVID-19 Vaccine: Interim Analysis of Phase 3 Clinical Data Released. National Institutes of Health (NIH).
<https://www.nih.gov/news-events/news-releases/janssen-investigational-covid-19-vaccine-interim-analysis-phase-3-clinical-data-released>. Retrieved 5 December 2022.
1576. 2021. Johnson & Johnson Announces Single-Shot Janssen COVID-19 Vaccine Candidate Met Primary Endpoints in Interim Analysis of its Phase 3 ENSEMBLE Trial. Janssen.
https://www.janssen.com/emea/sites/www.janssen.com_emea/files/johnson_johnson_announces_single-shot_janssen_covid-19_vaccine_candidate_met_primary_endpoints_in_interim_analysis_of_its_phase_3_ensemble_trial.pdf.
1577. Burki TK. 2020. [The Russian vaccine for COVID-19](#). The Lancet Respiratory Medicine 8:e85–e86.
1578. Cohen J. 2020. Russia's claim of a successful COVID-19 vaccine doesn't pass the 'smell test,' critics say. Science
<https://doi.org/10.1126/science.abf6791>.
1579. Callaway E. 2020. Russia announces positive COVID-vaccine results from controversial trial. Nature <https://doi.org/10.1038/d41586-020-03209-0>.
1580. Thorp HH. 2020. [A dangerous rush for vaccines](#). Science 369:885–885.
1581. Jr BL. Scientists worry whether Russia's 'Sputnik V' coronavirus vaccine is safe and effective. CNBC.
<https://www.cnbc.com/2020/08/11/scientists-worry-whether-russias->

[sputnik-v-coronavirus-vaccine-is-safe-and-effective.html](#). Retrieved 5 December 2022.

1582. Logunov DY, Dolzhikova IV, Zubkova OV, Tukhvatulin AI, Shcheblyakov DV, Dzharullaeva AS, Grousova DM, Erokhova AS, Kovyrshina AV, Botikov AG, Izhaeva FM, Popova O, Ozharovskaya TA, Esmagambetov IB, Favorskaya IA, Zrelkin DI, Voronina DV, Shcherbinin DN, Semikhin AS, Simakova YV, Tokarskaya EA, Lubenets NL, Egorova DA, Shmarov MM, Nikitenko NA, Morozova LF, Smolyarchuk EA, Kryukov EV, Babira VF, Borisevich SV, Naroditsky BS, Gintsburg AL. 2020. [Safety and immunogenicity of an rAd26 and rAd5 vector-based heterologous prime-boost COVID-19 vaccine in two formulations: two open, non-randomised phase 1/2 studies from Russia](#). The Lancet 396:887–897.
1583. Clinical Trials. <https://sputnikvaccine.com/about-vaccine/clinical-trials/>. Retrieved 5 December 2022.
1584. Phillips N, Cyranoski D, Mallapaty S. 2020. A leading coronavirus vaccine trial is on hold: scientists react. Nature <https://doi.org/10.1038/d41586-020-02594-w>.
1585. Cyranoski D, Mallapaty S. 2020. [Scientists relieved as coronavirus vaccine trial restarts — but question lack of transparency](#). Nature 585:331–332.
1586. Robbins R, LaFraniere S, Weiland N, Kirkpatrick DD, Mueller B. 2020. [Blunders Eroded U.S. Confidence in Early Vaccine Front-Runner](#). The New York Times.
1587. 2020. [Oxford/AstraZeneca Covid vaccine 'dose error' explained](#). BBC News.
1588. Sanchez S, Palacio N, Dangi T, Ciucci T, Penaloza-MacMaster P. 2021. [Fractionating a COVID-19 Ad5-vectored vaccine improves virus-specific immunity](#). Sci Immunol 6.
1589. Wolf ME, Luz B, Niehaus L, Bhogal P, Bänzner H, Henkes H. 2021. [Thrombocytopenia and Intracranial Venous Sinus Thrombosis after “COVID-19 Vaccine AstraZeneca” Exposure](#). JCM 10:1599.
1590. Wise J. 2021. [Covid-19: European countries suspend use of Oxford-AstraZeneca vaccine after reports of blood clots](#). BMJ n699.
1591. Mahase E. 2021. [Covid-19: AstraZeneca vaccine is not linked to increased risk of blood clots, finds European Medicine Agency](#). BMJ n774.
1592. Mahase E. 2021. [Covid-19: US suspends Johnson and Johnson vaccine rollout over blood clots](#). BMJ n970.
1593. Tanne JH. 2021. [Covid-19: US authorises Johnson and Johnson vaccine again, ending pause in rollout](#). BMJ n1079.
1594. Mahase E. 2021. [Covid-19: Unusual blood clots are “very rare side effect” of Janssen vaccine, says EMA](#). BMJ n1046.

1595. Oliver SE, Wallace M, See I, Mbaeyi S, Godfrey M, Hadler SC, Jatlaoui TC, Twentyman E, Hughes MM, Rao AK, Fiore A, Su JR, Broder KR, Shimabukuro T, Lale A, Shay DK, Markowitz LE, Wharton M, Bell BP, Brooks O, McNally V, Lee GM, Talbot HK, Daley MF. 2022. [Use of the Janssen \(Johnson & Johnson\) COVID-19 Vaccine: Updated Interim Recommendations from the Advisory Committee on Immunization Practices — United States, December 2021](#). MMWR Morb Mortal Wkly Rep 71:90–95.
1596. Shay DK, Gee J, Su JR, Myers TR, Marquez P, Liu R, Zhang B, Licata C, Clark TA, Shimabukuro TT. 2021. [Safety Monitoring of the Janssen \(Johnson & Johnson\) COVID-19 Vaccine — United States, March–April 2021](#). MMWR Morb Mortal Wkly Rep 70:680–684.
1597. Rosenblum HG, Hadler SC, Moulia D, Shimabukuro TT, Su JR, Tepper NK, Ess KC, Woo EJ, Mba-Jonas A, Alimchandani M, Nair N, Klein NP, Hanson KE, Markowitz LE, Wharton M, McNally VV, Romero JR, Talbot HK, Lee GM, Daley MF, Mbaeyi SA, Oliver SE. 2021. [Use of COVID-19 Vaccines After Reports of Adverse Events Among Adult Recipients of Janssen \(Johnson & Johnson\) and mRNA COVID-19 Vaccines \(Pfizer-BioNTech and Moderna\): Update from the Advisory Committee on Immunization Practices — United States, July 2021](#). MMWR Morb Mortal Wkly Rep 70:1094–1099.
1598. Pottegård A, Lund LC, Karlstad Ø, Dahl J, Andersen M, Hallas J, Lidegaard Ø, Tapia G, Gulseth HL, Ruiz PL-D, Wattle SV, Mikkelsen AP, Pedersen L, Sørensen HT, Thomsen RW, Hviid A. 2021. [Arterial events, venous thromboembolism, thrombocytopenia, and bleeding after vaccination with Oxford-AstraZeneca ChAdOx1-S in Denmark and Norway: population based cohort study](#). BMJ n1114.
1599. Chan B, Odutayo A, Juni P, Stall NM, Bobos P, Brown AD, Grill A, Ivers N, Maltsev A, McGeer A, Miller KJ, Niel U, Razak F, Sander B, Sholzberg M, Slutsky AS, Morris AM, Pai M. 2021. [Risk of Vaccine-Induced Thrombotic Thrombocytopenia \(VITT\) following the AstraZeneca/COVISHIELD Adenovirus Vector COVID-19 Vaccines](#). Ontario COVID-19 Science Advisory Table.
1600. Baker AT, Boyd RJ, Sarkar D, Teijeira-Crespo A, Chan CK, Bates E, Waraich K, Vant J, Wilson E, Truong CD, Lipka-Lloyd M, Fromme P, Vermaas J, Williams D, Machiesky L, Heurich M, Nagalo BM, Coughlan L, Umlauf S, Chiu P-L, Rizkallah PJ, Cohen TS, Parker AL, Singharoy A, Borad MJ. 2021. [ChAdOx1 interacts with CAR and PF4 with implications for thrombosis with thrombocytopenia syndrome](#). Sci Adv 7.
1601. Schultz NH, Sørvoll IH, Michelsen AE, Munthe LA, Lund-Johansen F, Ahlen MT, Wiedmann M, Aamodt A-H, Skattør TH, Tjønnfjord GE, Holme PA. 2021. [Thrombosis and Thrombocytopenia after ChAdOx1 nCoV-19 Vaccination](#). N Engl J Med 384:2124–2130.
1602. 2021. EMA recommends COVID-19 Vaccine AstraZeneca for authorisation in the EU. <https://www.ema.europa.eu/en/news/ema-recommends-covid-19-vaccine-astrazeneca-authorisation-eu>.

1603. Miesbach W, Makris M. 2020. [COVID-19: Coagulopathy, Risk of Thrombosis, and the Rationale for Anticoagulation](#). Clin Appl Thromb Hemost 26:107602962093814.
1604. Verbeke R, Lentacker I, De Smedt SC, Dewitte H. 2019. [Three decades of messenger RNA vaccine development](#). Nano Today 28:100766.
1605. Schlake T, Thess A, Fotin-Mleczek M, Kallen K-J. 2012. [Developing mRNA-vaccine technologies](#). RNA Biology 9:1319–1330.
1606. Martinon F, Krishnan S, Lenzen G, Magné R, Gomard E, Guillet J-G, Lévy J-P, Meulien P. 1993. [Induction of virus-specific cytotoxic T lymphocytes in vivo by liposome-entrapped mRNA](#). European Journal of Immunology 23:1719–1722.
1607. Reichmuth AM, Oberli MA, Jaklenec A, Langer R, Blankschtein D. 2016. [mRNA vaccine delivery using lipid nanoparticles](#). Therapeutic Delivery 7:319–334.
1608. Iavarone C, O'hagan DT, Yu D, Delahaye NF, Ulmer JB. 2017. [Mechanism of action of mRNA-based vaccines](#). Expert Review of Vaccines 16:871–881.
1609. RNA vaccines: an introduction. PHG Foundation. <https://www.phgfoundation.org/briefing/rna-vaccines>. Retrieved 8 February 2021.
1610. Crotty S. 2014. [T Follicular Helper Cell Differentiation, Function, and Roles in Disease](#). Immunity 41:529–542.
1611. Pardi N, Hogan MJ, Porter FW, Weissman D. 2018. [mRNA vaccines — a new era in vaccinology](#). Nature Reviews Drug Discovery 17:261–279.
1612. Stuart LM. 2021. [In Gratitude for mRNA Vaccines](#). N Engl J Med 385:1436–1438.
1613. Pardi N, Hogan MJ, Weissman D. 2020. [Recent advances in mRNA vaccine technology](#). Current Opinion in Immunology 65:14–20.
1614. Amanat F, Krammer F. 2020. [SARS-CoV-2 Vaccines: Status Report](#). Immunity 52:583–589.
1615. Fonteilles-Drabek S, Reddy D, Wells TNC. 2017. [Managing intellectual property to develop medicines for the world's poorest](#). Nat Rev Drug Discov 16:223–224.
1616. Study in Healthy Adults to Evaluate Gene Activation After Vaccination With GlaxoSmithKline (GSK) Biologicals' Candidate Tuberculosis (TB) Vaccine GSK 692342 - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT01669096>. Retrieved 8 February 2021.
1617. Pardi N, Parkhouse K, Kirkpatrick E, McMahon M, Zost SJ, Mui BL, Tam YK, Karikó K, Barbosa CJ, Madden TD, Hope MJ, Krammer F, Hensley SE, Weissman D. 2018. [Nucleoside-modified mRNA immunization elicits](#)

[influenza virus hemagglutinin stalk-specific antibodies](#). Nature Communications 9:3361.

1618. Veiga N, Goldsmith M, Granot Y, Rosenblum D, Dammes N, Kedmi R, Ramishetti S, Peer D. 2018. [Cell specific delivery of modified mRNA expressing therapeutic proteins to leukocytes](#). Nature Communications 9:4493.
1619. Vabret N, Britton GJ, Gruber C, Hegde S, Kim J, Kuksin M, Levantovsky R, Malle L, Moreira A, Park MD, Pia L, Risson E, Saffern M, Salomé B, Esai Selvan M, Spindler MP, Tan J, van der Heide V, Gregory JK, Alexandropoulos K, Bhardwaj N, Brown BD, Greenbaum B, Gümüş ZH, Homann D, Horowitz A, Kamphorst AO, Curotto de Lafaille MA, Mehandru S, Merad M, Samstein RM, Agrawal M, Aleynick M, Belabed M, Brown M, Casanova-Acebes M, Catalan J, Centa M, Charap A, Chan A, Chen ST, Chung J, Bozkus CC, Cody E, Cossarini F, Dalla E, Fernandez N, Grout J, Ruan DF, Hamon P, Humblin E, Jha D, Kodysh J, Leader A, Lin M, Lindblad K, Lozano-Ojalvo D, Lubitz G, Magen A, Mahmood Z, Martinez-Delgado G, Mateus-Tique J, Meritt E, Moon C, Noel J, O'Donnell T, Ota M, Plitt T, Pothula V, Redes J, Reyes Torres I, Roberto M, Sanchez-Paulete AR, Shang J, Schanoski AS, Suprun M, Tran M, Vaninov N, Wilk CM, Aguirre-Ghiso J, Bogunovic D, Cho J, Faith J, Grasset E, Heeger P, Kenigsberg E, Krammer F, Laserson U. 2020. [Immunology of COVID-19: Current State of the Science](#). Immunity 52:910–941.
1620. Chien KR, Zangi L, Lui KO. 2014. [Synthetic Chemically Modified mRNA \(modRNA\): Toward a New Technology Platform for Cardiovascular Biology and Medicine](#). Cold Spring Harbor Perspectives in Medicine 5:a014035–a014035.
1621. Pfizer and BioNTech Announce Early Positive Data from an Ongoing Phase 1/2 study of mRNA-based Vaccine Candidate Against SARS-CoV-2 | Pfizer. <https://www.pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-announce-early-positive-data-ongoing-0>. Retrieved 8 February 2021.
1622. National Institute of Allergy and Infectious Diseases (NIAID). 2020. [Phase I, Open-Label, Dose-Ranging Study of the Safety and Immunogenicity of 2019-nCoV Vaccine \(mRNA-1273\) in Healthy Adults](#). NCT04283461. Clinical trial registration. clinicaltrials.gov.
1623. Funk CD, Laferrière C, Ardakani A. 2020. [A Snapshot of the Global Race for Vaccines Targeting SARS-CoV-2 and the COVID-19 Pandemic](#). Frontiers in Pharmacology 11:937.
1624. Ledford H. 2021. [What the Moderna–NIH COVID vaccine patent fight means for research](#). Nature 600:200–201.
1625. Diamond D. 2021. [Moderna halts patent fight over coronavirus vaccine with federal government](#). Washington Post.
1626. Goodman AS Brenda. 2022. Moderna files patent infringement lawsuits against Pfizer and BioNTech over mRNA Covid-19 vaccines.

CNN. <https://www.cnn.com/2022/08/26/health/moderna-pfizer-mrna-patent-lawsuit/index.html>. Retrieved 5 December 2022.

1627. Baden LR, El Sahly HM, Essink B, Kotloff K, Frey S, Novak R, Diemert D, Spector SA, Roupheal N, Creech CB, McGettigan J, Khetan S, Segall N, Solis J, Brosz A, Fierro C, Schwartz H, Neuzil K, Corey L, Gilbert P, Janes H, Follmann D, Marovich M, Mascola J, Polakowski L, Ledgerwood J, Graham BS, Bennett H, Pajon R, Knightly C, Leav B, Deng W, Zhou H, Han S, Ivarsson M, Miller J, Zaks T. 2020. [Efficacy and Safety of the mRNA-1273 SARS-CoV-2 Vaccine](#). New England Journal of Medicine NEJMoa2035389.
1628. Commissioner O of the. 2020. FDA Takes Key Action in Fight Against COVID-19 By Issuing Emergency Use Authorization for First COVID-19 Vaccine. FDA. <https://www.fda.gov/news-events/press-announcements/fda-takes-key-action-fight-against-covid-19-issuing-emergency-use-authorization-first-covid-19>. Retrieved 8 February 2021.
1629. Oliver SE. 2021. [The Advisory Committee on Immunization Practices' Interim Recommendation for Use of Moderna COVID-19 Vaccine — United States, December 2020](#). MMWR Morbidity and Mortality Weekly Report 69.
1630. Puranik A, Lenehan PJ, Silvert E, Niesen MJM, Corchado-Garcia J, O'Horo JC, Virk A, Swift MD, Halamka J, Badley AD, Venkatakrishnan AJ, Soundararajan V. 2021. [Comparison of two highly-effective mRNA vaccines for COVID-19 during periods of Alpha and Delta variant prevalence](#). Cold Spring Harbor Laboratory.
1631. Tseng HF, Ackerson BK, Luo Y, Sy LS, Talarico CA, Tian Y, Bruxvoort KJ, Tubert JE, Florea A, Ku JH, Lee GS, Choi SK, Takhar HS, Aragonés M, Qian L. 2022. [Effectiveness of mRNA-1273 against SARS-CoV-2 Omicron and Delta variants](#). Nat Med 28:1063–1071.
1632. Collie S, Champion J, Moultrie H, Bekker L-G, Gray G. 2022. [Effectiveness of BNT162b2 Vaccine against Omicron Variant in South Africa](#). N Engl J Med 386:494–496.
1633. Andrews N, Stowe J, Kirsebom F, Toffa S, Rickeard T, Gallagher E, Gower C, Kall M, Groves N, O'Connell A-M, Simons D, Blomquist PB, Zaidi A, Nash S, Iwani Binti Abdul Aziz N, Thelwall S, Dabrera G, Myers R, Amirthalingam G, Gharbia S, Barrett JC, Elson R, Ladhani SN, Ferguson N, Zambon M, Campbell CNJ, Brown K, Hopkins S, Chand M, Ramsay M, Lopez Bernal J. 2022. [Covid-19 Vaccine Effectiveness against the Omicron \(B.1.1.529\) Variant](#). N Engl J Med 386:1532–1546.
1634. Coronavirus in the U.S.: Latest Map and Case Count. The New York Times. <https://www.nytimes.com/interactive/2021/us/covid-cases.html>. Retrieved 11 March 2022.
1635. Tan S. Analysis | Four charts that analyze how omicron's wave compares to previous coronavirus peaks. Washington Post. <https://www.washingtonpost.com/health/interactive/2022/omicron->

[comparison-cases-deaths-hospitalizations/](#). Retrieved 5 December 2022.

1636. Abu Mouch S, Roguin A, Hellou E, Ishai A, Shoshan U, Mahamid L, Zoabi M, Aisman M, Goldschmid N, Berar Yanay N. 2021. [Myocarditis following COVID-19 mRNA vaccination](#). Vaccine 39:3790–3793.
1637. Kim HW, Jenista ER, Wendell DC, Azevedo CF, Campbell MJ, Darty SN, Parker MA, Kim RJ. 2021. [Patients With Acute Myocarditis Following mRNA COVID-19 Vaccination](#). JAMA Cardiol 6:1196.
1638. Mevorach D, Anis E, Cedar N, Bromberg M, Haas EJ, Nadir E, Olsha-Castell S, Arad D, Hasin T, Levi N, Asleh R, Amir O, Meir K, Cohen D, Dichtiar R, Novick D, HersHKovitz Y, Dagan R, Leitersdorf I, Ben-Ami R, Miskin I, Saliba W, Muhsen K, Levi Y, Green MS, Keinan-Boker L, Alroy-Preis S. 2021. [Myocarditis after BNT162b2 mRNA Vaccine against Covid-19 in Israel](#). N Engl J Med 385:2140–2149.
1639. Goddard K, Hanson KE, Lewis N, Weintraub E, Fireman B, Klein NP. 2022. Incidence of Myocarditis/Pericarditis Following mRNA COVID-19 Vaccination Among Children and Younger Adults in the United States. Ann Intern Med <https://doi.org/10.7326/m22-2274>.
1640. Matta A, Kunadharaju R, Osman M, Jesme C, McMiller Z, Johnson EM, Matta D, Kallamadi R, Bande D. 2021. Clinical Presentation and Outcomes of Myocarditis Post mRNA Vaccination: A Meta-Analysis and Systematic Review. Cureus <https://doi.org/10.7759/cureus.19240>.
1641. Commissioner O of the. 2022. Coronavirus (COVID-19) Update: FDA Recommends Inclusion of Omicron BA.4/5 Component for COVID-19 Vaccine Booster Doses. FDA. <https://www.fda.gov/news-events/press-announcements/coronavirus-covid-19-update-fda-recommends-inclusion-omicron-ba45-component-covid-19-vaccine-booster>. Retrieved 5 December 2022.
1642. 2022. FDA Panel Gives Thumbs Up to Omicron-Containing COVID Boosters. <https://www.medpagetoday.com/infectiousdisease/covid19vaccine/99493>. Retrieved 5 December 2022.
1643. 2022. Fall 2022 COVID-19 Vaccine Strain Composition Selection Recommendation. US Food and Drug Administration. <https://www.fda.gov/media/159597/download>.
1644. Erman M. 2022. [U.S. FDA to use existing Omicron booster data to review shots targeting new subvariants -official](#). Reuters.
1645. Tseng HF, Ackerson BK, Bruxvoort KJ, Sy LS, Tubert JE, Lee GS, Ku JH, Florea A, Luo Y, Qiu S, Choi SK, Takhar HS, Aragones M, Paila YD, Chavers S, Talarico CA, Qian L. 2022. [Effectiveness of mRNA-1273 against infection and COVID-19 hospitalization with SARS-CoV-2 Omicron subvariants: BA.1, BA.2, BA.2.12.1, BA.4, and BA.5](#). Cold Spring Harbor Laboratory.
1646. Hardt K, Vandebosch A, Sadoff J, Gars ML, Truysers C, Lowson D, Van Dromme I, Vingerhoets J, Kamphuis T, Scheper G, Ruiz-Guiñazú J, Faust

SN, Spinner CD, Schuitemaker H, Van Hoof J, Douoguih M, Struyf F. 2022. [Efficacy and Safety of a Booster Regimen of Ad26.COV2.S Vaccine against Covid-19](#). Cold Spring Harbor Laboratory.

1647. Dolzhikova I, Iliukhina A, Kovyrshina A, Kuzina A, Gushchin V, Siniavin A, Pochtovyi A, Shidlovskaya E, Kuznetsova N, Megeryan M, Dzharullaeva A, Erokhova A, Izhaeva F, Grousova D, Botikov A, Shcheblyakov D, Tikhvatulin A, Zubkova O, Logunov D, Gintsburg A. 2021. [Sputnik Light booster after Sputnik V vaccination induces robust neutralizing antibody response to B.1.1.529 \(Omicron\) SARS-CoV-2 variant](#). Cold Spring Harbor Laboratory.
1648. Flaxman A, Marchevsky NG, Jenkin D, Aboagye J, Aley PK, Angus B, Belij-Rammerstorfer S, Bibi S, Bittaye M, Cappuccini F, Cicconi P, Clutterbuck EA, Davies S, Dejnirattisai W, Dold C, Ewer KJ, Folegatti PM, Fowler J, Hill AVS, Kerridge S, Minassian AM, Mongkolsapaya J, Mujadidi YF, Plested E, Ramasamy MN, Robinson H, Sanders H, Sheehan E, Smith H, Snape MD, Song R, Woods D, Screatton G, Gilbert SC, Voysey M, Pollard AJ, Lambe T, Adlou S, Aley R, Ali A, Anslow R, Baker M, Baker P, Barrett JR, Bates L, Beadon K, Beckley R, Bell J, Bellamy D, Beveridge A, Bissett C, Blackwell L, Bletchly H, Boyd A, Bridges-Webb A, Brown C, Byard N, Camara S, Cifuentes Gutierrez L, Collins AM, Cooper R, Crocker WEM, Darton TC, Davies H, Davies J, Demissie T, Di Maso C, Dinesh T, Donnellan FR, Douglas AD, Drake-Brockman R, Duncan CJA, Elias SC, Emary KRW, Ghulam Farooq M, Faust SN, Felle S, Ferreira D, Ferreira Da Silva C, Finn A, Ford KJ, Francis E, Furze J, Fuskova M, Galiza E, Gibertoni Cruz A, Godfrey L, Goodman AL, Green C, Green CA, Greenwood N, Harrison D, Hart TC, Hawkins S, Heath PT, Hill H, Hillson K, Horsington B, Hou MM, Howe E, Howell N, Joe C, Jones E, Kasanyinga M, Keen J, Kelly S, Kerr D, Khan L, Khozoe B, Kinch J, Kinch P, Koleva S, Kwok J, Larkworthy CW, Lawrie AM, Lazarus R, Lees EA, Li G, Libri V, Lillie PJ, Linder A, Long F, Lopez Ramon R, Mabbett R, Makinson R, Marinou S, Marlow E, Marshall JL, Mazur O, McEwan J, McGregor AC, Mokaya J, Morey E, Morshead G, Morter R, Muller J, Mweu P, Noristani R, Owino N, Polo Peralta Alvarez M, Platt A, Pollock KM, Poulton I, Provstgaard-Morys S, Pulido-Gomez D, Rajan M, Ramos Lopez F, Ritchie A, Roberts H, Rollier C, Rudiansyah I, Sanders K, Saunders JE, Seddiqi S, Sharpe HR, Shaw R, Silva-Reyes L, Singh N, Smith DJ, Smith CC, Smith A, Spencer AJ, Stuart ASV, Sutherland R, Szigeti A, Tang K, Thomas M, Thomas TM, Thompson A, Thomson EC, Török EM, Toshner M, Tran N, Trivett R, Turnbull I, Turner C, Turner DPJ, Ulaszewska M, Vichos I, Walker L, Watson ME, Whelan C, White R, Williams SJ, Williams CJA, Wright D, Yao A. 2021. [Reactogenicity and immunogenicity after a late second dose or a third dose of ChAdOx1 nCoV-19 in the UK: a substudy of two randomised controlled trials \(COV001 and COV002\)](#). The Lancet 398:981–990.
1649. Regev-Yochay G, Gonen T, Gilboa M, Mandelboim M, Indenbaum V, Amit S, Meltzer L, Asraf K, Cohen C, Fluss R, Biber A, Nemet I, Kliker L, Joseph G, Doolman R, Mendelson E, Freedman LS, Harats D, Kreiss Y, Lustig Y. 2022. [4th Dose COVID mRNA Vaccines' Immunogenicity & Efficacy Against Omicron VOC](#). Cold Spring Harbor Laboratory.

1650. Chiu N-C, Chi H, Tu Y-K, Huang Y-N, Tai Y-L, Weng S-L, Chang L, Huang DT-N, Huang F-Y, Lin C-Y. 2021. [To mix or not to mix? A rapid systematic review of heterologous prime-boost covid-19 vaccination](#). Expert Review of Vaccines 20:1211–1220.
1651. Atmar RL, Lyke KE, Deming ME, Jackson LA, Branche AR, El Sahly HM, Rostad CA, Martin JM, Johnston C, Rupp RE, Mulligan MJ, Brady RC, Frenck RW Jr., Bäcker M, Kottkamp AC, Babu TM, Rajakumar K, Edupuganti S, Dobrzynski D, Coler RN, Posavad CM, Archer JI, Crandon S, Nayak SU, Szydlo D, Zemanek JA, Dominguez Islas CP, Brown ER, Suthar MS, McElrath MJ, McDermott AB, O'Connell SE, Montefiori DC, Eaton A, Neuzil KM, Stephens DS, Roberts PC, Beigel JH. 2022. [Homologous and Heterologous Covid-19 Booster Vaccinations](#). N Engl J Med 386:1046–1057.
1652. Jara A, Undurraga EA, Zubizarreta JR, González C, Pizarro A, Acevedo J, Leo K, Paredes F, Bralic T, Vergara V, Mosso M, Leon F, Parot I, Leighton P, Suárez P, Rios JC, García-Escorza H, Araos R. 2022. [Effectiveness of homologous and heterologous booster doses for an inactivated SARS-CoV-2 vaccine: a large-scale prospective cohort study](#). The Lancet Global Health 10:e798–e806.
1653. Sapkota B, Saud B, Shrestha R, Al-Fahad D, Sah R, Shrestha S, Rodriguez-Morales AJ. 2021. Heterologous prime-boost strategies for COVID-19 vaccines. Journal of Travel Medicine <https://doi.org/10.1093/jtm/taab191>.
1654. Assawakosri S, Kanokudom S, Suntronwong N, Auphimai C, Nilyanimit P, Vichaiwattana P, Thongmee T, Duangchinda T, Chantima W, Pakchotanont P, Srimuan D, Thatsanatorn T, Klinfueng S, Yorsaeng R, Sudhinaraset N, Wanlapakorn N, Mongkolsapaya J, Honsawek S, Poovorawan Y. 2022. [Neutralizing Activities Against the Omicron Variant After a Heterologous Booster in Healthy Adults Receiving Two Doses of CoronaVac Vaccination](#). The Journal of Infectious Diseases 226:1372–1381.
1655. Accorsi EK, Britton A, Shang N, Fleming-Dutra KE, Link-Gelles R, Smith ZR, Derado G, Miller J, Schrag SJ, Verani JR. 2022. [Effectiveness of Homologous and Heterologous Covid-19 Boosters against Omicron](#). N Engl J Med 386:2433–2435.
1656. Munro APS, Janani L, Cornelius V, Aley PK, Babbage G, Baxter D, Bula M, Cathie K, Chatterjee K, Dodd K, Enever Y, Gokani K, Goodman AL, Green CA, Harndahl L, Haughney J, Hicks A, van der Klaauw AA, Kwok J, Lambe T, Libri V, Llewelyn MJ, McGregor AC, Minassian AM, Moore P, Mughal M, Mujadidi YF, Murira J, Osanlou O, Osanlou R, Owens DR, Pacurar M, Palfreeman A, Pan D, Rampling T, Regan K, Saich S, Salkeld J, Saralaya D, Sharma S, Sheridan R, Sturdy A, Thomson EC, Todd S, Twelves C, Read RC, Charlton S, Hallis B, Ramsay M, Andrews N, Nguyen-Van-Tam JS, Snape MD, Liu X, Faust SN, Riordan A, Ustianowski A, Rogers CA, Hughes S, Longshaw L, Stockport J, Hughes R, Grundy L, Tudor Jones L, Guha A, Snashall E, Eadsforth T, Reeder S, Storton K, Munusamy M, Tandy B, Egbo A, Cox S, Ahmed NN, Shenoy A, Bousfield R, Wixted D, Gutteridge H, Mansfield B, Herbert C, Holliday K, Calderwood J, Barker D, Brandon J, Tulloch H, Colquhoun S, Thorp H,

Radford H, Evans J, Baker H, Thorpe J, Batham S, Hailstone J, Phillips R, Kumar D, Westwell F, Makia F, Hopkins N, Barcella L, Mpelembue M, dabagh M, lang M, khan F, Adebambo O, Chita S, Corrah T, Whittington A, John L, Roche S, Wagstaff L, Farrier A, Bisnauthsing K, Serafimova T, Nanino E, Cooney E, Wilson-Goldsmith J, Nguyen H, Mazzella A, Jackson B, Aslam S, Bawa T, Broadhead S, Farooqi S, Piper J, Weighell R, Pickup L, Shamtally D, Domingo J, Kourampa E, Hale C, Gibney J, Stackpoole M, Rashid-Gardner Z, Lyon R, McDonnell C, Cole C, Stewart A, McMillan G, Savage M, Beckett H, Moorbey C, Desai A, Brown C, Naker K, Qureshi E, Trinhnam C, Sabine C, Moore S, Hurdover S, Justice E, Smith D, Plested E, Ferreira Da Silva C, White R, Robinson H, Cifuentes L, Morshead G, Drake-Brockman R, Kinch P, Kasanyinga M, Clutterbuck EA, Bibi S, Stuart AS, Shaw RH, Singh M, Champaneri T, Irwin M, Khan M, Kownacka A, Nabunjo M, Osuji C, Hladkiwskyj J, Galvin D, Patel G, Moulard J, Longhurst B, Moon M, Giddins B, Pereira Dias Alves C, Richmond L, Minnis C, Baryschpolec S, Elliott S, Fox L, Graham V, Baker N, Godwin K, Buttigieg K, Knight C, Brown P, Lall P, Shaik I, Chiplin E, Brunt E, Leung S, Allen L, Thomas S, Fraser S, Choi B, Gouriet J, Freedman A, Perkins J, Gowland A, Macdonald J, Seenan JP, Starinskij I, Seaton A, Peters E, Singh S, Gardside B, Bonnaud A, Davies C, Gordon E, Keenan S, Hall J, Wilkins S, Tasker S, James R, Seath I, Littlewood K, Newman J, Boubriak I, Suggitt D, Haydock H, Bennett S, Woodyatt W, Hughes K, Bell J, Coughlan T, van Welsenens D, Kamal M, Cooper C, Tunstall S, Ronan N, Cutts R, Dare T, Yim YTN, Whittlely S, Ricamara M, Hamal S, Adams K, Baker H, Driver K, Turner N, Rawlins T, Roy S, Merida-Morillas M, Sakagami Y, Andrews A, Goncalves cordeiro L, Stokes M, Ambihapathy W, Spencer J, Parungao N, Berry L, Cullinane J, Presland L, Ross-Russell A, Warren S, Baker J, Oliver A, Buadi A, Lee K, Haskell L, Romani R, Bentley I, Whitbred T, Fowler S, Gavin J, Magee A, Watson T, Nightingale K, Marius P, Summerton E, Locke E, Honey T, Lingwood A, de la Haye A, Elliott RS, Underwood K, King M, Davies-Dear S, Horsfall E, Chalwin O, Burton H, Edwards CJ, Welham B, Garrahy S, Hall F, Ladikou E, Mullan D, Hansen D, Campbell M, Dos Santos F, Habash-Bailey H, Lakeman N, Branney D, Vamplew L, Hogan A, Frankham J, Wiselka M, Vail D, Wenn V, Renals V, Ellis K, Lewis-Taylor J, Magan J, Hardy A, Appleby K. 2021. [Safety and immunogenicity of seven COVID-19 vaccines as a third dose \(booster\) following two doses of ChAdOx1 nCov-19 or BNT162b2 in the UK \(COV-BOOST\): a blinded, multicentre, randomised, controlled, phase 2 trial](#). The Lancet 398:2258–2276.

1657. 2021. Overview of EU/EEA country recommendations on COVID-19 vaccination with Vaxzevria, and a scoping review of evidence to guide decision-making. European Centre for Disease Prevention and Control. <https://www.ecdc.europa.eu/en/publications-data/overview-eueea-country-recommendations-covid-19-vaccination-vaxzevria-and-scoping>. Retrieved 5 December 2022.
1658. Duarte-Salles T, Prieto-Alhambra D. 2021. [Heterologous vaccine regimens against COVID-19](#). The Lancet 398:94–95.
1659. Moderna Announces Omicron-Containing Bivalent Booster Candidate mRNA-1273.214 Demonstrates Superior Antibody Response Against Omicron. <https://investors.modernatx.com/news/news->

[details/2022/Moderna-Announces-Omicron-Containing-Bivalent-Booster-Candidate-mRNA-1273.214-Demonstrates-Superior-Antibody-Response-Against-Omicron/default.aspx](#). Retrieved 5 December 2022.

1660. Choi A, Koch M, Wu K, Chu L, Ma L, Hill A, Nunna N, Huang W, Oestreicher J, Colpitts T, Bennett H, Legault H, Paila Y, Nestorova B, Ding B, Montefiori D, Pajon R, Miller JM, Leav B, Carfi A, McPhee R, Edwards DK. 2021. [Safety and immunogenicity of SARS-CoV-2 variant mRNA vaccine boosters in healthy adults: an interim analysis](#). Nat Med 27:2025–2031.
1661. Pajon R, Doria-Rose NA, Shen X, Schmidt SD, O'Dell S, McDanal C, Feng W, Tong J, Eaton A, Maglinao M, Tang H, Manning KE, Edara V-V, Lai L, Ellis M, Moore KM, Floyd K, Foster SL, Posavad CM, Atmar RL, Lyke KE, Zhou T, Wang L, Zhang Y, Gaudinski MR, Black WP, Gordon I, Guech M, Ledgerwood JE, Misasi JN, Widge A, Sullivan NJ, Roberts PC, Beigel JH, Korber B, Baden LR, El Sahly H, Chalkias S, Zhou H, Feng J, Girard B, Das R, Aunins A, Edwards DK, Suthar MS, Mascola JR, Montefiori DC. 2022. [SARS-CoV-2 Omicron Variant Neutralization after mRNA-1273 Booster Vaccination](#). N Engl J Med 386:1088–1091.
1662. Pfizer and BioNTech Announce Omicron-Adapted COVID-19 Vaccine Candidates Demonstrate High Immune Response Against Omicron | Pfizer. <https://www.pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-announce-omicron-adapted-covid-19>. Retrieved 5 December 2022.
1663. Moxon R, Reche PA, Rappuoli R. 2019. [Editorial: Reverse Vaccinology](#). Front Immunol 10.
1664. Kudchodkar SB, Choi H, Reuschel EL, Esquivel R, Jin-Ah Kwon J, Jeong M, Maslow JN, Reed CC, White S, Kim JJ, Kobinger GP, Tebas P, Weiner DB, Muthumani K. 2018. [Rapid response to an emerging infectious disease – Lessons learned from development of a synthetic DNA vaccine targeting Zika virus](#). Microbes and Infection 20:676–684.
1665. Olena A. 2020. Newer Vaccine Technologies Deployed to Develop COVID-19 Shot. The Scientist Magazine. <https://www.the-scientist.com/news-opinion/newer-vaccine-technologies-deployed-to-develop-covid-19-shot-67152>.
1666. Pulliam JRC, van Schalkwyk C, Govender N, Gottberg A von, Cohen C, Groome MJ, Dushoff J, Mlisana K, Moultrie H. 2021. [Increased risk of SARS-CoV-2 reinfection associated with emergence of Omicron in South Africa](#). Cold Spring Harbor Laboratory.
1667. 2022-07-15 12:20 | Archive of CDC Covid Pages. <https://public4.pagefreezer.com/browse/CDC%20Covid%20Pages/15-07-2022T12:20/https://www.cdc.gov/coronavirus/2019-ncov/science/science-briefs/scientific-brief-omicron-variant.html>. Retrieved 5 December 2022.
1668. Planas D, Bruel T, Grzelak L, Guivel-Benhassine F, Staropoli I, Porrot F, Planchais C, Buchrieser J, Rajah MM, Bishop E, Albert M, Donati F, Prot M, Behillil S, Enouf V, Maquart M, Smati-Lafarge M, Varon E, Schortgen

- F, Yahyaoui L, Gonzalez M, De Sèze J, Péré H, Veyer D, Sève A, Simon-Lorière E, Fafi-Kremer S, Stefic K, Mouquet H, Hocqueloux L, van der Werf S, Prazuck T, Schwartz O. 2021. [Sensitivity of infectious SARS-CoV-2 B.1.1.7 and B.1.351 variants to neutralizing antibodies](#). Nat Med 27:917–924.
1669. Wang P, Nair MS, Liu L, Iketani S, Luo Y, Guo Y, Wang M, Yu J, Zhang B, Kwong PD, Graham BS, Mascola JR, Chang JY, Yin MT, Sobieszczyk M, Kyratsous CA, Shapiro L, Sheng Z, Huang Y, Ho DD. 2021. [Antibody resistance of SARS-CoV-2 variants B.1.351 and B.1.1.7](#). Nature 593:130–135.
1670. Edara VV, Hudson WH, Xie X, Ahmed R, Suthar MS. 2021. [Neutralizing Antibodies Against SARS-CoV-2 Variants After Infection and Vaccination](#). JAMA 325:1896.
1671. Classification of Omicron (B.1.1.529): SARS-CoV-2 Variant of Concern. [https://www.who.int/news/item/26-11-2021-classification-of-omicron-\(b.1.1.529\)-sars-cov-2-variant-of-concern](https://www.who.int/news/item/26-11-2021-classification-of-omicron-(b.1.1.529)-sars-cov-2-variant-of-concern). Retrieved 5 December 2022.
1672. Kuhlmann C, Mayer CK, Claassen M, Maponga TG, Sutherland AD, Suliman T, Shaw M, Preiser W. 2021. Breakthrough Infections with SARS-CoV-2 Omicron Variant Despite Booster Dose of mRNA Vaccine. SSRN Journal <https://doi.org/10.2139/ssrn.3981711>.
1673. Self WH, Tenforde MW, Rhoads JP, Gaglani M, Ginde AA, Douin DJ, Olson SM, Talbot HK, Casey JD, Mohr NM, Zepeski A, McNeal T, Ghamande S, Gibbs KW, Files DC, Hager DN, Shehu A, Prekker ME, Erickson HL, Gong MN, Mohamed A, Henning DJ, Steingrub JS, Peltan ID, Brown SM, Martin ET, Monto AS, Khan A, Hough CL, Busse LW, ten Lohuis CC, Duggal A, Wilson JG, Gordon AJ, Qadir N, Chang SY, Mallow C, Rivas C, Babcock HM, Kwon JH, Exline MC, Halasa N, Chappell JD, Luring AS, Grijalva CG, Rice TW, Jones ID, Stubblefield WB, Baughman A, Womack KN, Lindsell CJ, Hart KW, Zhu Y, Mills L, Lester SN, Stumpf MM, Naioti EA, Kobayashi M, Verani JR, Thornburg NJ, Patel MM, Calhoun N, Murthy K, Herrick J, McKillop A, Hoffman E, Zayed M, Smith M, Seattle N, Ettlinger J, Priest E, Thomas J, Arroliga A, Beeram M, Kindle R, Kozikowski L-A, De Souza L, Ouellette S, Thornton-Thompson S, Mehkri O, Ashok K, Gole S, King A, Poynter B, Stanley N, Hendrickson A, Maruggi E, Scharber T, Jorgensen J, Bowers R, King J, Aston V, Armbruster B, Rothman RE, Nair R, Chen J-TT, Karow S, Robart E, Maldonado PN, Khan M, So P, Levitt J, Perez C, Visweswaran A, Roque J, Rivera A, Angeles L, Frankel T, Angeles L, Goff J, Huynh D, Howell M, Friedel J, Tozier M, Driver C, Carricato M, Foster A, Nassar P, Stout L, Sibenaller Z, Walter A, Mares J, Olson L, Clinansmith B, Rivas C, Gershengorn H, McSpadden E, Truscon R, Kanicldes A, Thomas L, Bielak R, Valvano WD, Fong R, Fitzsimmons WJ, Blair C, Valesano AL, Gilbert J, Crider CD, Steinbock KA, Paulson TC, Anderson LA, Kampe C, Johnson J, McHenry R, Blair M, Conway D, LaRose M, Landreth L, Hicks M, Parks L, Bongu J, McDonald D, Cass C, Seiler S, Park D, Hink T, Wallace M, Burnham C-A, Arter OG. 2021. [Comparative Effectiveness of Moderna, Pfizer-BioNTech, and Janssen \(Johnson & Johnson\) Vaccines in Preventing COVID-19 Hospitalizations Among Adults Without Immunocompromising Conditions — United States, March–August 2021](#). MMWR Morb Mortal Wkly Rep 70:1337–1343.

1674. Mueller B, Robbins R. 2021. [Where a Vast Global Vaccination Program Went Wrong](#). The New York Times.
1675. Sheridan C. 2021. [Innovators target vaccines for variants and shortages in global South](#). Nat Biotechnol 39:393–396.
1676. Nohynek H, Wilder-Smith A. 2022. [Does the World Still Need New Covid-19 Vaccines?](#) N Engl J Med 386:2140–2142.
1677. Bernal JL, Andrews N, Gower C, Stowe J, Robertson C, Tessier E, Simmons R, Cottrell S, Roberts R, O'Doherty M, Brown K, Cameron C, Stockton D, McMenamin J, Ramsay M. 2021. [Early effectiveness of COVID-19 vaccination with BNT162b2 mRNA vaccine and ChAdOx1 adenovirus vector vaccine on symptomatic disease, hospitalisations and mortality in older adults in England](#). Cold Spring Harbor Laboratory.
1678. Balakrishnan VS. 2020. [The arrival of Sputnik V](#). The Lancet Infectious Diseases 20:1128.
1679. Tumban E. 2020. [Lead SARS-CoV-2 Candidate Vaccines: Expectations from Phase III Trials and Recommendations Post-Vaccine Approval](#). Viruses 13:54.
1680. Mahase E. 2021. [Covid-19: Russian vaccine efficacy is 91.6%, show phase III trial results](#). BMJ n309.
1681. 2021. Russia cuts size of COVID-19 vaccine study, stops enrollment. AP NEWS. <https://apnews.com/article/coronavirus-pandemic-russia-163068f551e080939b982389b576320a>. Retrieved 5 December 2022.
1682. About Sputnik V. <https://sputnikvaccine.com/about-vaccine/>. Retrieved 5 December 2022.
1683. Pagotto V, Ferloni A, Soriano MM, Díaz M, Golde MB, González MI, Asprea V, Staneloni I, Vidal G, Silveira M, Zingoni P, Aliperti V, Michelangelo H, Figar S. 2021. [ACTIVE SURVEILLANCE OF THE SPUTNIK V VACCINE IN HEALTH WORKERS](#). Cold Spring Harbor Laboratory.
1684. 2021. [UPDATE 1-Russia's Sputnik V vaccine found safe in India mid-stage trial -Dr.Reddy's](#). Reuters.
1685. Fund (RDIF) TRDI. RDIF, The Gamaleya National Center, AstraZeneca and R-Pharm sign an agreement to cooperate on COVID-19 vaccine development. <https://www.prnewswire.com/ae/news-releases/rdif-the-gamaleya-national-center-astrazeneca-and-r-pharm-sign-an-agreement-to-cooperate-on-covid-19-vaccine-development-301196874.html>. Retrieved 5 December 2022.
1686. R-Pharm. 2022. [Open-label, Non-randomized, Non-comparative, Phase II Study in Adult Subjects to Assess Safety and Immunogenicity of Combination of AZD1222, a Non-replicating ChAdOx1 Vector Vaccine, and rAd26-S, a Recombinant Adenovirus Type 26 Component of Gam-COVID-Vac Vaccine, for COVID-19 Prevention](#). NCT04686773. Clinical trial registration. clinicaltrials.gov.

1687. The first registered COVID-19 vaccine. <https://sputnikvaccine.com/>. Retrieved 5 December 2022.
1688. Johnson & Johnson Initiates Pivotal Global Phase 3 Clinical Trial of Janssen's COVID-19 Vaccine Candidate | Johnson & Johnson. Content Lab US. <https://www.jnj.com/johnson-johnson-initiates-pivotal-global-phase-3-clinical-trial-of-janssens-covid-19-vaccine-candidate>. Retrieved 5 December 2022.
1689. He X, Chandrashekar A, Zahn R, Wegmann F, Yu J, Mercado NB, McMahan K, Martinot AJ, Piedra-Mora C, Beecy S, Ducat S, Chamanza R, Huber SR, van der Fits L, Borducchi EN, Lifton M, Liu J, Nampanya F, Patel S, Peter L, Tostanoski LH, Pessaint L, Van Ry A, Finneyfrock B, Velasco J, Teow E, Brown R, Cook A, Andersen H, Lewis MG, Schuitemaker H, Barouch DH. 2021. [Low-Dose Ad26.COV2.S Protection Against SARS-CoV-2 Challenge in Rhesus Macaques](#). Cold Spring Harbor Laboratory.
1690. Sadoff J, Gars ML, Shukarev G, Heerwegh D, Truyers C, de Groot AM, Stoop J, Tete S, Van Damme W, Leroux-Roels I, Berghmans P-J, Kimmel M, Van Damme P, de Hoon J, Smith W, Stephenson KE, Barouch DH, De Rosa SC, Cohen KW, McElrath MJ, Cormier E, Scheper G, Hendriks J, Struyf F, Douoguih M, Van Hoof J, Schuitemaker H. 2020. [Safety and immunogenicity of the Ad26.COV2.S COVID-19 vaccine candidate: interim results of a phase 1/2a, double-blind, randomized, placebo-controlled trial](#). Cold Spring Harbor Laboratory.
1691. Stephenson KE, Le Gars M, Sadoff J, de Groot AM, Heerwegh D, Truyers C, Atyeo C, Loos C, Chandrashekar A, McMahan K, Tostanoski LH, Yu J, Gebre MS, Jacob-Dolan C, Li Z, Patel S, Peter L, Liu J, Borducchi EN, Nkolola JP, Souza M, Tan CS, Zash R, Julg B, Nathavitharana RR, Shapiro RL, Azim AA, Alonso CD, Jaegle K, Ansel JL, Kanjilal DG, Guiney CJ, Bradshaw C, Tyler A, Makoni T, Yanosick KE, Seaman MS, Lauffenburger DA, Alter G, Struyf F, Douoguih M, Van Hoof J, Schuitemaker H, Barouch DH. 2021. [Immunogenicity of the Ad26.COV2.S Vaccine for COVID-19](#). JAMA 325:1535.
1692. McMahan K, Yu J, Mercado NB, Loos C, Tostanoski LH, Chandrashekar A, Liu J, Peter L, Atyeo C, Zhu A, Bondzie EA, Dagotto G, Gebre MS, Jacob-Dolan C, Li Z, Nampanya F, Patel S, Pessaint L, Van Ry A, Blade K, Yalley-Ogunro J, Cabus M, Brown R, Cook A, Teow E, Andersen H, Lewis MG, Lauffenburger DA, Alter G, Barouch DH. 2020. [Correlates of protection against SARS-CoV-2 in rhesus macaques](#). Nature 590:630–634.
1693. Janssen Vaccines & Prevention B.V. 2022. [A Randomized, Double-blind, Placebo-controlled Phase 3 Study to Assess the Efficacy and Safety of Ad26.COV2.S for the Prevention of SARS-CoV-2-mediated COVID-19 in Adults Aged 18 Years and Older](#). NCT04505722. Clinical trial registration. clinicaltrials.gov.
1694. Johnson & Johnson Prepares to Resume Phase 3 ENSEMBLE Trial of its Janssen COVID-19 Vaccine Candidate in the U.S. | Johnson & Johnson. Content Lab US. <https://www.jnj.com/our-company/johnson-johnson->

[prepares-to-resume-phase-3-ensemble-trial-of-its-janssen-covid-19-vaccine-candidate-in-the-us](#). Retrieved 5 December 2022.

1695. Johnson & Johnson Initiates Second Global Phase 3 Clinical Trial of its Janssen COVID-19 Vaccine Candidate | Johnson & Johnson. Content Lab US. <https://www.jnj.com/johnson-johnson-initiates-second-global-phase-3-clinical-trial-of-its-janssen-covid-19-vaccine-candidate>. Retrieved 5 December 2022.
1696. Commissioner O of the. 2022. Coronavirus (COVID-19) Update: FDA Authorizes Moderna and Pfizer-BioNTech COVID-19 Vaccines for Children Down to 6 Months of Age. FDA. <https://www.fda.gov/news-events/press-announcements/coronavirus-covid-19-update-fda-authorizes-moderna-and-pfizer-biontech-covid-19-vaccines-children>. Retrieved 5 December 2022.
1697. Moderna's COVID-19 Vaccine Candidate Meets its Primary Efficacy Endpoint in the First Interim Analysis of the Phase 3 COVE Study | Moderna, Inc. <https://investors.modernatx.com/news-releases/news-release-details/modernas-covid-19-vaccine-candidate-meets-its-primary-efficacy/>. Retrieved 8 February 2021.
1698. 2021. Vaccines and Related Biological Products Advisory Committee December 17, 2020 Meeting Announcement - 12/17/2020 - 12/17/2020. FDA. <https://www.fda.gov/advisory-committees/advisory-committee-calendar/vaccines-and-related-biological-products-advisory-committee-december-17-2020-meeting-announcement>. Retrieved 8 February 2021.
1699. FDA/CBER. 2020. Vaccines and Related Biological Products Advisory Committee December 17, 2020 Meeting Briefing Document - FDA . <https://www.fda.gov/media/144434/download>.
1700. Moderna Has Completed Case Accrual for First Planned Interim Analysis of its mRNA Vaccine Against COVID-19 (mRNA-1273) | Moderna, Inc. <https://investors.modernatx.com/news-releases/news-release-details/moderna-has-completed-case-accrual-first-planned-interim/>. Retrieved 8 February 2021.
1701. CBER. 2018. Guidance for Industry: Toxicity Grading Scale for Healthy Adult and Adolescent Volunteers Enrolled in Preventive Vaccine Clinical Trials . <https://www.fda.gov/media/73679/download>.
1702. Health Canada Authorizes Moderna COVID-19 Vaccine in Canada | Moderna, Inc. <https://investors.modernatx.com/news-releases/news-release-details/health-canada-authorizes-moderna-covid-19-vaccine-canada/>. Retrieved 8 February 2021.
1703. GLANVILLE D. 2021. EMA recommends COVID-19 Vaccine Moderna for authorisation in the EU. European Medicines Agency. Text. <https://www.ema.europa.eu/en/news/ema-recommends-covid-19-vaccine-moderna-authorisation-eu>. Retrieved 8 February 2021.
1704. Pfizer and BioNTech Announce Vaccine Candidate Against COVID-19 Achieved Success in First Interim Analysis from Phase 3 Study | Pfizer. <https://www.pfizer.com/news/press-release/press-release->

[detail/pfizer-and-biontech-announce-vaccine-candidate-against.](#)

Retrieved 8 February 2021.

1705. Mulligan MJ, Lyke KE, Kitchin N, Absalon J, Gurtman A, Lockhart S, Neuzil K, Raabe V, Bailey R, Swanson KA, Li P, Koury K, Kalina W, Cooper D, Fontes-Garfias C, Shi P-Y, Türeci Ö, Tompkins KR, Walsh EE, Frenck R, Falsey AR, Dormitzer PR, Gruber WC, Şahin U, Jansen KU. 2020. [Phase I/II study of COVID-19 RNA vaccine BNT162b1 in adults.](#) Nature 586:589–593.
1706. Sahin U, Muik A, Derhovanessian E, Vogler I, Kranz LM, Vormehr M, Baum A, Pascal K, Quandt J, Maurus D, Brachtendorf S, Lörks V, Sikorski J, Hilker R, Becker D, Eller A-K, Grützner J, Boesler C, Rosenbaum C, Kühnle M-C, Luxemburger U, Kemmer-Brück A, Langer D, Bexon M, Bolte S, Karikó K, Palanche T, Fischer B, Schultz A, Shi P-Y, Fontes-Garfias C, Perez JL, Swanson KA, Loschko J, Scully IL, Cutler M, Kalina W, Kyratsous CA, Cooper D, Dormitzer PR, Jansen KU, Türeci Ö. 2020. [COVID-19 vaccine BNT162b1 elicits human antibody and TH1 T cell responses.](#) Nature 586:594–599.
1707. Coronavirus COVID-19 Vaccine Update: Latest Developments | Pfizer. <https://www.pfizer.com/science/coronavirus/vaccine>. Retrieved 8 February 2021.
1708. Pfizer and BioNTech Conclude Phase 3 Study of COVID-19 Vaccine Candidate, Meeting All Primary Efficacy Endpoints | Pfizer. <https://www.pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-conclude-phase-3-study-covid-19-vaccine>. Retrieved 8 February 2021.
1709. Mahase E. 2020. [Covid-19: UK approves Pfizer and BioNTech vaccine with rollout due to start next week.](#) BMJ m4714.
1710. 2020. [Covid-19 vaccine: First person receives Pfizer jab in UK.](#) BBC News.
1711. Commissioner O of the. 2021. FDA Approves First COVID-19 Vaccine. FDA. <https://www.fda.gov/news-events/press-announcements/fda-approves-first-covid-19-vaccine>. Retrieved 5 December 2022.
1712. Aggarwal A, Stella AO, Walker G, Akerman A, Milogiannakis V, Brilot F, Amatayakul-Chantler S, Roth N, Coppola G, Schofield P, Jackson J, Henry JY, Mazigi O, Langley D, Lu Y, Forster C, McAllery S, Mathivanan V, Fichter C, Hoppe AC, Munier ML, Jack H-M, Cromer D, Darley D, Matthews G, Christ D, Khoury D, Davenport M, Rawlinson W, Kelleher AD, Turville S. 2021. [SARS-CoV-2 Omicron: evasion of potent humoral responses and resistance to clinical immunotherapeutics relative to viral variants of concern.](#) Cold Spring Harbor Laboratory.
1713. Khoury DS, Steain M, Triccas JA, Sigal A, Davenport MP, Cromer D. 2021. [A meta-analysis of Early Results to predict Vaccine efficacy against Omicron.](#) Cold Spring Harbor Laboratory.
1714. Ferdinands JM, Rao S, Dixon BE, Mitchell PK, DeSilva MB, Irving SA, Lewis N, Natarajan K, Stenehjem E, Grannis SJ, Han J, McEvoy C, Ong TC, Naleway AL, Reese SE, Embi PJ, Dascomb K, Klein NP, Griggs EP,

Konatham D, Kharbanda AB, Yang D-H, Fadel WF, Grisel N, Goddard K, Patel P, Liao I-C, Birch R, Valvi NR, Reynolds S, Arndorfer J, Zerbo O, Dickerson M, Murthy K, Williams J, Bozio CH, Blanton L, Verani JR, Schrag SJ, Dalton AF, Wondimu MH, Link-Gelles R, Azziz-Baumgartner E, Barron MA, Gaglani M, Thompson MG, Fireman B. 2022. [Waning 2-Dose and 3-Dose Effectiveness of mRNA Vaccines Against COVID-19–Associated Emergency Department and Urgent Care Encounters and Hospitalizations Among Adults During Periods of Delta and Omicron Variant Predominance — VISION Network, 10 States, August 2021–January 2022](#). MMWR Morb Mortal Wkly Rep 71:255–263.

1715. Zeng C, Evans JP, Chakravarthy K, Qu P, Reisinger S, Song N-J, Rubinstein MP, Shields PG, Li Z, Liu S-L. 2022. [COVID-19 mRNA booster vaccines elicit strong protection against SARS-CoV-2 Omicron variant in patients with cancer](#). Cancer Cell 40:117–119.
1716. Gilbert PB, Montefiori DC, McDermott AB, Fong Y, Benkeser D, Deng W, Zhou H, Houchens CR, Martins K, Jayashankar L, Castellino F, Flach B, Lin BC, O'Connell S, McDanal C, Eaton A, Sarzotti-Kelsoe M, Lu Y, Yu C, Borate B, van der Laan LWP, Hejazi NS, Huynh C, Miller J, El Sahly HM, Baden LR, Baron M, De La Cruz L, Gay C, Kalams S, Kelley CF, Andrasik MP, Kublin JG, Corey L, Neuzil KM, Carpp LN, Pajon R, Follmann D, Donis RO, Koup RA. 2022. [Immune correlates analysis of the mRNA-1273 COVID-19 vaccine efficacy clinical trial](#). Science 375:43–50.
1717. Goldblatt D, Fiore-Gartland A, Johnson M, Hunt A, Bengt C, Zavadska D, Snipe HD, Brown JS, Workman L, Zar HJ, Montefiori D, Shen X, Dull P, Plotkin S, Siber G, Ambrosino D. 2022. [Towards a population-based threshold of protection for COVID-19 vaccines](#). Vaccine 40:306–315.
1718. Earle KA, Ambrosino DM, Fiore-Gartland A, Goldblatt D, Gilbert PB, Siber GR, Dull P, Plotkin SA. 2021. [Evidence for antibody as a protective correlate for COVID-19 vaccines](#). Vaccine 39:4423–4428.
1719. Castillo-Olivares J, Wells DA, Ferrari M, Chan A, Smith P, Nadesalingam A, Paloniemi M, Carnell G, Ohlendorf L, Cantoni D, Mayora-Neto M, Palmer P, Tonks P, Temperton N, Wagner R, Neckermann P, Peterhoff D, Doffinger R, Kempster S, Otter A, Semper A, Brooks T, Page M, Albecka A, James LC, Briggs J, Schwaeble W, Baxendale H, Heeney J. 2021. [Towards Internationally standardised humoral Immune Correlates of Protection from SARS-CoV-2 infection and COVID-19 disease](#). Cold Spring Harbor Laboratory.
1720. Legros V, Denolly S, Vogrig M, Boson B, Siret E, Rigail J, Pillet S, Grattard F, Gonzalo S, Verhoeven P, Allatif O, Berthelot P, Pélissier C, Thierry G, Botelho-Nevers E, Millet G, Morel J, Paul S, Walzer T, Cosset F-L, Bourlet T, Pozzetto B. 2021. [A longitudinal study of SARS-CoV-2-infected patients reveals a high correlation between neutralizing antibodies and COVID-19 severity](#). Cell Mol Immunol 18:318–327.
1721. 2022. Vaccine development and approval in Canada. <https://www.canada.ca/en/health-canada/services/drugs-health-products/covid19-industry/drugs-vaccines-treatments/vaccines/development-approval-infographic.html>.

1722. 2022. Vaccines and treatments for COVID-19: Safety after authorization. Government of Canada.
<https://www.canada.ca/en/public-health/services/diseases/2019-novel-coronavirus-infection/prevention-risks/covid-19-vaccine-treatment/safety-after-authorization.html>. Retrieved 5 December 2022.
1723. . <https://www.canada.ca/en/health-canada/services/drugs-health-products/covid19-industry/drugs-vaccines-treatments/authorization/applications.html>.
1724. News · JPT·C. An earlier end date for vaccination campaign is 'possible', Trudeau says | CBC News. CBC.
<https://www.cbc.ca/news/politics/trudeau-possible-vaccination-campaign-ends-sooner-1.5934994>. Retrieved 5 December 2022.
1725. Dooling K, McClung N, Chamberland M, Marin, M, Wallace M, Bell BP, Lee GM, Talbot HK, Romero JR, Oliver SE. 2020. [The Advisory Committee on Immunization Practices' Interim Recommendation for Allocating Initial Supplies of COVID-19 Vaccine — United States, 2020](#). MMWR Morb Mortal Wkly Rep 69:1857–1859.
1726. Oliver Sara E, Gargano Julia W, Marin M, Wallace M, Curran KG, Chamberland M, McClung N, Campos-Outcalt D, Morgan Rebecca L, Mbaeyi S, Romero José R, Talbot H Keipp, Lee Grace M, Bell Beth P, Dooling K. 2020. [The Advisory Committee on Immunization Practices' Interim Recommendation for Use of Pfizer-BioNTech COVID-19 Vaccine — United States, December 2020](#). MMWR Morb Mortal Wkly Rep 69:1922–1924.
1727. News ABC. US administers 1st doses of Pfizer coronavirus vaccine. ABC News. <https://abcnews.go.com/US/us-administer-1st-doses-pfizer-coronavirus-vaccine/story?id=74703018>. Retrieved 5 December 2022.
1728. Oliver Sara E, Gargano Julia W, Marin M, Wallace M, Curran KG, Chamberland M, McClung N, Campos-Outcalt D, Morgan Rebecca L, Mbaeyi S, Romero José R, Talbot H Keipp, Lee Grace M, Bell Beth P, Dooling K. 2021. [The Advisory Committee on Immunization Practices' Interim Recommendation for Use of Moderna COVID-19 Vaccine — United States, December 2020](#). MMWR Morb Mortal Wkly Rep 69:1653–1656.
1729. Dooling K, Marin M, Wallace M, McClung N, Chamberland M, Lee GM, Talbot HK, Romero JR, Bell BP, Oliver SE. 2021. [The Advisory Committee on Immunization Practices' Updated Interim Recommendation for Allocation of COVID-19 Vaccine — United States, December 2020](#). MMWR Morb Mortal Wkly Rep 69:1657–1660.
1730. Almasry MH Holly Yan, Steve. 2020. The Moderna vaccine is now in some Americans' arms as Covid-19 cases in the US pass 18 million. CNN. <https://www.cnn.com/2020/12/21/health/us-coronavirus-monday/index.html>. Retrieved 5 December 2022.
1731. Commissioner O of the. 2022. [Janssen COVID-19 Vaccine](#). FDA.

1732. Oliver SE, Gargano JW, Scobie H, Wallace M, Hadler SC, Leung J, Blain AE, McClung N, Campos-Outcalt D, Morgan RL, Mbaeyi S, MacNeil J, Romero JR, Talbot HK, Lee GM, Bell BP, Dooling K. 2021. [The Advisory Committee on Immunization Practices' Interim Recommendation for Use of Janssen COVID-19 Vaccine — United States, February 2021](#). MMWR Morb Mortal Wkly Rep 70:329–332.
1733. WHO adds Janssen vaccine to list of safe and effective emergency tools against COVID-19. <https://www.who.int/news/item/12-03-2021-who-adds-janssen-vaccine-to-list-of-safe-and-effective-emergency-tools-against-covid-19>. Retrieved 5 December 2022.
1734. Affairs (ASPA) AS for P. 2020. COVID-19 Vaccines. HHSgov. Text. <https://www.hhs.gov/coronavirus/covid-19-vaccines/index.html>. Retrieved 5 December 2022.
1735. CDC. 2020. COVID Data Tracker. Centers for Disease Control and Prevention. <https://covid.cdc.gov/covid-data-tracker>. Retrieved 5 December 2022.
1736. Harwood KL Jeff Zeleny, John. 2021. Biden now says US will have enough vaccine for every adult by the end of May | CNN Politics. CNN. <https://www.cnn.com/2021/03/02/politics/biden-merck-johnson--johnson-vaccine/index.html>. Retrieved 5 December 2022.
1737. 2021. [Covid-19: Was US vaccine rollout a 'dismal failure' under Trump?](#) BBC News.
1738. Camacho AE, Glicksman RL. 2021. [Structured to Fail: Lessons from the Trump Administration's Faulty Pandemic Planning and Response](#). 3770368. SSRN Scholarly Paper. Social Science Research Network, Rochester, NY.
1739. Arthur R. 2021. South Africa starts administering Janssen COVID-19 vaccine to health workers. BioPharma-Reporter. <https://www.biopharma-reporter.com/Article/2021/02/18/South-Africa-starts-administering-Janssen-COVID-19-vaccine-to-health-workers>.
1740. 2021. EMA receives application for conditional marketing authorisation of COVID-19 Vaccine Janssen. <https://www.ema.europa.eu/en/news/ema-receives-application-conditional-marketing-authorisation-covid-19-vaccine-janssen>.
1741. Rowland C, McGinley L. 2021. [Merck will help make Johnson & Johnson coronavirus vaccine as rivals team up to help Biden accelerate shots](#). Washington Post.
1742. Ledford H, Cyranoski D, Van Noorden R. 2020. [The UK has approved a COVID vaccine — here's what scientists now want to know](#). Nature 588:205–206.
1743. 2020. EMA recommends first COVID-19 vaccine for authorisation in the EU. <https://www.ema.europa.eu/en/news/ema-recommends-first-covid-19-vaccine-authorisation-eu>.

1744. Regulatory approval of COVID-19 Vaccine AstraZeneca. GOVUK. <https://www.gov.uk/government/publications/regulatory-approval-of-covid-19-vaccine-astrazeneca>. Retrieved 5 December 2022.
1745. 2021. [Covid: Brian Pinker, 82, first to get Oxford-AstraZeneca vaccine](#). BBC News.
1746. 2022. Spikevax (previously COVID-19 Vaccine Moderna). Europeans Medicine Agency. <https://www.ema.europa.eu/en/medicines/human/EPAR/spikevax>. Retrieved 13 January 2023.
1747. Regulatory approval of Spikevax (formerly COVID-19 Vaccine Moderna). GOVUK. <https://www.gov.uk/government/publications/regulatory-approval-of-covid-19-vaccine-moderna>. Retrieved 5 December 2022.
1748. Arkhipov I, Kravchenko S. 2020. Putin Orders Start of Mass Covid-19 Shots Hours After U.K. News. Bloomberg. <https://www.bloomberg.com/news/articles/2020-12-02/within-hours-of-u-k-putin-orders-start-of-mass-covid-19-shots>.
1749. Rodgers J. Facing Record Covid-19 Case Rise, Russia Rolls Out Sputnik V Vaccine. Forbes. <https://www.forbes.com/sites/jamesrodgerseurope/2020/12/05/facing-record-covid-19-case-rise-russia-rolls-out-sputnik-v-vaccine/>. Retrieved 5 December 2022.
1750. Ellyatt H. Russia's coronavirus vaccine is alluring for Eastern Europe, creating a headache for the EU. CNBC. <https://www.cnbc.com/2021/03/02/russias-sputnik-vaccine-is-luring-eastern-europe-worrying-the-eu.html>. Retrieved 5 December 2022.
1751. 2021. Clarification on Sputnik V vaccine in the EU approval process. <https://www.ema.europa.eu/en/news/clarification-sputnik-v-vaccine-eu-approval-process>.
1752. Countries are lining up for Russia's once-scorned Sputnik vaccine after strong efficacy results. Fortune. <https://fortune.com/2021/02/08/international-sputnik-russia-demand/>. Retrieved 5 December 2022.
1753. RDIF announces delivery of the first batch of Sputnik V vaccine to Venezuela for clinical trials. <https://sputnikvaccine.com/newsroom/pressreleases/rdif-announces-delivery-of-the-first-batch-of-sputnik-v-vaccine-to-venezuela-for-clinical-trials/>. Retrieved 5 December 2022.
1754. Heath R. Unable to get U.S. vaccines, world turns to Russia and China. POLITICO. <https://www.politico.com/news/2021/02/25/global-vaccine-public-relations-war-471665>. Retrieved 5 December 2022.
1755. 2021. Germany moves to bring Russian vaccine into EU orbit. France 24. <https://www.france24.com/en/live-news/20210203-germany-moves-to-bring-russian-vaccine-into-eu-orbit>. Retrieved 5 December 2022.

1756. 2021. [Russia approves its third COVID-19 vaccine, CoviVac](#). Reuters.
1757. Zabetakis I, Lordan R, Norton C, Tsoupras A. 2020. [COVID-19: The Inflammation Link and the Role of Nutrition in Potential Mitigation](#). *Nutrients* 12:1466.
1758. James PT, Ali Z, Armitage AE, Bonell A, Cerami C, Drakesmith H, Jobe M, Jones KS, Liew Z, Moore SE, Morales-Berstein F, Nabwera HM, Nadjm B, Pasricha S-R, Scheelbeek P, Silver MJ, Teh MR, Prentice AM. 2020. Could nutrition modulate COVID-19 susceptibility and severity of disease? A systematic review. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.10.19.20214395>.
1759. Silverio R, Gonçalves DC, Andrade MF, Seelaender M. 2020. [Coronavirus Disease 2019 \(COVID-19\) and Nutritional Status: The Missing Link?](#) *Advances in Nutrition* nmaa125.
1760. Im JH, Je YS, Baek J, Chung M-H, Kwon HY, Lee J-S. 2020. [Nutritional status of patients with COVID-19](#). *International Journal of Infectious Diseases* 100:390–393.
1761. Calder P, Carr A, Gombart A, Eggersdorfer M. 2020. [Optimal Nutritional Status for a Well-Functioning Immune System Is an Important Factor to Protect against Viral Infections](#). *Nutrients* 12:1181.
1762. Peak dietary supplement sales leveling off during COVID-19 pandemic, but growth still remains strong over last year, market researchers report during webcast. Nutritional Outlook. <https://www.nutritionaloutlook.com/view/peak-dietary-supplement-sales-leveling-during-covid-19-pandemic-growth-still-remains-strong>. Retrieved 8 February 2021.
1763. Zhao A, Li Z, Ke Y, Huo S, Ma Y, Zhang Y, Zhang J, Ren Z. 2020. [Dietary Diversity among Chinese Residents during the COVID-19 Outbreak and Its Associated Factors](#). *Nutrients* 12:1699.
1764. nutraingredients-asia.com. Lockdown impact: Grocery stores bolstered NZ supplements sales as pharmacies slumped. nutraingredients-asia.com. <https://www.nutraingredients-asia.com/Article/2020/07/06/Lockdown-impact-Grocery-stores-bolstered-NZ-supplements-sales-as-pharmacies-slumped>. Retrieved 8 February 2021.
1765. COVID-19 temporarily bolsters European interest in supplements. nutritioninsight.com/. <https://ni.cnsmedia.com/a/EHHJsDOG2oc=>. Retrieved 8 February 2021.
1766. nutraingredients.com. India's immune health surge: Nation leads APAC in number of new product launches – new data. nutraingredients.com. <https://www.nutraingredients.com/Article/2020/07/21/India-s-immune-health-surge-Nation-leads-APAC-in-number-of-new-product-launches-new-data>. Retrieved 8 February 2021.
1767. Ayseli YI, Aytekin N, Buyukkayhan D, Aslan I, Ayseli MT. 2020. [Food policy, nutrition and nutraceuticals in the prevention and management](#)

[of COVID-19: Advice for healthcare professionals](#). Trends in Food Science & Technology 105:186–199.

1768. 5 Food and Beverage Trends in Europe During COVID-19. <https://kerry.com/insights/kerrydigest/2020/5-food-and-beverage-trends-in-europe-during-covid-19>. Retrieved 8 February 2021.
1769. McClements DJ, Decker EA, Park Y, Weiss J. 2009. [Structural Design Principles for Delivery of Bioactive Components in Nutraceuticals and Functional Foods](#). Critical Reviews in Food Science and Nutrition 49:577–606.
1770. Moss JWE, Ramji DP. 2016. [Nutraceutical therapies for atherosclerosis](#). Nature Reviews Cardiology 13:513–532.
1771. Kalra EK. 2015. [Nutraceutical-definition and introduction](#). AAPS PharmSci 5:27–28.
1772. Dietary Supplement Health and Education Act of 1994. National Institutes of Health Office of Dietary Supplements. https://ods.od.nih.gov/About/DSHEA_Wording.aspx. Retrieved 25 October 1994.
1773. Commissioner O of the. 2018. Food and Drug Administration Modernization Act (FDAMA) of 1997. FDA. <https://www.fda.gov/regulatory-information/selected-amendments-fdc-act/food-and-drug-administration-modernization-act-fdama-1997>. Retrieved 8 February 2021.
1774. Santini A, Novellino E. 2018. [Nutraceuticals - shedding light on the grey area between pharmaceuticals and food](#). Expert Review of Clinical Pharmacology 11:545–547.
1775. 2002. [Directive 2002/46/EC of the European Parliament and of the Council of 10 June 2002 on the approximation of the laws of the Member States relating to food supplements \(Text with EEA relevance\)](#)OJ L.
1776. 2004. [Directive 2004/27/EC of the European Parliament and of the Council of 31 March 2004 amending Directive 2001/83/EC on the Community code relating to medicinal products for human use \(Text with EEA relevance\)](#)OJ L.
1777. EU Register of nutrition and health claims made on foods (v.3.5). https://ec.europa.eu/food/safety/labelling_nutrition/claims/register/public/. Retrieved 8 February 2021.
1778. Santini A, Cammarata SM, Capone G, Ianaro A, Tenore GC, Pani L, Novellino E. 2018. [Nutraceuticals: opening the debate for a regulatory framework](#). British Journal of Clinical Pharmacology 84:659–672.
1779. Bröring S, Khedkar S, Ciliberti S. 2016. [Reviewing the Nutrition and Health Claims Regulation \(EC\) No. 1924/2006: What do we know about its challenges and potential impact on innovation?](#) International Journal of Food Sciences and Nutrition 68:1–9.

1780. Dwyer J, Coates P, Smith M. 2018. [Dietary Supplements: Regulatory Challenges and Research Resources](#). *Nutrients* 10:41.
1781. Research C for DE and. 2020. Noetic Nutraceuticals - 607572 - 05/15/2020. Center for Drug Evaluation and Research. <https://www.fda.gov/inspections-compliance-enforcement-and-criminal-investigations/warning-letters/noetic-nutraceuticals-607572-05152020>. Retrieved 8 February 2021.
1782. Regulations.gov. <https://beta.regulations.gov/document/FDA-2020-S-0023-0068>. Retrieved 8 February 2021.
1783. Research C for DE and. 2020. Spartan Enterprises Inc. dba Watershed Wellness Center - 610876 - 10/30/2020. Center for Drug Evaluation and Research. <https://www.fda.gov/inspections-compliance-enforcement-and-criminal-investigations/warning-letters/spartan-enterprises-inc-dba-watershed-wellness-center-610876-10302020>. Retrieved 8 February 2021.
1784. 2020. FTC Sues California Marketer of \$23,000 COVID-19 “Treatment” Plan. Federal Trade Commission. <https://www.ftc.gov/news-events/press-releases/2020/07/ftc-sues-california-marketer-23000-covid-19-treatment-plan>. Retrieved 8 February 2021.
1785. . <https://cen.acs.org/biological-chemistry/natural-products/oleandrin-compound-touted-possible-COVID/98/web/2020/08>. Retrieved 8 February 2021.
1786. Zumla A, Hui DS, Azhar EI, Memish ZA, Maeurer M. 2020. [Reducing mortality from 2019-nCoV: host-directed therapies should be an option](#). *The Lancet* 395:e35–e36.
1787. Infusino F, Marazzato M, Mancone M, Fedele F, Mastroianni CM, Severino P, Ceccarelli G, Santinelli L, Cavarretta E, Marullo AGM, Miraldi F, Carnevale R, Nocella C, Biondi-Zoccai G, Pagnini C, Schiavon S, Pugliese F, Frati G, d’Ettorre G. 2020. [Diet Supplementation, Probiotics, and Nutraceuticals in SARS-CoV-2 Infection: A Scoping Review](#). *Nutrients* 12:1718.
1788. Zhang L, Liu Y. 2020. [Potential interventions for novel coronavirus in China: A systematic review](#). *Journal of Medical Virology* 92:479–490.
1789. McCarty MF, DiNicolantonio JJ. 2020. [Nutraceuticals have potential for boosting the type 1 interferon response to RNA viruses including influenza and coronavirus](#). *Progress in Cardiovascular Diseases* 63:383–385.
1790. Lordan R, Redfern S, Tsoupras A, Zabetakis I. 2020. [Inflammation and cardiovascular disease: are marine phospholipids the answer?](#) *Food & Function* 11:2861–2885.
1791. Szabó Z, Marosvölgyi T, Szabó É, Bai P, Figler M, Verzár Z. 2020. [The Potential Beneficial Effect of EPA and DHA Supplementation Managing Cytokine Storm in Coronavirus Disease](#). *Frontiers in Physiology* 11:752.

1792. Saha S, Murray P. 2018. [Exploitation of Microalgae Species for Nutraceutical Purposes: Cultivation Aspects](#). Fermentation 4:46.
1793. Ratha SK, Renuka N, Rawat I, Bux F. 2021. [Prospective options of algae-derived nutraceuticals as supplements to combat COVID-19 and human coronavirus diseases](#). Nutrition 83:111089.
1794. Schmidt EB, Møller JM, Svaneborg N, Dyerberg J. 2012. [Safety Aspects of Fish Oils](#). Drug Investigation 7:215–220.
1795. Update on Seafood Consumption During Pregnancy. [https://www.acog.org/en/Clinical/Clinical Guidance/Practice Advisory/Articles/2017/01/Update on Seafood Consumption During Pregnancy](https://www.acog.org/en/Clinical/Clinical%20Guidance/Practice%20Advisory/Articles/2017/01/Update%20on%20Seafood%20Consumption%20During%20Pregnancy). Retrieved 8 February 2021.
1796. Greenberg JA, Bell SJ, Ausdal WV. 2008. [Omega-3 Fatty Acid supplementation during pregnancy](#). Rev Obstet Gynecol 1:162–9.
1797. Calder PC. 2013. [Omega-3 polyunsaturated fatty acids and inflammatory processes: nutrition or pharmacology?](#) British Journal of Clinical Pharmacology 75:645–662.
1798. Calder PC. 2015. [Marine omega-3 fatty acids and inflammatory processes: Effects, mechanisms and clinical relevance](#). Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids 1851:469–484.
1799. Whelan J, Gowdy KM, Shaikh SR. 2016. [N-3 polyunsaturated fatty acids modulate B cell activity in pre-clinical models: Implications for the immune response to infections](#). European Journal of Pharmacology 785:10–17.
1800. Asher A, Tintle NL, Myers M, Lockshon L, Bacareza H, Harris WS. 2021. [Blood omega-3 fatty acids and death from COVID-19: A pilot study](#). Prostaglandins, Leukotrienes and Essential Fatty Acids 166:102250.
1801. Tian T, Zhao Y, Huang Q, Li J. 2016. [n-3 Polyunsaturated Fatty Acids Improve Inflammation via Inhibiting Sphingosine Kinase 1 in a Rat Model of Parenteral Nutrition and CLP-Induced Sepsis](#). Lipids 51:271–278.
1802. Das UN. 2019. [Polyunsaturated fatty acids and sepsis](#). Nutrition 65:39–43.
1803. Sabater J, Masclans J, Sacanell J, Chacon P, Sabin P, Planas M. 2011. [Effects of an omega-3 fatty acid-enriched lipid emulsion on eicosanoid synthesis in acute respiratory distress syndrome \(ARDS\): A prospective, randomized, double-blind, parallel group study](#). Nutrition & Metabolism 8:22.
1804. Dushianthan A, Cusack R, Burgess VA, Grocott MP, Calder P. 2020. [Immunonutrition for Adults With ARDS: Results From a Cochrane Systematic Review and Meta-Analysis](#). Respiratory Care 65:99–110.
1805. Chen H, Wang S, Zhao Y, Luo Y, Tong H, Su L. 2018. [Correlation analysis of omega-3 fatty acids and mortality of sepsis and sepsis-induced](#)

[ARDS in adults: data from previous randomized controlled trials.](#)
Nutrition Journal 17:57.

1806. Buckley Christopher D, Gilroy Derek W, Serhan Charles N. 2014. [Proresolving Lipid Mediators and Mechanisms in the Resolution of Acute Inflammation.](#) Immunity 40:315–327.
1807. Basil MC, Levy BD. 2015. [Specialized pro-resolving mediators: endogenous regulators of infection and inflammation.](#) Nature Reviews Immunology 16:51–67.
1808. Sandhaus S, Swick AG. 2020. Specialized mediators in infection and lung injury. BioFactors <https://doi.org/10.1002/biof.1691>.
1809. Serhan CN. 2014. [Pro-resolving lipid mediators are leads for resolution physiology.](#) Nature 510:92–101.
1810. Ramon S, Baker SF, Sahler JM, Kim N, Feldsott EA, Serhan CN, Martínez-Sobrido L, Topham DJ, Phipps RP. 2014. [The Specialized Proresolving Mediator 17-HDHA Enhances the Antibody-Mediated Immune Response against Influenza Virus: A New Class of Adjuvant?](#) The Journal of Immunology 193:6031–6040.
1811. Morita M, Kuba K, Ichikawa A, Nakayama M, Katahira J, Iwamoto R, Watanebe T, Sakabe S, Daidoji T, Nakamura S, Kadowaki A, Ohto T, Nakanishi H, Taguchi R, Nakaya T, Murakami M, Yoneda Y, Arai H, Kawaoka Y, Penninger Josef M, Arita M, Imai Y. 2013. [The Lipid Mediator Protectin D1 Inhibits Influenza Virus Replication and Improves Severe Influenza.](#) Cell 153:112–125.
1812. Panigrahy D, Gilligan MM, Huang S, Gartung A, Cortés-Puch I, Sime PJ, Phipps RP, Serhan CN, Hammock BD. 2020. [Inflammation resolution: a dual-pronged approach to averting cytokine storms in COVID-19?](#) Cancer and Metastasis Reviews 39:337–340.
1813. Regidor P-A, Santos FG, Rizo JM, Egea FM. 2020. [Pro resolving inflammatory effects of the lipid mediators of omega 3 fatty acids and its implication in SARS COVID-19.](#) Medical Hypotheses 145:110340.
1814. Pal A, Gowdy KM, Oestreich KJ, Beck M, Shaikh SR. 2020. [Obesity-Driven Deficiencies of Specialized Pro-resolving Mediators May Drive Adverse Outcomes During SARS-CoV-2 Infection.](#) Frontiers in Immunology 11:1997.
1815. Schwerbrock NMJ, Karlsson EA, Shi Q, Sheridan PA, Beck MA. 2009. [Fish Oil-Fed Mice Have Impaired Resistance to Influenza Infection.](#) The Journal of Nutrition 139:1588–1594.
1816. Husson M-O, Ley D, Portal C, Gottrand M, Hueso T, Desseyn J-L, Gottrand F. 2016. [Modulation of host defence against bacterial and viral infections by omega-3 polyunsaturated fatty acids.](#) Journal of Infection 73:523–535.
1817. Skarke C, Alamuddin N, Lawson JA, Li X, Ferguson JF, Reilly MP, FitzGerald GA. 2015. [Bioactive products formed in humans from fish oils.](#) Journal of Lipid Research 56:1808–1820.

1818. Bäck M. 2020. [Resolving Inflammatory Storm in COVID-19 Patients by Omega-3 Polyunsaturated Fatty Acids - A Single-blind, Randomized, Placebo-controlled Feasibility Study](#). NCT04647604. Clinical trial registration. clinicaltrials.gov.
1819. Arnardottir H, Pawelzik S-C, Öhlund Wistbacka U, Artiach G, Hofmann R, Reinholdsson I, Braunschweig F, Tornvall P, Religa D, Bäck M. 2021. [Stimulating the Resolution of Inflammation Through Omega-3 Polyunsaturated Fatty Acids in COVID-19: Rationale for the COVID-Omega-F Trial](#). Frontiers in Physiology 11:624657.
1820. Connors JM, Levy JH. 2020. [COVID-19 and its implications for thrombosis and anticoagulation](#). Blood 135:2033–2040.
1821. Bikdeli B, Madhavan MV, Jimenez D, Chuich T, Dreyfus I, Driggin E, Nigoghossian CD, Ageno W, Madjid M, Guo Y, Tang LV, Hu Y, Giri J, Cushman M, Quéré I, Dimakakos EP, Gibson CM, Lippi G, Favaloro EJ, Fareed J, Caprini JA, Tafur AJ, Burton JR, Francese DP, Wang EY, Falanga A, McLintock C, Hunt BJ, Spyropoulos AC, Barnes GD, Eikelboom JW, Weinberg I, Schulman S, Carrier M, Piazza G, Beckman JA, Steg PG, Stone GW, Rosenkranz S, Goldhaber SZ, Parikh SA, Monreal M, Krumholz HM, Konstantinides SV, Weitz JI, Lip GYH. 2020. [COVID-19 and Thrombotic or Thromboembolic Disease: Implications for Prevention, Antithrombotic Therapy, and Follow-Up](#). Journal of the American College of Cardiology 75:2950–2973.
1822. Tsoupras A, Lordan R, Zabetakis I. 2020. [Thrombosis and COVID-19: The Potential Role of Nutrition](#). Frontiers in Nutrition 7:583080.
1823. Adili R, Hawley M, Holinstat M. 2018. [Regulation of platelet function and thrombosis by omega-3 and omega-6 polyunsaturated fatty acids](#). Prostaglandins & Other Lipid Mediators 139:10–18.
1824. Lordan R, Tsoupras A, Zabetakis I. 2020. [Platelet activation and prothrombotic mediators at the nexus of inflammation and atherosclerosis: Potential role of antiplatelet agents](#). Blood Reviews 100694.
1825. An Investigation on the Effects of Icosapent Ethyl (Vascepa™) on Inflammatory Biomarkers in Individuals With COVID-19 - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04412018>. Retrieved 8 February 2021.
1826. Bhatt DL, Steg PG, Miller M, Brinton EA, Jacobson TA, Ketchum SB, Doyle RT, Juliano RA, Jiao L, Granowitz C, Tardif J-C, Ballantyne CM. 2019. [Cardiovascular Risk Reduction with Icosapent Ethyl for Hypertriglyceridemia](#). New England Journal of Medicine 380:11–22.
1827. S.L.A. Pharma AG. 2020. [A Randomised, Double-blind, Placebo Controlled Study of Eicosapentaenoic Acid \(EPA-FFA\) Gastro-resistant Capsules to Treat Hospitalised Subjects With Confirmed SARS-CoV-2](#). NCT04335032. Clinical trial registration. clinicaltrials.gov.
1828. FACN MA M. D. 2020. [Anti-inflammatory/Antioxidant Oral Nutrition Supplementation on the Cytokine Storm and Progression of COVID-19:](#)

[A Randomized Controlled Trial](#). NCT04323228. Clinical trial registration. clinicaltrials.gov.

1829. Messina G, Polito R, Monda V, Cipolloni L, Di Nunno N, Di Mizio G, Murabito P, Carotenuto M, Messina A, Pisanelli D, Valenzano A, Cibelli G, Scarinci A, Monda M, Sessa F. 2020. [Functional Role of Dietary Intervention to Improve the Outcome of COVID-19: A Hypothesis of Work](#). International Journal of Molecular Sciences 21:3104.
1830. Maares M, Haase H. 2016. [Zinc and immunity: An essential interrelation](#). Archives of Biochemistry and Biophysics 611:58–65.
1831. von Bülow V, Dubben S, Engelhardt G, Hebel S, Plümäkers B, Heine H, Rink L, Haase H. 2007. [Zinc-Dependent Suppression of TNF- \$\alpha\$ Production Is Mediated by Protein Kinase A-Induced Inhibition of Raf-1, I \$\kappa\$ B Kinase \$\beta\$, and NF- \$\kappa\$ B](#). The Journal of Immunology 179:4180–4186.
1832. Prasad AS, Bao B, Beck FWJ, Sarkar FH. 2001. [Zinc activates NF- \$\kappa\$ B in HUT-78 cells](#). Journal of Laboratory and Clinical Medicine 138:250–256.
1833. Vivier E, Raulet DH, Moretta A, Caligiuri MA, Zitvogel L, Lanier LL, Yokoyama WM, Ugolini S. 2011. [Innate or Adaptive Immunity? The Example of Natural Killer Cells](#). Science 331:44–49.
1834. Prasad AS, Beck FW, Bao B, Fitzgerald JT, Snell DC, Steinberg JD, Cardozo LJ. 2007. [Zinc supplementation decreases incidence of infections in the elderly: effect of zinc on generation of cytokines and oxidative stress](#). The American Journal of Clinical Nutrition 85:837–844.
1835. Read SA, Obeid S, Ahlenstiel C, Ahlenstiel G. 2019. [The Role of Zinc in Antiviral Immunity](#). Advances in Nutrition 10:696–710.
1836. Hulisz D. 2004. [Efficacy of Zinc Against Common Cold Viruses: An Overview](#). Journal of the American Pharmacists Association 44:594–603.
1837. Hemilä H. 2011. [Zinc Lozenges May Shorten the Duration of Colds: A Systematic Review](#). The Open Respiratory Medicine Journal 5:51–58.
1838. Jothimani D, Kailasam E, Danielraj S, Nallathambi B, Ramachandran H, Sekar P, Manoharan S, Ramani V, Narasimhan G, Kaliamoorthy I, Rela M. 2020. [COVID-19: Poor outcomes in patients with zinc deficiency](#). International Journal of Infectious Diseases 100:343–349.
1839. te Velthuis AJW, van den Worm SHE, Sims AC, Baric RS, Snijder EJ, van Hemert MJ. 2010. [Zn²⁺ Inhibits Coronavirus and Arterivirus RNA Polymerase Activity In Vitro and Zinc Ionophores Block the Replication of These Viruses in Cell Culture](#). PLoS Pathogens 6:e1001176.
1840. Báez-Santos YM, St. John SE, Mesecar AD. 2015. [The SARS-coronavirus papain-like protease: Structure, function and inhibition by designed antiviral compounds](#). Antiviral Research 115:21–38.
1841. MD AT. 2020. [A Randomized Study Evaluating the Safety and Efficacy of Hydroxychloroquine and Zinc in Combination With Either Azithromycin](#)

[or Doxycycline for the Treatment of COVID-19 in the Outpatient Setting](#). NCT04370782. Clinical trial registration. clinicaltrials.gov.

1842. A Study of Hydroxychloroquine and Zinc in the Prevention of COVID-19 Infection in Military Healthcare Workers - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04377646>. Retrieved 8 February 2021.
1843. National Institute of Integrative Medicine, Australia. 2020. [Therapies to Prevent Progression of COVID-19, Including Hydroxychloroquine, Azithromycin, Zinc, Vitamin D, Vitamin B12 With or Without Vitamin C, a Multi-centre, International, Randomized Trial: The International ALLIANCE Study](#). NCT04395768. Clinical trial registration. clinicaltrials.gov.
1844. Early Intervention in COVID-19: Favipiravir Verses Standard Care - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04373733>. Retrieved 8 February 2021.
1845. Cavalcanti AB, Zampieri FG, Rosa RG, Azevedo LCP, Veiga VC, Avezum A, Damiani LP, Marcadenti A, Kawano-Dourado L, Lisboa T, Junqueira DLM, de Barros e Silva PGM, Tramuja L, Abreu-Silva EO, Laranjeira LN, Soares AT, Echenique LS, Pereira AJ, Freitas FGR, Gebara OCE, Dantas VCS, Furtado RHM, Milan EP, Golin NA, Cardoso FF, Maia IS, Hoffmann Filho CR, Kormann APM, Amazonas RB, Bocchi de Oliveira MF, Serpa-Neto A, Falavigna M, Lopes RD, Machado FR, Berwanger O. 2020. [Hydroxychloroquine with or without Azithromycin in Mild-to-Moderate Covid-19](#). New England Journal of Medicine 383:2041–2052.
1846. Lewis K, Chaudhuri D, Alshamsi F, Carayannopoulos L, Dearness K, Chagla Z, Alhazzani W, for the GUIDE Group. 2021. [The efficacy and safety of hydroxychloroquine for COVID-19 prophylaxis: A systematic review and meta-analysis of randomized trials](#). PLOS ONE 16:e0244778.
1847. Fiolet T, Guihur A, Rebeaud ME, Mulot M, Peiffer-Smadja N, Mahamat-Saleh Y. 2021. [Effect of hydroxychloroquine with or without azithromycin on the mortality of coronavirus disease 2019 \(COVID-19\) patients: a systematic review and meta-analysis](#). Clinical Microbiology and Infection 27:19–27.
1848. Carlucci PM, Ahuja T, Petrilli C, Rajagopalan H, Jones S, Rahimian J. 2020. [Zinc sulfate in combination with a zinc ionophore may improve outcomes in hospitalized COVID-19 patients](#). Journal of Medical Microbiology 69:1228–1234.
1849. Yao JS, Paguio JA, Dee EC, Tan HC, Moulick A, Milazzo C, Jurado J, Della Penna N, Celi LA. 2021. [The Minimal Effect of Zinc on the Survival of Hospitalized Patients With COVID-19](#). Chest 159:108–111.
1850. Desai M. 2021. [Coronavirus Disease 2019- Using Ascorbic Acid and Zinc Supplementation \(COVIDAtoZ\) Research Study A Randomized, Open Label Single Center Study](#). NCT04342728. Clinical trial registration. clinicaltrials.gov.

1851. Narayanan N, Nair DT. 2020. Vitamin B12 May Inhibit RNA-Dependent-RNA Polymerase Activity of nsp12 from the COVID-19 Virus. Preprints <https://doi.org/10.20944/preprints202003.0347.v1>.
1852. Cerullo G, Negro M, Parimbelli M, Pecoraro M, Perna S, Liguori G, Rondanelli M, Cena H, D'Antona G. 2020. [The Long History of Vitamin C: From Prevention of the Common Cold to Potential Aid in the Treatment of COVID-19](#). Frontiers in Immunology 11:574029.
1853. Carr AC, Rowe S. 2020. [The Emerging Role of Vitamin C in the Prevention and Treatment of COVID-19](#). Nutrients 12:3286.
1854. Chen Y, Luo G, Yuan J, Wang Y, Yang X, Wang X, Li G, Liu Z, Zhong N. 2014. [Vitamin C Mitigates Oxidative Stress and Tumor Necrosis Factor-Alpha in Severe Community-Acquired Pneumonia and LPS-Induced Macrophages](#). Mediators of Inflammation 2014:1–11.
1855. Hagel AF, Layritz CM, Hagel WH, Hagel H-J, Hagel E, Dauth W, Kressel J, Regnet T, Rosenberg A, Neurath MF, Molderings GJ, Raithel M. 2013. [Intravenous infusion of ascorbic acid decreases serum histamine concentrations in patients with allergic and non-allergic diseases](#). Naunyn-Schmiedeberg's Archives of Pharmacology 386:789–793.
1856. Carr A, Maggini S. 2017. [Vitamin C and Immune Function](#). Nutrients 9:1211.
1857. Hume R, Weyers E. 2016. [Changes in Leucocyte Ascorbic Acid during the Common Cold](#). Scottish Medical Journal 18:3–7.
1858. Wilson CWM. 1975. [ASCORBIC ACID FUNCTION AND METABOLISM DURING COLDS](#). Annals of the New York Academy of Sciences 258:529–539.
1859. Davies JEW, Hughes RE, Jones E, Reed SE, Craig JW, Tyrrell DAJ. 1979. [Metabolism of ascorbic acid \(vitamin C\) in subjects infected with common cold viruses](#). Biochemical Medicine 21:78–85.
1860. Hemilä H. 2017. [Vitamin C and Infections](#). Nutrients 9:339.
1861. Hemilä H. 2007. [Vitamin C and the common cold](#). British Journal of Nutrition 67:3–16.
1862. Hemilä H, Chalker E. 2019. [Vitamin C Can Shorten the Length of Stay in the ICU: A Meta-Analysis](#). Nutrients 11:708.
1863. Arvinte C, Singh M, Marik PE. 2020. [Serum Levels of Vitamin C and Vitamin D in a Cohort of Critically Ill COVID-19 Patients of a North American Community Hospital Intensive Care Unit in May 2020: A Pilot Study](#). Medicine in Drug Discovery 8:100064.
1864. Chiscano-Camón L, Ruiz-Rodriguez JC, Ruiz-Sanmartin A, Roca O, Ferrer R. 2020. [Vitamin C levels in patients with SARS-CoV-2-associated acute respiratory distress syndrome](#). Critical Care 24:522.
1865. José RJ, Williams A, Manuel A, Brown JS, Chambers RC. 2020. [Targeting coagulation activation in severe COVID-19 pneumonia: lessons from](#)

[bacterial pneumonia and sepsis](#). European Respiratory Review 29:200240.

1866. Tymk K. 2017. [Vitamin C and Microvascular Dysfunction in Systemic Inflammation](#). Antioxidants 6:49.
1867. Hiedra R, Lo KB, Elbashabsheh M, Gul F, Wright RM, Albano J, Azmaiparashvili Z, Patarroyo Aponte G. 2020. [The use of IV vitamin C for patients with COVID-19: a case series](#). Expert Review of Anti-infective Therapy 18:1259–1261.
1868. Hemilä H, Chalker E. 2013. Vitamin C for preventing and treating the common cold. Cochrane Database of Systematic Reviews <https://doi.org/10.1002/14651858.cd000980.pub4>.
1869. HEMILÄ H. 1997. [Vitamin C intake and susceptibility to pneumonia](#). The Pediatric Infectious Disease Journal 16:836–837.
1870. Fowler AA, Truitt JD, Hite RD, Morris PE, DeWilde C, Priday A, Fisher B, Thacker LR, Natarajan R, Brophy DF, Sculthorpe R, Nanchal R, Syed A, Sturgill J, Martin GS, Sevransky J, Kashiouris M, Hamman S, Egan KF, Hastings A, Spencer W, Tench S, Mehkri O, Bindas J, Duggal A, Graf J, Zellner S, Yanny L, McPolin C, Hollrith T, Kramer D, Ojielo C, Damm T, Cassity E, Wieliczko A, Halquist M. 2019. [Effect of Vitamin C Infusion on Organ Failure and Biomarkers of Inflammation and Vascular Injury in Patients With Sepsis and Severe Acute Respiratory Failure](#). JAMA 322:1261.
1871. Liu F, Zhu Y, Zhang J, Li Y, Peng Z. 2020. [Intravenous high-dose vitamin C for the treatment of severe COVID-19: study protocol for a multicentre randomised controlled trial](#). BMJ Open 10:e039519.
1872. Zhang J, Rao X, Li Y, Zhu Y, Liu F, Guo G, Luo G, Meng Z, Backer DD, Xiang H, Peng Z-Y. 2020. Pilot Trial of High-dose vitamin C in critically ill COVID-19 patients. Research Square <https://doi.org/10.21203/rs.3.rs-52778/v2>.
1873. Zhang J, Rao X, Li Y, Zhu Y, Liu F, Guo G, Luo G, Meng Z, Backer DD, Xiang H, Peng Z-Y. 2020. High-dose vitamin C infusion for the treatment of critically ill COVID-19. Research Square <https://doi.org/10.21203/rs.3.rs-52778/v1>.
1874. Panel on Dietary Antioxidants and Related Compounds, Subcommittee on Upper Reference Levels of Nutrients, Subcommittee on Interpretation and Uses of Dietary Reference Intakes, Standing Committee on the Scientific Evaluation of Dietary Reference Intakes, Food and Nutrition Board, Institute of Medicine. 2000. Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium, and Carotenoids. The National Academies Press. <https://doi.org/ghvtvqx>.
1875. Gois P, Ferreira D, Olenski S, Seguro A. 2017. [Vitamin D and Infectious Diseases: Simple Bystander or Contributing Factor?](#) Nutrients 9:651.
1876. Gruber–Bzura BM. 2018. [Vitamin D and Influenza—Prevention or Therapy?](#) International Journal of Molecular Sciences 19:2419.

1877. Charoenngam N, Holick MF. 2020. [Immunologic Effects of Vitamin D on Human Health and Disease](#). *Nutrients* 12:2097.
1878. Hughes DA, Norton R. 2009. [Vitamin D and respiratory health](#). *Clinical & Experimental Immunology* 158:20–25.
1879. Vanherwegen A-S, Gysemans C, Mathieu C. 2017. [Regulation of Immune Function by Vitamin D and Its Use in Diseases of Immunity](#). *Endocrinology and Metabolism Clinics of North America* 46:1061–1094.
1880. Aranow C. 2015. [Vitamin D and the Immune System](#). *Journal of Investigative Medicine* 59:881–886.
1881. Jolliffe DA, Griffiths CJ, Martineau AR. 2013. [Vitamin D in the prevention of acute respiratory infection: Systematic review of clinical studies](#). *The Journal of Steroid Biochemistry and Molecular Biology* 136:321–329.
1882. Baeke F, Takiishi T, Korf H, Gysemans C, Mathieu C. 2010. [Vitamin D: modulator of the immune system](#). *Current Opinion in Pharmacology* 10:482–496.
1883. Grant WB, Lahore H, McDonnell SL, Baggerly CA, French CB, Aliano JL, Bhattoa HP. 2020. [Evidence that Vitamin D Supplementation Could Reduce Risk of Influenza and COVID-19 Infections and Deaths](#). *Nutrients* 12:988.
1884. Rhodes JM, Subramanian S, Laird E, Griffin G, Kenny RA. 2020. [Perspective: Vitamin D deficiency and COVID-19 severity – plausibly linked by latitude, ethnicity, impacts on cytokines, ACE2 and thrombosis](#). *Journal of Internal Medicine* 289:97–115.
1885. Whittemore PB. 2020. [COVID-19 fatalities, latitude, sunlight, and vitamin D](#). *American Journal of Infection Control* 48:1042–1044.
1886. Rhodes JM, Subramanian S, Laird E, Kenny RA. 2020. [Editorial: low population mortality from COVID-19 in countries south of latitude 35 degrees North supports vitamin D as a factor determining severity](#). *Alimentary Pharmacology & Therapeutics* 51:1434–1437.
1887. D'Avolio A, Avataneo V, Manca A, Cusato J, De Nicolò A, Lucchini R, Keller F, Cantù M. 2020. [25-Hydroxyvitamin D Concentrations Are Lower in Patients with Positive PCR for SARS-CoV-2](#). *Nutrients* 12:1359.
1888. De Smet D, De Smet K, Herroelen P, Gryspeerdt S, Martens GA. 2020. Vitamin D deficiency as risk factor for severe COVID-19: a convergence of two pandemics. *Cold Spring Harbor Laboratory* <https://doi.org/10.1101/2020.05.01.20079376>.
1889. Maghbooli Z, Sahraian MA, Ebrahimi M, Pazoki M, Kafan S, Tabriz HM, Hadadi A, Montazeri M, Nasiri M, Shirvani A, Holick MF. 2020. [Vitamin D sufficiency, a serum 25-hydroxyvitamin D at least 30 ng/mL reduced risk for adverse clinical outcomes in patients with COVID-19 infection](#). *PLOS ONE* 15:e0239799.
1890. Ali N. 2020. [Role of vitamin D in preventing of COVID-19 infection, progression and severity](#). *Journal of Infection and Public Health*

1891. Merzon E, Tworowski D, Gorohovski A, Vinker S, Golan Cohen A, Green I, Frenkel-Morgenstern M. 2020. [Low plasma 25\(OH\) vitamin D level is associated with increased risk of COVID-19 infection: an Israeli population-based study](#). The FEBS Journal 287:3693–3702.
1892. Meltzer DO, Best TJ, Zhang H, Vokes T, Arora V, Solway J. 2020. [Association of Vitamin D Status and Other Clinical Characteristics With COVID-19 Test Results](#). JAMA Network Open 3:e2019722.
1893. Hernández JL, Nan D, Fernandez-Ayala M, García-Unzueta M, Hernández-Hernández MA, López-Hoyos M, Muñoz-Cacho P, Olmos JM, Gutiérrez-Cuadra M, Ruiz-Cubillán JJ, Crespo J, Martínez-Taboada VM. 2020. [Vitamin D Status in Hospitalized Patients with SARS-CoV-2 Infection](#). The Journal of Clinical Endocrinology & Metabolism dgaa733.
1894. Jain A, Chaurasia R, Sengar NS, Singh M, Mahor S, Narain S. 2020. [Analysis of vitamin D level among asymptomatic and critically ill COVID-19 patients and its correlation with inflammatory markers](#). Scientific Reports 10:20191.
1895. Vassiliou AG, Jahaj E, Pratikaki M, Orfanos SE, Dimopoulou I, Kotanidou A. 2020. [Low 25-Hydroxyvitamin D Levels on Admission to the Intensive Care Unit May Predispose COVID-19 Pneumonia Patients to a Higher 28-Day Mortality Risk: A Pilot Study on a Greek ICU Cohort](#). Nutrients 12:3773.
1896. Carpagnano GE, Di Lecce V, Quaranta VN, Zito A, Buonamico E, Capozza E, Palumbo A, Di Gioia G, Valerio VN, Resta O. 2020. Vitamin D deficiency as a predictor of poor prognosis in patients with acute respiratory failure due to COVID-19. Journal of Endocrinological Investigation <https://doi.org/10.1007/s40618-020-01370-x>.
1897. Radujkovic A, Hippchen T, Tiwari-Heckler S, Dreher S, Boxberger M, Merle U. 2020. [Vitamin D Deficiency and Outcome of COVID-19 Patients](#). Nutrients 12:2757.
1898. Pizzini A, Aichner M, Sahanic S, Böhm A, Egger A, Hoermann G, Kurz K, Widmann G, Bellmann-Weiler R, Weiss G, Tancevski I, Sonnweber T, Löffler-Ragg J. 2020. [Impact of Vitamin D Deficiency on COVID-19—A Prospective Analysis from the CovILD Registry](#). Nutrients 12:2775.
1899. Ye K, Tang F, Liao X, Shaw BA, Deng M, Huang G, Qin Z, Peng X, Xiao H, Chen C, Liu X, Ning L, Wang B, Tang N, Li M, Xu F, Lin S, Yang J. 2020. [Does Serum Vitamin D Level Affect COVID-19 Infection and Its Severity? -A Case-Control Study](#). Journal of the American College of Nutrition 1–8.
1900. Padhi S, Suvankar S, Panda VK, Pati A, Panda AK. 2020. [Lower levels of vitamin D are associated with SARS-CoV-2 infection and mortality in the Indian population: An observational study](#). International Immunopharmacology 88:107001.
1901. Luo X, Liao Q, Shen Y, Li H, Cheng L. 2021. [Vitamin D Deficiency Is Associated with COVID-19 Incidence and Disease Severity in Chinese](#)

[People](#). The Journal of Nutrition 151:98–103.

1902. Hastie CE, Mackay DF, Ho F, Celis-Morales CA, Katikireddi SV, Niedzwiedz CL, Jani BD, Welsh P, Mair FS, Gray SR, O'Donnell CA, Gill JMR, Sattar N, Pell JP. 2020. [Vitamin D concentrations and COVID-19 infection in UK Biobank](#). Diabetes & Metabolic Syndrome: Clinical Research & Reviews 14:561–565.
1903. Hastie CE, Pell JP, Sattar N. 2020. Vitamin D and COVID-19 infection and mortality in UK Biobank. European Journal of Nutrition <https://doi.org/10.1007/s00394-020-02372-4>.
1904. Panagiotou G, Tee SA, Ihsan Y, Athar W, Marchitelli G, Kelly D, Boot CS, Stock N, Macfarlane J, Martineau AR, Burns G, Quinton R. 2020. [Low serum 25-hydroxyvitamin D \(25\[OH\]D\) levels in patients hospitalized with COVID-19 are associated with greater disease severity](#). Clinical Endocrinology 93:508–511.
1905. Grant WB, McDonnell SL. 2020. [Letter in response to the article: Vitamin D concentrations and COVID-19 infection in UK biobank \(Hastie et al.\)](#). Diabetes & Metabolic Syndrome: Clinical Research & Reviews 14:893–894.
1906. Martín Giménez VM, Inserra F, Ferder L, García J, Manucha W. 2020. Vitamin D deficiency in African Americans is associated with a high risk of severe disease and mortality by SARS-CoV-2. Journal of Human Hypertension <https://doi.org/10.1038/s41371-020-00398-z>.
1907. Daneshkhah A, Agrawal V, Eshein A, Subramanian H, Roy HK, Backman V. 2020. [Evidence for possible association of vitamin D status with cytokine storm and unregulated inflammation in COVID-19 patients](#). Aging Clinical and Experimental Research 32:2141–2158.
1908. Rastogi A, Bhansali A, Khare N, Suri V, Yaddanapudi N, Sachdeva N, Puri GD, Malhotra P. 2020. [Short term, high-dose vitamin D supplementation for COVID-19 disease: a randomised, placebo-controlled, study \(SHADE study\)](#). Postgraduate Medical Journal postgradmedj-2020-139065.
1909. Entrenas Castillo M, Entrenas Costa LM, Vaquero Barrios JM, Alcalá Díaz JF, López Miranda J, Bouillon R, Quesada Gomez JM. 2020. [“Effect of calcifediol treatment and best available therapy versus best available therapy on intensive care unit admission and mortality among patients hospitalized for COVID-19: A pilot randomized clinical study”](#). The Journal of Steroid Biochemistry and Molecular Biology 203:105751.
1910. COVID-19 rapid evidence summary: vitamin D for COVID-19 | Advice | NICE. <https://www.nice.org.uk/advice/es28>. Retrieved 8 February 2021.
1911. Jungreis I, Kellis M. 2020. Mathematical analysis of Córdoba calcifediol trial suggests strong role for Vitamin D in reducing ICU admissions of hospitalized COVID-19 patients. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.11.08.20222638>.

1912. Ling SF, Broad E, Murphy R, Pappachan JM, Pardesi-Newton S, Kong M-F, Jude EB. 2020. [High-Dose Cholecalciferol Booster Therapy is Associated with a Reduced Risk of Mortality in Patients with COVID-19: A Cross-Sectional Multi-Centre Observational Study](#). *Nutrients* 12:3799.
1913. Murai IH, Fernandes AL, Sales LP, Pinto AJ, Goessler KF, Duran CSC, Silva CBR, Franco AS, Macedo MB, Dalmolin HHH, Baggio J, Balbi GGM, Reis BZ, Antonangelo L, Caparbo VF, Gualano B, Pereira RMR. 2020. Effect of Vitamin D ³ Supplementation vs Placebo on Hospital Length of Stay in Patients with Severe COVID-19: A Multicenter, Double-blind, Randomized Controlled Trial. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.11.16.20232397>.
1914. Garzón MC. 2020. [Effect of Vitamin D Administration on Prevention and Treatment of Mild Forms of Suspected Covid-19](#). NCT04334005. Clinical trial registration. clinicaltrials.gov.
1915. Montano-Loza A. 2020. [Improving Vitamin D Status in the Management of COVID-19](#). NCT04385940. Clinical trial registration. clinicaltrials.gov.
1916. Cholecalciferol to Improve the Outcomes of COVID-19 Patients - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04411446>. Retrieved 8 February 2021.
1917. COvid-19 and Vitamin D Supplementation: a Multicenter Randomized Controlled Trial of High Dose Versus Standard Dose Vitamin D3 in High-risk COVID-19 Patients (CoVitTrial) - Full Text View - ClinicalTrials.gov. <https://clinicaltrials.gov/ct2/show/NCT04344041>. Retrieved 8 February 2021.
1918. Louisiana State University Health Sciences Center in New Orleans. 2020. [The LEAD COVID-19 Trial: Low-risk, Early Aspirin and Vitamin D to Reduce COVID-19 Hospitalizations](#). NCT04363840. Clinical trial registration. clinicaltrials.gov.
1919. MD MM. 2020. [Randomized Double-Blind Placebo-Controlled Proof-of-Concept Trial of a Plant Polyphenol for the Outpatient Treatment of Mild Coronavirus Disease \(COVID-19\)](#). NCT04400890. Clinical trial registration. clinicaltrials.gov.
1920. Lips P, Cashman KD, Lamberg-Allardt C, Bischoff-Ferrari HA, Obermayer-Pietsch B, Bianchi ML, Stepan J, El-Hajj Fuleihan G, Bouillon R. 2019. [Current vitamin D status in European and Middle East countries and strategies to prevent vitamin D deficiency: a position statement of the European Calcified Tissue Society](#). *European Journal of Endocrinology* 180:P23–P54.
1921. Communiqué de l'Académie nationale de Médecine : Vitamine D et Covid-19 – Académie nationale de médecine | Une institution dans son temps. <https://www.academie-medecine.fr/communiqué-de-lacademie-nationale-de-medecine-vitamine-d-et-covid-19/>. Retrieved 8 February 2021.
1922. Iacobucci G. 2020. [Covid-19: NHS bosses told to assess risk to ethnic minority staff who may be at greater risk](#). *BMJ* m1820.

1923. Torjesen I. 2020. [Covid-19: Public health agencies review whether vitamin D supplements could reduce risk](#). BMJ m2475.
1924. Kohlmeier M. 2020. [Avoidance of vitamin D deficiency to slow the COVID-19 pandemic](#). BMJ Nutrition, Prevention & Health 3:67–73.
1925. National Institute for Health and Care Excellence (NICE). COVID-19 rapid guideline: vitamin D. <https://www.nice.org.uk/guidance/ng187/resources/covid19-rapid-guideline-vitamin-d-pdf-66142026720709>.
1926. Parva NR, Tadepalli S, Singh P, Qian A, Joshi R, Kandala H, Nookala VK, Cheriya P. 2018. Prevalence of Vitamin D Deficiency and Associated Risk Factors in the US Population (2011-2012). Cureus <https://doi.org/10.7759/cureus.2741>.
1927. Vitamin D. COVID-19 Treatment Guidelines. <https://www.covid19treatmentguidelines.nih.gov/adjunctive-therapy/vitamin-d/>. Retrieved 8 February 2021.
1928. Hamulka J, Jeruszka-Bielak M, Górnicka M, Drywień ME, Zielinska-Pukos MA. 2020. [Dietary Supplements during COVID-19 Outbreak. Results of Google Trends Analysis Supported by PLifeCOVID-19 Online Studies](#). Nutrients 13:54.
1929. 2021. Court Orders Georgia Defendants to Stop Selling Vitamin D Products as Treatments for Covid-19 and Other Diseases. <https://www.justice.gov/opa/pr/court-orders-georgia-defendants-stop-selling-vitamin-d-products-treatments-covid-19-and-other>. Retrieved 8 February 2021.
1930. Hill C, Guarner F, Reid G, Gibson GR, Merenstein DJ, Pot B, Morelli L, Canani RB, Flint HJ, Salminen S, Calder PC, Sanders ME. 2014. [The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic](#). Nature Reviews Gastroenterology & Hepatology 11:506–514.
1931. Kang E-J, Kim SY, Hwang I-H, Ji Y-J. 2013. [The Effect of Probiotics on Prevention of Common Cold: A Meta-Analysis of Randomized Controlled Trial Studies](#). Korean Journal of Family Medicine 34:2.
1932. Kanauchi O, Andoh A, AbuBakar S, Yamamoto N. 2018. [Probiotics and Paraprobiotics in Viral Infection: Clinical Application and Effects on the Innate and Acquired Immune Systems](#). Current Pharmaceutical Design 24:710–717.
1933. Baud D, Dimopoulou Agri V, Gibson GR, Reid G, Giannoni E. 2020. [Using Probiotics to Flatten the Curve of Coronavirus Disease COVID-2019 Pandemic](#). Frontiers in Public Health 8:186.
1934. O'Toole PW, Marchesi JR, Hill C. 2017. [Next-generation probiotics: the spectrum from probiotics to live biotherapeutics](#). Nature Microbiology 2:17057.
1935. Plaza-Diaz J, Ruiz-Ojeda FJ, Gil-Campos M, Gil A. 2019. [Mechanisms of Action of Probiotics](#). Advances in Nutrition 10:S49–S66.

1936. Halloran K, Underwood MA. 2019. [Probiotic mechanisms of action](#). Early Human Development 135:58–65.
1937. Bermudez-Brito M, Plaza-Díaz J, Muñoz-Quezada S, Gómez-Llorente C, Gil A. 2012. [Probiotic Mechanisms of Action](#). Annals of Nutrition and Metabolism 61:160–174.
1938. BOTIC T, KLINGBERG T, WEINGARTL H, CENCIC A. 2007. [A novel eukaryotic cell culture model to study antiviral activity of potential probiotic bacteria](#). International Journal of Food Microbiology 115:227–234.
1939. Waki N, Yajima N, Suganuma H, Buddle BM, Luo D, Heiser A, Zheng T. 2014. [Oral administration of <i>Lactobacillus brevis</i> KB290 to mice alleviates clinical symptoms following influenza virus infection](#). Letters in Applied Microbiology 58:87–93.
1940. Mastromarino P, Cacciotti F, Masci A, Mosca L. 2011. [Antiviral activity of Lactobacillus brevis towards herpes simplex virus type 2: Role of cell wall associated components](#). Anaerobe 17:334–336.
1941. Percopo CM, Ma M, Brenner TA, Krumholz JO, Break TJ, Laky K, Rosenberg HF. 2019. [Critical Adverse Impact of IL-6 in Acute Pneumovirus Infection](#). The Journal of Immunology 202:871–882.
1942. Biliavska, Pankivska, Povnitsa, Zagorodnya. 2019. [Antiviral Activity of Exopolysaccharides Produced by Lactic Acid Bacteria of the Genera Pediococcus, Leuconostoc and Lactobacillus against Human Adenovirus Type 5](#). Medicina 55:519.
1943. Eguchi K, Fujitani N, Nakagawa H, Miyazaki T. 2019. [Prevention of respiratory syncytial virus infection with probiotic lactic acid bacterium Lactobacillus gasseri SBT2055](#). Scientific Reports 9:4812.
1944. Turner RB, Woodfolk JA, Borish L, Steinke JW, Patrie JT, Muehling LM, Lahtinen S, Lehtinen MJ. 2017. [Effect of probiotic on innate inflammatory response and viral shedding in experimental rhinovirus infection – a randomised controlled trial](#). Beneficial Microbes 8:207–215.
1945. Zelaya H, Tsukida K, Chiba E, Marranzino G, Alvarez S, Kitazawa H, Agüero G, Villena J. 2014. [Immunobiotic lactobacilli reduce viral-associated pulmonary damage through the modulation of inflammation-coagulation interactions](#). International Immunopharmacology 19:161–173.
1946. Tonetti FR, Islam MdA, Vizoso-Pinto MG, Takahashi H, Kitazawa H, Villena J. 2020. [Nasal priming with immunobiotic lactobacilli improves the adaptive immune response against influenza virus](#). International Immunopharmacology 78:106115.
1947. Olaimat AN, Aolymat I, Al-Holy M, Ayyash M, Abu Ghoush M, Al-Nabulsi AA, Osaili T, Apostolopoulos V, Liu S-Q, Shah NP. 2020. [The potential application of probiotics and prebiotics for the prevention and treatment of COVID-19](#). npj Science of Food 4:17.

1948. Keely S, Talley NJ, Hansbro PM. 2011. [Pulmonary-intestinal cross-talk in mucosal inflammatory disease](#). *Mucosal Immunology* 5:7–18.
1949. Dumas A, Bernard L, Poquet Y, Lugo-Villarino G, Neyrolles O. 2018. [The role of the lung microbiota and the gut-lung axis in respiratory infectious diseases](#). *Cellular Microbiology* 20:e12966.
1950. Dhar D, Mohanty A. 2020. [Gut microbiota and Covid-19- possible link and implications](#). *Virus Research* 285:198018.
1951. Bao L, Zhang C, Dong J, Zhao L, Li Y, Sun J. 2020. [Oral Microbiome and SARS-CoV-2: Beware of Lung Co-infection](#). *Frontiers in Microbiology* 11:1840.
1952. Khatiwada S, Subedi A. 2020. [Lung microbiome and coronavirus disease 2019 \(COVID-19\): Possible link and implications](#). *Human Microbiome Journal* 17:100073.
1953. Lehtoranta L, Pitkäranta A, Korpela R. 2014. [Probiotics in respiratory virus infections](#). *European Journal of Clinical Microbiology & Infectious Diseases* 33:1289–1302.
1954. Hao Q, Dong BR, Wu T. 2015. Probiotics for preventing acute upper respiratory tract infections. *Cochrane Database of Systematic Reviews* <https://doi.org/10.1002/14651858.cd006895.pub3>.
1955. Vouloumanou EK, Makris GC, Karageorgopoulos DE, Falagas ME. 2009. [Probiotics for the prevention of respiratory tract infections: a systematic review](#). *International Journal of Antimicrobial Agents* 34:197.e1–197.e10.
1956. King S, Glanville J, Sanders ME, Fitzgerald A, Varley D. 2014. [Effectiveness of probiotics on the duration of illness in healthy children and adults who develop common acute respiratory infectious conditions: a systematic review and meta-analysis](#). *British Journal of Nutrition* 112:41–54.
1957. Zeng J, Wang C-T, Zhang F-S, Qi F, Wang S-F, Ma S, Wu T-J, Tian H, Tian Z-T, Zhang S-L, Qu Y, Liu L-Y, Li Y-Z, Cui S, Zhao H-L, Du Q-S, Ma Z, Li C-H, Li Y, Si M, Chu Y-F, Meng M, Ren H-S, Zhang J-C, Jiang J-J, Ding M, Wang Y-P. 2016. [Effect of probiotics on the incidence of ventilator-associated pneumonia in critically ill patients: a randomized controlled multicenter trial](#). *Intensive Care Medicine* 42:1018–1028.
1958. Morrow LE, Kollef MH, Casale TB. 2010. [Probiotic Prophylaxis of Ventilator-associated Pneumonia](#). *American Journal of Respiratory and Critical Care Medicine* 182:1058–1064.
1959. Shimizu K, Yamada T, Ogura H, Mohri T, Kiguchi T, Fujimi S, Asahara T, Yamada T, Ojima M, Ikeda M, Shimazu T. 2018. [Synbiotics modulate gut microbiota and reduce enteritis and ventilator-associated pneumonia in patients with sepsis: a randomized controlled trial](#). *Critical Care* 22:239.
1960. Su M, Jia Y, Li Y, Zhou D, Jia J. 2020. [Probiotics for the Prevention of Ventilator-Associated Pneumonia: A Meta-Analysis of Randomized](#)

[Controlled Trials](#). Respiratory Care 65:673–685.

1961. Helvécio Cardoso Corrêa Póvoa, Chianca GC, Iorio NLPP. 2020. [COVID-19: An Alert to Ventilator-Associated Bacterial Pneumonia](#). Adis Journals.
1962. François B, Laterre P-F, Luyt C-E, Chastre J. 2020. [The challenge of ventilator-associated pneumonia diagnosis in COVID-19 patients](#). Critical Care 24:289.
1963. Perceval C, Szajewska H, Indrio F, Weizman Z, Vandenplas Y. 2019. [Prophylactic use of probiotics for gastrointestinal disorders in children](#). The Lancet Child & Adolescent Health 3:655–662.
1964. Zhou Z, Zhao N, Shu Y, Han S, Chen B, Shu X. 2020. [Effect of Gastrointestinal Symptoms in Patients With COVID-19](#). Gastroenterology 158:2294–2297.
1965. Zhang H, Kang Z, Gong H, Xu D, Wang J, Li Z, Cui X, Xiao J, Meng T, Zhou W, Liu J, Xu H. 2020. The digestive system is a potential route of 2019-nCoV infection: a bioinformatics analysis based on single-cell transcriptomes. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.01.30.927806>.
1966. Gui M, Song W, Zhou H, Xu J, Chen S, Xiang Y, Wang X. 2016. [Cryo-electron microscopy structures of the SARS-CoV spike glycoprotein reveal a prerequisite conformational state for receptor binding](#). Cell Research 27:119–129.
1967. Yeo C, Kaushal S, Yeo D. 2020. [Enteric involvement of coronaviruses: is faecal–oral transmission of SARS-CoV-2 possible?](#) The Lancet Gastroenterology & Hepatology 5:335–337.
1968. Xu Y, Li X, Zhu B, Liang H, Fang C, Gong Y, Guo Q, Sun X, Zhao D, Shen J, Zhang H, Liu H, Xia H, Tang J, Zhang K, Gong S. 2020. [Characteristics of pediatric SARS-CoV-2 infection and potential evidence for persistent fecal viral shedding](#). Nature Medicine 26:502–505.
1969. Gonzalez-Ochoa G, Flores-Mendoza LK, Icedo-Garcia R, Gomez-Flores R, Tamez-Guerra P. 2017. [Modulation of rotavirus severe gastroenteritis by the combination of probiotics and prebiotics](#). Archives of Microbiology 199:953–961.
1970. Freedman SB, Williamson-Urquhart S, Farion KJ, Gouin S, Willan AR, Poonai N, Hurley K, Sherman PM, Finkelstein Y, Lee BE, Pang X-L, Chui L, Schnadower D, Xie J, Gorelick M, Schuh S. 2018. [Multicenter Trial of a Combination Probiotic for Children with Gastroenteritis](#). New England Journal of Medicine 379:2015–2026.
1971. Medical University of Graz. 2021. [Synbiotic Therapy of Gastrointestinal Symptoms During Covid-19 Infection: A Randomized, Double-blind, Placebo Controlled, Telemedicine Study \(SynCov Study\)](#). NCT04420676. Clinical trial registration. clinicaltrials.gov.
1972. Biosearch S.A. 2020. [Multicentric Study to Assess the Effect of Consumption of Lactobacillus Coryniformis K8 on Healthcare](#)

[Personnel Exposed to COVID-19](#). NCT04366180. Clinical trial registration. [clinicaltrials.gov](#).

1973. Bioithas SL. 2021. [The Intestinal Microbiota as a Therapeutic Target in Hospitalized Patients With COVID-19 Infection](#). NCT04390477. Clinical trial registration. [clinicaltrials.gov](#).
1974. Reid G. 2016. [Probiotics: definition, scope and mechanisms of action](#). Best Practice & Research Clinical Gastroenterology 30:17–25.
1975. Rijkers GT, de Vos WM, Brummer R-J, Morelli L, Corthier G, Marteau P. 2011. [Health benefits and health claims of probiotics: bridging science and marketing](#). British Journal of Nutrition 106:1291–1296.
1976. Mak JWY, Chan FKL, Ng SC. 2020. [Probiotics and COVID-19: one size does not fit all](#). The Lancet Gastroenterology & Hepatology 5:644–645.
1977. Bloomberg - Are you a robot? <https://www.bloomberg.com/tosv2.html?vid=&uuid=b91b9b90-6a34-11eb-a07d-15fd64b6d7f0&url=L3ByZXNzLXJlbGVhc2VzLzlwMjAtMDgtMDMvchJvYmlvdGljcy1tYXJrZXQtd29ydGgtNzYtNy1iaWxsaW9uLWJ5LTlwMjctZXhjbHVzaXZILXJlcG9ydC1jb3ZlcmluZy1wcmUtYW5kLXBvc3QtY292aWQtMTktbWFya2V0LWFuYWx5c2lzLWJ5LW1ldGljdWxvdXM=>. Retrieved 8 February 2021.
1978. Pereira M, Dantas Damascena A, Galvão Azevedo LM, de Almeida Oliveira T, da Mota Santana J. 2020. [Vitamin D deficiency aggravates COVID-19: systematic review and meta-analysis](#). Critical Reviews in Food Science and Nutrition 1–9.
1979. Galland L. 2010. [Diet and Inflammation](#). Nutrition in Clinical Practice 25:634–640.
1980. Kain V, Van Der Pol W, Mariappan N, Ahmad A, Eipers P, Gibson DL, Gladine C, Vigor C, Durand T, Morrow C, Halade GV. 2019. [Obesogenic diet in aging mice disrupts gut microbe composition and alters neutrophil:lymphocyte ratio, leading to inflamed milieu in acute heart failure](#). The FASEB Journal 33:6456–6469.
1981. Colloidal Silver. NCCIH. <https://www.nccih.nih.gov/health/colloidal-silver>. Retrieved 8 February 2021.
1982. Zhou Y, Hou Y, Shen J, Huang Y, Martin W, Cheng F. 2020. [Network-based drug repurposing for novel coronavirus 2019-nCoV/SARS-CoV-2](#). Cell Discovery 6:14.
1983. Wongchitrat P, Shukla M, Sharma R, Govitrapong P, Reiter RJ. 2021. [Role of Melatonin on Virus-Induced Neuropathogenesis—A Concomitant Therapeutic Strategy to Understand SARS-CoV-2 Infection](#). Antioxidants 10:47.
1984. McCarty MF, Iloki Assanga SB, Lewis Luján L, O’Keefe JH, DiNicolantonio JJ. 2020. [Nutraceutical Strategies for Suppressing NLRP3 Inflammasome Activation: Pertinence to the Management of COVID-19 and Beyond](#). Nutrients 13:47.

1985. Cohen J. 2020. Update: Here's what is known about Trump's COVID-19 treatment. Science <https://doi.org/10.1126/science.abf0974>.
1986. Louca P, Murray B, Klaser K, Graham MS, Mazidi M, Leeming ER, Thompson E, Bowyer R, Drew DA, Nguyen LH, Merino J, Gomez M, Mompeo O, Costeira R, Sudre CH, Gibson R, Steves CJ, Wolf J, Franks PW, Ourselin S, Chan AT, Berry SE, Valdes AM, Calder PC, Spector TD, Menni C. 2020. Dietary supplements during the COVID-19 pandemic: insights from 1.4M users of the COVID Symptom Study app - a longitudinal app-based community survey. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.11.27.20239087>.
1987. Barazzoni R, Bischoff SC, Breda J, Wickramasinghe K, Krznaric Z, Nitzan D, Pirlich M, Singer P. 2020. [ESPEN expert statements and practical guidance for nutritional management of individuals with SARS-CoV-2 infection](#). Clinical Nutrition 39:1631–1638.
1988. Haraj NE, El Aziz S, Chadli A, Dafir A, Mjabber A, Aissaoui O, Barrou L, El Kettani El Hamidi C, Nsiri A, AL Harrar R, Ezzouine H, Charra B, Abdallaoui MS, El Kebbaï N, Kamal N, Bennouna GM, El Filali KM, Ramdani B, El Mdaghri N, Gharbi MB, Afif MH. 2021. [Nutritional status assessment in patients with Covid-19 after discharge from the intensive care unit](#). Clinical Nutrition ESPEN 41:423–428.
1989. Berger MM. 2020. [Nutrition Status Affects COVID-19 Patient Outcomes](#). Journal of Parenteral and Enteral Nutrition 44:1166–1167.
1990. Zhao X, Li Y, Ge Y, Shi Y, Lv P, Zhang J, Fu G, Zhou Y, Jiang K, Lin N, Bai T, Jin R, Wu Y, Yang X, Li X. 2020. [Evaluation of Nutrition Risk and Its Association With Mortality Risk in Severely and Critically Ill COVID-19 Patients](#). Journal of Parenteral and Enteral Nutrition 45:32–42.
1991. Ahmed M, Advani S, Moreira A, Zoretic S, Martinez J, Chorath K, Acosta S, Naqvi R, Burmeister-Morton F, Burmeister F, Tarriela A, Petershack M, Evans M, Hoang A, Rajasekaran K, Ahuja S, Moreira A. 2020. [Multisystem inflammatory syndrome in children: A systematic review](#). EClinicalMedicine 26:100527.
1992. Brugliera L, Spina A, Castellazzi P, Cimino P, Arcuri P, Negro A, Houdayer E, Alemanno F, Giordani A, Mortini P, Iannaccone S. 2020. [Nutritional management of COVID-19 patients in a rehabilitation unit](#). European Journal of Clinical Nutrition 74:860–863.
1993. Domínguez Díaz L, Fernández-Ruiz V, Cámara M. 2019. [The frontier between nutrition and pharma: The international regulatory framework of functional foods, food supplements and nutraceuticals](#). Critical Reviews in Food Science and Nutrition 60:1738–1746.
1994. Commissioner O of the. 2020. Coronavirus Update: FDA and FTC Warn Seven Companies Selling Fraudulent Products that Claim to Treat or Prevent COVID-19. FDA. <https://www.fda.gov/news-events/press-announcements/coronavirus-update-fda-and-ftc-warn-seven-companies-selling-fraudulent-products-claim-treat-or>. Retrieved 8 February 2021.

1995. CDC. 2021. COVID-19 and Your Health. Centers for Disease Control and Prevention. <https://www.cdc.gov/coronavirus/2019-ncov/prevent-getting-sick/prevention.html>. Retrieved 8 February 2021.
1996. Park SW, Sun K, Viboud C, Grenfell BT, Dushoff J. 2020. Potential roles of social distancing in mitigating the spread of coronavirus disease 2019 (COVID-19) in South Korea. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.27.20045815>.
1997. Matrajt L, Leung T. 2020. [Evaluating the Effectiveness of Social Distancing Interventions to Delay or Flatten the Epidemic Curve of Coronavirus Disease](#). Emerging Infectious Diseases 26:1740–1748.
1998. Yehia BR, Winegar A, Fogel R, Fakih M, Ottenbacher A, Jesser C, Bufalino A, Huang R-H, Cacchione J. 2020. [Association of Race With Mortality Among Patients Hospitalized With Coronavirus Disease 2019 \(COVID-19\) at 92 US Hospitals](#). JAMA Network Open 3:e2018039.
1999. Wu Z, McGoogan JM. 2020. [Characteristics of and Important Lessons From the Coronavirus Disease 2019 \(COVID-19\) Outbreak in China](#). JAMA 323:1239.
2000. Grasselli G, Pesenti A, Cecconi M. 2020. [Critical Care Utilization for the COVID-19 Outbreak in Lombardy, Italy](#). JAMA 323:1545.
2001. Garg S, Kim L, Whitaker M, O'Halloran A, Cummings C, Holstein R, Prill M, Chai SJ, Kirley PD, Alden NB, Kawasaki B, Yousey-Hindes K, Niccolai L, Anderson EJ, Openo KP, Weigel A, Monroe ML, Ryan P, Henderson J, Kim S, Como-Sabetti K, Lynfield R, Sosin D, Torres S, Muse A, Bennett NM, Billing L, Sutton M, West N, Schaffner W, Talbot HK, Aquino C, George A, Budd A, Brammer L, Langley G, Hall AJ, Fry A. 2020. [Hospitalization Rates and Characteristics of Patients Hospitalized with Laboratory-Confirmed Coronavirus Disease 2019 — COVID-NET, 14 States, March 1–30, 2020](#). MMWR Morbidity and Mortality Weekly Report 69:458–464.
2002. CDC. 2020. COVID-19 and Your Health. Centers for Disease Control and Prevention. <https://www.cdc.gov/coronavirus/2019-ncov/need-extra-precautions/older-adults.html>. Retrieved 8 February 2021.
2003. Azar KMJ, Shen Z, Romanelli RJ, Lockhart SH, Smits K, Robinson S, Brown S, Pressman AR. 2020. [Disparities In Outcomes Among COVID-19 Patients In A Large Health Care System In California](#). Health Affairs 39:1253–1262.
2004. Killerby ME, Link-Gelles R, Haight SC, Schrodtt CA, England L, Gomes DJ, Shamout M, Pettrone K, O'Laughlin K, Kimball A, Blau EF, Burnett E, Ladvá CN, Szablewski CM, Tobin-D'Angelo M, Oosmanally N, Drenzek C, Murphy DJ, Blum JM, Hollberg J, Lefkove B, Brown FW, Shimabukuro T, Midgley CM, Tate JE, Browning SD, Bruce BB, da Silva J, Gold JAW, Jackson BR, Bamrah Morris S, Natarajan P, Neblett Fanfair R, Patel PR, Rogers-Brown J, Rossow J, Wong KK, CDC COVID-19 Response Clinical Team, CDC COVID-19 Response Clinical Team. 2020. [Characteristics Associated with Hospitalization Among Patients with COVID-19 —](#)

[Metropolitan Atlanta, Georgia, March–April 2020](#). MMWR Morbidity and Mortality Weekly Report 69:790–794.

2005. Dowd JB, Andriano L, Brazel DM, Rotondi V, Block P, Ding X, Liu Y, Mills MC. 2020. [Demographic science aids in understanding the spread and fatality rates of COVID-19](#). Proceedings of the National Academy of Sciences 117:9696–9698.
2006. Shahid Z, Kalayanamitra R, McClafferty B, Kepko D, Ramgobin D, Patel R, Aggarwal CS, Vunnam R, Sahu N, Bhatt D, Jones K, Golamari R, Jain R. 2020. [-19 and Older Adults: What We Know](#). Journal of the American Geriatrics Society 68:926–929.
2007. Docherty AB, Harrison EM, Green CA, Hardwick HE, Pius R, Norman L, Holden KA, Read JM, Dondelinger F, Carson G, Merson L, Lee J, Plotkin D, Sigfrid L, Halpin S, Jackson C, Gamble C, Horby PW, Nguyen-Van-Tam JS, Ho A, Russell CD, Dunning J, Openshaw PJ, Baillie JK, Semple MG. 2020. [Features of 20 133 UK patients in hospital with covid-19 using the ISARIC WHO Clinical Characterisation Protocol: prospective observational cohort study](#). BMJ m1985.
2008. Gebhard C, Regitz-Zagrosek V, Neuhauser HK, Morgan R, Klein SL. 2020. [Impact of sex and gender on COVID-19 outcomes in Europe](#). Biology of Sex Differences 11:29.
2009. The Sex, Gender and COVID-19 Project | Global Health 50/50. <https://globalhealth5050.org/the-sex-gender-and-covid-19-project/>. Retrieved 8 February 2021.
2010. Klein SL, Dhakal S, Ursin RL, Deshpande S, Sandberg K, Mauvais-Jarvis F. 2020. [Biological sex impacts COVID-19 outcomes](#). PLOS Pathogens 16:e1008570.
2011. Meng Y, Wu P, Lu W, Liu K, Ma K, Huang L, Cai J, Zhang H, Qin Y, Sun H, Ding W, Gui L, Wu P. 2020. [Sex-specific clinical characteristics and prognosis of coronavirus disease-19 infection in Wuhan, China: A retrospective study of 168 severe patients](#). PLOS Pathogens 16:e1008520.
2012. Liu J, Ji H, Zheng W, Wu X, Zhu JJ, Arnold AP, Sandberg K. 2010. [Sex differences in renal angiotensin converting enzyme 2 \(ACE2\) activity are 17 \$\beta\$ -oestradiol-dependent and sex chromosome-independent](#). Biology of Sex Differences 1:6.
2013. Fallon A, Dukelow T, Kennelly SP, O'Neill D. 2020. [COVID-19 in nursing homes](#). QJM: An International Journal of Medicine 113:391–392.
2014. Herman JL, O'Neill K. 2020. [Vulnerabilities to COVID-19 Among Transgender Adults in the U.S.](#)
2015. Zhu L, She Z-G, Cheng X, Qin J-J, Zhang X-J, Cai J, Lei F, Wang H, Xie J, Wang W, Li H, Zhang P, Song X, Chen X, Xiang M, Zhang C, Bai L, Xiang D, Chen M-M, Liu Y, Yan Y, Liu M, Mao W, Zou J, Liu L, Chen G, Luo P, Xiao B, Zhang C, Zhang Z, Lu Z, Wang J, Lu H, Xia X, Wang D, Liao X, Peng G, Ye P, Yang J, Yuan Y, Huang X, Guo J, Zhang B-H, Li H. 2020. [Association of Blood Glucose Control and Outcomes in Patients with](#)

[COVID-19 and Pre-existing Type 2 Diabetes](#). Cell Metabolism 31:1068–1077.e3.

2016. Wu Z, Tang Y, Cheng Q. 2020. Diabetes increases the mortality of patients with COVID-19: a meta-analysis. Acta Diabetologica <https://doi.org/10.1007/s00592-020-01546-0>.
2017. Li J, Wang X, Chen J, Zuo X, Zhang H, Deng A. 2020. COVID-19 infection may cause ketosis and ketoacidosis. Diabetes, Obesity and Metabolism <https://doi.org/10.1111/dom.14057>.
2018. Muniyappa R, Gubbi S. 2020. [COVID-19 pandemic, coronaviruses, and diabetes mellitus](#). American Journal of Physiology-Endocrinology and Metabolism 318:E736–E741.
2019. Palaiodimos L, Kokkinidis DG, Li W, Karamanis D, Ognibene J, Arora S, Southern WN, Mantzoros CS. 2020. [Severe obesity, increasing age and male sex are independently associated with worse in-hospital outcomes, and higher in-hospital mortality, in a cohort of patients with COVID-19 in the Bronx, New York](#). Metabolism 108:154262.
2020. Docherty A, Harrison E, Green C, Hardwick H, Pius R, Norman L, Holden K, Read J, Dondelinger F, Carson G, Merson L, Lee J, Plotkin D, Sigfrid L, Halpin S, Jackson C, Gamble C, Horby P, Nguyen-Van-Tam J, Dunning J, Openshaw P, Baillie J, Semple M, ISARIC4C Investigators. 2020. Features of 16,749 hospitalised UK patients with COVID-19 using the ISARIC WHO Clinical Characterisation Protocol. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.04.23.20076042>.
2021. Lockhart SM, O’Rahilly S. 2020. [When Two Pandemics Meet: Why Is Obesity Associated with Increased COVID-19 Mortality?](#) Med 1:33–42.
2022. Nepomuceno MR, Acosta E, Alburez-Gutierrez D, Aburto JM, Gagnon A, Turra CM. 2020. [Besides population age structure, health and other demographic factors can contribute to understanding the COVID-19 burden](#). Proceedings of the National Academy of Sciences 117:13881–13883.
2023. . <https://www.who.int/publications/i/item/report-of-the-who-china-joint-mission-on-coronavirus-disease-2019>.
2024. Vincent J-L, Moreno R, Takala J, Willatts S, De Mendonça A, Bruining H, Reinhart CK, Suter PM, Thijs LG. 1996. [The SOFA \(Sepsis-related Organ Failure Assessment\) score to describe organ dysfunction/failure](#). Intensive Care Medicine 22:707–710.
2025. Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M, Bellomo R, Bernard GR, Chiche J-D, Coopersmith CM, Hotchkiss RS, Levy MM, Marshall JC, Martin GS, Opal SM, Rubenfeld GD, van der Poll T, Vincent J-L, Angus DC. 2016. [The Third International Consensus Definitions for Sepsis and Septic Shock \(Sepsis-3\)](#). JAMA 315:801.
2026. Yancy CW. 2020. [COVID-19 and African Americans](#). JAMA 323:1891.

2027. Webb Hooper M, Nápoles AM, Pérez-Stable EJ. 2020. [COVID-19 and Racial/Ethnic Disparities](#). JAMA 323:2466.
2028. Dyer O. 2020. [Covid-19: Black people and other minorities are hardest hit in US](#). BMJ m1483.
2029. Kakol M, Upson D, Sood A. 2020. [Susceptibility of Southwestern American Indian Tribes to Coronavirus Disease 2019 \(COVID-19\)](#). The Journal of Rural Health 37:197–199.
2030. Jr RAO, Gebeloff R, Lai KKR, Wright W, Smith M. 2020. [The Fullest Look Yet at the Racial Inequity of Coronavirus](#). The New York Times.
2031. Wang ML, Behrman P, Dulin A, Baskin ML, Buscemi J, Alcaraz KI, Goldstein CM, Carson TL, Shen M, Fitzgibbon M. 2020. [Addressing inequities in COVID-19 morbidity and mortality: research and policy recommendations](#). Translational Behavioral Medicine 10:516–519.
2032. Parkhurst NAD, Huyser KR, Horse AJY. 2020. [Historical Environmental Racism, Structural Inequalities, and Dik'os Ntsaaígíí-19 \(COVID-19\) on Navajo Nation](#). Journal of Indigenous Social Development 9:127–140.
2033. Ferrante L, Fearnside PM. 2020. [Protect Indigenous peoples from COVID-19](#). Science 368:251.1–251.
2034. Williamson EJ, Walker AJ, Bhaskaran K, Bacon S, Bates C, Morton CE, Curtis HJ, Mehrkar A, Evans D, Inglesby P, Cockburn J, McDonald HI, MacKenna B, Tomlinson L, Douglas IJ, Rentsch CT, Mathur R, Wong AYS, Grieve R, Harrison D, Forbes H, Schultze A, Croker R, Parry J, Hester F, Harper S, Perera R, Evans SJW, Smeeth L, Goldacre B. 2020. [Factors associated with COVID-19-related death using OpenSAFELY](#). Nature 584:430–436.
2035. Tishkoff SA, Kidd KK. 2004. [Implications of biogeography of human populations for 'race' and medicine](#). Nature Genetics 36:S21–S27.
2036. Campbell MC, Tishkoff SA. 2008. [African Genetic Diversity: Implications for Human Demographic History, Modern Human Origins, and Complex Disease Mapping](#). Annual Review of Genomics and Human Genetics 9:403–433.
2037. Yudell M, Roberts D, DeSalle R, Tishkoff S, 70 signatories. 2020. [NIH must confront the use of race in science](#). Science 369:1313–1314.
2038. Kuo C-L, Pilling LC, Atkins JL, Masoli JAH, Delgado J, Kuchel GA, Melzer D. 2020. [APOE e4 Genotype Predicts Severe COVID-19 in the UK Biobank Community Cohort](#). The Journals of Gerontology: Series A 75:2231–2232.
2039. Wei J, Alfajaro MM, Hanna RE, DeWeirdt PC, Strine MS, Lu-Culligan WJ, Zhang S-M, Graziano VR, Schmitz CO, Chen JS, Mankowski MC, Filler RB, Gasque V, de Miguel F, Chen H, Oguntuyo K, Abriola L, Surovtseva YV, Orchard RC, Lee B, Lindenbach B, Politi K, van Dijk D, Simon MD, Yan Q, Doench JG, Wilen CB. 2020. Genome-wide CRISPR screen reveals host genes that regulate SARS-CoV-2 infection. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.06.16.155101>.

2040. Hou Y, Zhao J, Martin W, Kallianpur A, Chung MK, Jehi L, Sharifi N, Erzurum S, Eng C, Cheng F. 2020. [New insights into genetic susceptibility of COVID-19: an ACE2 and TMPRSS2 polymorphism analysis](#). BMC Medicine 18:216.
2041. Uyoga S, M.O. Adetifa I, Karanja HK, Nyagwange J, Tuju J, Wanjiku P, Aman R, Mwangangi M, Amoth P, Kasera K, Ng'ang'a W, Rombo C, Yegon C, Kithi K, Odhiambo E, Rotich T, Orgut I, Kihara S, Otiende M, Bottomley C, N. Mupe Z, W. Kagucia E, E Gallagher K, Etyang A, Voller S, N. Gitonga J, Mugo D, N. Agoti C, Otieno E, Ndwiga L, Lambe T, Wright D, Barasa E, Tsofa B, Bejon P, I. Ochola-Oyier L, Agweyu A, Scott JAG, Warimwe GM. 2020. Seroprevalence of anti-SARS-CoV-2 IgG antibodies in Kenyan blood donors. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.07.27.20162693>.
2042. Chibwana MG, Jere KC, Kamn'gona R, Mandolo J, Katunga-Phiri V, Tembo D, Mitole N, Musasa S, Sichone S, Lakudzala A, Sibale L, Matambo P, Kadwala I, Byrne RL, Mbewe A, Morton B, Phiri C, Mallewa J, Mwandumba HC, Adams ER, Gordon SB, Jambo KC. 2020. High SARS-CoV-2 seroprevalence in Health Care Workers but relatively low numbers of deaths in urban Malawi. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.07.30.20164970>.
2043. Nordling L. 2020. [Africa's pandemic puzzle: why so few cases and deaths?](#) Science 369:756–757.
2044. Are some ethnic groups more vulnerable to COVID-19 than others? <https://www.ifs.org.uk/inequality/chapter/are-some-ethnic-groups-more-vulnerable-to-covid-19-than-others/>. Retrieved 8 February 2021.
2045. Dasgupta N, Funk MJ, Lazard A, White BE, Marshall SW. 2020. Quantifying the social distancing privilege gap: a longitudinal study of smartphone movement. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.05.03.20084624>.
2046. Fraiberger SP, Astudillo P, Candeago L, Chune A, Jones NKW, Khan MF, Lepri B, Gracia NL, Lucchini L, Massaro E, Montfort A. 2020. [Uncovering socioeconomic gaps in mobility reduction during the COVID-19 pandemic using location data](#). 2006.15195arXiv. arXiv.
2047. Chang S, Pierson E, Koh PW, Gerardin J, Redbird B, Grusky D, Leskovec J. 2020. [Mobility network models of COVID-19 explain inequities and inform reopening](#). Nature 589:82–87.
2048. Brown HJRF. 2020. A Basic Demographic Profile of Workers in Frontline Industries. <https://mronline.org/wp-content/uploads/2020/06/2020-04-Frontline-Workers.pdf>.
2049. Hawkins D. 2020. [Differential occupational risk for COVID-19 and other infection exposure according to race and ethnicity](#). American Journal of Industrial Medicine 63:817–820.
2050. Baker MG, Peckham TK, Seixas NS. 2020. [Estimating the burden of United States workers exposed to infection or disease: A key factor in containing risk of COVID-19 infection](#). PLOS ONE 15:e0232452.

2051. Windsor-Shellard B, Kaur J. 2020. Coronavirus (COVID-19) related deaths by occupation, England and Wales: deaths registered up to and including 20 April 2020.
<https://www.ons.gov.uk/peoplepopulationandcommunity/healthandsocialcare/causesofdeath/bulletins/coronaviruscovid19relateddeathsbyoccupationenglandandwales/deathsregistereduptoandincluding20april2020>.
2052. Office for National Statistics. 2020. Which occupations have the highest potential exposure to the coronavirus (COVID-19)?
<https://www.ons.gov.uk/employmentandlabourmarket/peopleinwork/employmentandemployeetypes/articles/whichoccupationshavethehighestpotentialexposuretothecoronaviruscovid19/2020-05-11>.
2053. Public Health England. 2020. Disparities in the risk and outcomes from COVID-19.
https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/892085/disparities_review.pdf.
2054. Cook T, Kursumovic E, Lennane. 2020-04-22T12:42:00+01:00 S. Exclusive: deaths of NHS staff from covid-19 analysed. Health Service Journal. <https://www.hsj.co.uk/exclusive-deaths-of-nhs-staff-from-covid-19-analysed/7027471.article>. Retrieved 8 February 2021.
2055. . <https://act.nationalnursesunited.org/page>.
2056. McLaren J. 2020. [Racial Disparity in COVID-19 Deaths: Seeking Economic Roots with Census data](#). w27407. National Bureau of Economic Research.
2057. Barnett ML, Hu L, Martin T, Grabowski DC. 2020. [Mortality, Admissions, and Patient Census at SNFs in 3 US Cities During the COVID-19 Pandemic](#). JAMA 324:507.
2058. Hawks L, Woolhandler S, McCormick D. 2020. [COVID-19 in Prisons and Jails in the United States](#). JAMA Internal Medicine 180:1041.
2059. Saloner B, Parish K, Ward JA, DiLaura G, Dolovich S. 2020. [COVID-19 Cases and Deaths in Federal and State Prisons](#). JAMA 324:602.
2060. nlizanna. 2018. State Rates of Incarceration Race & Ethnicity_updated2 . <https://www.issuelab.org/resources/695/695.pdf>.
2061. Keene JR, Batson CD. 2010. [Under One Roof: A Review of Research on Intergenerational Coresidence and Multigenerational Households in the United States](#). Sociology Compass 4:642–657.
2062. Evans GW, Eckenrode J, Marcynyszyn LA. 2010. [Chaos and the macrosetting: The role of poverty and socioeconomic status](#), p. 225–238. In. American Psychological Association (APA).
2063. Curtis MA, Geller AB. 2010. Housing insecurity among urban fathers. Columbia University <https://doi.org/10.7916/d8wh2w9t>.
2064. Desmond M, Gershenson C. 2016. [Housing and Employment Insecurity among the Working Poor](#). Social Problems 63:46–67.

2065. Khaodhiar L, McCowen KC, Blackburn GL. 1999. [Obesity and its comorbid conditions](#). Clinical Cornerstone 2:17–31.
2066. Mauvais-Jarvis F. 2020. [Aging, Male Sex, Obesity, and Metabolic Inflammation Create the Perfect Storm for COVID-19](#). Diabetes 69:1857–1863.
2067. Mendenhall E, Kohrt BA, Norris SA, Ndeti D, Prabhakaran D. 2017. [Non-communicable disease syndemics: poverty, depression, and diabetes among low-income populations](#). The Lancet 389:951–963.
2068. Żukiewicz-Sobczak W, Wróblewska P, Zwoliński J, Chmielewska-Badora J, Adamczuk P, Krasowska E, Zagórski J, Oniszcuk A, Piątek J, Silny W. 2014. [Obesity and poverty paradox in developed countries](#). Annals of Agricultural and Environmental Medicine 21:590–594.
2069. Ashby NJS. 2020. [Impact of the COVID-19 Pandemic on Unhealthy Eating in Populations with Obesity](#). Obesity 28:1802–1805.
2070. Myers CA, Broyles ST. 2020. [Fast Food Patronage and Obesity Prevalence During the COVID-19 Pandemic: An Alternative Explanation](#). Obesity 28:1796–1797.
2071. Huizar MI, Arena R, Laddu DR. 2020. [The global food syndemic: The impact of food insecurity, Malnutrition and obesity on the healthspan amid the COVID-19 pandemic](#). Progress in Cardiovascular Diseases S0033062020301390.
2072. Chrousos GP. 2000. [Stress, chronic inflammation, and emotional and physical well-being: Concurrent effects and chronic sequelae](#). Journal of Allergy and Clinical Immunology 106:S275–S291.
2073. Miller GE, Cohen S, Ritchey AK. 2002. [Chronic psychological stress and the regulation of pro-inflammatory cytokines: A glucocorticoid-resistance model](#). Health Psychology 21:531–541.
2074. Gouin J-P, Glaser R, Malarkey WB, Beversdorf D, Kiecolt-Glaser J. 2012. [Chronic stress, daily stressors, and circulating inflammatory markers](#). Health Psychology 31:264–268.
2075. Miller GE, Blackwell E. 2016. [Turning Up the Heat](#). Current Directions in Psychological Science 15:269–272.
2076. Sapolsky R. 2005. [Sick of Poverty](#). Scientific American 293:92–99.
2077. Wu X, Nethery RC, Sabath MB, Braun D, Dominici F. 2020. Exposure to air pollution and COVID-19 mortality in the United States: A nationwide cross-sectional study. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.04.05.20054502>.
2078. Burnett R, Chen H, Szyszkowicz M, Fann N, Hubbell B, Pope CA, Apte JS, Brauer M, Cohen A, Weichenthal S, Coggins J, Di Q, Brunekreef B, Frostad J, Lim SS, Kan H, Walker KD, Thurston GD, Hayes RB, Lim CC, Turner MC, Jerrett M, Krewski D, Gapstur SM, Diver WR, Ostro B, Goldberg D, Crouse DL, Martin RV, Peters P, Pinault L, Tjepkema M, van Donkelaar A, Villeneuve PJ, Miller AB, Yin P, Zhou M, Wang L, Janssen

- NAH, Marra M, Atkinson RW, Tsang H, Quoc Thach T, Cannon JB, Allen RT, Hart JE, Laden F, Cesaroni G, Forastiere F, Weinmayr G, Jaensch A, Nagel G, Concin H, Spadaro JV. 2018. [Global estimates of mortality associated with long-term exposure to outdoor fine particulate matter](#). *Proceedings of the National Academy of Sciences* 115:9592–9597.
2079. Kravitz-Wirtz N, Teixeira S, Hajat A, Woo B, Crowder K, Takeuchi D. 2018. [Early-Life Air Pollution Exposure, Neighborhood Poverty, and Childhood Asthma in the United States, 1990–2014](#). *International Journal of Environmental Research and Public Health* 15:1114.
2080. Olvera Alvarez HA, Kubzansky LD, Campen MJ, Slavich GM. 2018. [Early life stress, air pollution, inflammation, and disease: An integrative review and immunologic model of social-environmental adversity and lifespan health](#). *Neuroscience & Biobehavioral Reviews* 92:226–242.
2081. Belanger MJ, Hill MA, Angelidi AM, Dalamaga M, Sowers JR, Mantzoros CS. 2020. [Covid-19 and Disparities in Nutrition and Obesity](#). *New England Journal of Medicine* 383:e69.
2082. Gravlee CC. 2020. [Systemic racism, chronic health inequities, and -19: A syndemic in the making?](#) *American Journal of Human Biology* 32.
2083. Lopez L III, Hart LH III, Katz MH. 2021. [Racial and Ethnic Health Disparities Related to COVID-19](#). *JAMA* 325:719.
2084. Chavez S, Long B, Koyfman A, Liang SY. 2020. [Coronavirus Disease \(COVID-19\): A primer for emergency physicians](#). *The American Journal of Emergency Medicine* S0735675720301789.
2085. Insurers May Only Pay For Coronavirus Tests When They're 'Medically Necessary'. NPR.org. <https://www.npr.org/sections/health-shots/2020/06/19/880543755/insurers-may-only-pay-for-coronavirus-tests-when-theyre-medically-necessary>. Retrieved 8 February 2021.
2086. Private Health Insurance Coverage in the COVID-19 Public Health Emergency | Commonwealth Fund. <https://www.commonwealthfund.org/blog/2020/private-health-insurance-coverage-covid-19-public-health-emergency>. Retrieved 8 February 2021.
2087. Shah M, Sachdeva M, Dodiuk-Gad RP. 2020. [COVID-19 and racial disparities](#). *Journal of the American Academy of Dermatology* 83:e35.
2088. FAQs for COVID-19 Claims Reimbursement to Health Care Providers and Facilities for Testing, Treatment and Vaccine Administration. Official web site of the US Health Resources & Services Administration. Text. <https://www.hrsa.gov/coviduninsuredclaim/frequently-asked-questions>. Retrieved 8 February 2021.
2089. Ji Y, Ma Z, Peppelenbosch MP, Pan Q. 2020. [Potential association between COVID-19 mortality and health-care resource availability](#). *The Lancet Global Health* 8:e480.
2090. Wang Z, Tang K. 2020. [Combating COVID-19: health equity matters](#). *Nature Medicine* 26:458–458.

2091. chen0307. 2020. lockup black .
<https://www.census.gov/content/dam/Census/library/publications/2019/demo/p60-267.pdf>.
2092. Raluca D, Veronica C, Oana MC, Alexandra E. 2020. [An Ethical Dilemma in SARS-Cov-2 Pandemic : Who Gets the Ventilator?](#) European Scientific Journal ESJ 16.
2093. Raffensperger J, Brauner M, Briggs R. 2020. Planning Hospital Needs for Ventilators and Respiratory Therapists in the COVID-19 Crisis. Rand Corporation. <https://doi.org/ghfprp>.
2094. Pathak PA, Sönmez T, Ünver MU, Yenmez MB. 2021. [Fair Allocation of Vaccines, Ventilators and Antiviral Treatments: Leaving No Ethical Value Behind in Health Care Rationing](#). 2008.00374arXiv. arXiv.
2095. Chu Q, Correa R, Henry TL, McGregor KA, Stoklosa H, Robinson L, Jha S, Annamalai A, Hsu BS, Gupta R, Patton DU, Moreno-Walton LA, Butts C, Chai C, Kuy S. 2020. [Reallocating ventilators during the coronavirus disease 2019 pandemic: Is it ethical?](#) Surgery 168:388–391.
2096. Dudzinski DM, Hoisington BY, Brown CE. 2020. [Ethics Lessons From Seattle's Early Experience With COVID-19](#). The American Journal of Bioethics 20:67–74.
2097. Farrell TW, Francis L, Brown T, Ferrante LE, Widera E, Rhodes R, Rosen T, Hwang U, Witt LJ, Thothala N, Liu SW, Vitale CA, Braun UK, Stephens C, Saliba D. 2020. [Rationing Limited Healthcare Resources in the COVID-19 Era and Beyond: Ethical Considerations Regarding Older Adults](#). Journal of the American Geriatrics Society 68:1143–1149.
2098. Haward MF, Moore GP, Lantos J, Janvier A. 2020. [Paediatric ethical issues during the COVID-19 pandemic are not just about ventilator triage](#). Acta Paediatrica 109:1519–1521.
2099. McGuire AL, Aulisio MP, Davis FD, Erwin C, Harter TD, Jagsi R, Klitzman R, Macauley R, Racine E, Wolf SM, Wynia M, Wolpe PR, The COVID-19 Task Force of the Association of Bioethics Program Directors (ABPD). 2020. [Ethical Challenges Arising in the COVID-19 Pandemic: An Overview from the Association of Bioethics Program Directors \(ABPD\) Task Force](#). The American Journal of Bioethics 20:15–27.
2100. Sabatello M, Burke TB, McDonald KE, Appelbaum PS. 2020. [Disability, Ethics, and Health Care in the COVID-19 Pandemic](#). American Journal of Public Health 110:1523–1527.
2101. Wicclair M. 2020. [Allocating Ventilators During the COVID-19 Pandemic and Conscientious Objection](#). The American Journal of Bioethics 20:204–207.
2102. Williams JC, Anderson N, Mathis M, Sanford E, Eugene J, Isom J. 2020. [Colorblind Algorithms: Racism in the Era of COVID-19](#). Journal of the National Medical Association 112:550–552.
2103. Egede LE, Walker RJ. 2020. [Structural Racism, Social Risk Factors, and Covid-19 — A Dangerous Convergence for Black Americans](#). New

England Journal of Medicine 383:e77.

2104. Weber E, Bliton MJ. 2020. [Allocating Remdesivir Under Scarcity: Social Justice or More Systemic Racism](#). The American Journal of Bioethics 20:31–33.
2105. Ballantyne A, Rogers WA, Entwistle V, Towns C. 2020. [Revisiting the equity debate in COVID-19: ICU is no panacea](#). Journal of Medical Ethics 46:641–645.
2106. Fink S. 2020. [Ethical Dilemmas in Covid-19 Medical Care: Is a Problematic Triage Protocol Better or Worse than No Protocol at All?](#) The American Journal of Bioethics 20:1–5.
2107. Lim S, DeBruin DA, Leider JP, Sederstrom N, Lynfield R, Baker JV, Kline S, Kesler S, Rizza S, Wu J, Sharp RR, Wolf SM. 2020. [Developing an Ethics Framework for Allocating Remdesivir in the COVID-19 Pandemic](#). Mayo Clinic Proceedings 95:1946–1954.
2108. Webb J, Shah LD, Lynch HF. 2020. [Ethically Allocating COVID-19 Drugs Via Pre-approval Access and Emergency Use Authorization](#). The American Journal of Bioethics 20:4–17.
2109. 2011. https://www.cdc.gov/about/advisory/pdf/VentDocument_Release.pdf.
2110. Jefferson AA. 2020. [Adopting an Anti-Racist Model of COVID-19 Drug Allocation and Prioritization](#). The American Journal of Bioethics 20:33–36.
2111. Coleman CH. 2020. [Equitably Sharing the Benefits and Burdens of Research: Covid-19 Raises the Stakes](#). Ethics & Human Research 42:38–40.
2112. Yamey G, Schäferhoff M, Hatchett R, Pate M, Zhao F, McDade KK. 2020. [Ensuring global access to COVID-19 vaccines](#). The Lancet 395:1405–1406.
2113. Bollyky TJ, Gostin LO, Hamburg MA. 2020. [The Equitable Distribution of COVID-19 Therapeutics and Vaccines](#). JAMA 323:2462.
2114. Baquet CR, Commiskey P, Daniel Mullins C, Mishra SI. 2006. [Recruitment and participation in clinical trials: Socio-demographic, rural/urban, and health care access predictors](#). Cancer Detection and Prevention 30:24–33.
2115. Makoni M. 2020. [COVID-19 vaccine trials in Africa](#). The Lancet Respiratory Medicine 8:e79–e80.
2116. Langford AT, Resnicow K, Dimond EP, Denicoff AM, Germain DSt, McCaskill-Stevens W, Enos RA, Carrigan A, Wilkinson K, Go RS. 2014. [Racial/ethnic differences in clinical trial enrollment, refusal rates, ineligibility, and reasons for decline among patients at sites in the National Cancer Institute's Community Cancer Centers Program](#). Cancer 120:877–884.

2117. Murthy VH, Krumholz HM, Gross CP. 2004. [Participation in Cancer Clinical Trials](#). JAMA 291:2720.
2118. Stewart JH, Bertoni AG, Staten JL, Levine EA, Gross CP. 2007. [Participation in Surgical Oncology Clinical Trials: Gender-, Race/Ethnicity-, and Age-based Disparities](#). Annals of Surgical Oncology 14:3328–3334.
2119. Geller SE, Koch A, Pellettieri B, Carnes M. 2011. [Inclusion, Analysis, and Reporting of Sex and Race/Ethnicity in Clinical Trials: Have We Made Progress?](#) Journal of Women's Health 20:315–320.
2120. Falasinnu T, Chaichian Y, Bass MB, Simard JF. 2018. [The Representation of Gender and Race/Ethnic Groups in Randomized Clinical Trials of Individuals with Systemic Lupus Erythematosus](#). Current Rheumatology Reports 20:20.
2121. Chastain DB, Osae SP, Henao-Martínez AF, Franco-Paredes C, Chastain JS, Young HN. 2020. [Racial Disproportionality in Covid Clinical Trials](#). New England Journal of Medicine 383:e59.
2122. Johnson JL, Bottorff JL, Browne AJ, Grewal S, Hilton BA, Clarke H. 2004. [Othering and Being Othered in the Context of Health Care Services](#). Health Communication 16:255–271.
2123. Maina IW, Belton TD, Ginzberg S, Singh A, Johnson TJ. 2018. [A decade of studying implicit racial/ethnic bias in healthcare providers using the implicit association test](#). Social Science & Medicine 199:219–229.
2124. Dehon E, Weiss N, Jones J, Faulconer W, Hinton E, Sterling S. 2017. [A Systematic Review of the Impact of Physician Implicit Racial Bias on Clinical Decision Making](#). Academic Emergency Medicine 24:895–904.
2125. Penner LA, Dovidio JF, West TV, Gaertner SL, Albrecht TL, Dailey RK, Markova T. 2010. [Aversive racism and medical interactions with Black patients: A field study](#). Journal of Experimental Social Psychology 46:436–440.
2126. Johnson TJ. 2020. [Intersection of Bias, Structural Racism, and Social Determinants With Health Care Inequities](#). Pediatrics 146:e2020003657.
2127. Feldman WB, Hey SP, Kesselheim AS. 2018. [A Systematic Review Of The Food And Drug Administration's 'Exception From Informed Consent' Pathway](#). Health Affairs 37:1605–1614.
2128. Schmidt TA. 2003. [The legacy of the tuskegee syphilis experiments for emergency exception from informed consent](#). Annals of Emergency Medicine 41:79–81.
2129. Al-Arshani S. CDC officials are considering a plan to distribute COVID-19 vaccines to the most vulnerable first — including people of color. Business Insider. <https://www.businessinsider.com/cdc-official-considering-giving-covid-19-vaccine-most-vulnerable-first-2020-10>. Retrieved 8 February 2021.

2130. Federico MJ, Covar RA, Brown EE, Leung DYM, Spahn JD. 2005. [Racial Differences in T-Lymphocyte Response to Glucocorticoids](#). Chest 127:571–578.
2131. Charmandari E, Tsigos C, Chrousos G. 2005. [ENDOCRINOLOGY OF THE STRESS RESPONSE](#). Annual Review of Physiology 67:259–284.
2132. Cohen S, Janicki-Deverts D, Doyle WJ, Miller GE, Frank E, Rabin BS, Turner RB. 2012. [Chronic stress, glucocorticoid receptor resistance, inflammation, and disease risk](#). Proceedings of the National Academy of Sciences 109:5995–5999.
2133. Vlasschaert C, Topf JM, Hiremath S. 2020. [Proliferation of Papers and Preprints During the Coronavirus Disease 2019 Pandemic: Progress or Problems With Peer Review?](#) Advances in Chronic Kidney Disease 27:418–426.
2134. Zarocostas J. 2020. [How to fight an infodemic](#). The Lancet 395:676.
2135. Wang LL, Lo K, Chandrasekhar Y, Reas R, Yang J, Burdick D, Eide D, Funk K, Katsis Y, Kinney R, Li Y, Liu Z, Merrill W, Mooney P, Murdick D, Rishi D, Sheehan J, Shen Z, Stilson B, Wade A, Wang K, Wang NXR, Wilhelm C, Xie B, Raymond D, Weld DS, Etzioni O, Kohlmeier S. 2020. [CORD-19: The COVID-19 Open Research Dataset](#). 2004.10706arXiv. arXiv.
2136. Lever J, Altman RB. 2021. [Analyzing the vast coronavirus literature with CoronaCentral](#). Proc Natl Acad Sci USA 118.
2137. Eysenbach G. 2000. [The impact of preprint servers and electronic publishing on biomedical research](#). Current Opinion in Immunology 12:499–503.
2138. Yeo-Teh NSL, Tang BL. 2020. [An alarming retraction rate for scientific publications on Coronavirus Disease 2019 \(COVID-19\)](#). Accountability in Research 28:47–53.
2139. Abritis A, Marcus A, Oransky I. 2020. [An “alarming” and “exceptionally high” rate of COVID-19 retractions?](#) Accountability in Research 28:58–59.
2140. Agoramoorthy G, Hsu MJ, Shieh P. 2020. [Queries on the COVID-19 quick publishing ethics](#). Bioethics 34:633–634.
2141. Boodman C, Lee S, Bullard J. 2020. [Idle medical students review emerging COVID-19 research](#). Medical Education Online 25.
2142. Brainard J. 2020. Scientists are drowning in COVID-19 papers. Can new tools keep them afloat? Science <https://doi.org/10.1126/science.abc7839>.
2143. Vabret N, Samstein R, Fernandez N, Merad M, The Sinai Immunology Review Project, Trainees, Faculty. 2020. [Advancing scientific knowledge in times of pandemics](#). Nature Reviews Immunology 20:338–338.

2144. Sun J, He W-T, Wang L, Lai A, Ji X, Zhai X, Li G, Suchard MA, Tian J, Zhou J, Veit M, Su S. 2020. [COVID-19: Epidemiology, Evolution, and Cross-Disciplinary Perspectives](#). Trends in Molecular Medicine 26:483–495.
2145. Weissleder R, Lee H, Ko J, Pittet MJ. 2020. [COVID-19 diagnostics in context](#). Science Translational Medicine 12:eabc1931.
2146. Sanders JM, Monogue ML, Jodlowski TZ, Cutrell JB. 2020. Pharmacologic Treatments for Coronavirus Disease 2019 (COVID-19). JAMA <https://doi.org/10.1001/jama.2020.6019>.
2147. Carvalho T. 2020. [COVID-19 Research in Brief: December, 2019 to June, 2020](#). Nature Medicine 26:1152–1153.
2148. Wiersinga WJ, Rhodes A, Cheng AC, Peacock SJ, Prescott HC. 2020. [Pathophysiology, Transmission, Diagnosis, and Treatment of Coronavirus Disease 2019 \(COVID-19\)](#). JAMA 324:782.
2149. Himmelstein DS, Rubineti V, Slochower DR, Hu D, Malladi VS, Greene CS, Gitter A. 2019. [Open collaborative writing with Manubot](#). PLOS Computational Biology 15:e1007128.
2150. Tennant JP, Bielczyk N, Tzovaras BG, Masuzzo P, Steiner T. 2020. [Introducing Massively Open Online Papers \(MOOPs\)](#). KULA 4:1.
2151. Gog JR. 2020. [How you can help with COVID-19 modelling](#). Nature Reviews Physics 2:274–275.
2152. Bell J, LaToza TD, Baldmisi F, Stavrou A. 2017. [Advancing Open Science with Version Control and Blockchains](#) 2017 IEEE/ACM 12th International Workshop on Software Engineering for Science (SE4Science). IEEE.
2153. Vuorre M, Curley JP. 2018. [Curating Research Assets: A Tutorial on the Git Version Control System](#). Advances in Methods and Practices in Psychological Science 1:219–236.
2154. Ram K. 2013. [Git can facilitate greater reproducibility and increased transparency in science](#). Source Code Biol Med 8.
2155. Corcoran M. 2006. [Using the MAARIE Framework To Read the Research Literature](#). American Journal of Occupational Therapy 60:367–368.
2156. Riegelman RK. 2013. Studying a study & testing a test: reading evidence-based health research 6th ed. Wolters Kluwer/Lippincott Williams & Wilkins Heath, Philadelphia.
2157. Hunter JD. 2007. [Matplotlib: A 2D Graphics Environment](#). Computing in Science & Engineering 9:90–95.
2158. Nicholson JM, Mordaunt M, Lopez P, Uppala A, Rosati D, Rodrigues NP, Grabitz P, Rife SC. 2021. [scite: a smart citation index that displays the context of citations and classifies their intent using deep learning](#). Cold Spring Harbor Laboratory.
2159. Perkel JM. 2020. [Synchronized editing: the future of collaborative writing](#). Nature 580:154–155.

2160. Roser M, Ritchie H, Ortiz-Ospina E, Hasell J. 2020. [Coronavirus pandemic \(COVID-19\)](#). Our World in Data.
2161. Nicholson DN, Rubinetti V, Hu D, Thielk M, Hunter LE, Greene CS. 2021. [Linguistic Analysis of the bioRxiv Preprint Landscape](#). Cold Spring Harbor Laboratory.
2162. Tian X, Li C, Huang A, Xia S, Lu S, Shi Z, Lu L, Jiang S, Yang Z, Wu Y, Ying T. 2020. Potent binding of 2019 novel coronavirus spike protein by a SARS coronavirus-specific human monoclonal antibody. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.01.28.923011>.
2163. He X, Zhang L, Ran Q, Wang J, Xiong A, Wu D, Chen F, Li G. 2020. Integrative Bioinformatics Analysis Provides Insight into the Molecular Mechanisms of 2019-nCoV. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.03.20020206>.
2164. Liang W, Feng Z, Rao S, Xiao C, Lin Z-X, Zhang Q, Wei Q. 2020. Diarrhea may be underestimated: a missing link in 2019 novel coronavirus. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.03.20020289>.
2165. Chai X, Hu L, Zhang Y, Han W, Lu Z, Ke A, Zhou J, Shi G, Fang N, Fan J, Cai J, Fan J, Lan F. 2020. Specific ACE2 Expression in Cholangiocytes May Cause Liver Damage After 2019-nCoV Infection. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.03.931766>.
2166. Zhao B, Ni C, Gao R, Wang Y, Yang L, Wei J, Lv T, Liang J, Zhang Q, Xu W, Xie Y, Wang X, Yuan Z, Liang J, Zhang R, Lin X. 2020. Recapitulation of SARS-CoV-2 Infection and Cholangiocyte Damage with Human Liver Organoids. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.16.990317>.
2167. Wang J, Zhao S, Liu M, Zhao Z, Xu Y, Wang P, Lin M, Xu Y, Huang B, Zuo X, Chen Z, Bai F, Cui J, Lew AM, Zhao J, Zhang Y, Luo H-B, Zhang Y. 2020. ACE2 expression by colonic epithelial cells is associated with viral infection, immunity and energy metabolism. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.05.20020545>.
2168. Bao L, Deng W, Huang B, Gao H, Liu J, Ren L, Wei Q, Yu P, Xu Y, Qi F, Qu Y, Li F, Lv Q, Wang W, Xue J, Gong S, Liu M, Wang G, Wang S, Song Z, Zhao L, Liu P, Zhao L, Ye F, Wang H, Zhou W, Zhu N, Zhen W, Yu H, Zhang X, Guo L, Chen L, Wang C, Wang Y, Wang X, Xiao Y, Sun Q, Liu H, Zhu F, Ma C, Yan L, Yang M, Han J, Xu W, Tan W, Peng X, Jin Q, Wu G, Qin C. 2020. The Pathogenicity of SARS-CoV-2 in hACE2 Transgenic Mice. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.07.939389>.
2169. Li Z, Wu M, Yao J, Guo J, Liao X, Song S, Li J, Duan G, Zhou Y, Wu X, Zhou Z, Wang T, Hu M, Chen X, Fu Y, Lei C, Dong H, Xu C, Hu Y, Han M, Zhou Y, Jia H, Chen X, Yan J. 2020. Caution on Kidney Dysfunctions of COVID-19 Patients. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.08.20021212>.
2170. Chu KH, Tsang WK, Tang CS, Lam MF, Lai FM, To KF, Fung KS, Tang HL, Yan WW, Chan HWH, Lai TST, Tong KL, Lai KN. 2005. [Acute renal](#)

[impairment in coronavirus-associated severe acute respiratory syndrome](#). *Kidney International* 67:698–705.

2171. Lin W, Hu L, Zhang Y, Ooi JD, Meng T, Jin P, Ding X, Peng L, Song L, Xiao Z, Ao X, Xiao X, Zhou Q, Xiao P, Fan J, Zhong Y. 2020. Single-cell Analysis of ACE2 Expression in Human Kidneys and Bladders Reveals a Potential Route of 2019-nCoV Infection. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.08.939892>.
2172. Zhu J, Kim J, Xiao X, Wang Y, Luo D, Jiang S, Chen R, Xu L, Zhang H, Moise L, Gutierrez AH, De Groot AS, Xiao G, Schoggins JW, Zhan X, Wang T, Xie Y. 2020. The immune vulnerability landscape of the 2019 Novel Coronavirus, SARS-CoV-2. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.08.939553>.
2173. Arabi YM, Arifi AA, Balkhy HH, Najm H, Aldawood AS, Ghabashi A, Hawa H, Alothman A, Khaldi A, Al Raiy B. 2014. [Clinical Course and Outcomes of Critically Ill Patients With Middle East Respiratory Syndrome Coronavirus Infection](#). *Annals of Internal Medicine* 160:389–397.
2174. Liu J, Liu Y, Xiang P, Pu L, Xiong H, Li C, Zhang M, Tan J, Xu Y, Song R, Song M, Wang L, Zhang W, Han B, Yang L, Wang X, Zhou G, Zhang T, Li B, Wang Y, Chen Z, Wang X. 2020. Neutrophil-to-Lymphocyte Ratio Predicts Severe Illness Patients with 2019 Novel Coronavirus in the Early Stage. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.10.20021584>.
2175. Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, Xie C, Ma K, Shang K, Wang W, Tian D-S. 2020. [Dysregulation of Immune Response in Patients With Coronavirus 2019 \(COVID-19\) in Wuhan, China](#). *Clinical Infectious Diseases* 71:762–768.
2176. Wan S, Yi Q, Fan S, Lv J, Zhang X, Guo L, Lang C, Xiao Q, Xiao K, Yi Z, Qiang M, Xiang J, Zhang B, Chen Y. 2020. Characteristics of lymphocyte subsets and cytokines in peripheral blood of 123 hospitalized patients with 2019 novel coronavirus pneumonia (NCP). Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.10.20021832>.
2177. Liu J, Li S, Liu J, Liang B, Wang X, Li W, Wang H, Tong Q, Yi J, Zhao L, Xiong L, Guo C, Tian J, Luo J, Yao J, Pang R, Shen H, Peng C, Liu T, Zhang Q, Wu J, Xu L, Lu S, Wang B, Weng Z, Han C, Zhu H, Zhou R, Zhou H, Chen X, Ye P, Zhu B, He S, He Y, Jie S, Wei P, Zhang J, Lu Y, Wang W, Zhang L, Li L, Zhou F, Wang J, Dittmer U, Lu M, Hu Y, Yang D, Zheng X. 2020. Longitudinal Characteristics of Lymphocyte Responses and Cytokine Profiles in the Peripheral Blood of SARS-CoV-2 Infected Patients. SSRN Electronic Journal <https://doi.org/10.2139/ssrn.3539682>.
2178. Li J, Li S, Cai Y, Liu Q, Li X, Zeng Z, Chu Y, Zhu F, Zeng F. 2020. Epidemiological and Clinical Characteristics of 17 Hospitalized Patients with 2019 Novel Coronavirus Infections Outside Wuhan, China. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.11.20022053>.

2179. Fan C, Li K, Ding Y, Lu W, Wang J. 2020. ACE2 Expression in Kidney and Testis May Cause Kidney and Testis Damage After 2019-nCoV Infection. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.12.20022418>.
2180. Zhou Y, Fu B, Zheng X, Wang D, Zhao C, Qi Y, Sun R, Tian Z, Xu X, Wei H. 2020. Aberrant pathogenic GM-CSF⁺ T cells and inflammatory CD14⁺ CD16⁺ monocytes in severe pulmonary syndrome patients of a new coronavirus. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.12.945576>.
2181. Qian K, Deng Y, Tai Y-H, Peng J, Peng H, Jiang L-H. 2020. Clinical Characteristics of 2019 Novel Infected Coronavirus Pneumonia: A Systemic Review and Meta-analysis. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.14.20021535>.
2182. Liu J, Li S, Liu J, Liang B, Wang X, Wang H, Li W, Tong Q, Yi J, Zhao L, Xiong L, Guo C, Tian J, Luo J, Yao J, Pang R, Shen H, Peng C, Liu T, Zhang Q, Wu J, Xu L, Lu S, Wang B, Weng Z, Han C, Zhu H, Zhou R, Zhou H, Chen X, Ye P, Zhu B, He S, He Y, Jie S, Wei P, Zhang J, Lu Y, Wang W, Zhang L, Li L, Zhou F, Wang J, Dittmer U, Lu M, Hu Y, Yang D, Zheng X. 2020. Longitudinal characteristics of lymphocyte responses and cytokine profiles in the peripheral blood of SARS-CoV-2 infected patients. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.16.20023671>.
2183. Chen G, Wu D, Guo W, Cao Y, Huang D, Wang H, Wang T, Zhang X, Chen H, Yu H, Zhang X, Zhang M, Wu S, Song J, Chen T, Han M, Li S, Luo X, Zhao J, Ning Q. 2020. Clinical and immunologic features in severe and moderate forms of Coronavirus Disease 2019. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.16.20023903>.
2184. Chen Y, Wei Q, Li R, Gao H, Zhu H, Deng W, Bao L, Tong W, Cong Z, Jiang H, Qin C. 2020. Protection of Rhesus Macaque from SARS-Coronavirus challenge by recombinant adenovirus vaccine. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.17.951939>.
2185. Diao B, Wang C, Tan Y, Chen X, Liu Y, Ning L, Chen L, Li M, Liu Y, Wang G, Yuan Z, Feng Z, Wu Y, Chen Y. 2020. Reduction and Functional Exhaustion of T Cells in Patients with Coronavirus Disease 2019 (COVID-19). Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.18.20024364>.
2186. Li X, Wang L, Yan S, Yang F, Xiang L, Zhu J, Shen B, Gong Z. 2020. Clinical characteristics of 25 death cases with COVID-19: a retrospective review of medical records in a single medical center, Wuhan, China. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.19.20025239>.
2187. Wang L, Li X, Chen H, Yan S, Li Y, Li D, Gong Z. 2020. SARS-CoV-2 infection does not significantly cause acute renal injury: an analysis of 116 hospitalized patients with COVID-19 in a single hospital, Wuhan, China. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.19.20025288>.

2188. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, Wang B, Xiang H, Cheng Z, Xiong Y, Zhao Y, Li Y, Wang X, Peng Z. 2020. [Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China](#). JAMA 323:1061.
2189. Guan W, Ni Z, Hu Y, Liang W, Ou C, He J, Liu L, Shan H, Lei C, Hui DSC, Du B, Li L, Zeng G, Yuen K-Y, Chen R, Tang C, Wang T, Chen P, Xiang J, Li S, Wang J, Liang Z, Peng Y, Wei L, Liu Y, Hu Y, Peng P, Wang J, Liu J, Chen Z, Li G, Zheng Z, Qiu S, Luo J, Ye C, Zhu S, Zhong N. 2020. Clinical characteristics of 2019 novel coronavirus infection in China. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.02.06.20020974>.
2190. Fast E, Altman RB, Chen B. 2020. Potential T-cell and B-cell Epitopes of 2019-nCoV. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.02.19.955484>.
2191. Walls AC, Park Y-J, Tortorici MA, Wall A, McGuire AT, Veessler D. 2020. Structure, function and antigenicity of the SARS-CoV-2 spike glycoprotein. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.02.19.956581>.
2192. Thevarajan I, Nguyen TH, Koutsakos M, Druce J, Caly L, van de Sandt CE, Jia X, Nicholson S, Catton M, Cowie B, Tong SY, Lewin SR, Kedzierska K. 2020. Breadth of concomitant immune responses underpinning viral clearance and patient recovery in a non-severe case of COVID-19. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.02.20.20025841>.
2193. Liao M, Liu Y, Yuan J, Wen Y, Xu G, Zhao J, Chen L, Li J, Wang X, Wang F, Liu L, Zhang S, Zhang Z. 2020. The landscape of lung bronchoalveolar immune cells in COVID-19 revealed by single-cell RNA sequencing. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.02.23.20026690>.
2194. Scharenberg M, Vangeti S, Kekäläinen E, Bergman P, Al-Ameri M, Johansson N, Söndén K, Falck-Jones S, Färnert A, Ljunggren H-G, Michaëlsson J, Smed-Sörensen A, Marquardt N. 2019. [Influenza A Virus Infection Induces Hyperresponsiveness in Human Lung Tissue-Resident and Peripheral Blood NK Cells](#). Frontiers in Immunology 10:1116.
2195. Amanat F, Stadlbauer D, Strohmeier S, Nguyen THO, Chromikova V, McMahon M, Jiang K, Arunkumar GA, Jurczyszak D, Polanco J, Bermudez-Gonzalez M, Kleiner G, Aydiillo T, Miorin L, Fierer D, Lugo LA, Kojic EM, Stoeve J, Liu STH, Cunningham-Rundles C, Felgner PL, Moran T, Garcia-Sastre A, Caplivski D, Cheng A, Kedzierska K, Vapalahti O, Hepojoki JM, Simon V, Krammer F. 2020. A serological assay to detect SARS-CoV-2 seroconversion in humans. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.17.20037713>.
2196. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q, Wang J, Cao B. 2020.

[Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China](#). The Lancet 395:497–506.

2197. Allard B, Panariti A, Martin JG. 2018. [Alveolar Macrophages in the Resolution of Inflammation, Tissue Repair, and Tolerance to Infection](#). Frontiers in Immunology 9:1777.
2198. Huang S, Zhu B, Cheon IS, Goplen NP, Jiang L, Zhang R, Peebles RS, Mack M, Kaplan MH, Limper AH, Sun J. 2019. [PPAR- \$\gamma\$ in Macrophages Limits Pulmonary Inflammation and Promotes Host Recovery following Respiratory Viral Infection](#). Journal of Virology 93:e00030–19.
2199. Pan Y, Ye G, Zeng X, Liu G, Zeng X, Jiang X, Zhao J, Chen L, Guo S, Deng Q, Hong X, Yang Y, Li Y, Wang X. 2020. Can routine laboratory tests discriminate 2019 novel coronavirus infected pneumonia from other community-acquired pneumonia? Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.25.20024711>.
2200. Gong J, Dong H, Xia Q, Huang Z, Wang D, Zhao Y, Liu W, Tu S, Zhang M, Wang Q, Lu F. 2020. Correlation Analysis Between Disease Severity and Inflammation-related Parameters in Patients with COVID-19 Pneumonia. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.25.20025643>.
2201. Herst C, Burkholz S, Sidney J, Sette A, Harris P, Massey S, Brasel T, Cunha-Neto E, Rosa D, Chao W, Carback R, Hodge T, Wang L, Ciotlos S, Lloyd P, Rubsamen R. 2020. An Effective CTL Peptide Vaccine for Ebola Zaire Based on Survivors' CD8+ Targeting of a Particular Nucleocapsid Protein Epitope with Potential Implications for COVID-19 Vaccine Design. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.25.963546>.
2202. Li L, Sun T, He Y, Li W, Fan Y, Zhang J. 2020. Epitope-based peptide vaccine design and target site characterization against novel coronavirus disease caused by SARS-CoV-2. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.25.965434>.
2203. Wenjun W, Xiaoqing L, Sipei W, Puyi L, Liyan H, Yimin L, Linling C, Sibe C, Lingbo N, Yongping L, Jianxing H. 2020. The definition and risks of Cytokine Release Syndrome-Like in 11 COVID-19-Infected Pneumonia critically ill patients: Disease Characteristics and Retrospective Analysis. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.26.20026989>.
2204. Huang Y, Yang R, Xu Y, Gong P. 2020. Clinical characteristics of 36 non-survivors with COVID-19 in Wuhan, China. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.27.20029009>.
2205. Li L, Li S, Xu M, Yu P, Zheng S, Duan Z, Liu J, Chen Y, Li J. 2020. Risk factors related to hepatic injury in patients with corona virus disease 2019. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.28.20028514>.
2206. Chen X, Zhao B, Qu Y, Chen Y, Xiong J, Feng Y, Men D, Huang Q, Liu Y, Yang B, Ding J, Li F. 2020. Detectable serum SARS-CoV-2 viral load (RNAemia) is closely associated with drastically elevated interleukin 6

(IL-6) level in critically ill COVID-19 patients. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.29.20029520>.

2207. Chen W, Ware LB. 2015. [Prognostic factors in the acute respiratory distress syndrome](#). Clinical and Translational Medicine 4.
2208. Tan L, Wang Q, Zhang D, Ding J, Huang Q, Tang Y-Q, Wang Q, Miao H. 2020. Lymphopenia predicts disease severity of COVID-19: a descriptive and predictive study. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.01.20029074>.
2209. Liu T, Zhang J, Yang Y, Ma H, Li Z, Zhang J, Cheng J, Zhang X, Zhao Y, Xia Z, Zhang L, Wu G, Yi J. 2020. The potential role of IL-6 in monitoring severe case of coronavirus disease 2019. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.01.20029769>.
2210. Zhao Z, Xie J, Yin M, Yang Y, He H, Jin T, Li W, Zhu X, Xu J, Zhao C, Li L, Li Y, Mengist HM, Zahid A, Yao Z, Ding C, Qi Y, Gao Y, Ma X. 2020. Clinical and Laboratory Profiles of 75 Hospitalized Patients with Novel Coronavirus Disease 2019 in Hefei, China. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.01.20029785>.
2211. Yang Y, Shen C, Li J, Yuan J, Yang M, Wang F, Li G, Li Y, Xing L, Peng L, Wei J, Cao M, Zheng H, Wu W, Zou R, Li D, Xu Z, Wang H, Zhang M, Zhang Z, Liu L, Liu Y. 2020. Exuberant elevation of IP-10, MCP-3 and IL-1ra during SARS-CoV-2 infection is associated with disease severity and fatal outcome. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.02.20029975>.
2212. Zhao J, Yuan Q, Wang H, Liu W, Liao X, Su Y, Wang X, Yuan J, Li T, Li J, Qian S, Hong C, Wang F, Liu Y, Wang Z, He Q, Li Z, He B, Zhang T, Ge S, Liu L, Zhang J, Xia N, Zhang Z. 2020. Antibody responses to SARS-CoV-2 in patients of novel coronavirus disease 2019. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.02.20030189>.
2213. Chen X, Ling J, Mo P, Zhang Y, Jiang Q, Ma Z, Cao Q, Hu W, Zou S, Chen L, Yao L, Luo M, Chen T, Deng L, Liang K, Song S, Yang R, Zheng R, Gao S, Gui X, Ke H, Hou W, Lundkvist Å, Xiong Y. 2020. Restoration of leukomonocyte counts is associated with viral clearance in COVID-19 hospitalized patients. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.03.20030437>.
2214. Olnes MJ, Kotliarov Y, Biancotto A, Cheung F, Chen J, Shi R, Zhou H, Wang E, Tsang JS, Nussenblatt R, The CHI Consortium. 2016. [Effects of Systemically Administered Hydrocortisone on the Human Immunome](#). Scientific Reports 6:23002.
2215. Lippi G, Plebani M. 2020. [Procalcitonin in patients with severe coronavirus disease 2019 \(COVID-19\): A meta-analysis](#). Clinica Chimica Acta 505:190–191.
2216. Xu Y, Xu Z, Liu X, Cai L, Zheng H, Huang Y, Zhou L, Huang L, Lin Y, Deng L, Li J, Chen S, Liu D, Lin Z, Zhou L, He W, Liu X, Li Y. 2020. Clinical findings in critically ill patients infected with SARS-CoV-2 in Guangdong Province, China: a multi-center, retrospective, observational study.

2217. Feng Y, Qiu M, Liu L, Zou S, Li Y, Luo K, Guo Q, Han N, Sun Y, Wang K, Zhuang X, Zhang S, Chen S, Mo F. 2020. Multi-epitope vaccine design using an immunoinformatics approach for 2019 novel coronavirus (SARS-CoV-2). Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.03.962332>.
2218. Cao M, Zhang D, Wang Y, Lu Y, Zhu X, Li Y, Xue H, Lin Y, Zhang M, Sun Y, Yang Z, Shi J, Wang Y, Zhou C, Dong Y, Peng L, Liu P, Dudek SM, Xiao Z, Lu H. 2020. Clinical Features of Patients Infected with the 2019 Novel Coronavirus (COVID-19) in Shanghai, China. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.04.20030395>.
2219. Zhang J, Liu J, Li N, Liu Y, Ye R, Qin X, Zheng R. 2020. Serological detection of 2019-nCoV respond to the epidemic: A useful complement to nucleic acid testing. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.04.20030916>.
2220. Diao B, Wang C, Wang R, Feng Z, Tan Y, Wang H, Wang C, Liu L, Liu Y, Liu Y, Wang G, Yuan Z, Ren L, Wu Y, Chen Y. 2020. Human Kidney is a Target for Novel Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Infection. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.04.20031120>.
2221. Song C-Y, Xu J, He J-Q, Lu Y-Q. 2020. COVID-19 early warning score: a multi-parameter screening tool to identify highly suspected patients. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.05.20031906>.
2222. Pfaender S, Mar KB, Michailidis E, Kratzel A, Hirt D, V'kovski P, Fan W, Ebert N, Stalder H, Kleine-Weber H, Hoffmann M, Hoffmann HH, Saeed M, Dijkman R, Steinmann E, Wight-Carter M, Hanners NW, Pöhlmann S, Gallagher T, Todt D, Zimmer G, Rice CM, Schoggins JW, Thiel V. 2020. LY6E impairs coronavirus fusion and confers immune control of viral disease. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.05.979260>.
2223. Liu L, Liu W, Zheng Y, Jiang X, Kou G, Ding J, Wang Q, Huang Q, Ding Y, Ni W, Wu W, Tang S, Tan L, Hu Z, Xu W, Zhang Y, Zhang B, Tang Z, Zhang X, Li H, Rao Z, Jiang H, Ren X, Wang S, Zheng S. 2020. A preliminary study on serological assay for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in 238 admitted hospital patients. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.06.20031856>.
2224. Zheng Z, Monteil VM, Maurer-Stroh S, Yew CW, Leong C, Mohd-Ismail NK, Arularasu SC, Chow VTK, Pin RLT, Mirazimi A, Hong W, Tan Y-J. 2020. Monoclonal antibodies for the S2 subunit of spike of SARS-CoV cross-react with the newly-emerged SARS-CoV-2. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.06.980037>.
2225. Zeng Q, Li Y, Huang G, Wu W, Dong S, Xu Y. 2020. Mortality of COVID-19 is Associated with Cellular Immune Function Compared to Immune

Function in Chinese Han Population. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.08.20031229>.

2226. Fan H, Zhang L, Huang B, Zhu M, Zhou Y, Zhang H, Tao X, Cheng S, Yu W, Zhu L, Chen J. 2020. Retrospective Analysis of Clinical Features in 101 Death Cases with COVID-19. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.09.20033068>.
2227. Matsuyama S, Kawase M, Nao N, Shirato K, Ujike M, Kamitani W, Shimojima M, Fukushi S. 2020. The inhaled corticosteroid ciclesonide blocks coronavirus RNA replication by targeting viral NSP15. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.11.987016>.
2228. Zhang B, Zhou X, Zhu C, Feng F, Qiu Y, Feng J, Jia Q, Song Q, Zhu B, Wang J. 2020. Immune phenotyping based on neutrophil-to-lymphocyte ratio and IgG predicts disease severity and outcome for patients with COVID-19. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.12.20035048>.
2229. Bao L, Deng W, Gao H, Xiao C, Liu J, Xue J, Lv Q, Liu J, Yu P, Xu Y, Qi F, Qu Y, Li F, Xiang Z, Yu H, Gong S, Liu M, Wang G, Wang S, Song Z, Liu Y, Zhao W, Han Y, Zhao L, Liu X, Wei Q, Qin C. 2020. Lack of Reinfection in Rhesus Macaques Infected with SARS-CoV-2. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.13.990226>.
2230. Yuan M, Wu NC, Zhu X, Lee C-CD, So RTY, Lv H, Mok CKP, Wilson IA. 2020. A highly conserved cryptic epitope in the receptor-binding domains of SARS-CoV-2 and SARS-CoV. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.13.991570>.
2231. Dong L, Zhou J, Niu C, Wang Q, Pan Y, Sheng S, Wang X, Zhang Y, Yang J, Liu M, Zhao Y, Zhang X, Zhu T, Peng T, Xie J, Gao Y, Wang D, Zhao Y, Dai X, Fang X. 2020. Highly accurate and sensitive diagnostic detection of SARS-CoV-2 by digital PCR. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.14.20036129>.
2232. Wang K, Chen W, Zhou Y-S, Lian J-Q, Zhang Z, Du P, Gong L, Zhang Y, Cui H-Y, Geng J-J, Wang B, Sun X-X, Wang C-F, Yang X, Lin P, Deng Y-Q, Wei D, Yang X-M, Zhu Y-M, Zhang K, Zheng Z-H, Miao J-L, Guo T, Shi Y, Zhang J, Fu L, Wang Q-Y, Bian H, Zhu P, Chen Z-N. 2020. SARS-CoV-2 invades host cells via a novel route: CD147-spike protein. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.14.988345>.
2233. Kosugi T, Maeda K, Sato W, Maruyama S, Kadomatsu K. 2015. [CD147 \(EMMPRIN/Basigin\) in kidney diseases: from an inflammation and immune system viewpoint](#). Nephrology Dialysis Transplantation 30:1097–1103.
2234. Su H, Yang Y. 2018. [The roles of CyPA and CD147 in cardiac remodelling](#). Experimental and Molecular Pathology 104:222–226.
2235. Weidle UH, Scheuer W, Eggle D, Klostermann S, Stockinger H. [Cancer-related issues of CD147](#). Cancer Genomics Proteomics 7:157–69.
2236. Huang L, Shi Y, Gong B, Jiang L, Liu X, Yang J, Tang J, You C, Jiang Q, Long B, Zeng T, Luo M, Zeng F, Zeng F, Wang S, Yang X, Yang Z. 2020.

Blood single cell immune profiling reveals the interferon-MAPK pathway mediated adaptive immune response for COVID-19. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.15.20033472>.

2237. Lv H, Wu NC, Tsang OT-Y, Yuan M, Perera RAPM, Leung WS, So RTY, Chun Chan JM, Yip GK, Hong Chik TS, Wang Y, Chung Choi CY, Lin Y, Ng WW, Zhao J, Poon LLM, Malik Peiris JS, Wilson IA, Mok CKP. 2020. Cross-reactive antibody response between SARS-CoV-2 and SARS-CoV infections. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.15.993097>.
2238. Duan K, Liu B, Li C, Zhang H, Yu T, Qu J, Zhou M, Chen L, Meng S, Hu Y, Peng C, Yuan M, Huang J, Wang Z, Yu J, Gao X, Wang D, Yu X, Li L, Zhang J, Wu X, Li B, Xu Y, Chen W, Peng Y, Hu Y, Lin L, Liu X, Huang S, Zhou Z, Zhang L, Wang Y, Zhang Z, Deng K, Xia Z, Gong Q, Zhang W, Zheng X, Liu Y, Yang H, Zhou D, Yu D, Hou J, Shi Z, Saijuan C, Chen Z, Zhang X, Yang X. 2020. The feasibility of convalescent plasma therapy in severe COVID-19 patients: a pilot study. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.16.20036145>.
2239. Gautret P, Lagier J-C, Parola P, Hoang VT, Meddeb L, Mailhe M, Doudier B, Courjon J, Giordanengo V, Vieira VE, Dupont HT, Honoré S, Colson P, Chabrière E, Scola BL, Rolain J-M, Brouqui P, Raoult D. 2020. Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.16.20037135>.
2240. Thomé R, Lopes SCP, Costa FTM, Verinaud L. 2013. [Chloroquine: Modes of action of an undervalued drug](#). Immunology Letters 153:50–57.
2241. Lo B, Zhang K, Lu W, Zheng L, Zhang Q, Kanellopoulou C, Zhang Y, Liu Z, Fritz JM, Marsh R, Husami A, Kissell D, Nortman S, Chaturvedi V, Haines H, Young LR, Mo J, Filipovich AH, Bleesing JJ, Mustillo P, Stephens M, Rueda CM, Chougnet CA, Hoebe K, McElwee J, Hughes JD, Karakoc-Aydiner E, Matthews HF, Price S, Su HC, Rao VK, Lenardo MJ, Jordan MB. 2015. [Patients with LRBA deficiency show CTLA4 loss and immune dysregulation responsive to abatacept therapy](#). Science 349:436–440.
2242. Procko E. 2020. The sequence of human ACE2 is suboptimal for binding the S spike protein of SARS coronavirus 2. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.16.994236>.
2243. Rockx B, Kuiken T, Herfst S, Bestebroer T, Lamers MM, de Meulder D, van Amerongen G, van den Brand J, Okba NMA, Schipper D, van Run P, Leijten L, Verschoor E, Verstrepen B, Langermans J, Drosten C, van Vlisningen MF, Fouchier R, de Swart R, Koopmans M, Haagmans BL. 2020. Comparative Pathogenesis Of COVID-19, MERS And SARS In A Non-Human Primate Model. Cold Spring Harbor Laboratory
<https://doi.org/10.1101/2020.03.17.995639>.
2244. McCray PB, Pewe L, Wohlford-Lenane C, Hickey M, Manzel L, Shi L, Netland J, Jia HP, Halabi C, Sigmund CD, Meyerholz DK, Kirby P, Look

DC, Perlman S. 2007. [Lethal Infection of K18-hACE2 Mice Infected with Severe Acute Respiratory Syndrome Coronavirus](#). Journal of Virology 81:813–821.

2245. Aguilar JB, Faust JS, Westafer LM, Gutierrez JB. 2020. Modeling the Impact of Asymptomatic Carriers on COVID-19 Transmission Dynamics During Lockdown. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.18.20037994>.
2246. Long Q, Deng H, Chen J, Hu J, Liu B, Liao P, Lin Y, Yu L, Mo Z, Xu Y, Gong F, Wu G, Zhang X, Chen Y, Li Z, Wang K, Zhang X, Tian W, Niu C, Yang Q, Xiang J, Du H, Liu H, Lang C, Luo X-H, Wu S, Cui X, Zhou Z, Wang J, Xue C, Li X, Wang L, Tang X, Zhang Y, Qiu J, Liu X, Li J, Zhang D, Zhang F, Cai X, Wang D, Hu Y, Ren J, Tang N, Liu P, Li Q, Huang A. 2020. Antibody responses to SARS-CoV-2 in COVID-19 patients: the perspective application of serological tests in clinical practice. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.18.20038018>.
2247. Hu X, An T, Situ B, Hu Y, Ou Z, Li Q, He X, Zhang Y, Tian P, Sun D, Rui Y, Wang Q, Ding D, Zheng L. 2020. Heat inactivation of serum interferes with the immunoanalysis of antibodies to SARS-CoV-2. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.12.20034231>.
2248. Okba NMA, Müller MA, Li W, Wang C, GeurtsvanKessel CH, Corman VM, Lamers MM, Sikkema RS, de Bruin E, Chandler FD, Yazdanpanah Y, Le Hingrat Q, Descamps D, Houhou-Fidouh N, Reusken CBEM, Bosch B-J, Drosten C, Koopmans MPG, Haagmans BL. 2020. SARS-CoV-2 specific antibody responses in COVID-19 patients. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.18.20038059>.
2249. Belhadi D, Peiffer-Smadja N, Lescure F-X, Yazdanpanah Y, Mentré F, Laouénan C. 2020. A brief review of antiviral drugs evaluated in registered clinical trials for COVID-19. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.18.20038190>.
2250. Leung JM, Yang CX, Tam A, Shaipanich T, Hackett T-L, Singhera GK, Dorscheid DR, Sin DD. 2020. ACE-2 Expression in the Small Airway Epithelia of Smokers and COPD Patients: Implications for COVID-19. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.18.20038455>.
2251. Xu Y. 2020. Dynamic profile of severe or critical COVID-19 cases. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.18.20038513>.
2252. Fu H, Xu H, Zhang N, Xu H, Li Z, Chen H, Xu R, Sun R, Wen L, Xie L, Liu H, Zhang K, Fu C, Hou K, Yang Z, Yang M, Guo Y. 2020. Association between Clinical, Laboratory and CT Characteristics and RT-PCR Results in the Follow-up of COVID-19 patients. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.19.20038315>.
2253. Sheahan TP, Sims AC, Zhou S, Graham RL, Hill CS, Leist SR, Schäfer A, Dinnon KH, Montgomery SA, Agostini ML, Pruijssers AJ, Chapell JD, Brown AJ, Bluemling GR, Natchus MG, Saindane M, Kolykhalov AA, Painter G, Harcourt J, Tamin A, Thornburg NJ, Swanstrom R, Denison

MR, Baric RS. 2020. An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 and multiple endemic, epidemic and bat coronavirus. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.19.997890>.

2254. Jeon S, Ko M, Lee J, Choi I, Byun SY, Park S, Shum D, Kim S. 2020. Identification of antiviral drug candidates against SARS-CoV-2 from FDA-approved drugs. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.20.999730>.
2255. Munster VJ, Feldmann F, Williamson BN, van Doremalen N, Pérez-Pérez L, Schulz J, Meade-White K, Okumura A, Callison J, Brumbaugh B, Avanzato VA, Rosenke R, Hanley PW, Saturday G, Scott D, Fischer ER, de Wit E. 2020. Respiratory disease and virus shedding in rhesus macaques inoculated with SARS-CoV-2. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.21.001628>.
2256. Deng W, Bao L, Gao H, Xiang Z, Qu Y, Song Z, Gong S, Liu J, Liu J, Yu P, Qi F, Xu Y, Li F, Xiao C, Lv Q, Xue J, Wei Q, Liu M, Wang G, Wang S, Yu H, Liu X, Zhao W, Han Y, Qin C. 2020. Ocular conjunctival inoculation of SARS-CoV-2 can cause mild COVID-19 in Rhesus macaques. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.13.990036>.
2257. Pinto BGG, Oliveira AER, Singh Y, Jimenez L, Gonçalves ANA, Ogawa RLT, Creighton R, Schatzmann Peron JP, Nakaya HI. 2020. ACE2 Expression is Increased in the Lungs of Patients with Comorbidities Associated with Severe COVID-19. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.21.20040261>.
2258. Bian H, Zheng Z-H, Wei D, Zhang Z, Kang W-Z, Hao C-Q, Dong K, Kang W, Xia J-L, Miao J-L, Xie R-H, Wang B, Sun X-X, Yang X-M, Lin P, Geng J-J, Wang K, Cui H-Y, Zhang K, Chen X-C, Tang H, Du H, Yao N, Liu S-S, Liu L-N, Zhang Z, Gao Z-W, Nan G, Wang Q-Y, Lian J-Q, Chen Z-N, Zhu P. 2020. Meplazumab treats COVID-19 pneumonia: an open-labelled, concurrent controlled add-on clinical trial. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.21.20040691>.
2259. Pushkarsky T, Zybarth G, Dubrovsky L, Yurchenko V, Tang H, Guo H, Toole B, Sherry B, Bukrinsky M. 2001. [CD147 facilitates HIV-1 infection by interacting with virus-associated cyclophilin A](#). Proceedings of the National Academy of Sciences 98:6360–6365.
2260. Watanabe A, Yoneda M, Ikeda F, Terao-Muto Y, Sato H, Kai C. 2010. [CD147/EMMPRIN Acts as a Functional Entry Receptor for Measles Virus on Epithelial Cells](#). Journal of Virology 84:4183–4193.
2261. Crosnier C, Bustamante LY, Bartholdson SJ, Bei AK, Theron M, Uchikawa M, Mboup S, Ndir O, Kwiatkowski DP, Duraisingh MT, Rayner JC, Wright GJ. 2011. [Basigin is a receptor essential for erythrocyte invasion by Plasmodium falciparum](#). Nature 480:534–537.
2262. Chen Z, Mi L, Xu J, Yu J, Wang X, Jiang J, Xing J, Shang P, Qian A, Li Y, Shaw Peter X, Wang J, Duan S, Ding J, Fan C, Zhang Y, Yang Y, Yu X, Feng Q, Li B, Yao X, Zhang Z, Li L, Xue X, Zhu P. 2005. [Function of HAb18G/CD147 in Invasion of Host Cells by Severe Acute Respiratory](#)

[Syndrome Coronavirus](#). The Journal of Infectious Diseases 191:755–760.

2263. Yee C, Main NM, Terry A, Stevanovski I, Maczurek A, Morgan AJ, Calabro S, Potter AJ, Iemma TL, Bowen DG, Ahlenstiel G, Warner FJ, McCaughan GW, McLennan SV, Shackel NA. 2019. [CD147 mediates intrahepatic leukocyte aggregation and determines the extent of liver injury](#). PLOS ONE 14:e0215557.
2264. Davidson AD, Williamson MK, Lewis S, Shoemark D, Carroll MW, Heesom K, Zambon M, Ellis J, Lewis PA, Hiscox JA, Matthews DA. 2020. Characterisation of the transcriptome and proteome of SARS-CoV-2 using direct RNA sequencing and tandem mass spectrometry reveals evidence for a cell passage induced in-frame deletion in the spike glycoprotein that removes the furin-like cleavage site. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.22.002204>.
2265. Sun X, Tse LV, Ferguson AD, Whittaker GR. 2010. [Modifications to the Hemagglutinin Cleavage Site Control the Virulence of a Neurotropic H1N1 Influenza Virus](#). Journal of Virology 84:8683–8690.
2266. Kim D, Lee J-Y, Yang J-S, Kim JW, Kim VN, Chang H. 2020. The architecture of SARS-CoV-2 transcriptome. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.12.988865>.
2267. Gordon DE, Jang GM, Bouhaddou M, Xu J, Obernier K, O'Meara MJ, Guo JZ, Swaney DL, Tummino TA, Hüttenhain R, Kaake RM, Richards AL, Tutuncuoglu B, Foussard H, Batra J, Haas K, Modak M, Kim M, Haas P, Polacco BJ, Braberg H, Fabius JM, Eckhardt M, Soucheray M, Bennett MJ, Cakir M, McGregor MJ, Li Q, Naing ZZC, Zhou Y, Peng S, Kirby IT, Melnyk JE, Chorbha JS, Lou K, Dai SA, Shen W, Shi Y, Zhang Z, Barrio-Hernandez I, Memon D, Hernandez-Armenta C, Mathy CJP, Perica T, Pilla KB, Ganesan SJ, Saltzberg DJ, Ramachandran R, Liu X, Rosenthal SB, Calviello L, Venkataramanan S, Lin Y, Wankowicz SA, Bohn M, Trenker R, Young JM, Caverio D, Hiatt J, Roth T, Rathore U, Subramanian A, Noack J, Hubert M, Roesch F, Vallet T, Meyer B, White KM, Miorin L, Agard D, Emerman M, Ruggero D, García-Sastre A, Jura N, Zastrow M von, Taunton J, Schwartz O, Vignuzzi M, d'Enfert C, Mukherjee S, Jacobson M, Malik HS, Fujimori DG, Ideker T, Craik CS, Floor S, Fraser JS, Gross J, Sali A, Kortemme T, Beltrao P, Shokat K, Shoichet BK, Krogan NJ. 2020. A SARS-CoV-2-Human Protein-Protein Interaction Map Reveals Drug Targets and Potential Drug-Repurposing. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.22.002386>.
2268. Chen H, Zhang Z, Wang L, Huang Z, Gong F, Li X, Chen Y, Wu JJ. 2020. First Clinical Study Using HCV Protease Inhibitor Danoprevir to Treat Naïve and Experienced COVID-19 Patients. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.22.20034041>.
2269. Seiwert SD, Andrews SW, Jiang Y, Serebryany V, Tan H, Kossen K, Rajagopalan PTR, Misialek S, Stevens SK, Stoycheva A, Hong J, Lim SR, Qin X, Rieger R, Condroski KR, Zhang H, Do MG, Lemieux C, Hingorani GP, Hartley DP, Josey JA, Pan L, Beigelman L, Blatt LM. 2008. [Preclinical Characteristics of the Hepatitis C Virus NS3/4A Protease Inhibitor](#)

[ITMN-191 \(R7227\)](#). Antimicrobial Agents and Chemotherapy 52:4432–4441.

2270. Xu X, Feng B, Guan Y, Zheng S, Sheng J, Yang X, Ma Y, Huang Y, Kang Y, Wen X, Li J, Tan Y, He Q, Xie Q, Wang M, An P, Gong G, Liu H, Ning Q, Hua R, Ning B, Xie W, Zhang J, Huang W, Yang Y, Lin M, Zhao Y, Yu Y, Jia J, Yang D, Chen L, Ye Y, Nan Y, Gong Z, Zhang Q, Hu P, Wang F, Li Y, Li D, Jia Z, Hou J, Chen C, Wu JJ, Wei L. 2019. [Efficacy and Safety of All-oral, 12-week Ravidasvir Plus Ritonavir-boosted Danoprevir and Ribavirin in Treatment-naïve Noncirrhotic HCV Genotype 1 Patients: Results from a Phase 2/3 Clinical Trial in China](#). Journal of Clinical and Translational Hepatology 7:1–8.
2271. Nguyen DD, Gao K, Chen J, Wang R, Wei G-W. 2020. Potentially highly potent drugs for 2019-nCoV. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.02.05.936013>.
2272. Lou B, Li T-D, Zheng S-F, Su Y-Y, Li Z-Y, Liu W, Yu F, Ge S-X, Zou Q-D, Yuan Q, Lin S, Hong C-M, Yao X-Y, Zhang X-J, Wu D-H, Zhou G-L, Hou W-H, Li T-T, Zhang Y-L, Zhang S-Y, Fan J, Zhang J, Xia N-S, Chen Y. 2020. Serology characteristics of SARS-CoV-2 infection since the exposure and post symptoms onset. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.23.20041707>.
2273. Blanco-Melo D, Nilsson-Payant BE, Liu W-C, Møller R, Panis M, Sachs D, Albrecht RA, tenOever BR. 2020. SARS-CoV-2 launches a unique transcriptional signature from in vitro, ex vivo, and in vivo systems. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.24.004655>.
2274. Zhou Y, Yang Z, Guo Y, Geng S, Gao S, Ye S, Hu Y, Wang Y. 2020. A New Predictor of Disease Severity in Patients with COVID-19 in Wuhan, China. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.24.20042119>.
2275. Nie S, Zhao X, Zhao K, Zhang Z, Zhang Z, Zhang Z. 2020. Metabolic disturbances and inflammatory dysfunction predict severity of coronavirus disease 2019 (COVID-19): a retrospective study. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.24.20042283>.
2276. Tan W, Lu Y, Zhang J, Wang J, Dan Y, Tan Z, He X, Qian C, Sun Q, Hu Q, Liu H, Ye S, Xiang X, Zhou Y, Zhang W, Guo Y, Wang X-H, He W, Wan X, Sun F, Wei Q, Chen C, Pan G, Xia J, Mao Q, Chen Y, Deng G. 2020. Viral Kinetics and Antibody Responses in Patients with COVID-19. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.24.20042382>.
2277. Jiang H, Li Y, Zhang H, Wang W, Men D, Yang X, Qi H, Zhou J, Tao S. 2020. Global profiling of SARS-CoV-2 specific IgG/ IgM responses of convalescents using a proteome microarray. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.20.20039495>.
2278. Liu L, Wei Q, Lin Q, Fang J, Wang H, Kwok H, Tang H, Nishiura K, Peng J, Tan Z, Wu T, Cheung K-W, Chan K-H, Alvarez X, Qin C, Lackner A,

Perlman S, Yuen K-Y, Chen Z. 2019. [Anti-spike IgG causes severe acute lung injury by skewing macrophage responses during acute SARS-CoV infection](#). JCI Insight 4:e123158.

2279. Zhang D, Guo R, Lei L, Liu H, Wang Y, Wang Y, Qian H, Dai T, Zhang T, Lai Y, Wang J, Liu Z, Chen T, He A, O'Dwyer M, Hu J. 2020. COVID-19 infection induces readily detectable morphological and inflammation-related phenotypic changes in peripheral blood monocytes, the severity of which correlate with patient outcome. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.24.20042655>.
2280. Miller A, Reandelar MJ, Fasciglione K, Roumenova V, Li Y, Otazu GH. 2020. Correlation between universal BCG vaccination policy and reduced morbidity and mortality for COVID-19: an epidemiological study. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.24.20042937>.
2281. Moorlag SJCFM, Arts RJW, van Crevel R, Netea MG. 2019. [Non-specific effects of BCG vaccine on viral infections](#). Clinical Microbiology and Infection 25:1473–1478.
2282. BCG vaccination to reduce the impact of COVID-19 in healthcare workers (The BRACE Trial). Murdoch Children's Research Institute. <https://www.mcric.edu.au/BRACE>. Retrieved 31 July 2020.
2283. Brann DH, Tsukahara T, Weinreb C, Lipovsek M, Van den Berge K, Gong B, Chance R, Macaulay IC, Chou H, Fletcher R, Das D, Street K, de Bezieux HR, Choi Y-G, Risso D, Dudoit S, Purdom E, Mill JS, Hachem RA, Matsunami H, Logan DW, Goldstein BJ, Grubb MS, Ngai J, Datta SR. 2020. Non-neuronal expression of SARS-CoV-2 entry genes in the olfactory system suggests mechanisms underlying COVID-19-associated anosmia. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.25.009084>.
2284. Smith JC, Sausville EL, Girish V, Yuan ML, John KM, Sheltzer JM. 2020. Cigarette smoke exposure and inflammatory signaling increase the expression of the SARS-CoV-2 receptor ACE2 in the respiratory tract. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.28.013672>.
2285. Liu R, Liu X, Han H, Shereen MA, Niu Z, Li D, Liu F, Wu K, Luo Z, Zhu C. 2020. The comparative superiority of IgM-IgG antibody test to real-time reverse transcriptase PCR detection for SARS-CoV-2 infection diagnosis. Cold Spring Harbor Laboratory <https://doi.org/10.1101/2020.03.28.20045765>.

14 Appendix A

This appendix contains reviews produced by the Immunology Institute of the Icahn School of Medicine

14.1 Potent binding of 2019 novel coronavirus spike protein by a SARS coronavirus-specific human monoclonal antibody

Tian et al. *Emerg Microbes Infect* 2020 ([2162](#))

14.1.1 Keywords

- Monoclonal antibody
- Cross-reactivity
- receptor binding domain

14.1.2 Summary

Considering the relatively high identity of the receptor binding domain (RBD) of the spike proteins from 2019-nCoV and SARS-CoV (73%), this study aims to assess the cross-reactivity of several anti-SARS-CoV monoclonal antibodies with 2019-nCoV. The results showed that the SARS-CoV-specific antibody CR3022 can potently bind 2019-nCoV RBD.

14.1.3 Main Findings

The structure of the 2019-nCoV spike RBD and its conformation in complex with the receptor angiotensin-converting enzyme (ACE2) was modeled *in silico* and compared with the SARS-CoV RBD structure. The models predicted very similar RBD-ACE2 interactions for both viruses. The binding capacity of representative SARS-CoV-RBD specific monoclonal antibodies (m396, CR3014, and CR3022) to recombinant 2019-nCoV RBD was then investigated by ELISA and their binding kinetics studied using biolayer interferometry. The analysis showed that only CR3022 was able to bind 2019-nCoV RBD with high affinity (KD of 6.3 nM), however it did not interfere with ACE2 binding. Antibodies m396 and CR3014, which target the ACE2 binding site of SARS-CoV failed to bind 2019-nCoV spike protein.

14.1.4 Limitations

The 2019-nCoV RBD largely differ from the SARS-CoV at the C-terminus residues, which drastically impact the cross-reactivity of antibodies described for other B beta-coronaviruses, including SARS-CoV. This study claims that CR3022 antibody could be a potential candidate for therapy. However, none of the antibodies assayed in this work showed cross-reactivity with the ACE2 binding site of 2019-nCoV, essential for the replication of this virus. Furthermore, neutralization assays with 2019-nCoV virus or pseudovirus

were not performed. Although the use of neutralizing antibodies is an interesting approach, these results suggest that it is critical the development of novel monoclonal antibodies able to specifically bind 2019-nCoV spike protein.

14.1.5 Credit

Review by D.L.O as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.2 Integrative Bioinformatics Analysis Provides Insight into the Molecular Mechanisms of 2019-nCoV

He et al. *medRxiv* ([2163](#))

14.2.1 Keywords

- ACE2
- lungs
- smoking
- COPD
- asthma
- SARS-Cov
- IL-1
- IL-10
- IL-6
- IL-8

14.2.2 Main Findings

The authors used bioinformatics tools to identify features of ACE2 expression in the lungs of different patient groups: healthy, smokers, patients with chronic airway disease (i.e., COPD) or asthma. They used gene expression data publicly available from GEO that included lung tissues, bronchoalveolar lavage, bronchial epithelial cells, small airway epithelial cells, or SARS-Cov infected cells.

The authors describe no significant differences in ACE2 expression in lung tissues of Healthy, COPD, and Asthma groups ($p=0.85$); or in BAL of Healthy and COPD ($p=0.48$); or in epithelial brushings of Healthy and Mild/Moderate/Severe Asthma ($p=0.99$). ACE2 was higher in the small airway epithelium of long-term smokers vs non-smokers ($p<0.001$). Consistently, there was a trend of higher ACE2 expression in the bronchial airway epithelial cells 24h post-acute smoking exposure ($p=0.073$). Increasing ACE2 expression at 24h and 48h compared to 12h post SARS-Cov infection ($p=0.026$; $n=3$ at each time point) was also detected.

15 lung samples' data from healthy participants were separated into high and low ACE2 expression groups. "High" ACE2 expression was associated with the following GO pathways: innate and adaptive immune responses, B cell

mediated immunity, cytokine secretion, and IL-1, IL-10, IL-6, IL-8 cytokines. The authors speculate that a high basal ACE2 expression will increase susceptibility to SARS-CoV infection.

In 3 samples SARS-Cov infection was associated with IL-1, IL-10 and IL-6 cytokine production (GO pathways) at 24h. And later, at 48h, with T-cell activation and T-cell cytokine production. It is unclear whether those changes were statistically significant.

The authors describe a time course quantification of immune infiltrates in epithelial cells infected with SARS-Cov infection. They state that in healthy donors ACE2 expression did not correlate with the immune cell infiltration. However, in SARS-Cov samples, at 48h they found that ACE2 correlated with neutrophils, NK-, Th17-, Th2-, Th1- cells, and DCs. Again, while authors claim significance, the corresponding correlation coefficients and p-values are not presented in the text or figures. In addition, the source of the data for this analysis is not clear.

Using network analysis, proteins SRC, FN1, MAPK3, LYN, MBP, NLRC4, NLRP1 and PRKCD were found to be central (Hub proteins) in the regulating network of cytokine secretion after coronavirus infection. Authors conclude this indicates that these molecules were critically important in ACE2-induced inflammatory response. Additionally, authors speculate that the increased expression of ACE2 affected RPS3 and SRC, which were the two hub genes involved in viral replication and inflammatory response.

14.2.3 Limitations

The methods section is very limited and does not describe any of the statistical analyses; and description of the construction of the regulatory protein networks is also limited. For the findings in Figures 2 authors claim significance, which is not supported by p-values or coefficients. For the sample selection, would be useful if sample sizes and some of the patients' demographics (e.g. age) were described.

For the analysis of high vs low ACE2 expression in healthy subjects, it is not clear what was the cut off for 'high' expression and how it was determined. Additionally, further laboratory studies are warranted to confirm that high ACE2 gene expression would have high correlation with the amount of ACE2 protein on cell surface. For the GO pathway analysis significance was set at $p < 0.05$, but not adjusted for multiple comparisons.

There were no samples with SARS-CoV-2 infection. While SARS-Cov and SARS-CoV-2 both use ACE2 to enter the host cells, the analysis only included data on SARS-Cov and any conclusions about SARS-CoV2 are limited.

Upon checking GSE accession numbers of the datasets references, two might not be cited correctly: GSE37758 ("Aspergillus niger: Control (fructose) vs. steam-exploded sugarcane induction (SEB)"" was used in this paper as "lung tissue" data) and GSE14700 ("Steroid Pretreatment of Organ Donors to Prevent Postischemic Renal Allograft Failure: A Randomized, Controlled Trial" – was used as SARS-Cov infection data).

14.2.4 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.3 Diarrhea may be underestimated: a missing link in 2019 novel coronavirus

Liang et al. *medRxiv* ([2164](#))

14.3.1 Keywords

- SARS-CoV-2
- diarrhea
- ACE2
- scRNA-seq

14.3.2 Main Findings

This study examined the incidence of diarrhea in patients infected with SARS-CoV-2 across three recently published cohorts and found that there are statistically significant differences by Fisher's exact test. They report that this could be due to subjective diagnosis criterion for diarrhea or from patients first seeking medical care from gastroenterologist. In order to minimize nosocomial infections arising from unsuspected patients with diarrhea and gain comprehensive understanding of transmission routes for this viral pathogen, they compared the transcriptional levels of ACE2 of various human tissues from NCBI public database as well as in small intestine tissue from CD57BL/6 mice using single cell sequencing. They show that ACE2 expression is not only increased in the human small intestine, but demonstrate a particular increase in mice enterocytes positioned on the surface of the intestinal lining exposed to viral pathogens. Given that ACE2 is the viral receptor for SARS-CoV-2 and also reported to regulate diarrhea, their data suggests the small intestine as a potential transmission route and diarrhea as a potentially underestimated symptom in COVID19 patients that must be carefully monitored. Interestingly, however, they show that ACE2 expression level is not elevated in human lung tissue.

14.3.3 Limitations

Although this study demonstrates a statistical difference in the incidence of diarrhea across three separate COVID19 patient cohorts, their conclusions are limited by a small sample size. Specifically, the p-value computed by Fisher's exact test is based on a single patient cohort of only six cases of which 33% are reported to have diarrhea, while the remaining two larger cohorts with 41 and 99 cases report 3% and 2% diarrhea incidence, respectively. Despite showing significance, they would need to acquire larger sample sizes and cohorts to minimize random variability and draw meaningful conclusions. Furthermore, they do not address why ACE2 expression level is not elevated in human lung tissue despite it being a major

established route of transmission for SARS-CoV-2. It could be helpful to validate this result by looking at ACE2 expression in mouse lung tissue. Finally, although this study is descriptive and shows elevated ACE2 expression in small intestinal epithelial cells, it does not establish a mechanistic link to SARS-CoV-2 infection of the host. Overall, their claim that infected patients exhibiting diarrhea pose an increased risk to hospital staff needs to be further substantiated.

14.3.4 Significance

This study provides a possible transmission route and a potentially underappreciated clinical symptom for SARS-CoV-2 for better clinical management and control of COVID19.

14.3.5 Credit

Summary generated as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.4 Specific ACE2 Expression in Cholangiocytes May Cause Liver Damage After 2019-nCoV Infection

Chai et al. *bioRxiv* ([2165](#))

14.4.1 Keywords

- ACE2
- Cholangiocytes
- COVID-associated Liver Damage

14.4.2 Summary

Using both publicly available scRNA-seq dataset of liver samples from colorectal patients and scRNA-sequencing of four liver samples from healthy volunteers, the authors show that ACE2 is significantly enriched in the majority of cholangiocytes (59.7 %) but not in hepatocytes (2.6%).

14.4.3 Main Findings

Using bioinformatics approaches of RNASeq analysis, this study reveals that ACE2 dominates in cholangiocytes and is present at very low levels in hepatocytes.

14.4.4 Limitations

The study does not provide mechanistic insights into how SARS-CoV-2 can infect and replicate in cholangiocytes and the types of intrinsic anti-viral responses induced by cholangiocytes when infected. In addition, because the

study relies on the assumption that SARS-CoV-2 infects cells only through ACE2, it cannot discount the possibility that the virus can infect hepatocytes through mechanisms other than ACE2-mediated entry. Furthermore, because the scRNA-seq analysis were performed on healthy liver samples, one cannot draw any definitive conclusions about gene expression states (including ACE2 expression in liver cell types) in system-wide inflammatory contexts.

14.4.5 Significance

This article with other studies on liver damage in COVID patients suggests that liver damage observed in COVID patients is more due to inflammatory cytokines than direct infection of the liver. Even if cholangiocytes are infectable by SARS-CoV-2 (which was demonstrated by human liver ductal organoid study (([2166](#)))), published clinical data show no significant increase in bile duct injury related indexes (i.e. alkaline phosphatase, gamma-glutamyl transpeptidase and total bilirubin). In sum, it underscores the importance of future studies characterizing cellular responses of extra-pulmonary organs in the context of COVID or at least in viral lung infections..

14.4.6 Credit

Summary generated by Chang Moon as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.5 ACE2 expression by colonic epithelial cells is associated with viral infection, immunity and energy metabolism

Wang et al. *medRxiv.* ([2167](#))

14.5.1 Keywords

- single cell RNA seq
- ACE2 expression
- human colonic biopsy

14.5.2 Main Findings

Colonic enterocytes primarily express ACE2. Cellular pathways associated with ACE2 expression include innate immune signaling, HLA up regulation, energy metabolism and apoptotic signaling.

14.5.3 Limitations

This is a study of colonic biopsies taken from 17 children with and without IBD and analyzed using scRNAseq to look at ACE2 expression and identify gene families correlated with ACE2 expression. The authors find ACE2 expression to be primarily in colonocytes. It is not clear why both healthy and

IBD patients were combined for the analysis. Biopsies were all of children so extrapolation to adults is limited. The majority of genes found to be negatively correlated with ACE2 expression include immunoglobulin genes (IGs). IG expression will almost certainly be low in colonocytes irrespective of ACE2 expression.

14.5.4 Significance

This study performs a retrospective analysis of ACE2 expression using an RNAseq dataset from intestinal biopsies of children with and without IBD. The implications for the CoV-19 epidemic are modest, but do provide support that ACE2 expression is specific to colonocytes in the intestines. The ontological pathway analysis provides some limited insights into gene expression associated with ACE2.

14.5.5 Credit

Summary generated as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.6 The Pathogenicity of 2019 Novel Coronavirus in hACE2 Transgenic Mice

Bao et al. *bioRxiv* ([2168](#))

14.6.1 Keywords

- Covid-19 mouse model
- hACE2 mice
- 2019-nCoV model
- ACE2
- 2019-nCoV

14.6.2 Main Findings

Using a transgenic human Angiotensin-converting enzyme 2 (hACE2) mouse that has previously been shown susceptible to infection by SARS-CoV, Bao et al. create a model of pandemic 2019-nCoV strain coronavirus. The model includes interstitial hyperplasia in lung tissue, moderate inflammation in bronchioles and blood vessels, and histology consistent with viral pneumonia at 3 days post infection. Wildtype did not experience these symptoms. In addition, viral antigen and hACE2 receptor were found to co-localize the lung by immunofluorescence 3-10 days post infection only in the hACE2 infected mice.

14.6.3 Limitations

The characterization of the infection remains incomplete, as well as lacking characterization of the immune response other than the presence of a single antiviral antibody. Though they claim to fulfill Koch's postulates, they only

isolate the virus and re-infect Vero cells, rather than naive mice.

14.6.4 Significance

This paper establishes a murine model for 2019-nCoV infection with symptoms consistent with viral pneumonia. Though not fully characterized, this model allows in vivo analysis of viral entry and pathology that is important for the development of vaccines and antiviral therapeutics.

14.6.5 Credit

Review by Dan Fu Ruan, Evan Cody and Venu Pothula as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.7 Caution on Kidney Dysfunctions of 2019-nCoV Patients

Li et al. *medRxiv*. ([2169](#))

14.7.1 Keywords

CoVID-19, 2019-nCoV, SARS-CoV-2, kidney, clinical, creatinine, proteinuria, albuminuria, CT

14.7.2 Main Findings

- Retrospective study of 59 patients assayed key function indicators of the kidney—including urine protein, blood urea nitrogen (BUN), plasma creatinine (Cre), and renal CT scan data.
- Found that 34% of patients developed massive albuminuria on the first day of admission, and 63% developed proteinuria during their stay in hospital; and 19% of patients had high plasma creatinine, especially the terminal cases.
- CT analyses of 27 patients showed all patients to have abnormal kidney damage; indicate that inflammation and edema of the renal parenchyma very common.

14.7.3 Limitations

- No analysis of immunity-dependent damage and cytokines in blood/plasma/urine. Will be worth correlating disease progression with cytokine production, immune activity and kidney function.
- Extrapolating to earlier SARS-CoV studies provides the only rationale for viral-damage in kidney and resultant pathologic immune response (*understandable for this clinical study*).

14.7.4 Significance

- Multiple lines of evidence along this study's finding point to the idea that renal impairment/injury is a key risk factor in 2019-nCoV patients similar to what has been reported for SARS-CoV([2170](#)); this may be one of the major causes of virally-induced damage and contribute to multiorgan failure.
- ACE2 expression in kidney proximal tubule epithelia and bladder epithelia ([2171](#)) support these clinical findings.
- Study argues for closely monitoring kidney function, and applying potential interventions including continuous renal replacement therapies (CRRT) for protecting kidney functions as early as possible, particularly for those with rising plasma creatinine.

14.7.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.8 Profiling the immune vulnerability landscape of the 2019 Novel Coronavirus

Zhu et al. *bioRxiv* ([2172](#))

14.8.1 Keywords

- epitope prediction
- vaccine development.

14.8.2 Main Findings

This study harnesses bioinformatic profiling to predict the potential of COV2 viral proteins to be presented on MHC I and II and to form linear B-cell epitopes. These estimates suggest a T-cell antigenic profile distinct from SARS-CoV or MERS-CoV, identify focused regions of the virus with a high density of predicted epitopes, and provide preliminary evidence for adaptive immune pressure in the genetic evolution of the virus.

14.8.3 Limitations

While the study performs a comprehensive analysis of potential epitopes within the virus genome, the analysis relies solely on bioinformatic prediction to examine MHC binding affinity and B-cell epitope potential and does not capture the immunogenicity or recognition of these epitopes. Future experimental validation in data from patients infected with SARS-CoV-2 will be important to validate and refine these findings. Thus some of

the potential conclusions stated, including viral evolution toward lower immunogenicity or a dominant role for CD4+ T-cells rather than CD8+ T-cells in viral clearance, require further validation.

14.8.4 Significance

These findings may help direct peptide vaccine design toward relevant epitopes and provide intriguing evidence of viral evolution in response to immune pressure.

14.8.5 Credit

Summary generated as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.9 Single-cell Analysis of ACE2 Expression in Human Kidneys and Bladders Reveals a Potential Route of 2019-nCoV Infection

Lin et al. *bioRxiv* ([2171](#))

14.9.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- ACE2
- scRNAseq
- kidney
- bladder
- public dataset

14.9.2 Main Findings

- To investigate the possible cause of kidney damage in 2019-nCoV patients, authors used published kidney and bladder cell atlas data (GSE131685, GSE108097; 3 healthy donors each) as well as an unpublished kidney single-cell RNA-Seq data (in-house from 2 transplant donors) to evaluate ACE2 gene expressions in all cell types of healthy kidneys and bladders.
- They find enriched expression of ACE2 transcript in all subtypes of proximal tubule cells of kidney, with 5%-15% of both straight and convoluted proximal tubule cells expressing ACE2.
- They also find detectable levels of ACE2 in bladder epithelial cells, noting expression from around 1.5% of cells in the outer layer umbrella cells of the bladder epithelium and decreasing in the basal cells.
- Importantly endothelial or immune cells in kidney/bladder do not express ACE2.

14.9.3 Limitations

- This study primarily characterizes ACE2 expression (amongst other genes) from a small healthy-donor dataset, and will benefit from supporting data in (expired) patient samples to show functional viral damage. ACE2 transcript does not necessarily translate to viral permissiveness in kidney/bladder epithelia or cytokine release.
- This study focuses on only healthy tissue; it will be useful to analyze kidney/bladder epithelial ACE2 expression under inflammatory conditions or in patients with underlying kidney conditions.
- Given what is known about protease TMPRSS2 expression during SARS-CoV-2 infection, ACE2+TMPRSS2+ double-positive cell identification would be useful in these datasets.

14.9.4 Significance

- ACE2 protein is spatially restricted to brush border of proximal tubules and in bladder umbrella cells ([67](#)), such cells in direct contact with viral particles are likely to be highly sensitive to viral-induced damage.
- SARS-CoV and MERS-CoV have been shown to be detected in urine of patients and associate with higher mortality ([2170](#), [2173](#)), thus worth understanding kidney damage and resultant immune response in SARS-CoV-2 as well.
- This study argues for a potential mode of viral infectivity and resultant inflammatory responses in these tissue in addition to reported infectivity in the lung and digestive system, which is supported by clinical data showing acute and early kidney complications in 2019-nCoV patients ([2169](#)).
- Clinically, thus very important to track urinary CoVID-19 shedding as well as study acute kidney injury-related co-morbidities.

14.9.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.10 Neutrophil-to-Lymphocyte Ratio Predicts Severe Illness Patients with 2019 Novel Coronavirus in the Early Stage

Liu et al. *medRxiv* ([2174](#))

14.10.1 Keywords

- severe disease
- pneumonia
- lymphocytes
- neutrophils

14.10.2 Main Findings

This study aimed to find prognostic biomarkers of COVID-19 pneumonia severity. Sixty-one (61) patients with COVID-19 treated in January at a hospital in Beijing, China were included. On average, patients were seen within 5 days from illness onset. Samples were collected on admission; and then patients were monitored for the development of severe illness with a median follow-up of 10 days].

Patients were grouped as “mild” (N=44) or “moderate/severe” (N=17) according to symptoms on admission and compared for different clinical/laboratory features. “Moderate/severe” patients were significantly older (median of 56 years old, compared to 41 years old). Whereas comorbidities rates were largely similar between the groups, except for hypertension, which was more frequent in the severe group ($p=0.056$). ‘Severe’ patients had higher counts of neutrophils, and serum glucose levels; but lower lymphocyte counts, sodium and serum chlorine levels. The ratio of neutrophils to lymphocytes (NLR) was also higher for the ‘severe’ group. ‘Severe’ patients had a higher rate of bacterial infections (and antibiotic treatment) and received more intensive respiratory support and treatment.

26 clinical/laboratory variables were used to select NLR and age as the best predictors of the severe disease. Predictive cutoffs for a severe illness as $\text{NLR} \geq 3.13$ or $\text{age} \geq 50$ years.

14.10.3 Limitations

Identification of early biomarkers is important for making clinical decisions, but large sample size and validation cohorts are necessary to confirm findings. It is worth noting that patients classified as “mild” showed pneumonia by imaging and fever, and in accordance with current classifications this would be consistent with “moderate” cases. Hence it would be more appropriate to refer to the groups as “moderate” vs “severe/critical”. Furthermore, there are several limitations that could impact the interpretation of the results: e.g. classification of patients was based on symptoms presented on admission and not based on disease progression, small sample size, especially the number of ‘severe’ cases (with no deaths among these patients). Given the small sample size, the proposed NLR and age cut offs might not hold for a slightly different set of patients. For example, in a study of >400 patients, ‘non-severe’ and ‘severe’ NLR were 3.2 and 5.5, respectively ([2175](#)).

14.10.4 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.11 Characteristics of lymphocyte subsets and cytokines in peripheral blood of 123 hospitalized patients with 2019 novel coronavirus pneumonia (NCP)

Wan et. al. *medRxiv* ([2176](#))

14.11.1 Keywords

- Cytokines
- lymphocyte subsets
- CD8 + T
- B cells
- NK cells,
- PBMCs
- IL-6
- IL-10

14.11.2 Main Findings

The authors analyzed lymphocyte subsets and cytokines of 102 patients with mild disease and 21 with severe disease. CD8+T cells and CD4+T cells were significantly reduced in both cohort, particularly in severe patients. The cytokines IL6 and IL10 were significantly elevated in severe patients as compared to mild. No significant differences were observed in frequency of B cells and NK cells.

The authors argue that the measurement of T cell frequencies and cytokine levels of IL6 and IL10 can be used to predict progression of disease from Mild to severe Cov-2 infection.

14.11.3 Limitations

The study demonstrates in a limited cohort similar associations to several other reported studies. The authors didn't compare the changes in lymphocyte and cytokine with healthy individual (Covid-19 Negative) rather used an internal standard value. The recently preprint in LANCET shows The degree of lymphopenia and a pro-inflammatory cytokine storm is higher in severe COVID-19 patients than in mild cases, and is associated with the disease severity ([2177](#)).

14.11.4 Significance

This translational data identifies key cytokines and lymphopenia associated with disease severity although mechanism and key cellular players are still unknown. Higher level IL-6 production in severe patient suggests potential role of Tocilizumab (anti-IL6R) biologic although clinical trial will be necessary.

14.11.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.12 Epidemiological and Clinical Characteristics of 17 Hospitalized Patients with 2019 Novel Coronavirus Infections Outside Wuhan, China

Li et al. *medRxiv* ([2178](#))

14.12.1 Keywords

- epidemiology
- clinical characteristics

14.12.2 Major Findings

These authors looked at 17 hospitalized patients with COVID-19 confirmed by RT-PCR in Dazhou, Sichuan. Patients were admitted between January 22 and February 10 and the final data were collected on February 11. Of the 17 patients, 12 remained hospitalized while 5 were discharged after meeting national standards. The authors observed no differences based on the sex of the patients but found that the discharged patients were younger in age ($p = 0.026$) and had higher lymphocyte counts ($p = 0.005$) and monocyte counts ($p = 0.019$) upon admission.

14.12.3 Limitations

This study is limited in the sample size of the study and the last data collection point was only one day after some of the patients were admitted.

14.12.4 Significance

These findings have been somewhat supported by subsequent studies that show that older age and an immunocompromised state are more likely to result in a more severe clinical course with COVID-19. However, other studies have been published that report on larger numbers of cases.

14.12.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.13 ACE2 Expression in Kidney and Testis May Cause Kidney and Testis Damage After 2019-nCoV Infection

14.13.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- kidney
- testis
- ACE2
- scRNAseq

14.13.2 Main Findings

- Study used online datasets (scRNAseq GSE131685, scRNAseq GSE107585, Human Protein Atlas, GTEx portal, CCLE) to analyze ACE2 expression in different human organs.
- Study re-analyzed three clinical datasets (n=6, n=99, and n=41) to show 3~10% of 2019-nCoV patients present with abnormal renal function.
- results indicate ACE2 highly expressed in renal tubular cells, Leydig cells and seminiferous ductal cells of testis.

14.13.3 Limitations

- Very preliminary transcript/protein dataset analysis in healthy cohorts; does not necessarily translate to actual viral tropism and permissiveness.
- Clinically, would be important to determine with larger longitudinal dataset if SARS-CoV-2 infection changes sperm quality or testicular inflammation.
- Similarly, would be important to determine if simultaneous HBV or syphilis infection and orchitis impacts SARS-CoV-2 severity.
- Examination and follow-up of renal function and viral orchitis/sperm quality of CoVID-19 patients not done in this preliminary study.

14.13.4 Significance

- Kidney ACE2 result supports other concurrent sequencing studies ([2171](#)) and clinical reports of abnormal renal function or even kidney damage in patients infected with 2019-nCoV ([2169](#)).
- High ACE2 expression in testis suggests potential tropism of the virus to testicular tissues and indicates potential risks for male fertility. Viral orchitis reported for SARS-CoV previously [1], but no clear evidence so far of infertility in SARS, MERS or CoVID-19 patients.

14.13.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.14 Aberrant pathogenic GM-CSF+ T cells and inflammatory CD14+CD16+ monocytes in severe pulmonary syndrome patients of a new coronavirus

([2180](#))

14.14.1 Keywords

- immunopathology
- Th1
- inflammatory monocytes
- GM-CSF
- IFN- γ
- IL-6

14.14.2 Main Findings

The authors of this study sought to characterize the immune mechanism causing severe pulmonary disease and mortality in 2019-nCoV (COVID-19) patients. Peripheral blood was collected from hospitalized ICU (n=12) and non-ICU (n=21) patients with confirmed 2019-nCoV and from healthy controls (n=10) in The First Affiliated Hospital of University of Science and Technology China (Hefei, Anhui). Immune analysis was conducted by flow cytometry. 2019-nCoV patients had decreased lymphocyte, monocyte, and CD4 T cell counts compared to healthy controls. ICU patients had fewer lymphocytes than non-ICU patients. CD4 T cells of 2019-nCoV patients expressed higher levels of activation markers (OX40, CD69, CD38, CD44) and exhaustion markers (PD-1 and Tim3) than those of healthy controls. CD4 cells of ICU patients expressed significantly higher levels of OX40, PD-1, and Tim3 than those of non-ICU patients. 2019-nCoV patients had higher percentages of CD4 T cells co-expressing GM-CSF and IL-6 compared to healthy controls, while ICU patients had a markedly higher percentage of GM-CSF+ IFN- γ + CD4 T cells than non-ICU patients. The CD4 T cells of nCoV patients and healthy controls showed no differences in TNF- α secretion.

The CD8 T cells of 2019-nCoV patients also showed higher expression of activation markers CD69, CD38, and CD44, as well as exhaustion markers PD-1 and Tim3, compared to healthy controls. CD8 T cells of ICU patients expressed higher levels of GM-CSF than those of non-ICU patients and healthy controls. No IL-6 or TNF- α was found in the CD8 T cells of any group. There were no differences in numbers of NK cells or B cells in 2019-nCoV patients and healthy controls, nor was there any GM-CSF or IL-6 secretion from these cells in either group.

Percentages of CD14+ CD16+ GM-CSF+ and CD14+ CD16+ IL-6+ inflammatory monocytes were significantly increased in nCoV patients compared to healthy controls; in particular, patients in the ICU had greater percentages of CD14+ CD16+ IL-6+ monocytes than non-ICU patients. The authors suggest that in 2019-nCoV patients, pathogenic Th1 cells produce GM-CSF, recruiting CD14+ CD16+ inflammatory monocytes that secrete high levels of IL-6. These may enter pulmonary circulation and damage lung tissue while initiating the cytokine storm that causes mortality in severe cases. This is consistent with the cytokine storm seen in similar coronaviruses, as IL-6, IFN- γ , and GM-CSF are key inflammatory mediators seen in patients with SARS-CoV-1 and MERS-CoV.

14.14.3 Limitations

Though the results of this study open questions for further investigation, this is an early study on a small cohort of patients, and as such there are a number of limitations. The study included only 12 ICU patients and 21 non-ICU patients, and ideally would be repeated with a much larger patient cohort. Though the authors make claims about differences in lymphocyte and monocyte counts between patients and healthy controls, they did not report baseline laboratory findings for the control group. Additionally, severity of disease was classified based on whether or not patients were in the ICU. It would be interesting to contextualize the authors' immunological findings with more specific metrics of disease severity or time course. Noting mortality, time from disease onset, pre-existing conditions, or severity of lung pathology in post-mortem tissue samples would paint a fuller picture of how to assess risk level and the relationship between severity of disease and immunopathology. Another limitation is the selection of cytokines and immune markers for analysis, as the selection criteria were based on the cell subsets and cytokine storm typically seen in SARS-CoV-1 and MERS-CoV patients. Unbiased cytokine screens and immune profiling may reveal novel therapeutic targets that were not included in this study.

14.14.4 Significance

This study identifies potential therapeutic targets that could prevent acute respiratory disease syndrome (ARDS) and mortality in patients most severely affected by COVID-19. The authors propose testing monoclonal antibodies against IL6-R or GM-CSF to block recruitment of inflammatory monocytes and the subsequent cytokine storm in these patients.

14.14.5 Credit

Review by Gabrielle Lubitz as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.15 Clinical Characteristics of 2019 Novel Infected Coronavirus Pneumonia : A Systemic Review and Meta-analysis

14.15.1 Keywords

- White Blood Cells
- Lymphocytes
- Neutrophils

14.15.2 Main Findings

The authors performed a meta analysis of literature on clinical, laboratory and radiologic characteristics of patients presenting with pneumonia related to SARSCoV2 infection, published up to Feb 6 2020. They found that symptoms that were mostly consistent among studies were sore throat, headache, diarrhea and rhinorrhea. Fever, cough, malaise and muscle pain were highly variable across studies. Leukopenia (mostly lymphocytopenia) and increased white blood cells were highly variable across studies. They identified three most common patterns seen on CT scan, but there was high variability across studies. Consistently across the studies examined, the authors found that about 75% of patients need supplemental oxygen therapy, about 23% mechanical ventilation and about 5% extracorporeal membrane oxygenation (ECMO). The authors calculated a staggering pooled mortality incidence of 78% for these patients.

14.15.3 Limitations

The authors mention that the total number of studies included in this meta analysis is nine, however they also mentioned that only three studies reported individual patient data. It is overall unclear how many patients in total were included in their analysis. This is mostly relevant as they reported an incredibly high mortality (78%) and mention an absolute number of deaths of 26 cases overall. It is not clear from their report how the mortality rate was calculated.

The data is based on reports from China and mostly from the Wuhan area, which somewhat limits the overall generalizability and applicability of these results.

14.15.4 Significance

This meta analysis offers some important data for clinicians to refer to when dealing with patients with COVID-19 and specifically with pneumonia. It is very helpful to set expectations about the course of the disease.

14.15.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.16 Longitudinal characteristics of lymphocyte responses and cytokine profiles in the peripheral blood of SARS-CoV-2 infected patients

Liu et al. *medRxiv* ([2182](#))

14.16.1 Keywords

- Lymphopenia
- Neutrophil to CD8 T cell ratio (N8R)
- inflammatory cytokines

14.16.2 Main Findings

Liu et al. enrolled a cohort of 40 patients from Wuhan including 27 mild cases and 13 severe cases of COVID-19. They performed a 16-day kinetic analysis of peripheral blood from time of disease onset. Patients in the severe group were older (medium age of 59.7, compared to 48.7 in mild group) and more likely to have hypertension as a co-morbidity. Lymphopenia was observed in 44.4% of the mild patients and 84.6% of the severe patients. Lymphopenia was due to low T cell count, specially CD8 T cells. Severe patients showed higher neutrophil counts and an increase of cytokines in the serum (IL2, IL6, IL10 and IFN γ). The authors measured several other clinical laboratory parameters were also higher in severe cases compared to mild, but concluded that neutrophil to CD8 T cell ratio (N8R) as the best prognostic factor to identify the severe cases compared to other receiver operating characteristic (ROC).

14.16.3 Limitations

This was a small cohort (N=40), and two of the patients initially included in the severe group (N=13) passed away and were excluded from the analysis due to lack of longitudinal data. However, it would be most important to be able to identify patients with severe disease with higher odds of dying. It seems that the different time points analyzed relate to hospital admission, which the authors describe as disease onset. The time between first symptoms and first data points is not described. It would have been important to analyze how the different measured parameters change according to health condition, and not just time (but that would require a larger cohort). The predictive value of N8R compared to the more commonly used NLR needs to be assessed in other independent and larger cohorts. Lastly, it is important to note that pneumonia was detected in patients included in the “mild” group, but according to the Chinese Clinical Guidance for COVID-19 Pneumonia Diagnosis and Treatment (7th edition) this group should be considered “moderate”.

14.16.4 Significance

Lymphopenia and cytokine storm have been described to be detrimental in many other infections including SARS-CoV1 and MERS-CoV. However, it was necessary to confirm that this dramatic immune response was also observed in the SARS-CoV2 infected patients. These results and further validation of the N8R ratio as a predictor of disease severity will contribute for the management of COVID19 patients and potential development of therapies.

14.16.5 Credit

Review by Pauline Hamon as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.17 Clinical and immunologic features in severe and moderate forms of Coronavirus Disease 2019

Chen et al. *medRxiv* ([2183](#))

14.17.1 Keywords

- severe disease
- lymphocytes
- cytokines
- IFN γ
- CD4 Tcells
- HLA-DR CD8
- Tcells

14.17.2 Main Findings

This study retrospectively evaluated clinical, laboratory, hematological, biochemical and immunologic data from 21 subjects admitted to the hospital in Wuhan, China (late December/January) with confirmed SARS-CoV-2 infection. The aim of the study was to compare 'severe' (n=11, ~64 years old) and 'moderate' (n=10, ~51 years old) COVID-19 cases. Disease severity was defined by patients' blood oxygen level and respiratory output. They were classified as 'severe' if SpO₂ 93% or respiratory rates 30 per min.

In terms of the clinical laboratory measures, 'severe' patients had higher CRP and ferritin, alanine and aspartate aminotransferases, and lactate dehydrogenase but lower albumin concentrations.

The authors then compared plasma cytokine levels (ELISA) and immune cell populations (PBMCs, Flow Cytometry). 'Severe' cases had higher levels of IL-2R, IL-10, TNF α , and IL-6 (marginally significant). For the immune cell counts, 'severe' group had higher neutrophils, HLA-DR⁺ CD8 T cells and total B cells; and lower total lymphocytes, CD4 and CD8 T cells (except for HLA-DR⁺), CD45RA Tregs, and IFN γ -expressing CD4 T cells. No significant differences were observed for IL-8, counts of NK cells, CD45⁺RO Tregs, IFN γ -expressing CD8 T and NK cells.

14.17.3 Limitations

Several potential limitations should be noted: 1) Blood samples were collected 2 days post hospital admission and no data on viral loads were available; 2) Most patients were administered medications (e.g. corticosteroids), which could have affected lymphocyte counts. Medications are briefly mentioned in the text of the manuscript; authors should include medications as part of Table 1. 3) 'Severe' cases were significantly older and 4/11 'severe' patients died within 20 days. Authors should consider a sensitivity analysis of biomarkers with the adjustment for patients' age.

14.17.4 Significance

Although the sample size was small, this paper presented a broad range of clinical, biochemical, and immunologic data on patients with COVID-19. One of the main findings is that SARS-CoV-2 may affect T lymphocytes, primarily CD4+ T cells, resulting in decreased IFN γ production. Potentially, diminished T lymphocytes and elevated cytokines can serve as biomarkers of severity of COVID-19.

14.17.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.18 SARS-CoV-2 and SARS-CoV Spike-RBD Structure and Receptor Binding Comparison and Potential Implications on Neutralizing Antibody and Vaccine Development

Sun et al. *bioRxiv* ([913](#))

14.18.1 Keywords

- SARS-CoV
- SARS-CoV-2
- ACE2
- Spike (S) protein
- receptor binding domain (RBD)
- receptor binding motif (RBM)
- neutralizing antibody

14.18.2 Main Findings

This study compared the structure of SARS-CoV and SARS-CoV-2 Spike (S) protein receptor binding domain (RBD) and interactions with ACE2 using computational modeling, and interrogated cross-reactivity and cross-neutralization of SARS-CoV-2 by antibodies against SARS-CoV. While SARS-

CoV and SARS-CoV-2 have over 70 % sequence homology and share the same human receptor ACE2, the receptor binding motif (RBM) is only 50% homologous.

Computational prediction of the SARS-CoV-2 and ACE2 interactions based on the previous crystal structure data of SARS-CoV, and measurement of binding affinities against human ACE2 using recombinant SARS-CoV and SARS-CoV-2 S1 peptides, demonstrated similar binding of the two S1 peptides to ACE2, explaining the similar transmissibility of SARS-CoV and SARS-CoV-2 and consistent with previous data (Wall et al Cell 2020).

The neutralization activity of SARS-CoV-specific rabbit polyclonal antibodies were about two-order of magnitude less efficient to neutralize SARS-CoV-2 than SARS-CoV, and four potentially neutralizing monoclonal antibodies against SARS-CoV had poor binding and neutralizing activity against SARS-CoV-2. In contrast, 3 poor SARS-CoV-binding monoclonal antibodies show some efficiency to bind and neutralize SARS-CoV-2. The results suggest that antibodies to more conserved regions outside the RBM motif might possess better cross-protective neutralizing activities between two strains.

14.18.3 Limitations

It would have been helpful to show the epitopes recognized by the monoclonal antibodies tested on both SARS-CoV, SARS-CoV-2 to be able to make predictions for induction of broadly neutralizing antibodies. The data on monoclonal antibody competition with ACE2 for binding to SARS-CoV RBD should have also included binding on SARS-CoV2, especially for the three monoclonal antibodies that showed neutralization activity for SARS-CoV2. Because of the less homology in RBM sequences between viruses, it still may be possible that these antibodies would recognize the ACE2 RBD in SARS-CoV-2.

14.18.4 Significance

It is noteworthy that immunization to mice and rabbit with SARS-CoV S1 or RBD protein could induce monoclonal antibodies to cross-bind and cross-neutralize SARS-CoV-2 even if they are not ACE2-blocking. If these types of antibodies could be found in human survivors or in the asymptomatic populations as well, it might suggest that exposure to previous Coronavirus strains could have induced cross-neutralizing antibodies and resulted in the protection from severe symptoms in some cases of SARS-CoV2.

14.18.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.19 Protection of Rhesus Macaque from SARS-Coronavirus challenge by recombinant adenovirus vaccine

Chen et al. *bioRxiv* ([2184](#))

14.19.1 Keywords

- SARS-CoV-1
- rhesus macaque
- recombinant adenovirus vaccine

14.19.2 Main Findings

Rhesus macaques were immunized intramuscularly twice (week 0 and week 4) with SV8000 carrying the information to express a S1-orf8 fusion protein and the N protein from the BJ01 strain of SARS-CoV-1. By week 8, immunized animals had signs of immunological protection (IgG and neutralization titers) against SARS-CoV-1 and were protected against challenge with the PUMC-1 strain, with fewer detectable symptoms of respiratory distress, lower viral load, shorter periods of viral persistence, and less pathology in the lungs compared to non-immunized animals.

14.19.3 Limitations

The authors should write clearer descriptions of the methods used in this article. They do not describe how the IgG titers or neutralization titers were determined. There are some issues with the presentation of data, for example, in Figure 1a, y-axis should not be Vmax; forming cells and 1d would benefit from showing error bars. Furthermore, although I inferred that the animals were challenged at week 8, the authors did not explicitly detail when the animals were challenged. The authors should explain the design of their vaccine, including the choice of antigens and vector. The authors also do not include a description of the ethical use of animals in their study.

14.19.4 Significance

The authors describe a vaccine for SARS-CoV-1 with no discussion of possible implications for the current SARS-CoV-2 pandemic. Could a similar vaccine be designed to protect against SARS-CoV-2 and would the concerns regarding emerging viral mutations that the authors describe as a limitation for SARS-CoV-1 also be true in the context of SARS-CoV-2?

14.19.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.20 Reduction and Functional Exhaustion of T cells in Patients with Coronavirus Disease 2019 (COVID-19)

(2185)

14.20.1 Keywords

- T cell exhaustion
- T cell lymphopenia
- IL-6
- IL-10
- TNF- α

14.20.2 Main Findings

Based on a retrospective study of 522 COVID patients and 40 healthy controls from two hospitals in Wuhan, China, authors show both age-dependent and clinical severity-dependent decrease in T cell numbers with elderly patients and patients who are in ICU-care showing the most dramatic decrease in T cell counts. Cytokine profiling of COVID patients reveal that TNF- α , IL-6 and IL-10 are increased in infected patients with patients in the ICU showing the highest levels. Interestingly, these three cytokine levels were inversely correlated with T cell counts and such inverse relationship was preserved throughout the disease progression. Surface staining of exhaustion markers (PD-1 and Tim-3) and flow cytometry of stained peripheral blood of 14 patients and 3 healthy volunteers demonstrate that T cells of COVID patients have increased expression of PD-1 with patients in ICU having the highest number of CD8⁺PD-1⁺ cells than their counterparts in non-ICU groups.

14.20.3 Limitations

Compared to the number of patients, number of control (n= 40) is small and is not controlled for age. Additional data linking inflammatory cytokines and the quality of the adaptive response including humoral and antigen specific T cell response is much needed. T cell exhaustion study relies on marker-dependent labeling of T cell functionality of a very limited sample size (n=17) —a functional/mechanistic study of these T cells from PBMCs would have bolstered their claims.

14.20.4 Significance

Limited but contains interesting implications. It is already known in literature that in the context of acute respiratory viral infections CD8 T cells exhibit exhaustion-like phenotypes which further underscores the importance of mechanistic studies that can elucidate how COVID infection leads to lymphopenia and T cell exhaustion-like phenotype.

However, as authors have noted, the data does point to an interesting question: How these inflammatory cytokines (TNF- α , IL-6 and IL-10) correlate with or affect effective viral immunity and what types of cells produce these cytokines? Answering that question will help us refine our targets for immune-modulatory therapies especially in patients suffering from cytokine storms.

14.20.5 Credit

This review by Chang Moon was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.21 Clinical Characteristics of 25 death cases infected with COVID-19 pneumonia: a retrospective review of medical records in a single medical center, Wuhan, China

([2186](#))

14.21.1 Keywords

- COVID-19
- pneumonia
- hypertension
- diabetes
- biomarker
- neutrophilia
- lymphopenia

14.21.2 Main Findings

Most common chronic conditions among 25 patients that died from COVID-19 related respiratory failure were hypertension (64%) and diabetes (40%). Disease progression was marked by progressive organ failure, starting first with lung dysfunction, then heart (e.g. increased cTnI and pro-BNP), followed by kidney (e.g. increased BUN, Cr), and liver (e.g. ALT, AST). 72% of patients had neutrophilia and 88% also had lymphopenia. General markers of inflammation were also increased (e.g. PCT, D-Dimer, CRP, LDH, and SAA).

14.21.3 Limitations

The limitations of this study include small sample size and lack of measurements for some tests for several patients. This study would also have been stronger with comparison of the same measurements to patients suffering from less severe disease to further validate and correlate proposed biomarkers with disease severity.

14.21.4 Significance

This study identifies chronic conditions (i.e. hypertension and diabetes) that strongly correlates with disease severity. In addition to general markers of inflammation, the authors also identify concomitant neutrophilia and lymphopenia among their cohort of patients. This is a potentially interesting immunological finding because we would typically expect increased lymphocytes during a viral infection. Neutrophilia may also be contributing to cytokine storm. In addition, PCT was elevated in 90.5% of patients, suggesting a role for sepsis or secondary bacterial infection in COVID-19 related respiratory failure.

14.21.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.22 SARS-CoV-2 infection does not significantly cause acute renal injury: an analysis of 116 hospitalized patients with COVID-19 in a single hospital, Wuhan, China

[\(2187\)](#)

14.22.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- kidney
- clinical
- longitudinal

14.22.2 Main Findings

- Clinical data from 116 hospitalized CoVID-19 patients analyzed over 4 weeks for correlation with renal injury. Comorbidities included chronic renal failure (CRF) in 5 patients (4.3%).
- 10.8% of patients with no prior kidney disease showed elevations in blood urea or creatinine, and 7.2% of patients with no prior kidney disease showed albuminuria.
- Patients with pre-existing CRF underwent continuous renal replacement therapy (CRRT) alongside CoVID-19 treatment. Renal functions remained stable in these patients.
- All 5 patients with CRF survived CoVID-19 therapy without progression to ARDS or worsening of CRF.

14.22.3 Limitations

- Renal injury biomarkers in patients with incipient kidney abnormalities not tabulated separately, making overall data hard to interpret. It will be critical to separately examine kidney function (BUN, urine creatinine and eGFR) in patients that developed any kidney abnormalities (7.2~10.8% of cohort).
- No information on type of CoVID-19 therapy used across cohort; will be useful to correlate how treatment modality influences kidney function (and other parameters).
- Invokes previous clinical-correlation studies that indicate low instances of kidney damage([2188](#), [2189](#)), but those studies did not track longitudinal urine samples for acute renal injury markers and viral shedding.
- CRRT in patients with CRF is standard therapy irrespective of CoVID-19 status; it will be important to compare clinical parameters of these patients (n=5) with virus-naïve CRF patients (none in this study) to make any meaningful conclusions.

14.22.4 Significance

- This study argues that renal impairment is uncommon in CoVID-19 and not associated with high mortality, in stark contrast with a concurrent study ([2169](#)). If supported by further studies, this argues kidney impairment is secondary to cytokine storm/inflammation-induced organ failure, and not due to direct viral replication.
- Will be important to comprehensively characterize large-datasets of CoVID-19 patients to conclude if kidney function actively disrupted due to viral infection.

14.22.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.23 Potential T-cell and B-Cell Epitopes of 2019-nCoV

([2190](#))

14.23.1 Keywords

- COVID-19
- vaccine
- epitopes
- spike protein
- MHC-I
- MHC-II
- neutralizing antibodies

- ACE2

14.23.2 Main Findings

The authors use 2 neural network algorithms, NetMHCpan4 and MARIA, to identify regions within the COVID-19 genome that are presentable by HLA. They identify 405 viral epitopes that are presentable on MHC-I and MHC-II and validate using known epitopes from SARS-CoV. To determine whether immune surveillance drives viral mutations to evade MHC presentation, the authors analyzed 68 viral genomes from 4 continents. They identified 93 point mutations that occurred preferentially in regions predicted to be presented by MHC-I ($p=0.02$) suggesting viral evolution to evade CD8 T-cell mediated killing. 2 nonsense mutations were also identified that resulted in loss of presentation of an associated antigen (FGDSVEEVL) predicted to be good antigen for presentation across multiple HLA alleles.

To identify potential sites of neutralizing antibody binding, the authors used homology modeling to the SARS-CoV's spike protein (S protein) to determine the putative structure of the CoV2 spike protein. They used Discotope2 to identify antibody binding sites on the protein surface in both the down and up conformations of the S protein. The authors validate this approach by first identifying antibody binding site in SARS-CoV S protein. In both the down and up conformation of the CoV2 S protein, the authors identified a potential antibody binding site on the S protein receptor binding domain (RBD) of the ACE2 receptor (residues 440-460, 494-506). While RBDs in both SARS-CoV and CoV2 spike proteins may be important for antibody binding, the authors note that SARS-CoV has larger attack surfaces than CoV2. These results were later validated on published crystal structures of the CoV2 S protein RBD and human ACE2. Furthermore, analysis of 68 viral genomes did not identify any mutations in this potential antibody binding site in CoV2.

Finally, the authors compile a list of potential peptide vaccine candidates across the viral genome that can be presented by multiple HLA alleles. Several of the peptides showed homology to SARS-CoV T-cell and B-cell epitopes.

14.23.3 Limitations

While the authors used computational methods of validation, primarily through multiple comparisons to published SARS-CoV structures and epitopes, future work should include experimental validation of putative T-cell and B-cell epitopes.

14.23.4 Significance

The authors identified potential T-cell and B-cell epitopes that may be good candidates for peptide based vaccines against CoV2. They also made interesting observations in comparing SARS-CoV and CoV2 potential antibody binding sites, noting that SARS-CoV had larger attack surfaces for potential neutralizing antibody binding. One of the highlights of this paper was the authors' mutation analysis of 68 viral genomes from 4 continents. This analysis not only validated their computational method for identifying T-cell

epitopes, but showed that immune surveillance likely drives viral mutation in MHC-I binding peptides. The smaller attack surface may point to potential mechanisms of immune evasion by CoV2. However, absence of mutations in the RBD of CoV2 and the small number of mutations in peptides presentable to T cells suggests that vaccines against multiple epitopes could still elicit robust immunity against CoV2.

14.23.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.24 Structure, Function, and Antigenicity of the SARSCoV-2 Spike Glycoprotein

Walls et al. *bioRxiv.* ([2191](#)) now ([25](#))

14.24.1 Keywords

- binding affinity
- antigenicity
- neutralizing antibody

14.24.2 Main Findings

The authors highlight a human angiotensin-converting enzyme 2 (hACE2), as a potential receptor used by the current Severe Acute respiratory syndrome coronavirus-2 (SARS-CoV-2) as a host factor that allows the virus target human cells. This virus-host interaction facilitates the infection of human cells with a high affinity comparable with SARS-CoV. The authors propose this mechanism as a probable explanation of the efficient transmission of SARS-CoV-2 between humans. Besides, Walls and colleagues described SARS-CoV-2 S glycoprotein S by Cryo-EM along with neutralizing polyclonal response against SAR-CoV-2 S from mice immunized with SAR-CoV and blocking SAR-CoV-2 S-mediated entry into VeroE6 infected cells.**

14.24.3 Limitations

The SARS-CoV-2 depends on the cell factors ACE2 and TMPRSS2, this last, according to a recent manuscript by Markus Hoffman et al., *Cell*, 2020. The authors used green monkey (VeroE6) and hamster (BHK) cell lines in the experiments to drive its conclusions to humans; however, it is well known the caucasian colon adenocarcinoma human cell line (CaCo-2), highly express the hACE2 receptor as the TMPRSS2 protease as well. In humans, ACE2 protein is highly expressed in the gastrointestinal tract, which again, makes the CaCo-2 cell line suitable for the following SARS-CoV-2 studies.

14.24.4 Significance

The results propose a functional receptor used by SARS-CoV-2 to infect humans worldwide and defining two distinct conformations of spike (S) glycoprotein by cryogenic electron microscopy (Cryo-EM). This study might help establish a precedent for initial drug design and treatment of the current global human coronavirus epidemic.

14.24.5 Credit

Review by postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.25 Breadth of concomitant immune responses underpinning viral clearance and patient recovery in a non-severe case of COVID-19

Thevarajan et al. *medRxiv* ([2192](#))

14.25.1 Keywords

- IgG
- IgM
- T_H cells
- NK cells
- SNP

14.25.2 Main Findings

The authors characterized the immune response in peripheral blood of a 47-year old COVID-19 patient.

SARS-CoV2 was detected in nasopharyngeal swab, sputum and faeces samples, but not in urine, rectal swab, whole blood or throat swab. 7 days after symptom onset, the nasopharyngeal swab test turned negative, at day 10 the radiography infiltrates were cleared and at day 13 the patient became asymptomatic.

Immunofluorescence staining shows from day 7 the presence of **COVID-19-binding IgG and IgM** antibodies in plasma, that increase until day 20.

Flow cytometry on whole blood reveals a plasmablast peak at day 8, a gradual increase in T follicular helper cells, stable HLA-DR⁺ NK frequencies and decreased monocyte frequencies compared to healthy counterparts. The expression of CD38 and HLA-DR peaked on T cells at D9 and was associated with higher production of cytotoxic mediators by CD8⁺ T cells.

IL-6 and IL-8 were undetectable in plasma.

The authors further highlight the presence of the **IFITM3 SNP-rs12252-C/C variant** in this patient, which is associated with higher susceptibility to influenza virus.

14.25.3 Limitations

These results need to be confirmed in additional patients.

COVID-19 patients have increased infiltration of macrophages in their lungs ([2193](#)). Monitoring monocyte proportions in blood earlier in the disease might help to evaluate their eventual migration to the lungs.

The stable concentration of HLA-DR⁺ NK cells in blood from day 7 is not sufficient to rule out NK cell activation upon SARS-CoV2 infection. In response to influenza A virus, NK cells express higher levels of activation markers CD69 and CD38, proliferate better and display higher cytotoxicity ([2194](#)). Assessing these parameters in COVID-19 patients is required to better understand NK cell role in clearing this infection.

Neutralization potential of the COVID-19-binding IgG and IgM antibodies should be assessed in future studies.

This patient was able to clear the virus, while presenting a SNP associated with severe outcome following influenza infection. The association between this SNP and outcome upon SARS-CoV2 infection should be further investigated.

14.25.4 Significance

This study is among the first to describe the appearance of COVID-19-binding IgG and IgM antibodies upon infection. The emergence of new serological assays might contribute to monitor more precisely the seroconversion kinetics of COVID-19 patients ([2195](#)). Further association studies between IFITM3 SNP-rs12252-C/C variant and clinical data might help to refine the COVID-19 outcome prediction tools.

14.25.5 Credit

Review by Bérengère Salomé as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.26 The landscape of lung bronchoalveolar immune cells in COVID-19 revealed by single-cell RNA sequencing

Liao et al. *medRxiv* ([2193](#))

14.26.1 Keywords

- COVID-19
- SARS-CoV-2
- Broncho-alveolar lavage
- macrophages
- NK cells
- T cells
- cytokine storm
- scRNAseq

14.26.2 Main Findings

The authors performed single-cell RNA sequencing (scRNAseq) on bronchoalveolar lavage fluid (BAL) from 6 COVID-19 patients (n=3 mild cases, n=3 severe cases). Data was compared to previously generated scRNAseq data from healthy donor lung tissue (n=8).

Clustering analysis of the 6 patients revealed distinct immune cell organization between mild and severe disease. Specifically, they found that transcriptional clusters annotated as tissue resident alveolar macrophages were strongly reduced while monocytes-derived FCN1⁺SPP1⁺ inflammatory macrophages dominated the BAL of patients with severe COVID19 diseases. They show that inflammatory macrophages upregulated interferon-signaling genes, monocytes recruiting chemokines including CCL2, CCL3, CCL4 as well as IL-6, TNF, IL-8 and profibrotic cytokine TGF- β , while alveolar macrophages expressed lipid metabolism genes, such as PPARG.

The lymphoid compartment was overall enriched in lungs from patients. Clonally expanded CD8 T cells were enriched in mild cases suggesting that CD8 T cells contribute to viral clearance as in Flu infection, whereas proliferating T cells were enriched in severe cases.

SARS-CoV-2 viral transcripts were detected in severe patients, but considered here as ambient contaminations.

14.26.3 Limitations

These results are based on samples from 6 patients and should therefore be confirmed in the future in additional patients. Longitudinal monitoring of BAL during disease progression or resolution would have been most useful.

The mechanisms underlying the skewing of the macrophage compartment in patients towards inflammatory macrophages should be investigated in future studies.

Deeper characterization of the lymphoid subsets is required. The composition of the “proliferating” cluster and how these cells differ from conventional T cell clusters should be assessed. NK and CD8 T cell transcriptomic profile, in particular the expression of cytotoxic mediator and immune checkpoint transcripts, should be compared between healthy and diseased lesions.

14.26.4 Significance

COVID-19 induces a robust inflammatory cytokine storm in patients that contributes to severe lung tissue damage and ARDS ([2196](#)). Accumulation of monocyte-derived inflammatory macrophages at the expense of Alveolar macrophages known to play an anti-inflammatory role following respiratory viral infection, in part through the PPAR γ pathway ([2197](#), [2198](#)) are likely contributing to lung tissue injuries. These data suggest that reduction of monocyte accumulation in the lung tissues could help modulate COVID-19-induced inflammation. Further analysis of lymphoid subsets is required to understand the contribution of adaptive immunity to disease outcome.

14.26.5 Credit

Review by Bérengère Salomé and Assaf Magen as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.27 Can routine laboratory tests discriminate 2019 novel coronavirus infected pneumonia from other community-acquired pneumonia?

Pan et al. *medRxiv* ([2199](#))

14.27.1 Keywords

- Routine laboratory testing

14.27.2 Main Findings

In an attempt to use standard laboratory testing for the discrimination between “Novel Coronavirus Infected Pneumonia” (NCIP) and a usual community acquired pneumonia (CAP), the authors compared laboratory testing results of 84 NCIP patients with those of a historical group of 316 CAP patients from 2018 naturally COVID-19 negative. The authors describe significantly lower white blood- as well as red blood- and platelet counts in NCIP patients. When analyzing differential blood counts, lower absolute counts were measured in all subsets of NCIP patients. With regard to clinical chemistry parameters, they found increased AST and bilirubin in NCIP patients as compared to CAP patients.

14.27.3 Limitations

The authors claim to describe a simple method to rapidly assess a pre-test probability for NCIP. However, the study has substantial weakpoints. The deviation in clinical laboratory values in NCIP patients described here can usually be observed in severely ill patients. The authors do not comment on how severely ill the patients tested here were in comparison to the historical control. Thus, the conclusion that the tests discriminate between CAP and NCIP lacks justification.

14.27.4 Significance

The article strives to compare initial laboratory testing results in patients with COVID-19 pneumonia as compared to patients with a usual community acquired pneumonia. The implications of this study for the current clinical situation seem restricted due to a lack in clinical information and the use of a control group that might not be appropriate.

14.27.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.28 Correlation Analysis Between Disease Severity and Inflammation-related Parameters in Patients with COVID-19 Pneumonia

(2200)

14.28.1 Keywords

- cytokine
- COVID-19 pneumonia
- severity
- disease progression

14.28.2 Main Findings

This study is a cross-sectional analysis of 100 patients with COVID-19 pneumonia, divided into mild (n = 34), severe (n = 34), and critical (n = 32) disease status based on clinical definitions.

The criteria used to define disease severity are as follows:

1. *Severe* – any of the following: respiratory distress or respiratory rate ≥ 30 respirations/minute; oxygen saturation $\leq 93\%$ at rest; oxygen partial pressure (PaO₂)/oxygen concentration (FiO₂) in arterial blood $\leq 300\text{mmHg}$, progression of disease on imaging to $>50\%$ lung involvement in the short term.
2. *Critical* – any of the following: respiratory failure that requires mechanical ventilation; shock; other organ failure that requires treatment in the ICU.
3. Patients with pneumonia who test positive for COVID-19 who do not have the symptoms delineated above are considered *mild*.

Peripheral blood inflammatory markers were correlated to disease status. Disease severity was significantly associated with levels of IL-2R, IL-6, IL-8, IL-10, TNF- α , CRP, ferroprotein, and procalcitonin. Total WBC count, lymphocyte count, neutrophil count, and eosinophil count were also significantly correlated with disease status. Since this is a retrospective, cross-sectional

study of clinical laboratory values, these data may be extrapolated for clinical decision making, but without studies of underlying cellular causes of these changes this study does not contribute to a deeper understanding of SARS-CoV-2 interactions with the immune system.

It is also notable that the mean age of patients in the mild group was significantly different from the mean ages of patients designated as severe or critical ($p < 0.001$). The mean patient age was not significantly different between the severe and critical groups. However, IL-6, IL-8, procalcitonin (Table 2), CRP, ferroprotein (Figure 3A, 3B), WBC count, and neutrophil count (Figure 4A, 4B) were all significantly elevated in the critical group compared to severe. These data suggest underlying differences in COVID-19 progression that is unrelated to age.

14.28.3 Significance

Given the inflammatory profile outlined in this study, patients who have mild or severe COVID-19 pneumonia, who *also* have any elevations in the inflammatory biomarkers listed above, should be closely monitored for potential progression to critical status.

14.28.4 Credit

This review by JJF was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.29 An Effective CTL Peptide Vaccine for Ebola Zaire Based on Survivors' CD8+ Targeting of a Particular Nucleocapsid Protein Epitope with Potential Implications for COVID-19 Vaccine Design

Herst et al. *bioRxiv* ([2201](#))

14.29.1 Keywords

- Peptide vaccine
- Ebolavirus
- nucleocapsid
- epitope
- vaccine design
- microsphere

14.29.2 Main Findings

Vaccination of mice with a single dose of a 9-amino-acid peptide NP44-52 located in a conserved region of ebolavirus (EBOV) nucleocapsid protein (NP) confers CD8+ T-cell-mediated immunity against mouse adapted EBOV

(maEBOV). Bioinformatic analyses predict multiple conserved CD8+ T cell epitopes in the SARS-CoV-2 NP, suggesting that a similar approach may be feasible for vaccine design against SARS-CoV-2.

The authors focus on a site within a 20-peptide region of EBOV NP which was commonly targeted by CD8+ T cells in a group of EBOV survivors carrying the HLA-A*30:01:01 allele. To justify the testing of specific vaccine epitopes in a mouse challenge setting, the authors cite known examples of human pathogen-derived peptide antigens that are also recognized by C57BL/6 mice, as well as existing data surrounding known mouse immunogenicity of peptides related to this EBOV NP region. Testing 3 distinct 9mer peptides over an 11 amino-acid window and comparing to vaccination with the 11mer with a T-cell reactivity readout demonstrated that optimizing peptide length and position for immunogenicity may be crucial, likely due to suboptimal peptide processing and MHC-class-I loading.

Vaccines for maEBOV challenge studies were constructed by packaging NP44-52 in d,l poly(lactic-co-glycolic) acid microspheres. CpG was also packaged within the microspheres, while Monophosphoryl Lipid A (a TLR4 ligand) was added to the injectate solution. A second peptide consisting of a predicted MHC-II epitope from the EBOV VG19 protein was added using a separate population of microspheres, and the formulation was injected by intraperitoneal administration. The vaccine was protective against a range of maEBOV doses up to at least 10,000 PFU. Survival was anticorrelated with levels of IL6, MCP-1 (CCL2), IL9, and GM-CSF, which recapitulated trends seen in human EBOV infection.

While HLA-A*30:01:01 is only present in a minority of humans, the authors state that MHC binding algorithms predict NP44-52 to be a strong binder of a set of more common HLA-A*02 alleles. The authors predict that a peptide vaccine based on the proposed formulation could elicit responses in up to 50% of people in Sudan or 30% of people in North America.

SARS-CoV-2 NP, meanwhile, has conserved regions which may provide peptide-vaccine candidates. Scanning the SARS-CoV-2 NP sequence for HLA-binding 9mers identified 53 peptides with predicted binding affinity < 500nM, including peptides that are predicted to bind to HLA-class-I alleles of 97% of humans, 7 of which have previously been tested *in-vitro*.

The results support previously appreciated correlations between certain cytokines and disease severity, specifically IL6 which relates to multiple trial therapies. Prediction of HLA-class-I binding of SARS-CoV-2 NP peptides suggests the plausibility of a peptide vaccine targeting conserved regions of SARS-CoV-2 NP although further validation in previously infected patient samples will be essential.

14.29.3 Credit

Review by Andrew M. Leader as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.30 Epitope-based peptide vaccines predicted against novel coronavirus disease caused by SARS-CoV-2

Li et al. *bioRxiv*. ([2202](#))

14.30.1 Keywords

- SARS-CoV-2
- immune-informatics
- vaccine design
- T cell epitope
- B cell epitope

14.30.2 Main Findings

This study employs a series of bioinformatic pipelines to identify T and B cell epitopes on spike (S) protein of SARS-CoV-2 and assess their properties for vaccine potential. To identify B cell epitopes, they assessed structural accessibility, hydrophilicity, and beta-turn and flexibility which are all factors that promote their targeting by antibodies. To identify T cell epitopes, they filtered for peptides with high antigenicity score and capacity to bind 3 or more MHC alleles. Using the protein digest server, they also demonstrated that their identified T and B cell epitopes are stable, having multiple non-digesting enzymes per epitope. Epitopes were also determined to be non-allergenic and non-toxin as assessed by Allergen FP 1.0 and ToxinPred, respectively. For T cell epitopes, they assessed the strength of epitope-HLA interaction via PepSite. Overall, they predict four B cell and eleven T cell epitopes (two MHC I and nine MHC II binding) to pass stringent computational thresholds as candidates for vaccine development. Furthermore, they performed sequence alignment between all identified SARS-CoV-2 S protein mutations and predicted epitopes, and showed that the epitopes are conserved across 134 isolates from 38 locations worldwide. However, they report that these conserved epitopes may soon become obsolete given the known mutation rate of related SARS-CoV is estimated to be 4×10^{-4} /site/year, underscoring the urgency of anti-viral vaccine development.

14.30.3 Limitations

While spike (S) protein may have a critical role in viral entry into host cells and their epitope prediction criterion were comprehensive, this study did not examine other candidate SARS-CoV-2 proteins. This point is particularly important given that a single epitope may not be sufficient to induce robust immune memory, and recent approaches involve multi-epitope vaccine design. Furthermore, their study only included a direct implementation of various published methods, but did not validate individual bioinformatic tools with controls to demonstrate robustness. Finally, it is critical that these predicted epitopes are experimentally validated before any conclusions can be drawn about their potential as vaccine candidates or their clinical efficacy.

14.30.4 Significance

This study provides a computational framework to rapidly identify epitopes that may serve as potential vaccine candidates for treating SARS-CoV-2.

14.30.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.31 The definition and risks of Cytokine Release Syndrome-Like in 11 COVID-19-Infected Pneumonia critically ill patients: Disease Characteristics and Retrospective Analysis

Wang Jr. et al. *medRxiv.* ([2203](#))

14.31.1 Keywords

- Cytokine release syndrome (CRS)
- biomarkers
- ARDS
- IL-6
- lymphopenia

14.31.2 Main Findings

This study describes the occurrence of a cytokine release syndrome-like (CRSL) toxicity in ICU patients with COVID-19 pneumonia. The median time from first symptom to acute respiratory distress syndrome (ARDS) was 10 days. All patients had decreased CD3, CD4 and CD8 cells, and a significant increase of serum IL-6. Furthermore, 91% had decreased NK cells. The changes in IL-6 levels preceded those in CD4 and CD8 cell counts. All of these parameters correlated with the area of pulmonary inflammation in CT scan images. Mechanical ventilation increased the numbers of CD4 and CD8 cells, while decreasing the levels of IL-6, and improving the immunological parameters.

14.31.3 Limitations

The number of patients included in this retrospective single center study is small (n=11), and the follow-up period very short (25 days). Eight of the eleven patients were described as having CRSL, and were treated by intubation (7) or ECMO (2). Nine patients were still in the intensive care unit at the time of publication of this article, so their disease outcome is unknown.

14.31.4 Significance

The authors define a cytokine release syndrome-like toxicity in patients with COVID-19 with clinical radiological and immunological criteria: 1) decrease of circulating CD4, CD8 and NK cells; 2) substantial increase of IL-6 in peripheral blood; 3) continuous fever; 4) organ and tissue damage. This event seems to occur very often in critically ill patients with COVID-19 pneumonia. Interestingly, the increase of IL-6 in the peripheral blood preceded other laboratory alterations, thus, IL-6 might be an early biomarker for the severity of COVID-19 pneumonia. The manuscript will require considerable editing for organization and clarity.

14.31.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.32 Clinical characteristics of 36 non-survivors with COVID-19 in Wuhan, China

Huang et al. *medRxiv.* ([2204](#))

14.32.1 Keywords

- Clinical Characteristics
- Non Survivors
- retrospective study

14.32.2 Main Findings

This is a simple study reporting clinical characteristics of patients who did not survive COVID-19. All patients (mean age=69.22 years) had acute respiratory distress syndrome (ARDS) and their median time from onset to ARDS was 11 days. The median time from onset to death was 17 days. Most patients were older male (70% male) with co-morbidities and only 11 % were smokers. 75% patients showed bilateral pneumonia. Many patients had chronic diseases, including hypertension (58.33%), cardiovascular disease (22.22%) and diabetes (19.44%). Typical clinical feature measured in these patients includes lymphopenia and elevated markers of inflammation.

14.32.3 Limitations

As noted by the authors, the conclusions of this study are very limited because this is single-centered study focusing on a small cohort of patients who did not survive. Many clinical parameters observed by the authors (such* as increase levels of serum CRP, PCT, IL-6) have also been described in other COVID19 patients who survived the infection

14.32.4 Significance

This study is essentially descriptive and may be useful for clinical teams monitoring COVID19 patients.

14.32.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.33 Risk Factors Related to Hepatic Injury in Patients with Corona Virus Disease 2019

(2205)

14.33.1 Keywords

- COVID-related Hepatic Injury

14.33.2 Main Findings

Based on a retrospective study of 85 hospitalized COVID patients in a Beijing hospital, authors showed that patients with elevated ALT levels ($n = 33$) were characterized by significantly higher levels of lactic acid and CRP as well as lymphopenia and hypoalbuminemia compared to their counterparts with normal ALT levels. Proportion of severe and critical patients in the ALT elevation group was significantly higher than that of normal ALT group. Multivariate logistic regression performed on clinical factors related to ALT elevation showed that $\text{CRP} \geq 20\text{mg/L}$ and low lymphocyte count ($<1.1 \times 10^9$ cells/L) were independently related to ALT elevation—a finding that led the authors to suggest cytokine storm as a major mechanism of liver damage.

14.33.3 Limitations

The article's most attractive claim that liver damage seen in COVID patients is caused by cytokine storm (rather than direct infection of the liver) hinges solely on their multivariate regression analysis. Without further mechanistic studies a) demonstrating how high levels of inflammatory cytokines can induce liver damage and b) contrasting types of liver damage incurred by direct infection of the liver vs. system-wide elevation of inflammatory cytokines, their claim remains thin. It is also worth noting that six of their elevated ALT group ($n=33$) had a history of liver disease (i.e. HBV infection, alcoholic liver disease, fatty liver) which can confound their effort to pin down the cause of hepatic injury to COVID.

14.33.4 Significance

Limited. This article confirms a rich body of literature describing liver damage and lymphopenia in COVID patients.

14.33.5 Credit

14.34 Detectable serum SARS-CoV-2 viral load (RNAemia) is closely associated with drastically elevated interleukin 6 (IL-6) level in critically ill COVID-19 patients

([2206](#))

14.34.1 Keywords

- ARDS
- interleukin-6 (IL-6)
- procalcitonin (PCT)
- pro-inflammatory cytokines
- SARS-CoV-2 RNAemia

14.34.2 Main Findings

48 adult patients diagnosed with Covid19 according to Chinese guidelines for Covid19 diagnosis and treatment version 6 were included in this study. Patients were further sub-divided into three groups based on clinical symptoms and disease severity: (1) mild, positive Covid19 qPCR with no or mild clinical symptoms (fever; respiratory; radiological abnormalities); (2) severe, at least one of the following: shortness of breath/respiratory rate >30/min, oxygen saturation $\text{SaO}_2 < 93\%$, Horowitz index $\text{paO}_2/\text{FiO}_2 < 300$ mmHg (indicating moderate pulmonary damage); and (3) critically ill, at least one additional complicating factor: respiratory failure with need for mechanical ventilation; systemic shock; multi-organ failure and transfer to ICU. Serum samples and throat-swabs were collected from all 48 patients enrolled. SARS-CoV-2 RNA was assessed by qPCR with positive results being defined as Ct values < 40, and serum interleukin-6 (IL-6) was quantified using a commercially available detection kit. Briefly, patient characteristics in this study confirm previous reports suggesting that higher age and comorbidities are significant risk factors of clinical severity. Of note, 5 out of 48 of patients (10.41%), all in the critically ill category, were found to have detectable serum SARS-CoV-2 RNA levels, so-called RNAemia. Moreover, serum IL-6 levels in these patients were found to be substantially higher and this correlated with the presence of detectable SARS-CoV-2 RNA levels. The authors hypothesize that viral RNA might be released from acutely damages tissues in moribund patients during the course of Covid19 and that RNAemia along with IL-6 could potentially be used as a prognostic marker.

14.34.3 Limitations

While this group's report generally confirms some of the major findings of a more extensive study, published in early February 2020, ([2196](#)), there are limitations that should be taken into account. First, the number of patients enrolled is relatively small; second, interpretation of these data would benefit

from inclusion of information about study specifics as well as providing relevant data on the clinical course of these patients other than the fact that some were admitted to ICU (i.e. demographics on how many patients needed respiratory support, dialysis, APACHE II/III or other standard ICU scores as robust prognostic markers for mortality etc). It also remains unclear at which time point the serum samples were taken, i.e. whether at admission, when the diagnosis was made or during the course of the hospital stay (and potentially after onset of therapy, which could have affected both IL-6 and RNA levels). The methods section lacks important information on the qPCR protocol employed, including primers and cycling conditions used. From a technical point of view, Ct values >35 seem somewhat non-specific (although Ct <40 was defined as the CDC cutoff as well) indicating that serum RNA levels are probably very low, therefore stressing the need for highly specific primers and high qPCR efficiency. In addition, the statistical tests used (t-tests, according to the methods section) do not seem appropriate as the organ-specific data such as BUN and troponin T values seem to be not normally distributed across groups (n= 5 RNAemia+ vs. n= 43 RNAemia-). Given the range of standard deviations and the differences in patient sample size, it is difficult to believe that these data are statistically significantly different.

14.34.4 Significance

This study is very rudimentary and lacks a lot of relevant clinical details. However, it corroborates some previously published observations regarding RNAemia and IL-6 by another group. Generally, regarding future studies, it would be important to address the question of IL-6 and other inflammatory cytokine dynamics in relation to Covid19 disease kinetics (high levels of IL-6, IL-8 and plasma leukotriene were shown to have prognostic value at the onset of ARDS ; serum IL-2 and IL-15 have been associated with mortality; reviewed by Chen W & Ware L, Clin Transl Med. 2015 ([2207](#))).

14.34.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.35 Lymphopenia predicts disease severity of COVID-19: a descriptive and predictive study

([2208](#))

14.35.1 Keywords

- Lymphopenia

14.35.2 Main Findings

Based on a retrospective study of 162 COVID patients from a local hospital in Wuhan, China, the authors show an inverse correlation between lymphocyte % (LYM%) of patients and their disease severity. The authors have also tracked LYM% of 70 cases (15 deaths; 15 severe; 40 moderate) throughout the disease progression with fatal cases showing no recovery of lymphocytes (<5%) even after 17-19 days post-onset. The temporal data of LYM % in COVID patients was used to construct a Time-Lymphocyte% model which is used to categorize and predict patients' disease severity and progression. The model was validated using 92 hospitalized cases and kappa statistic test was used to assess agreement between predicted disease severity and the assigned clinical severity ($k = 0.49$).

14.35.3 Limitations

Time-Lymphocyte % Model (TLM) that authors have proposed as a predictive model for clinical severity is very simple in its construction and derives from correlative data of 162 patients. In order for the model to be of use, it needs validation using a far more robust data set and possibly a mechanistic study on how COVID leads to lymphopenia in the first place. In addition, it should be noted that no statistical test assessing significance of LYM % values between disease severities was performed.

14.35.4 Significance

This article is of limited significance as it simply reports similar descriptions of COVID patients made in previous literature that severe cases are characterized by lymphopenia.

14.35.5 Credit

Review by Chang Moon as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.36 The potential role of IL-6 in monitoring severe case of coronavirus disease 2019

Liu et al. *medRxiv.* ([2209](#))

14.36.1 Keywords

- Cytokine Release Syndrome
- lymphocytopenia
- IL-6
- CRP
- COVID19
- pneumonia

14.36.2 Main Findings

Study on blood biomarkers on 80 COVID19 patients (69 severe and 11 non-severe). Patients with severe symptoms at admission (baseline) showed obvious lymphocytopenia and significantly increased interleukin-6 (IL-6) and CRP, which was positively correlated with symptoms severity. IL-6 at baseline positively correlates with CRP, LDH, ferritin and D-Dimer abundance in blood.

Longitudinal analysis of 30 patients (before and after treatment) showed significant reduction of IL-6 in remission cases.

14.36.3 Limitations

Limited sample size at baseline, especially for the non-severe leads to question on representativeness. The longitudinal study method is not described in detail and suffers from non-standardized treatment. Limited panel of pro-inflammatory cytokine was analyzed. Patients with severe disease show a wide range of altered blood composition and biomarkers of inflammation, as well as differences in disease course (53.6% were cured, about 10% developed acute respiratory distress syndrome). The authors comment on associations between IL-6 levels and outcomes, but these were not statistically significant (maybe due to the number of patients, non-standardized treatments, etc.) and data is not shown. Prognostic biomarkers could have been better explored. Study lacks multivariate analysis.

14.36.4 Significance

IL-6 could be used as a pharmacodynamic marker of disease severity. Cytokine Release Syndrome (CRS) is a well-known side effect for CAR-T cancer therapy and there are several effective drugs to manage CRS. Drugs used to manage CRS could be tested to treat the most severe cases of COVID19.

14.36.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.37 Clinical and Laboratory Profiles of 75 Hospitalized Patients with Novel Coronavirus Disease 2019 in Hefei, China

Zhao et al. *medRxiv*. ([2210](#))

14.37.1 Keywords

- Routine laboratory testing

14.37.2 Main Findings

The authors of this study provide a comprehensive analysis of clinical laboratory assessments in 75 patients (median age 47 year old) hospitalized for Corona virus infection in China measuring differential blood counts including T-cell subsets (CD4, CD8), coagulation function, basic blood chemistry, of infection-related biomarkers including CRP, Procalcitonin (PCT) (Precursor of calcitonin that increases during bacterial infection or tissue injury), IL-6 and erythrocyte sedimentation rate as well as clinical parameters. Among the most common hematological changes they found increased neutrophils, reduced CD4 and CD8 lymphocytes, increased LDH, CRP and PCT

When looking at patients with elevated IL-6, the authors describe significantly reduced CD4 and CD8 lymphocyte counts and elevated CRP and PCT levels were significantly increased in infected patients suggesting that increased IL-6 may correlate well with disease severity in COVID-19 infections

14.37.3 Limitations

The authors performed an early assessment of clinical standard parameters in patients infected with COVID-19. Overall, the number of cases (75) is rather low and the snapshot approach does not inform about dynamics and thus potential relevance in the assessment of treatment options in this group of patients.

14.37.4 Significance

The article summarizes provides a good summary of some of the common changes in immune cells inflammatory cytokines in patients with a COVID-19 infection and. Understanding how these changes can help predict severity of disease and guide therapy including IL-6 cytokine receptor blockade using Tocilizumab or Sarilumab will be important to explore.

14.37.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.38 Exuberant elevation of IP-10, MCP-3 and IL-1ra during SARS-CoV-2 infection is associated with disease severity and fatal outcome

Yang et al. *medRxiv* ([2211](#))

14.38.1 Keywords

- cytokine
- IP-10
- MCP-3
- IL-1Ra
- lymphocyte

- neutrophil
- stratification
- disease severity
- viral load
- lung function
- complications
- clinical data

14.38.2 Summary

Plasma cytokine analysis (48 cytokines) was performed on COVID-19 patient plasma samples, who were sub-stratified as severe (N=34), moderate (N=19), and compared to healthy controls (N=8). Patients were monitored for up to 24 days after illness onset: viral load (qRT-PCR), cytokine (multiplex on subset of patients), lab tests, and epidemiological/clinical characteristics of patients were reported.

14.38.3 Main Findings

- Many elevated cytokines with COVID-19 onset compared to healthy controls (IFN γ , IL-1Ra, IL-2Ra, IL-6, IL-10, IL-18, HGF, MCP-3, MIG, M-CSF, G-CSF, MIG-1a, and IP-10).
- IP-10, IL-1Ra, and MCP-3 (esp. together) were associated with disease severity and fatal outcome.
- IP-10 was correlated to patient viral load ($r=0.3006$, $p=0.0075$).
- IP-10, IL-1Ra, and MCP-3 were correlated to loss of lung function (PaO₂/FaO₂ (arterial/atmospheric O₂) and Murray Score (lung injury) with MCP-3 being the most correlated ($r=0.4104$ $p<0.0001$ and $r=0.5107$ $p<0.0001$ respectively).
- Viral load (Lower Ct Value from qRT-PCR) was associated with upregulated IP-10 only (not IL-1Ra or MCP-3) and was mildly correlated with decreased lung function: PaO₂/FaO₂ (arterial/atmospheric O₂) and Murray Score (lung injury).
- Lymphopenia (decreased CD4 and CD8 T cells) and increased neutrophil correlated w/ severe patients.
- Complications were associated with COVID severity (ARDS, hepatic insufficiency, renal insufficiency).

14.38.4 Limitations

Collection time of clinical data and lab results not reported directly (likely 4 days (2,6) after illness onset), making it very difficult to determine if cytokines were predictive of patient outcome or reflective of patient compensatory immune response (likely the latter). Small N for cytokine analysis (N=2 fatal and N=5 severe/critical, and N=7 moderate or discharged). Viral treatment strategy not clearly outlined.

14.38.5 Expanded Results

NOTE: Moderate COVID-19 was classified by fever, respiratory manifestations, and radiological findings consistent with pneumonia while severe patients had one or more of the following: 1) respiratory distraction, resting O₂ saturation, or 3) arterial PaO₂/FiO₂ <300 mmHg.

Cytokine Results (Human Cytokine Screening Panel, Bio-Rad):

- **Significant elevation of cytokines observed in COVID patients compared to healthy controls: IFN γ , IL-1Ra, IL-2Ra, IL-6, IL-10, IL-18, HGF, MCP-3, MIG, M-CSF, G-CSF, MIG-1a, and IP-10.**
- Severity was correlated **with increase in measured IP-10, MCP-3, and IL-1Ra** as measure by area under the curve analysis during sample timecourse (2-24 days after illness onset).
- IL-1Ra incr. significant 0-7 days after onset, MCP-3 signif. upregulated throughout observation timecourse, and IP-10 increased and upregulated throughout (trending downwards over time).
- **The three cytokines together (IP-10, IL-1Ra, and MCP-3 AUC) served as the best predictors of disease deterioration and fatal outcome.**
- No significant differences between moderate/severe observed between groups in IL-2Ra, IL-6, IL-10, IL-18, CTACK, G-CSF, HGF, M-CSF, MIP-1a, MIG, and IFN γ at any timepoints.
- **Viral load (Lower Ct Value from qRT-PCR) was associated with upregulated IP-10 only (not IL-1Ra or MCP-3) and was highly correlated with decreased lung function: PaO₂/FaO₂ (arterial/atmospheric O₂) and Murray Score (lung injury).**
- **Antibodies against these cytokines (esp. anti-IP-10) may serve as a potential treatment for amelioration of COVID-19 (and associated ARDS).**

Lab results:

- **Decreased lymphocytes (%) in all patients – lymphopenia corr. w/ severe patients**
 - **Decreased CD4 and CD8 T cells** – no monocyte or eosino/basophil % measured
- **Increased neutrophils (%)**
- Increased BUN (mmol/L) – other kidney markers, liver markers, and LDH were not significantly different between groups and were not compared to healthy controls.

Clinical features (between moderate vs. severe patient groups):

- **Complications were associated with severity (ARDS, hepatic insufficiency, renal insufficiency).**
- Coexisting conditions between groups were not significantly different (chronic heart/lung/renal/liver disease, diabetes, or cancer) and patient time courses (onset to admission and onset to viral tx) also not significantly different – 4 days (2, 6) on average for admission and 4 (3,7) for antiviral.
- Increased corticosteroids and mechanical/ invasive mechanical ventilation in severe patients.
- Increased median age in severe group (Median (Range = 63.5 (42-74) vs. 51 (22-78)) and patients > 60 yrs had higher ratio of severe patients as compared patients 16-59 yrs.
- Higher incidence of fever in severe patients (91.2 vs. 68.4%), myalgia (57.7 vs. 48.1%), and chill (17.6% vs. 0%).
- No differences in cough, headache, nausea/vomiting, or diarrhea.

14.38.6 Significance

Outline of pathological time course (implicating innate immunity esp.) and identification key cytokines associated with disease severity and prognosis (+ comorbidities). Anti-IP-10 as a possible therapeutic intervention (ex: Eldelumab).

14.38.7 Credit

Review by Natalie Vaninov as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.39 Antibody responses to SARS-CoV-2 in patients of novel coronavirus disease 2019

Zhao Jr. et al. *medRxiv*. ([2212](#))

14.39.1 Keywords

- SARS-CoV-2 IgG
- seroconversion rate
- total Ab
- Ig
- IgM

14.39.2 Main Findings

This study examined antibody responses in the blood of COVID-19 patients during the early SARS CoV2 outbreak in China. Total 535 plasma samples were collected from 173 patients (51.4% female) and were tested for seroconversion rate using ELISA. Authors also compared the sensitivity of RNA and antibody tests over the course of the disease . The key findings are:

- Among 173 patients, the seroconversion rates for total antibody (Ab), IgM and IgG were 93.1% (161/173), 82.7% (143/173) and 64.7% (112/173), respectively.
- The seroconversion sequentially appeared for Ab, IgM and then IgG, with a median time of 11, 12 and 14 days, respectively. Overall, the seroconversion of Ab was significantly quicker than that of IgM ($p = 0.012$) and IgG ($p < 0.001$). Comparisons of seroconversion rates between critical and non-critical patients did not reveal any significant differences.
- RNA tests had higher sensitivity in early phase and within 7 days of disease onset than antibody assays (66.7% Vs 38.3% respectively).
- The sensitivity of the Ab assays was higher 8 days after disease onset, reached 90% at day 13 and 100% at later time points (15-39 days). In contrast, RNA was only detectable in 45.5% of samples at days 15-39.
- In patients with undetectable RNA in nasal samples collected during day 1-3, day 4-7, day 8-14 and day 15-39 since disease onset, 28.6% (2/7), 53.6% (15/28), 98.2% (56/57) and 100% (30/30) had detectable total Ab titers respectively. Combining RNA and antibody tests significantly raised the sensitivity for detecting COVID-19 patients in different stages of the disease ($p < 0.001$).
- There was a strong positive correlation between clinical severity and antibody titer 2-weeks after illness onset.
- Dynamic profiling of viral RNA and antibodies in representative COVID-19 patients ($n=9$) since onset of disease revealed that antibodies may not be sufficient to clear the virus. It should be noted that increases in of antibody titers were not always accompanied by RNA clearance.

14.39.3 Limitations

Because different types of ELISA assays were used for determining antibody concentrations at different time points after disease onset, sequential seroconversion of total Ab, IgM and IgG may not represent actual temporal differences but rather differences in the affinities of the assays used. Also, due to the lack of blood samples collected from patients in the later stage of illness, how long the antibodies could last remain unknown. For investigative dynamics of antibodies, more samples were required.

14.39.4 Significance

Total and IgG antibody titers could be used to understand the epidemiology of SARS CoV-2 infection and to assist in determining the level of humoral immune response in patients.

The findings provide strong clinical evidence for routine serological and RNA testing in the diagnosis and clinical management of COVID-19 patients. The understanding of antibody responses and their half-life during and after SARS CoV2 infection is important and warrants further investigations.

14.39.5 Credit

This review was undertaken by Zafar Mahmood and edited by K Alexandropoulos as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.40 Restoration of leukomonocyte counts is associated with viral clearance in COVID-19 hospitalized patients

Chen et al. *medRxiv* ([2213](#))

14.40.1 Keywords

- COVID-19
- T cell
- B cell
- NK cell
- IL-6
- pro-calcitonin
- cytokine storm

14.40.2 Main Findings

The authors collected data on 25 COVID-19 patients (n=11 men, n=14 women) using standard laboratory tests and flow cytometry. All patients were treated with antibiotics. Twenty-four of the 25 patients were also treated with anti-viral Umefinovir and 14 of the patients were treated with corticosteroids. 14 patients became negative for the virus after 8-14 days of treatment. The same treatment course was extended to 15-23 days for patients who were still positive for the virus at day 14.

The authors found a negative association between age and resolution of infection. Patients with hypertension, diabetes, malignancy or chronic liver disease were all unable to clear the virus at day 14, though not statistically significant.

Elevated procalcitonin and a trend for increased IL-6 were also found in peripheral blood prior to the treatment.

A trend for lower NK cell, T cell and B cell counts in patients was also reported. B cell, CD4 and CD8 T cell counts were only increased upon treatment in patients who cleared the virus. NK cell frequencies remained unchanged after treatment in all the patients.

14.40.3 Limitations

73% of the patients who remained positive for SARS-CoV2 after the 1st treatment, and 43% of all patients who cleared the virus were treated with corticosteroids. Corticosteroids have strong effects on the immune compartment in blood ([2214](#)). The authors should have accounted for corticosteroid treatment when considering changes in T, NK and B cell frequencies.

Assessing if IL-6 concentrations were back to baseline levels following treatment would have provided insights into the COVID-19 cytokine storm biology. Patients with higher baseline levels of IL-6 have been reported to have lower CD8 and CD4 T cell frequencies ([2210](#)). Correlating IL-6 with cell counts before and after treatment would thus have also been of interest. The report of the laboratory measures in table 2 is incomplete and should include the frequencies of patients with increased/decreased levels for each parameter.

Correction is needed for the 1st paragraph of the discussion as data does not support NK cell restoration upon treatment in patients who cleared the virus. NK cells remain unchanged after the 1st treatment course and only seem to increase in 2 out of 6 donors after the 2nd treatment course in those patients.

14.40.4 Significance

Previous reports suggest an association between disease severity and elevated IL-6 or pro-calcitonin concentrations in COVID-19 patients ([2206](#), [2215](#)). IL-6 receptor blockade is also being administered to patients enrolled in clinical trials (NCT04317092). This report thus contributes to highlight elevated concentrations of these analytes in COVID-19 patients. Mechanisms underlying the association between viral clearance and restoration of the T cell and B cell frequencies suggests viral-driven immune dysregulation, which needs to be investigated in further studies.

14.40.5 Credit

Review by Bérengère Salomé as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.41 Clinical findings in critically ill patients infected with SARS-CoV-2 in Guangdong Province, China: a multi-center, retrospective, observational study

14.41.1 Keywords

- clinical outcomes
- prognosis
- critically ill patients
- ICU
- lymphopenia
- LDH

14.41.2 Main Findings:

This work analyses laboratory and clinical data from 45 patients treated in the in ICU in a single province in China. Overall, 44% of the patients were intubated within 3 days of ICU admission with only 1 death.

Lymphopenia was noted in 91% of patient with an inverse correlation with LDH.

Lymphocyte levels are negatively correlated with Sequential Organ Failure Assessment (SOFA) score (clinical score, the higher the more critical state), LDH levels are positively correlated to SOFA score. Overall, older patients (>60yo), with high SOFA score, high LDH levels and low lymphocytes levels at ICU admission are at higher risk of intubation.

Of note, convalescent plasma was administered to 6 patients but due to limited sample size no conclusion can be made.

14.41.3 Limitations

While the study offers important insights into disease course and clinical lab correlates of outcome, the cohort is relatively small and is likely skewed towards a less-severe population compared to other ICU reports given the outcomes observed. Analysis of laboratory values and predictors of outcomes in larger cohorts will be important to make triage and treatment decisions. As with many retrospective analyses, pre-infection data is limited and thus it is not possible to understand whether lymphopenia was secondary to underlying comorbidities or infection.

Well-designed studies are necessary to evaluate the effect of convalescent plasma administration.

14.41.4 Significance

This clinical data enables the identification of at-risk patients and gives guidance for research for treatment options. Indeed, further work is needed to better understand the causes of the lymphopenia and its correlation with outcome.

14.41.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.42 Immune Multi-epitope vaccine design using an immunoinformatics approach for 2019 novel coronavirus in China (SARS-CoV-2)

([2217](#))

14.42.1 Keywords

- Vaccine
- in silico
- B cell epitopes
- T cell epitopes

14.42.2 Main Findings

Using in silico bioinformatic tools, this study identified putative antigenic B-cell epitopes and HLA restricted T-cell epitopes from the spike, envelope and membrane proteins of SARS-CoV-2, based on the genome sequence available on the NCBI database. T cell epitopes were selected based on predicted affinity for the more common HLA-I alleles in the Chinese population. Subsequently, the authors designed vaccine peptides by bridging selected B-cell epitopes and adjacent T-cell epitopes. Vaccine peptides containing only T-cell epitopes were also generated.

From 61 predicted B-cell epitopes, only 19 were exposed on the surface of the virion and had a high antigenicity score. A total of 499 T-cell epitopes were predicted. Based on the 19 B-cell epitopes and their 121 adjacent T-cell epitopes, 17 candidate vaccine peptides were designed. Additionally, another 102 vaccine peptides containing T-cell epitopes only were generated. Based on the epitope counts and HLA score, 13 of those were selected. Thus, a total of 30 peptide vaccine candidates were designed.

14.42.3 Limitations

While this study provides candidates for the development of vaccines against SARS-CoV-2, in vitro and in vivo trials are required to validate the immunogenicity of the selected B and T cell epitopes. This could be done using serum and cells from CoV-2-exposed individuals, and in preclinical studies. The implication of this study for the current epidemic are thus limited. Nevertheless, further research on this field is greatly needed.

14.42.4 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.43 Clinical Features of Patients Infected with the 2019 Novel Coronavirus (COVID-19) in Shanghai, China

Cao et al. *medRxiv* ([2218](#))

14.43.1 Keywords

- Disease severity
- clinical features
- laboratory abnormalities

14.43.2 Main Findings

This single-center cohort study analyzes the clinical and laboratory features of 198 patients with confirmed COVID-19 infection in Shanghai, China and correlated these parameters with clinical disease severity, including subsequent intensive care unit (ICU) admission. 19 cases (9.5%) required ICU admission after developing respiratory failure or organ dysfunction. Age, male sex, underlying cardiovascular disease, and high symptom severity (high fever, dyspnea) were all significantly correlated with ICU admission. Additionally, ICU admission was more common in patients who presented with lymphopenia and elevated neutrophil counts, among other laboratory abnormalities. Flow cytometric analysis revealed that patients admitted to the ICU had significantly reduced circulating CD3+ T cell, CD4+ T cell, CD8+ T cell, and CD45+ leukocyte populations compared to the cohort of patients not requiring ICU admission.

14.43.3 Limitations

The limitations of this study include the relatively small sample size and lack of longitudinal testing. The authors also did not assess whether respiratory comorbidity – such as asthma or chronic obstructive lung disease – in addition to immunosuppression affected ICU admission likelihood.

14.43.4 Significance

COVID-19 has already sickened thousands across the globe, though the severity of these infections is markedly diverse, ranging from mild symptoms to respiratory failure requiring maximal intervention. Understanding what clinical, laboratory, and immunologic factors predict the clinical course of COVID-19 infection permits frontline providers to distribute limited medical resources more effectively.

14.43.5 Credit

Review by Andrew Charap as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine at Mount Sinai.

14.44 Serological detection of 2019-nCoV respond to the epidemic: A useful complement to nucleic acid testing

Zhang et al. *medRxiv.* ([2219](#))

14.44.1 Keywords

- Immunoassay
- serum IgM and IgG
- specific antibodies

14.44.2 Main Finding

This study showed that both anti-2019-nCoV IgM and IgG were detected by automated chemiluminescent immunoassay in the patients who had been already confirmed as positive by nucleic acid detection, while single positivity of IgM or IgG were detected in a very few cases in the other population including 225 non-COVID-19 cases. In addition to the increase of anti-2019-nCoV IgM 7-12 days after morbidity, the increase of IgG was detected in three patients with COVID-19 within a very short of time (0-1 day).

14.44.3 Limitations

The limitation of this study is only 3 confirmed COVID-19 cases were included, so that the relationship between anti-2019-nCoV antibodies and disease progression might not be clearly defined. Another limitation is that they did not show the course of 2019-nCoV specific antibodies in the cases with positive for COVID-19 but without clinical symptoms.

14.44.4 Significance

The detection of anti-2019-nCoV antibodies can be an alternative method to diagnose and treat COVID-19 more comprehensively by distinguish non COVID-19 patients. It may be helpful to understand the course of individual cases with COVID-19 to predict the prognosis if more cases will be evaluated.

14.44.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.45 Human Kidney is a Target for Novel Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Infection

([2220](#))

14.45.1 Keywords

- Kidney/Renal Failure
- Macrophage Infiltration
- Complement Activation

14.45.2 Main Finding

Analyzing the eGFR (effective glomerular flow rate) of 85 Covid-19 patients and characterizing tissue damage and viral presence in post-mortem kidney samples from 6 Covid-19 patients, the authors conclude that significant damage occurs to the kidney, following Covid-19 infection. This is in contrast to the SARS infection from the 2003 outbreak. They determine this damage to be more prevalent in patients older than 60 years old, as determined by analysis of eGFR. H&E and IHC analysis in 6 Covid-19 patients revealed that damage was in the tubules, not the glomeruli of the kidneys and suggested that macrophage accumulation and C5b-9 deposition are key to this process.

14.45.3 Limitations

Severe limitations include that the H&E and IHC samples were performed on post-mortem samples of unknown age, thus we cannot assess how/if age correlates with kidney damage, upon Covid-19 infection. Additionally, eGFR was the only *in-vivo* measurement. Blood urea nitrogen and proteinuria are amongst other measurements that could have been obtained from patient records. An immune panel of the blood was not performed to assess immune system activation. Additionally, patients are only from one hospital.

14.45.4 Significance

This report makes clear that kidney damage is prevalent in Covid-19 patients and should be accounted for.

14.45.5 Credit

Review by Dan Fu Ruan, Evan Cody and Venu Pothula as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine at Mount Sinai.

14.46 COVID-19 early warning score: a multi-parameter screening tool to identify highly suspected patients

Song et al. *medRxiv*. ([2221](#))

14.46.1 Keywords

- retrospective
- electronic health records
- blood counts

- diagnostic
- prognostic
- modeling

14.46.2 Main Findings

The aim of this study was to identify diagnostic or prognostic criteria which could identify patients with COVID-19 and predict patients who would go on to develop severe respiratory disease. The authors use EMR data from individuals taking a COVID-19 test at Zhejiang hospital, China in late January/Early February. A large number of clinical parameters were different between individuals with COVID-19 and also between 'severe' and 'non-severe' infections and the authors combine these into a multivariate linear model to derive a weighted score, presumably intended for clinical use.

14.46.3 Limitations

Unfortunately, the paper is lacking a lot of crucial information, making it impossible to determine the importance or relevance of the findings. Most importantly, the timings of the clinical measurements are not described relative to the disease course, so it is unclear if the differences between 'severe' and 'non-severe' infections are occurring before progression to severe disease (which would make them useful prognostic markers), or after (which would not).

14.46.4 Significance

This paper is one of many retrospective studies coming from hospitals in China studying individuals with COVID-19. Because of the sparse description of the study design, this paper offers little new information. However, studies like this could be very valuable and we would strongly encourage the authors to revise this manuscript to include more information about the timeline of clinical measurements in relation to disease onset and more details of patient outcomes.

14.46.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.47 LY6E impairs coronavirus fusion and confers immune control of viral disease

Pfaender et al. *bioRxiv.* ([2222](#))

14.47.1 Keywords

- interferon-stimulated genes
- antiviral interferons

- human coronaviruses (CoV)
- murine hepatitis virus (MHV)

14.47.2 Main Findings

Screening a cDNA library of >350 human interferon-stimulated genes for antiviral activity against endemic human coronavirus HCoV-229E (associated with the common cold), Pfaender S & Mar K *et al.* identify lymphocyte antigen 6 complex, locus E (Ly6E) as an inhibitor of cellular infection of Huh7 cells, a human hepatoma cell line susceptible to HCoV-229E and other coronaviruses. In a series of consecutive *in vitro* experiments including both stable Ly6E overexpression and CRISPR-Cas9-mediated knockout the authors further demonstrate that Ly6E reduces cellular infection by various other coronaviruses including human SARS-CoV and SARS-CoV-2 as well as murine CoV mouse hepatitis virus (MHV). Their experiments suggest that this effect is dependent on Ly6E inhibition of CoV strain-specific spike protein-mediated membrane fusion required for viral cell entry.

To address the function of Ly6E *in vivo*, hematopoietic stem cell-specific Ly6E knock-out mice were generated by breeding Ly6E^{fl/fl} mice (referred to as functional wild-type mice) with transgenic *Vav-iCre* mice (offspring referred to as Ly6E HSC ko mice); wild-type and Ly6E HSC ko mice of both sexes were infected intraperitoneally with varying doses of the natural murine coronavirus MHV, generally causing a wide range of diseases in mice including hepatitis, enteritis and encephalomyelitis. Briefly, compared to wild-type controls, mice lacking hematopoietic cell-expressed Ly6E were found to present with a more severe disease phenotype as based on serum ALT levels (prognostic of liver damage), liver histopathology, and viral titers in the spleen. Moreover, bulk RNAseq analysis of infected liver and spleen tissues indicated changes in gene expression pathways related to tissue damage and antiviral immune responses as well as a reduction of genes associated with type I IFN response and inflammation. Finally, the authors report substantial differences in the numbers of hepatic and splenic APC subsets between wild-type and knockout mice following MHV infection and show that Ly6E-deficient B cells and to a lesser extent also DCs are particularly susceptible to MHV infection *in vitro*.

14.47.3 Limitations

Experiments and data in this study are presented in an overall logical and coherent fashion; however, some observations and the conclusions drawn are problematic and should be further addressed & discussed by the authors. Methodological & formal limitations include relatively low replicate numbers as well as missing technical replicates for some *in vitro* experiments (*cf.* Fig. legend 1; Fig. legend 2e); the omission of “outliers” in Fig. legend 2 without an apparent rationale as to why this approach was chosen; the lack of detection of actual Ly6E protein levels in Ly6E HSC ko or wild-type mice; and most importantly, missing information on RNAseq data collection & analysis in the method section and throughout the paper. A more relevant concern though is that the interpretation of the experimental data presented and the language used tend to overrate and at times overgeneralize findings: for example, while the authors demonstrate statistically significant, Ly6E-mediated reduction of coronavirus titers in stable cells lines *in vitro*, it

remains unclear whether a viral titer reduction by one log decade would be of actual biological relevance in face of high viral titers *in vivo*. After high-dose intraperitoneal MHV infection *in vivo*, early viral titers in Ly6E HSC knockout vs. wt mice only showed an elevation in the spleen (~1.5 log decades) but not liver of the ko mice (other tissue not evaluated), and while ko mice presented with only modestly increased liver pathology, both male and female ko mice exhibited significantly higher mortality. Thus, the manuscript title statement that “Ly6E ... confers immune control of viral disease” is supported by only limited *in vivo* data, and gain-of-function experiments (eg. Ly6E overexpression) were not performed. Of additional note here, tissue tropism and virulence differ greatly among various MHV strains and isolates whereas dose, route of infection, age, genetic background and sex of the mice used may additionally affect disease outcome and phenotype (*cf.* Taguchi F & Hirai-Yuki A, <https://doi.org/10.3389/fmicb.2012.00068>; Kanolkhar A et al, <https://jvi.asm.org/content/83/18/9258>). Observations attributed to hematopoietic stem cell-specific Ly6E deletion could therefore be influenced by the different genetic backgrounds of floxed and cre mice used, and although it appears that littermates wt and ko littermates were used in the experiments, the potentially decisive impact of strain differences should at least have been discussed. Along these lines, it should also be taken into account that the majority of human coronaviruses cause respiratory symptoms, which follow a different clinical course engaging other primary cellular mediators than the hepatotropic murine MHV disease studied here. It therefore remains highly speculative how the findings reported in this study will translate to human disease and it would therefore be important to test other routes of MHV infection and doses that have been described to produce a more comparable phenotype to human coronavirus disease (*cf.* Kanolkhar A et al, <https://jvi.asm.org/content/83/18/9258>). Another important shortcoming of this study is the lack of any information on functional deficits or changes in Ly6E-deficient immune cells and how this might relate to the phenotype observed. Overall, the *in vitro* experiments are more convincing than the *in vivo* studies which appear somewhat limited.

14.47.4 Significance

Despite some shortcomings, the experiments performed in this study suggest a novel and somewhat unexpected role of Ly6E in the protection against coronaviruses across species. These findings are of relevance and should be further explored in ongoing research on potential coronavirus therapies. Yet an important caveat pertains to the authors' suggestion that “therapeutic mimicking of Ly6E action” may constitute a first line of defense against novel coronaviruses since their own prior work demonstrated that Ly6E can enhance rather than curtail infection with influenza A and other viruses.

14.47.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.48 A preliminary study on serological assay for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in 238 admitted hospital patients

Liu et al. *medRxiv.* ([2223](#))

14.48.1 Keywords

- diagnosis
- serological assay
- ELISA
- RT-PCR

14.48.2 Main Findings

While RT-PCR is being used currently to routinely diagnose infection with SARS-CoV-2, there are significant limitations to the use of a nucleic acid test that lead to a high false-negative rate. This article describes ELISAs that can measure IgM and IgG antibodies against the N protein of SARS-CoV-2 to test samples from 238 patients (153 positive by RT-PCR and 85 negative by RT-PCR) at different times after symptom onset. The positivity rate of the IgM and/or IgG ELISAs was greater than that of the RT-PCR (81.5% compared to 64.3%) with similar positive rates in the confirmed and suspected cases (83% and 78.8%, respectively), suggesting that many of the suspected but RT-PCR-negative cases were also infected. The authors also found that the ELISAs have higher positive rates later after symptom onset while RT-PCR is more effective as a diagnostic test early during the infection.

14.48.3 Limitations

I cannot identify any limitations to this study.

14.48.4 Significance

The authors make a strong case for using a combination of ELISA and RT-PCR for diagnosis of infection with SARS-CoV-2, especially considering the dynamics of positivity rates of RT-PCR and ELISA. Fewer false-negative diagnoses would improve infection control and patient management.

14.48.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.49 Monoclonal antibodies for the S2 subunit of spike of SARS-CoV cross-react with the newly-emerged SARS-CoV-2

([2224](#))

14.49.1 Keywords

- Coronavirus Disease 2019 (COVID-19)
- SARS-CoV-2
- Spike protein
- Cross- reactive antibodies

14.49.2 Main Findings

Whole genome sequencing-based comparisons of the 2003 Severe Acute Respiratory Syndrome Coronavirus (SARS-CoV) and the 2019 SARS-CoV-2 revealed conserved receptor binding domain (RBD) and host cell receptor, angiotensin-converting enzyme 2 (ACE2). In line with this, the authors tested cross-reactivity of murine monoclonal antibodies (mAbs) previously generated against the SARS-CoV spike (S) glycoprotein involved in viral entry. One of the screened mAb, 1A9, was able to bind and cross-neutralize multiple strains of SARS-CoV, as well as, detect the S protein in SARS-CoV-2-infected cells. mAb 1A9 was generated using an immunogenic fragment in the S2 subunit of SARS-CoV and binds through a novel epitope within the S2 subunit at amino acids 1111-1130. It is important to note that CD8+ T lymphocyte epitopes overlap with these residues, suggesting that S2 subunit could be involved in inducing both, humoral and cell-mediated immunity.

14.49.3 Limitations

The authors used previously generated mouse mAbs against the S protein in SARS-CoV expressed in mammalian cell line. Future experimental validation using COVID-19 patient samples is needed to validate these findings. In addition, the results of these studies are predominantly based on in vitro experiments and so, evaluating the effects of the mAb 1A9 in an animal model infected with this virus will help us better understand the host immune responses in COVID-19 and potential therapeutic vaccines.

14.49.4 Significance

This study identified mAbs that recognize the new coronavirus, SARS-Cov-2. These cross-reactive mAbs will help in developing diagnostic assays for COVID-19.

14.49.5 Credit

This review was undertaken by Tamar Plitt and Katherine Lindblad as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.50 Mortality of COVID-19 is Associated with Cellular Immune Function Compared to Immune Function in Chinese Han Population

Zeng et al. *medRxiv.* ([2225](#))

14.50.1 Keywords

- WBC
- peripheral blood
- CD4
- CD8 T cells

14.50.2 Main Findings

Retrospective study of the clinical characteristics of 752 patients infected with COVID-19 at Chinese PLA General Hospital, Peking Union Medical College Hospital, and affiliated hospitals at Shanghai University of medicine & Health Sciences. This study is the first one that compares PB from healthy controls from the same regions in Shanghai and Beijing, and infected COVID-19 patients to standardize a reference range of WBCs of people at high risk.

14.50.3 Limitations

Lower levels of leukocyte counts -B cells, CD4 and CD8 T cells- correlated with mortality (WBCs are significantly lower in severe or critical UCI patients vs mild ones). Based on 14,117 normal controls in Chinese Han population (ranging in age from 18-86) it is recommended that reference ranges of people at high risk of COVID-19 infection are CD3+ lymphocytes below 900 cells/mm³, CD4+ lymphocytes below 500 cells/mm³, and CD8+ lymphocytes below 300 cells/mm³. Importantly, this study also reported that the levels of D-dimer, C-reactive protein and IL-6 were elevated in COVID-19 pts., indicating clot formation, severe inflammation and cytokine storm.

14.50.4 Significance

This study sets a threshold to identify patients at risk by analyzing their levels of leukocytes, which is an easy and fast approach to stratify individuals that require hospitalization. Although the study is limited (only counts of WBC are analyzed and not its profile) the data is solid and statistically robust to correlate levels of lymphopenia with mortality.

14.50.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.51 Retrospective Analysis of Clinical Features in 101 Death Cases with COVID-19

Chen et al. *medRxiv*. ([2226](#))

14.51.1 Keywords

- death biomarkers
- cardiac damage
- Troponin
- Blood type
- respiratory failure
- hypertension

14.51.2 Main Findings

This is a retrospective study involving 101 death cases with COVID-19 in Wuhan Jinyintan Hospital. The aim was to describe clinical, epidemiological and laboratory features of fatal cases in order to identify the possible primary mortality causes related to COVID-19.

Among 101 death cases, 56.44% were confirmed by RT-PCR and 43.6% by clinical diagnostics. Males dominated the number of deaths and the average age was 65.46 years. All patients died of respiratory failure and multiple organs failure, except one (acute coronary syndrome). The predominant comorbidities were hypertension (42.57%) and diabetes (22.77%). 25.74% of the patients presented more than two underlying diseases. 82% of patients presented myocardial enzymes abnormalities at admission and further increase in myocardial damage indicators with disease progression: patients with elevated Troponin I progressed faster to death. Alterations in coagulation were also detected. Indicators of liver and kidney damage increased 48 hours before death. The authors studied the deceased patients' blood type and presented the following results: type A (44.44%), type B (29.29%), type AB (8.08%) and type O (18.19%), which is inconsistent with the distribution in Han population in Wuhan.

Clinical analysis showed that the most common symptom was fever (91.9%), followed by cough and dyspnea. The medium time from onset of symptoms to acute respiratory distress syndrome (ARDS) development was 12 days. Unlike SARS, only 2 patients with COVID-19 had diarrhea. 98% presented abnormal lung imaging at admission and most had double-lung abnormalities. Related to the laboratorial findings some inflammatory indicators gradually increased during the disease progression, such as IL-6 secretion in the circulation, procalcitonin (PCT) and C-reactive protein (CRP), while platelets numbers decreased. The authors also reported an initial lymphopenia that was followed by an increase in the lymphocytes numbers. Neutrophil count increased with disease progression.

The patients received different treatments such as antiviral drugs (60.40%), glucocorticoids, thymosin and immunoglobulins. All patients received antibiotic treatment and some received antifungal drugs. All patients received oxygen therapy (invasive or non-invasive ones).

14.51.3 Limitations

This study involves just fatal patients, lacking comparisons with other groups of patients e.g. patients that recovered from COVID-19. The authors didn't discuss the different approaches used for treatments and how these may affect the several parameters measured. The possible relationship between the increase of inflammatory indicators and morbidities of COVID-19 are not discussed.

14.51.4 Significance

This study has the largest cohort of fatal cases reported so far. The authors show that COVID-19 causes fatal respiratory distress syndrome and multiple organ failure. This study highlights prevalent myocardial damage and indicates that cardiac function of COVID-19 patients should be carefully monitored. The data suggest that Troponin I should be further investigated as an early indicator of patients with high risk of accelerated health deterioration. Secondary bacterial and fungal infections were frequent in critically ill patients and these need to be carefully monitored in severe COVID-19 patients. Differences in blood type distribution were observed, suggesting that type A is detrimental while type O is protective – but further studies are needed to confirm these findings and elucidate if blood type influences infection or disease severity. Several inflammatory indicators (neutrophils, PCT, CRP and IL-6, D-dimer) increased according to disease severity and should be assessed as biomarkers and to better understand the biology of progression to severe disease.

14.51.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.52 Relationship between the ABO Blood Group and the COVID-19 Susceptibility

Zhao et al. *medRxiv*. ([452](#))

14.52.1 Keywords

- ABO blood group
- COVID-19 susceptibility

14.52.2 Main Findings

These authors compared the ABO blood group of 2,173 patients with RT-PCR-confirmed COVID-19 from hospitals in Wuhan and Shenzhen with the ABO blood group distribution in unaffected people in the same cities from previous studies (2015 and 2010 for Wuhan and Shenzhen, respectively). They found that people with blood group A are statistically over-represented

in the number of those infected and who succumb to death while those with blood group O are statistically underrepresented with no influence of age or sex.

14.52.3 Limitations

This study compares patients with COVID-19 to the general population but relies on data published 5 and 10 years ago for the control. The mechanisms that the authors propose may underlie the differences they observed require further study.

14.52.4 Significance

Risk stratification based on blood group may be beneficial for patients and also healthcare workers in infection control. Additionally, investigating the mechanism behind these findings could lead to better developing prophylactic and therapeutic targets for COVID-19.

14.52.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.53 The inhaled corticosteroid ciclesonide blocks coronavirus RNA replication by targeting viral NSP15

Matsuyama et al. *bioRxiv* ([2227](#))

14.53.1 Keywords

- Corticosteroids
- ciclesonide
- mometasone
- NSP15
- MERS-CoV

14.53.2 Main Findings

This study reconsiders the use of inhaled corticosteroids in the treatment of pneumonia by coronavirus. Corticosteroids were associated with increased mortality for SARS in 2003 and for MERS in 2013, probably due to that fact that systemic corticosteroids suppress the innate immune system, resulting in increased viral replication. However, some steroid compounds might block coronavirus replication. The authors screened steroids from a chemical library and assessed the viral growth suppression and drug cytotoxicity. Ciclesonide demonstrated low cytotoxicity and potent suppression of MERS-CoV viral growth. The commonly used systemic steroids cortisone, prednisolone and dexamethasone did not suppress viral growth, nor did the

commonly used inhaled steroid fluticasone. To identify the drug target of virus replication, the authors conducted 11 consecutive MERS-CoV passages in the presence of ciclesonide or mometasone, and they could generate a mutant virus that developed resistance to ciclesonide, but not to mometasone. Afterwards, they performed next-generation sequencing and identified an amino acid substitution in nonstructural protein 15 (NSP15) as the predicted mechanism for viral resistance to ciclesonide. The authors were able to successfully generate a recombinant virus carrying that amino acid substitution, which overcome the antiviral effect of ciclesonide, suggesting that ciclesonide interacts with NSP15. The mutant virus was inhibited by mometasone, suggesting that the antiviral target of mometasone is different from that of ciclesonide. Lastly, the effects of ciclesonide and mometasone on suppressing the replication of SARS-CoV-2 were evaluated. Both compounds were found to suppress viral replication with a similar efficacy to lopinavir.

14.53.3 Limitations

Most of the experiments, including the identification of the mutation in NSP15 were conducted with MERS-CoV. This is not the closest related virus to SARS-CoV-2, as that would be SARS-CoV. Thus, to repeat the initial experiments with SARS-CoV, or preferably SARS-CoV-2, is essential. The manuscript should address this and, therefore, it will require considerable editing for organization and clarity. Also, in terms of cell immunogenic epitopes, while SARS-CoV-2 spike protein contains several predicted B and T cell immunogenic epitopes that are shared with other coronaviruses, some studies have shown critical differences between MERS-CoV, SARS-CoV and SARS-CoV-2. A main criticism is that the authors only used VeroE6/TMPRSS2 cells to gauge the direct cytotoxic effects of viral replication. To evaluate this in other cell lines, including human airway epithelial cells, is crucial, as the infectivity of coronavirus strains greatly varies in different cell lines,

14.53.4 Significance

Nevertheless, these findings encourage evaluating ciclesonide and mometasone as better options for patients with COVID-19 in need of inhaled steroids, especially as an alternative to other corticosteroids that have been shown to increase viral replication in vitro. This should be evaluated in future clinical studies.

14.53.5 Credit

This review was undertaken by Alvaro Moreira, MD as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.54 A human monoclonal antibody blocking SARS-CoV-2 infection **

14.54.1 Keywords

- Monoclonal antibodies
- SARS-CoV2
- cross-neutralization
- potential treatment
- spike receptor

14.54.2 Main Findings

The authors reported a human monoclonal antibody that neutralizes SARS-CoV-2 and SARS-CoV which belong to same family of corona viruses. For identifying mAbs, supernatants of a collection of 51 hybridomas raised against the spike protein of SARS-CoV (SARS-S) were screened by ELISA for cross-reactivity against the spike protein of SARS-CoN2 (SARS2-S). Hybridomas were derived from immunized transgenic H2L2 mice (chimeric for fully human VH-VL and rat constant region). Four SARS-S hybridomas displayed cross-reactivity with SARS2-S, one of which (47D11) exhibited cross-neutralizing activity for SARS-S and SARS2-S pseudotyped VSV infection. A recombinant, fully human IgG1 isotype antibody was generated and used for further characterization.

The humanized 47D11 antibody inhibited infection of VeroE6 cells with SARS-CoV and SARS-CoV-2 with IC50 values of 0.19 and 0.57 µg/ml respectively. 47D11 mAb bound a conserved epitope on the spike receptor binding domain (RBD) explaining its ability to cross-neutralize SARS-CoV and SARS-CoV-2. 47D11 was shown to target the S1B RBD of SARS-S and SARS2-S with similar affinities. Interestingly, binding of 47D11 to SARS-S1B and SARS2-S1B did not interfere with S1B binding to ACE2 receptor-expressing cells assayed by flow cytometry.

14.54.3 Limitations

These results show that the human 47D11 antibody neutralizes SARS-CoV and SARS-CoV2 infectivity via an as yet unknown mechanism that is different from receptor binding interference. Alternative mechanisms were proposed but these as yet remain to be tested in the context of SARS-CoV2. From a therapeutic standpoint and in the absence of in vivo data, it is unclear whether the 47D11 ab can alter the course of infection in an infected host through virus clearance or protect an uninfected host that is exposed to the virus. There is a precedent for the latter possibility as it relates to SARS-CoV that was cited by the authors and could turn out to be true for SARS-CoV2.

14.54.4 Significance

This study enabled the identification of novel neutralizing antibody against COV-that could potentially be used as first line of treatment in the near future to reduce the viral load and adverse effects in infected patients. In addition, neutralizing antibodies such as 47D11 represent promising reagents for developing antigen-antibody-based detection test kits and assays.

14.54.5 Credit

This review was edited by K. Alexandropoulos as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

Heat inactivation of serum interferes with the immunoanalysis of antibodies to SARS-CoV-2

Heat inactivation, immunochromatography, diagnosis, serum antibodies, IgM, IgG

Summary

The use of heat inactivation to neutralize pathogens in serum samples collected from suspected COVID-19 patients reduces the sensitivity of a fluorescent immunochromatographic assay to detect anti-SARS-CoV-2 IgM and IgG.

Major findings

Coronaviruses can be killed by heat inactivation, and this is an important safety precaution in laboratory manipulation of clinical samples. However, the effect of this step on downstream SARS-CoV-2-specific serum antibody assays has not been examined. The authors tested the effect of heat inactivation (56 deg C for 30 minutes) versus no heat inactivation on a fluorescence immunochromatography assay. Heat inactivation reduced all IgM measurements by an average of 54% and most IgG measurements (22/36 samples, average reduction of 50%), consistent with the lower thermal stability of IgM than that of IgG. Heat inactivation caused a subset of IgM but not IgG readings to fall below a specified positivity threshold.

Limitations

Limitations included the use of only one type of assay for testing heat inactivated vs non-inactivated sera, and the use of the same baseline for heat inactivated and non-inactivated sera. The results indicate that heat inactivation affects the quantification of SARS-CoV-2-antibody response, specially IgM, but still allows to distinguish positive specific IgG. Therefore, the effect of heat inactivation should be studied when designing assays that quantitatively associate immunoglobulin levels (especially IgM) to immune state.

Review by Andrew M. Leader as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn school of medicine, Mount Sinai.

14.55 Immune phenotyping based on neutrophil-to-lymphocyte ratio and IgG predicts disease severity and outcome for patients with

COVID-19

Zhang et al. *medRxiv* ([2228](#))

14.55.1 Keywords

- Biomarkers
- cytokines
- IgG
- immune cells

14.55.2 Main Findings

In a cohort of 222 patients, anti-SARS-CoV-2 IgM and IgG levels were analyzed during acute and convalescent phases (up to day 35) and correlated to the diseases' severity. The same was done with neutrophil-to-lymphocyte ratio. High IgG levels and high neutrophil-to-lymphocyte ratio in convalescence were both independently associated to the severity of the disease. The simultaneous occurrence of both of these laboratory findings correlated even stronger to the diseases' severity.

Severe cases with high neutrophil-to-lymphocyte ratios had clearly higher levels of IL-6. The authors propose that a robust IgG response leads to immune-mediated tissue damage, thus explaining the worse outcome in patients with overexuberant antibody response.

14.55.3 Limitations

A main criticism is that the criteria for stratifying patients in severe vs. non-severe are not described. The only reference related to this is the difference between the percentage of patients who needed mechanical ventilation, which was greater in patients with both high IgG levels and high neutrophil-to-lymphocyte ratio. No patient with both low IgG levels and low neutrophil-to-lymphocyte ratio was treated with mechanical ventilation.

The proposed correlation of severity with IL-2 and IL-10 levels is not very strong.

Furthermore, although mostly ignored in the paper's discussion, one of the most interesting findings is that an early increase in anti-SARS-CoV-2 IgM levels also seems to correlate with severe disease. However, as only median values are shown for antibody kinetics curves, the extent of variation in acute phase cannot be assessed.

14.55.4 Significance

Anti-SARS-CoV-2 IgG levels and with neutrophil-to-lymphocyte ratio predict severity of COVID-19 independently of each other. An additive predictive value of both variables is noticeable. Importantly, an early-on increase in anti-SARS-CoV-2 IgM levels also seem to predict outcome.

14.55.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.56 Reinfection could not occur in SARS-2 CoV-2 infected rhesus macaques

Bao et al. *bioRxiv* ([2229](#))

14.56.1 Keywords

- SARS-CoV-2
- viral load
- reinfection
- relapse
- non-human primate model

14.56.2 Main Findings

This study addresses the issue of acquired immunity after a primary COVID-19 infection in rhesus monkeys. Four Chinese rhesus macaques were intratracheally infected with SARS-CoV-2 and two out of the four were re-infected at 28 days post initial infection (dpi) with the same viral dose after confirming the recovery by the absence of clinical symptoms, radiological abnormalities and viral detection (2 negative RT-PCR tests). While the initial infection led the viral loads in nasal and pharyngeal swabs that reach approximately $6.5 \log_{10}$ RNA copies/ml at 3 dpi in all four monkeys, viral loads in the swabs tested negative after reinfection in the two reinfected monkeys. In addition, the necropsies from a monkey (M1) at 7 days after primary infection, and another monkey (M3) at 5 days post reinfection, revealed the histopathological damages and viral replication in the examined tissues from M1, while no viral replication as well as no histological damages were detected in the tissues from M3. Furthermore, sera from three monkeys at 21 and 28 dpi exhibited neutralizing activity against SARS-CoV-2 in vitro, suggesting the production of protective neutralizing antibodies in these monkeys. Overall, this study indicates that primary infection with SARS-CoV-2 may protect from subsequent exposure to the same virus.

14.56.3 Limitations

In human, virus has been detected by nasopharyngeal swabs until 9 to 15 days after the onset of symptoms. In the infected monkeys in this study, virus were detected from day 1 after the infection, declining to undetectable level by day 15 post infection. It may suggest that there is a faster viral clearance mechanism in monkeys, therefore the conclusions of reinfection protection for humans need to be carefully considered. In addition, only two monkeys were re-infected in this study and the clinical signs of these monkeys were not similar: M3 did not show weight loss and M4 showed relatively higher fever on the day of infection and the day of re-challenge.

14.56.4 Significance

This study showed clear viral clearance and no indications of relapse or viremia after a secondary infection with SARS-CoV-2 in a Chinese rhesus macaque model. These results support the idea that patients with full recovery (two negative RT-PCR results) may also be protected from secondary SARS-CoV-2 infection. Recovered patients may be able to reintegrate to normal public life and provide protective serum perhaps even if having had a mild infection. The results are also encouraging for successful vaccine development against SARS-CoV-2.

14.56.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.57 A highly conserved cryptic epitope in the receptor-binding domains of SARS-CoV-2 and SARS-CoV

(2230)

14.57.1 Keywords

- neutralizing antibody
- cross-reactivity

14.57.2 Main Findings

Given the sequence similarity of the surface spike glycoprotein (S) of SARS-CoV-2 and SAR-CoV, Yuan et al. (2020) propose that neutralizing antibodies isolated from convalescent SARS-CoV patients may offer insight into cross-reactive antibodies targeting SARS-CoV-2. In particular, they find that the receptor-binding domain (RBD) of SARS-CoV-2 S protein shares 86% sequence similarity with the RBD of SARS-CoV S protein that binds to the CR3022 neutralizing antibody. CR3022 also displays increased affinity for the “up” conformation of the SARS-CoV-2 S protein compared to the “down” conformation as it does for the SARS-CoV S protein. Therefore, the authors propose that this cross-reactive antibody may confer some degree of protection *in vivo* even if it fails to neutralize *in vitro*.

14.57.3 Limitations

Although the authors offer a logical rationale for identifying cross-reactive neutralizing antibodies derived from SARS-CoV, their study using only CR3022 failed to demonstrate whether this approach will be successful. After all, CR3022 failed to neutralize *in vitro* despite the binding affinity to a similar

epitope on SARS-CoV-2. They would benefit from testing more candidates and using an *in vivo* model to demonstrate their claim that protection may be possible in the absence neutralization if combinations are used *in vivo*.

14.57.4 Significance

The ability to make use of previously characterized neutralizing antibodies for conserved epitopes can expedite drug design and treatment options.

14.57.5 Credit

This review was undertaken by Dan Fu Ruan, Evan Cody and Venu Pothula as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.58 Highly accurate and sensitive diagnostic detection of SARS-CoV-2 by digital PCR

Dong et al. *medRxiv* ([2231](#))

14.58.1 Keywords

- Diagnosis
- digital PCR

14.58.2 Main Findings

The authors present a digital PCR (dPCR) diagnostic test for SARS-CoV-2 infection. In 103 individuals that were confirmed in a follow-up to be infected, the standard qPCR test had a positivity rate of 28.2% while the dPCR test detected 87.4% of the infections by detecting an additional 61 positive cases. The authors also tested samples from close contacts (early in infection stage) and convalescing individuals (late in infection stage) and were able to detect SARS-CoV-2 nucleic acid in many more samples using dPCR compared to qPCR.

14.58.3 Limitations

I did not detect limitations.

14.58.4 Significance

The authors make a strong case for the need for a highly sensitive and accurate confirmatory method for diagnosing COVID-19 during this outbreak and present a potential addition to the diagnostic arsenal. They propose a dPCR test that they present has a dramatically lower false negative rate than the standard RT-qPCR tests and can be especially beneficial in people with low viral load, whether they are in the earlier or later stages of infection.

14.58.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.59 SARS-CoV-2 invades host cells via a novel route: CD147-spike protein

Wang et al. *bioRxiv* ([2232](#))

14.59.1 Keywords

- spike protein
- viral entry
- CD147
- SARS-CoV-2

14.59.2 Main Findings

The authors propose a novel mechanism of SARS-CoV-2 viral entry through the interaction of the viral spike protein (SP) and the immunoglobulin superfamily protein CD147 (also known as Basigin). Using an in-house developed humanized antibody against CD147 (maplazumab), they show that blocking CD147 decreases viral replication in Vero E6 cells. Using surface plasmon resonance (SPR), ELISA, and Co-IP assays, they show that the spike protein of SARS-CoV-2 directly interacts with CD147. Lastly, they utilize immune-electron microscopy to show spike protein and CD147 localize to viral inclusion bodies of Vero E6 cells.

14.59.3 Limitations

The authors claim that an anti-CD147 antibody (Meplazumab) inhibits SARS-CoV-2 replication by testing cell growth and viral load in cells infected with SARS-CoV-2, however there are key pieces of this experiment that are missing. First, the authors fail to use a non-specific antibody control. Second, the authors claim that viral replication is inhibited, and that they test this by qPCR, however this data is **not shown**. To further prove specificity, the authors should introduce CD147 to non-susceptible cells and show that they become permissive.

The authors claim that there is a direct interaction between CD147 and SP through SPR, ELISA, and Co-IP, and this data seems generally convincing. The electron microscopy provides further correlative evidence that SARS-CoV-2 may interact with CD147 as they are both found in the same viral inclusion body. A quantification of this data would make the findings more robust.

Finally, the data in this paper lacks replicates, error bars, and statistics to show that the data are reproducible and statistically significant.

14.59.4 Significance

It has been shown in various studies that SARS-CoV-2 binds to the cell surface protein ACE2 for cell entry, yet ACE2 is highly expressed in heart, kidney, and intestinal cells, raising the concern that blocking ACE2 would result in harmful side effects ([2233](#)) CD147 on the other hand is highly expressed in various tumor types, inflamed tissues, and pathogen infected cells, suggesting that the inhibition of CD147 would not result in major side effects ([2234](#), [2235](#)) The research in this paper has resulted in an ongoing clinical trail in China to test the safety and efficacy of anti-CD147 Meplazumab to treat COVID-19. (ClinicalTrials.gov identifier NCT04275245).

14.59.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.60 Blood single cell immune profiling reveals that interferon-MAPK pathway mediated adaptive immune response for COVID-19

Huang et al. *medRxiv* ([2236](#))

14.60.1 Keywords

- COVID-19
- SARS-CoV-2
- PBMC
- single cell
- MAPK

14.60.2 Main Findings

The authors performed single-cell RNA sequencing (scRNAseq) of peripheral blood mononuclear cells isolated from whole blood samples of COVID-19 patients (n=10). Data was compared to scRNAseq of samples collected from patients with influenza A (n=1), acute pharyngitis (n=1), and cerebral infarction (n=1), as well as, three healthy controls. COVID-19 patients were categorized into those with moderate (n=6), severe (n=1), critical (n=1), and cured (n=2) disease. Analysis across all COVID-19 disease levels revealed 56 different cellular subtypes, among 17 immune cell types; comparisons between each category to the normal controls revealed **increased proportions of CD1c⁺ dendritic cells, CD8⁺ CTLs, and plasmacytoid dendritic cells and a decrease in proportions of B cells and CD4⁺ T cells.**

TCR sequencing revealed that greater clonality is associated with milder COVID-19 disease; BCR sequencing revealed that COVID-19 patients have circulating antibodies against known viral antigens, including EBV, HIV,

influenza A, and other RNA viruses. This may suggest that the immune response to SARS-CoV-2 infection elicits production of antibodies against known RNA viruses.

Excluding enriched pathways shared by COVID-19 patients and patients with other conditions (influenza A, acute pharyngitis, and cerebral infarction), the authors identified the **interferon-MAPK signaling pathway as a major response to SARS-CoV-2 infection**. The authors performed quantitative real-time reverse transcriptase polymerase chain reaction (RT-PCR) for interferon-MAPK signaling genes: *IRF27*, *BST2*, and *FOS*. These samples were collected from a separate cohort of COVID-19 patients (critical, n=3; severe, n=3; moderate, n=19; mild, n=3; and cured, n=10; and healthy controls, n=5). Notably, consistent with the original scRNAseq data, *FOS* showed up-regulation in COVID-19 patients and down-regulation in cured patients. **The authors propose that *FOS* may be a candidate marker gene for curative COVID-19 disease.**

14.60.3 Limitations

The sample size of this study is limited. To further delineate differences in the immune profile of peripheral blood of COVID-19 patients, a greater sample size is needed, and longitudinal samples are needed, as well. A better understanding of the immunological interactions in cured patients, for example, would require a profile before and after improvement.

Moreover, the conclusions drawn from this scRNAseq study point to potential autoimmunity and immune deficiency to distinguish different severities of COVID-19 disease. However, this requires an expanded number of samples and a more robust organization of specific immune cell subtypes that can be compared across different patients. Importantly, this criterion is likely needed to ensure greater specificity in identifying markers for COVID-19 infection and subsequent immune response.

14.60.4 Significance

At the single-cell level, COVID-19 disease has been characterized in the lung, but a greater understanding of systemic immunological responses is furthered in this study. Type I interferon is an important signaling molecule for the anti-viral response. The identification of the interferon-MAPK signaling pathway and the differential expression of MAPK regulators between patients of differing COVID-19 severity and compared to cured patients may underscore the importance of either immune deficiency or autoimmunity in COVID-19 disease.

14.60.5 Credit

This review was undertaken by Matthew D. Park as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.61 Cross-reactive antibody response between SARS-CoV-2 and SARS-CoV infection.

Lv et al. *bioRxiv* ([2237](#))

14.61.1 Keywords

SARS-CoV-2, SARS-CoV, spike protein, RBD, cross-reactivity, cross-neutralization, antibody, human patients, mouse

14.61.2 Main Findings

The authors explore the antigenic differences between SARS-CoV-2 and SARS-CoV by analyzing plasma samples from SARS-CoV-2 (n = 15) and SARS-CoV (n = 7) patients. Cross-reactivity in antibody binding to the spike protein between SARS-CoV-2 and SARS-CoV was found to be common, mostly targeting non-RBD regions in plasma from SARS-CoV-2 patients. Only one SARS-CoV-2 plasma sample was able to cross-neutralize SARS-CoV, with low neutralization activity. No cross-neutralization response was detected in plasma from SARS-CoV patients.

To further investigate the cross-reactivity of antibody responses to SARS-CoV-2 and SARS-CoV, the authors analyzed the antibody response of plasma collected from mice infected or immunized with SARS-CoV-2 or SARS-CoV (n = 5 or 6 per group). Plasma from mice immunized with SARS-CoV-2 displayed cross-reactive responses to SARS-CoV S ectodomain and, to a lesser extent, SARS-CoV RBD. Similarly, plasma from mice immunized with SARS-CoV displayed cross-reactive responses to SARS-CoV-2 S ectodomain. Cross-neutralization activity was not detected in any of the mouse plasma samples.

14.61.3 Limitations

The size of each patient cohort is insufficient to accurately determine the frequency of cross-reactivity and cross-neutralization in the current SARS-CoV-2 pandemic. Recruitment of additional patients from a larger range of geographical regions and time points would also enable exploration into the effect of the genetic diversity and evolution of the SARS-CoV-2 virus on cross-reactivity. This work would also benefit from the mapping of specific epitopes for each sample. Future studies may determine whether the non-neutralizing antibody responses can confer *in vitro* protection or lead to antibody-dependent disease enhancement.

14.61.4 Significance

The cross-reactive antibody responses to S protein in the majority of SARS-CoV-2 patients is an important consideration for development of serological assays and vaccine development during the current outbreak. The limited extent of cross-neutralization demonstrated in this study indicates that vaccinating to cross-reactive conserved epitopes may have limited efficacy,

presenting a key concern for the development of a more universal coronavirus vaccine to address the global health risk of novel coronavirus outbreaks.

14.61.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.62 The feasibility of convalescent plasma therapy in severe COVID-19 patients: a pilot study

Duan et al. *medRxiv* ([2238](#))

14.62.1 Keywords

- COVID-19
- SARS-CoV-2
- convalescent plasma
- treatment outcome
- pilot
- therapy
- transfusion

14.62.2 Main Findings

This is the first report to date of convalescent plasma therapy as a therapeutic against COVID-19 disease. This is a feasibility pilot study. The authors report the administration and clinical benefit of 200 mL of convalescent plasma (CP) (1:640 titer) derived from recently cured donors (CP selected among 40 donors based on high neutralizing titer and ABO compatibility) to 10 severe COVID-19 patients with confirmed viremia. The primary endpoint was the safety of CP transfusion. The secondary endpoint were clinical signs of improvement based on symptoms and laboratory parameters.

The authors reported use of methylene blue photochemistry to inactivate any potential residual virus in the plasma samples, without compromising neutralizing antibodies, and no virus was detected before transfusion.

The authors report the following:

- No adverse events were observed in all patients, except 1 patient who exhibited transient facial red spotting.
- All patients showed significant improvement in or complete disappearance of clinical symptoms, including fever, cough, shortness of breath, and chest pain after 3 days of CP therapy.

- Reduction of pulmonary lesions revealed by chest CT.
- Elevation of lymphocyte counts in patients with lymphocytopenia.
- Increase in SaO₂ in all patients, indicative of recuperating lung function.
- Resolution of SARS-CoV-2 viremia in 7 patients and increase in neutralizing antibody titers in 5 patients. Persistence of neutralizing antibody levels in 4 patients.

14.62.3 Limitations

It is important to note that most recipients had high neutralization titers of antibodies before plasma transfusion and even without transfusion it would be expected to see an increase in neutralizing antibodies over time. In addition to the small sample set number (n=10), there are additional limitations to this pilot study:

1. All patients received concurrent therapy, in addition to the CP transfusion. Therefore, it is unclear whether a combinatorial or synergistic effect between these standards of care and CP transfusion contributed to the clearance of viremia and improvement of symptoms in these COVID-19 patients.
2. The kinetics of viral clearance was not investigated, with respect to the administration of CP transfusion. So, the definitive impact of CP transfusion on immune dynamics and subsequent viral load is not well defined.
3. Comparison with a small historical control group is not ideal.

14.62.4 Significance

For the first time, a pilot study provides promising results involving the use of convalescent plasma from cured COVID-19 patients to treat others with more severe disease. The authors report that the administration of a single, high-dose of neutralizing antibodies is safe. In addition, there were encouraging results with regards to the reduction of viral load and improvement of clinical outcomes. It is, therefore, necessary to expand this type of study with more participants, in order to determine optimal dose and treatment kinetics. It is important to note that CP has been studied to treat H1N1 influenza, SARS-CoV-1, and MERS-CoV, although it has not been proven to be effective in treating these infections.

14.62.5 Credit

Review by Matthew D. Park and revised by Alice O. Kamphorst and Maria A. Curotto de Lafaille as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.63 Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open label non-randomized clinical trial

([2239](#))

14.63.1 Keywords

- hydroxychloroquine
- clearance
- viral load
- clinical trial

14.63.2 Main Findings

This study was a single-arm, open label clinical trial with 600 mg hydroxychloroquine (HCQ) in the treatment arm (n = 20). Patients who refused participation or patients from another center not treated with HCQ were included as negative controls (n = 16). Among the patients in the treatment arm, 6 received concomitant azithromycin to prevent superimposed bacterial infection. The primary endpoint was respiratory viral loads on day 6 post enrollment, measured by nasopharyngeal swab followed by real-time reverse transcription-PCR.

HCQ alone was able to significantly reduce viral loads by day 6 (n = 8/14, 57.1% complete clearance, $p < 0.001$); azithromycin appears to be synergistic with HCQ, as 6/6 patients receiving combined treatment had complete viral clearance ($p < 0.001$).

14.63.3 Limitations

Despite what is outlined above, this study has a number of limitations that must be considered. First, there were originally n = 26 patients in the treatment arm, with 6 lost to follow up for the following reasons: 3 transferred to ICU, 1 discharge, 1 self-discontinued treatment d/t side effects, and 1 patient expired. Total length of clinical follow up was 14 days, but the data beyond day 6 post-inclusion are not shown.

Strikingly, in supplementary table 1, results of the real-time RT-PCR are listed for the control and treatment arms from D0 – D6. However, the data are not reported in a standard way, with a mix of broadly positive or negative result delineation with Ct (cycle threshold) values, the standard output of real time PCR. It is impossible to compare what is defined as a positive value between the patients in the control and treatment arms without a standardized threshold for a positive test. Further, the starting viral loads reported at D0 in the groups receiving HCQ or HCQ + azithromycin were significantly different (ct of 25.3 vs 26.8 respectively), which could explain in part the differences observed in the response to treatment between 2 groups. Finally, patients in the control arm from outside the primary medical center in this study

(Marseille) did not actually have samples tested by PCR daily. Instead, positive test results from every other day were extrapolated to mean positive results on the day before and after testing as well (Table 2, footnote ^a).

Taken together, the results of this study suggest that HCQ represents a promising treatment avenue for COVID-19 patients. However, the limited size of the trial, and the way in which the results were reported does not allow for other medical centers to extrapolate a positive or negative result in the treatment of their own patients with HCQ +/- azithromycin. Further larger randomized clinical trials will be required to ascertain the efficacy of HCQ +/- azithromycin in the treatment of COVID-19.

14.63.4 Significance

Chloroquine is thought to inhibit viral infection, including SARS-Cov-2, by increasing pH within endosomes and lysosomes, altering the biochemical conditions required for viral fusion ([806](#), [2240](#)). However, chloroquine also has immuno-modulatory effects that I think may play a role. Chloroquine has been shown to increase CTLA-4 expression at the cell surface by decreasing its degradation in the endo-lysosome pathway; AP-1 traffics the cytoplasmic tail of CTLA-4 to lysosomes, but in conditions of increased pH, the protein machinery required for degradation is less functional ([2241](#)). As such, more CTLA-4 remains in endosomes and is trafficked back to the cell surface. It is possible that this may also contribute to patient recovery via reduction of cytokine storm, in addition to the direct anti-viral effects of HCQ.

14.63.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.64 Recapitulation of SARS-CoV-2 Infection and Cholangiocyte Damage with Human Liver Organoids

([2166](#))

14.64.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- liver
- organoids
- Cholangiocyte

14.64.2 Main Findings

- Used human liver ductal organoids to determine ACE2+ cholangiocytes in healthy liver (2.92% of all cells) are infectable and support SARS-CoV-2 viral replication.
- Plaque-purified SARS-CoV-2 viral infection disrupted organoid barrier and bile transporting functions of cholangiocytes through dysregulation of genes involved in tight junction formation (CLDN1) and bile acid transportation (ASBT and CFTR).

14.64.3 Limitations:

- Unclear if liver damage observed in patients due to direct cholangiocyte infection or due to secondary immune/cytokine effects. This study argues for direct damage as it lacks immune contexture; but further studies needed with autopsy samples or organoid-immune cell co-culture to conclude strongly.
- Would be important to measure cholangiocyte-intrinsic anti-viral response and alarmins secreted upon infection, and furthermore study tropism of various immune cells to conditioned media from organoids infected with SARS-CoV-2.
- Does not address how cirrhotic liver or alcohol/smoking/obesity-associated liver organoids respond to SARS-CoV-2 infectivity and replication, worth pursuing to experimentally address clinical data indicating co-morbidities.

14.64.4 Significance

- Useful model to rapidly study drug activity against SARS-CoV-2 infection in liver, while monitoring baseline liver damage.
- Liver abnormality observed in >50% of CoVID-19 patients; the results from this study could explain the bile acid accumulation and consequent liver damage observed.

14.64.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.65 The sequence of human ACE2 is suboptimal for binding the S spike protein of SARS coronavirus 2

(2242)

14.65.1 Keywords

- Coronavirus Disease 2019 (COVID-19)
- SARS-CoV-2
- Spike protein S
- ACE2

14.65.2 Main Findings

Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) infects cells through S spike glycoprotein binding angiotensin-converting enzyme (ACE2) on host cells. S protein can bind both membrane-bound ACE2 and soluble ACE2 (sACE2), which can serve as a decoy that neutralizes infection. Recombinant sACE2 is now being tested in clinical trials for COVID-19. To determine if a therapeutic sACE2 with higher affinity for S protein could be designed, authors generated a library containing every amino acid substitution possible at the 117 sites spanning the binding interface with S protein. The ACE2 library was expressed in human Expi293F cells and cells were incubated with medium containing the receptor binding domain (RBD) of SARS-CoV-2 fused to GFP. Cells with high or low affinity mutant ACE2 receptor compared to affinity of wild type ACE2 for the RBD were FACS sorted and transcripts from these sorted populations were deep sequenced. Deep mutagenesis identified numerous mutations in ACE2 that enhance RBD binding. This work serves to identify putative high affinity ACE2 therapeutics for the treatment of CoV-2.

14.65.3 Limitations

The authors generated a large library of mutated ACE2, expressed them in human Expi293F cells, and performed deep mutagenesis to identify enhanced binders for the RBD of SARS-CoV-2 S protein. While these data serve as a useful resource, the ability of the high affinity ACE2 mutants identified to serve as therapeutics needs further validation in terms of conformational stability when purified as well as efficacy/safety both *in vitro* and *in vivo*. Additionally, authors mentioned fusing the therapeutic ACE2 to Fc receptors to elicit beneficial host immune responses, which would need further design and validation.

14.65.4 Significance

This study identified structural ACE2 mutants that have potential to serve as therapeutics in the treatment of SARS-CoV-2 upon further testing and validation.

14.65.5 Credit

This review was undertaken by Katherine Lindblad and Tamar Plitt as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

Title: A serological assay to detect SARS-Cov-2 seroconversion in humans

Immunology keywords: specific serological assay - ELISA - seroconversion - antibody titers

Note: the authors of this review work in the same institution as the authors of the study

Main findings:

Production of recombinant whole Spike (S) protein and the smaller Receptor Binding Domain (RBD) based on the sequence of Wuhan-Hu-1 SARS-CoV-2 isolate. The S protein was modified to allow trimerization and increase stability. The authors compared the antibody reactivity of 59 banked human serum samples (non-exposed) and 3 serum samples from confirmed SARS-CoV-2 infected patients. All Covid-19 patient sera reacted to the S protein and RBD domain compared to the control sera.

The authors also characterized the antibody isotypes from the Covid-19 patients, and observed stronger IgG3 response than IgG1. IgM and IgA responses were also prevalent.

Limitations of the study:

The authors analyzed a total of 59 control human serum samples, and samples from only three different patients to test for reactivity against the RBD domain and full-length spike protein. It will be important to follow up with a larger number of patient samples to confirm the data obtained. Furthermore, it would be interesting to assess people at different age groups and determine whether unexposed control kids have a higher “background”.

Applications of the assay described in this study in diagnosis are limited, since antibody response should start to be detectable only one to two weeks after infection. Future studies will be required to assess how long after infection this assay allow to detect anti-CoV2 antibodies. Finally, while likely, the association of seroconversion with protective immunity against SARS-Cov-2 infection still needs to be fully established.

Relevance:

This study has strong implications in the research against SARS-Cov-2. First, it is now possible to perform serosurveys and determine who has been infected, allowing a more accurate estimate of infection prevalence and death rate. Second, if it is confirmed that re-infection does not happen (or is rare), this assay can be used as a tool to screen healthcare workers and prioritize immune ones to work with infected patients. Third, potential convalescent plasma donors can now be screened to help treating currently infected patients. Of note, this assay does not involve live virus handling. experimentally, this is an advantage as the assay does not require the precautions required by manipulation of live virus. Finally, the recombinant proteins described in this study represent new tools that can be used for further applications, including vaccine development.

14.66 COMPARATIVE PATHOGENESIS OF COVID-19, MERS AND SARS IN A NON-HUMAN PRIMATE MODEL

14.66.1 Keywords

- SARS-CoV2
- cynomolgus macaque
- SARS-CoV

14.66.2 Main Findings

This work assesses SARS-CoV-2 infection in young adult and aged cynomolgus macaques. 4 macaques per age group were infected with low-passage clinical sample of SARS-CoV-2 by intranasal and intratracheal administration. Viral presence was assessed in nose, throat and rectum through RT-PCR and viral culture. SARS-CoV-2 replication was confirmed in the respiratory track (including nasal samples), and it was also detected in ileum. Viral nucleocapsid detection by IHC showed infection of type I and II pneumocytes and epithelia. Virus was found to peak between 2 and 4 days after administration and reached higher levels in aged vs. young animals. The early peak is consistent with data in patients and contrasts to SARS-CoV replication. SARS-CoV-2 reached levels below detection between 8 and 21 days after inoculation and macaques established antibody immunity against the virus by day 14. There were histopathological alteration in lung, but no overt clinical signs. At day 4 post inoculation of SARS-CoV-2, two of four animals presented foci of pulmonary consolidation, with limited areas of alveolar edema and pneumonia, as well as immune cell infiltration. In sum, cynomolgus macaques are permissive to SARS-CoV-2 and develop lung pathology (less severe than SARS-CoV, but more severe than MERS-CoV).

14.66.3 Limitations

Even though cynomolgus macaques were permissive to SARS-CoV-2 replication, it is unclear if the viral load reaches levels comparable to humans and there wasn't overt clinical pathology.

14.66.4 Significance

The development of platforms in which to carry out relevant experimentation on SARS-CoV-2 pathophysiology is of great urgency. Cynomolgus macaques offer an environment in which viral replication can happen, albeit in a limited way and without translating into clinically relevant symptoms. Other groups are contributing to SARS-CoV2 literature using this animal model (2229), potentially showing protection against reinfection in cured macaques. Therefore, this platform could be used to examine SARS-CoV2 pathophysiology while studies in other animal models are also underway (2168, 2244).

14.66.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.67 Investigating the Impact of Asymptomatic Carriers on COVID-19 Transmission

(2245)

14.67.1 Keywords

- Coronavirus Disease 2019 (COVID-19)
- SARS-CoV-2
- asymptomatic carriers
- mathematical model
- transmission

14.67.2 Main Findings

Multiple studies reported the same level of infectiousness between symptomatic and asymptomatic carriers of SARS-CoV-2. Given that asymptomatic and undocumented carriers escape public health surveillance systems, a better mathematical model of transmission is needed to determine a more accurate estimate of the basic reproductive number (R_0) of the virus to assess the contagiousness of virus. The authors developed a SEYAR dynamical model for transmission of the new coronavirus that takes into account asymptomatic and undocumented carriers. The model was validated using data reported from thirteen countries during the first three weeks of community transmission. While current studies estimate R_0 to be around 3, this model indicates that the value could range between 5.5 to 25.4.

14.67.3 Limitations

The SEYAR model realistically depicts transmission of the virus only during the initial stages of the disease. More data is necessary to better fit the model with current trends. In addition, multiple factors (e.g. behavioral patterns, surveillance capabilities, environmental and socioeconomic factors) affect transmission of the virus and so, these factors must be taken into consideration when estimating the R_0 .

14.67.4 Significance

Public health authorities use the basic reproductive number to determine the severity of disease. An accurate estimate of R_0 will inform intervention strategies. This model can be applied to different locations to assess the potential impact of COVID-19.

14.67.5 Credit

This review was undertaken by Tamar Plitt and Katherine Lindblad as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.68 Antibody responses to SARS-CoV-2 in COVID-19 patients: the perspective application of serological tests in clinical practice

Long et al. *medRxiv* ([2246](#))

14.68.1 Keywords

- Serum antibodies
- IgM
- IgG
- immunoassay
- diagnosis
- seroconversion

14.68.2 Main Findings

This study investigated the profile of the acute antibody response against SARS-CoV-2 and provided proposals for serologic tests in clinical practice. Magnetic Chemiluminescence Enzyme Immunoassay was used to evaluate IgM and IgG seroconversion in 285 hospital admitted patients who tested positive for SARS-CoV-2 by RT-PCR and in 52 COVID-19 suspected patients that tested negative by RT-PCR. A follow up study with 63 patients was performed to investigate longitudinal effects. In addition, IgG and IgM titers were evaluated in a cohort of close contacts (164 persons) of an infected couple.

The median day of seroconversion for both IgG and IgM was 13 days after symptom onset. Patients varied in the order of IgM/ IgG seroconversion and there was no apparent correlation of order with age, severity, or hospitalization time. This led the authors to conclude that for diagnosis IgM and IgG should be detected simultaneously at the early phase of infection.

IgG titers, but not IgM titers were higher in severe patients compared to non-severe patients after controlling for days post-symptom onset. Importantly, 12% of COVID-19 patients (RT-PCR confirmed) did not meet the WHO serological diagnosis criterion of either seroconversion or > 4-fold increase in IgG titer in sequential samples. This suggests the current serological criteria may be too stringent for COVID-19 diagnosis.

Of note, 4 patients from a group of 52 suspects (negative RT-PCR test) had anti-SARS-Cov-2 IgM and IgG. Similarly, 4.3% (7/162) of "close contacts" who had negative RT-PCR tests were positive for IgG and/or IgM. This highlights the usefulness of a serological assay to identify asymptomatic infections and/or infections that are missed by RT-PCR.

14.68.3 Limitations

This group's report generally confirms the findings of others that have evaluated the acute antibody response to SARS-Cov-2. However, these data would benefit from inclusion of data on whether the participants had a

documented history of viral infection. Moreover, serum samples that were collected prior to SARS-CoV-2 outbreak from patients with other viral infections would serve as a useful negative control for their assay. Methodological limitations include that only one serum sample per case was tested as well as the heat inactivation of serum samples prior to testing. It has previously been reported that heat inactivation interferes with the level of antibodies to SARS-CoV-2 and their protocol may have resulted in diminished quantification of IgM, specifically ([2247](#)).

14.68.4 Significance

Understanding the features of the antibody responses against SARS-CoV is useful in the development of a serological test for the diagnosis of COVID-19. This paper addresses the need for additional screening methods that can detect the presence of infection despite lower viral titers. Detecting the production of antibodies, especially IgM, which are produced rapidly after infection can be combined with PCR to enhance detection sensitivity and accuracy and map the full spread of infection in communities. Moreover, serologic assays would be useful to screen health care workers in order to identify those with immunity to care for patients with COVID19.

14.68.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.69 SARS-CoV-2 specific antibody responses in COVID-19 patients

([2248](#))

14.69.1 Keywords

- immunoassay
- antibody specificity
- serology
- cross-reactivity

14.69.2 Main findings

Antibodies specific to SARS-CoV-2 S protein, the S1 subunit and the RBD (receptor-binding domain) were detected in all SARS-CoV-2 patient sera by 13 to 21 days post onset of disease. Antibodies specific to SARS-CoV N protein (90% similarity to SARS-CoV-2) were able to neutralize SARS-CoV-2 by PRNT (plaque reduction neutralizing test). SARS-CoV-2 serum cross-reacted with SARS-CoV S and S1 proteins, and to a lower extent with MERS-CoV S protein, but not with the MERS-CoV S1 protein, consistent with an analysis of genetic similarity. No reactivity to SARS-CoV-2 antigens was observed in serum from patients with ubiquitous human CoV infections (common cold) or to non-CoV viral respiratory infections.

14.69.3 Limitations

Authors describe development of a serological ELISA based assay for the detection of neutralizing antibodies towards regions of the spike and nucleocapsid domains of the SARS-CoV-2 virus. Serum samples were obtained from PCR-confirmed COVID-19 patients. Negative control samples include a cohort of patients with confirmed recent exposure to non-CoV infections (i.e. adenovirus, bocavirus, enterovirus, influenza, RSV, CMV, EBV) as well as a cohort of patients with confirmed infections with ubiquitous human CoV infections known to cause the common cold. The study also included serum from patients with previous MERS-CoV and SARS-CoV zoonotic infections. This impressive patient cohort allowed the authors to determine the sensitivity and specificity of the development of their in-house ELISA assay. Of note, seroconversion was observed as early as 13 days following COVID-19 onset but the authors were not clear how disease onset was determined.

14.69.4 Significance

Validated serological tests are urgently needed to map the full spread of SARS-CoV-2 in the population and to determine the kinetics of the antibody response to SARS-CoV-2. Furthermore, clinical trials are ongoing using plasma from patients who have recovered from SARS-CoV-2 as a therapeutic option. An assay such as the one described in this study could be used to screen for strong antibody responses in recovered patients. Furthermore, the assay could be used to screen health care workers for antibody responses to SARS-CoV-2 as personal protective equipment continues to dwindle. The challenge going forward will be to standardize and scale-up the various in-house ELISA's being developed in independent laboratories across the world.

14.70 A brief review of antiviral drugs evaluated in registered clinical trials for COVID-19

Belhadi et al. ([2249](#))

14.70.1 Keywords

- Clinical trials
- COVID-19
- SARS CoV-2
- 2019-nCoV
- SARS Cov-2
- Hcov-19
- novel coronal virus
- new corona virus
- antiviral drugs

14.70.2 Main Findings

Summary of clinical trials registered as of March 7, 2020 from U.S, Chinese, Korean, Iranian and European registries. Out of the 353 studies identified, 115 were selected for data extraction. 80% of the trials were randomized with parallel assignment and the median number of planned inclusions was 63 (IRQ, 36-120). Most frequent therapies in the trials included; 1) antiviral drugs [lopinavir/ritonavir (n=15); umifenovir (n=9); favipiravir (n=7); remdesivir (n=5)]; 2) anti-malaria drugs [chloroquine (n=11); hydroxychloroquine (n=7)]; immunosuppressant drugs [methylprednisolone (n=5)]; and stem cell therapies (n=23). Medians of the total number of planned inclusions per trial for these therapies were also included. Stem cells and lopinavir/ritonavir were the most frequently evaluated candidate therapies (23 and 15 trials respectively), whereas remdesivir was only tested in 5 trials but these trials had the highest median number of planned inclusions per trial (400, IQR 394-453). Most of the agents used in the different trials were chosen based on preclinical assessments of antiviral activity against SARS CoV and MERS CoV corona viruses.

The primary outcomes of the studies were clinical (66%); virological (23%); radiological (8%); or immunological (3%). The trials were classified as those that included patients with severe disease only; trials that included patients with moderate disease; and trials that included patients with severe or moderate disease.

14.70.3 Limitations

The trials evaluated provided incomplete information: 23% of these were phase IV trials but the bulk of the trials (54%) did not describe the phase of the study. Only 52% of the trials (n=60) reported treatment dose and only 34% (n=39) reported the duration. A lot of the trials included a small number of patients and the trials are still ongoing, therefore no insight was provided on the outcome of the trials.

14.70.4 Significance

Nonetheless, this review serves as framework for identifying COVID-19 related trials, which can be expanded upon as new trials begin at an accelerated rate as the disease spreads around the world.

14.70.5 Credit

This review was undertaken by K Alexandropoulos as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.71 ACE-2 Expression in the Small Airway Epithelia of Smokers and COPD Patients: Implications for COVID-19

14.71.1 Keywords

- chronic obstructive pulmonary disease
- COPD
- smokingE-2
- risk factors

14.71.2 Main Findings

In bronchial epithelial samples from 3 different cohorts of individuals, ACE-2 gene expression was found to be significantly increased in both COPD patients and smokers relative to healthy controls. Across all test subjects, ACE-2 gene expression was also highly correlated with decreased forced expiratory volume in 1 second (FEV1), which may explain the increased COVID-19 disease severity in COPD patients. Former smokers were also found to show decreased ACE2 expression relative to current smokers and had no significant difference when compared to non-smokers.

14.71.3 Limitations

While the upregulation of ACE-2 is an interesting hypothesis for COVID-19 disease severity in COPD patients, this study leaves many more unanswered questions than it addresses. Further studies are required to show whether the specific cell type isolated in these studies is relevant to the pathophysiology of COVID-19. Furthermore, there is no attempt to show whether that increased ACE-2 expression contributes to greater disease severity. Does the increased ACE-2 expression lead to greater infectivity with SARS-CoV-2? There is no mechanistic explanation for why ACE-2 levels are increased in COPD patients. The authors could also have considered the impact of co-morbidities and interventions such as corticosteroids or bronchodilators on ACE-2 expression. Finally, given the extensive sequencing performed, the authors could have conducted significantly more in-depth analyses into gene signature differences.

14.71.4 Significance

This study attempts to address an important clinical finding that both smokers and COPD patients show increased mortality from COVID-19. The novel finding that ACE-2 expression is induced in smokers and COPD patients suggests not only a mechanism for the clinical observation, but also highlights the potential benefit of smoking cessation in reducing the risk of severe COVID-19 disease.

14.71.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.72 Dynamic profile of severe or critical COVID-19 cases

14.72.1 Keywords

- Coronavirus Disease 2019 (COVID-19)
- SARS-CoV-2
- progressive lymphopenia (PLD)
- T-lymphocytes
- clinical data
- co-infection
- influenza A

14.72.2 Main Findings

Authors evaluate clinical correlates of 10 patients (6 male and 4 female) hospitalized for severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). All patients required oxygen support and received broad spectrum antibiotics and 6 patients received anti-viral drugs. Additionally, 40% of patients were co-infected with influenza A. All 10 patients developed lymphopenia, two of which developed progressive lymphopenia (PLD) and died. Peripheral blood (PB) lymphocytes were analyzed – low CD4 and CD8 counts were noted in most patients, though CD4:CD8 ratio remained normal.

14.72.3 Limitations

The authors evaluated a small cohort of severe SARS-CoV-2 cases and found an association between T cell lymphopenia and adverse outcomes. However, this is an extremely small and diverse cohort (40% of patients were co-infected with influenza A). These findings need to be validated in a larger cohort. Additionally, the value of this data would be greatly increased by adding individual data points for each patient as well as by adding error bars to each of the figures.

14.72.4 Significance

This study provides a collection of clinical data and tracks evolution of T lymphocyte in 10 patients hospitalized for SARS-CoV-2, of which 4 patients were co-infected with influenza A.

14.72.5 Credit

This review was undertaken by Katherine Lindblad and Tamar Plitt as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.73 Association between Clinical, Laboratory and CT Characteristics and RT-PCR Results in the Follow-up of COVID-19 patients

14.73.1 Keywords

- COVID-19
- clinical
- lymphocyte
- CRP
- LDH
- HSST TNT
- PCR test
- readmission
- CT
- GGO
- disease progression

14.73.2 Study description

Data analyzed from 52 COVID-19 patients admitted and then discharged with COVID-19. Clinical, laboratory, and radiological data were longitudinally recorded with illness timecourse (PCR + to PCR-) and 7 patients (13.5%) were readmitted with a follow up positive test (PCR+) within two weeks of discharge.

14.73.3 Main Findings

- At admission:
 - The majority of patients had increased CRP at admission (63.5%).
 - LDH, and HSST TNT were significantly increased at admission.
 - Radiographic signs via chest CT showed increased involvement in lower lobes: right lower lobe (47 cases, 90.4%), left lower lobe (37 cases, 71.2%).
 - GGO (90.4%), interlobular septal thickening (42.3%), vascular enlargement (42.3%), and reticulation (11.5%) were most commonly observed.
- After negative PCR test (discharge):
 - CRP levels decreased lymphocyte counts (#/L) increased significantly (CD3+, CD3+/8+ and CD3+/4+) after negative PCR.
 - Consolidation and mixed GGO observed in longitudinal CT imaging w different extents of inflammatory exudation in lungs, with overall tendency for improvement (except 2/7 patients that were readmitted after discharge with re-positive test) after negative PCR.
- Seven patients repeated positive RT-PCR test and were readmitted to the hospital (9 to 17 day after initial discharge).

- Follow up CT necessary to monitor improvement during recovery and patients with lesion progression should be given more attention.
- Dynamic CT in addition to negative test essential in clinical diagnosis due to nasal swab PCR sampling bias (false-negatives).
- Increase in CRP occurred in 2 readmitted patients (and decr. in lymphocytes in one patient), but was not correlated with new lesions or disease progression vs. improvement (very low N).
- Patients readmitted attributed to false-negative PCR vs. re-exposure.

14.73.4 Limitations

Patients sampled in this study were generally younger (65.4% < 50 yrs) and less critically ill/all discharged. Small number of recovered patients (N=18). Time of follow up was relatively short.. Limited clinical information available about patients with re-positive test (except CRP and lymph tracking).

14.73.5 Extended Results

NOTE: Patients sampled in this study were generally younger (65.4% < 50 yrs) and less critically ill/all discharged. After two consecutive negative PCR tests, patients were discharged.

Clinical Results at Admission

- Median interval disease onset to admission (5 days, IQR: 3-7)
- Most common symptoms included fever, fatigue, dry cough, and expectoration.
- Fifteen patients had reduced lymphocyte counts (28.8%).
- No change in WBC or Neutrophil counts.
- **The majority of patients had increased CRP at admission (63.5%).**
- **LDH, and HSST TNT were significantly increased at admission.**
- Fibrinogen was trending high though not significant.
- No major changes in liver function observed.
- **Radiographic signs via chest CT showed increased involvement in lower lobes: right lower lobe (47 cases, 90.4%), left lower lobe (37 cases, 71.2%).**
- **GGO (90.4%), interlobular septal thickening (42.3%), vascular enlargement (42.3%),** and reticulation (11.5%) were most commonly observed.

Change in Clinical Results following Negative Test

- **CRP levels decreased after negative PCR.**
- **Lymphocyte counts (#/L) increased significantly (CD3+, CD3+/8+ and CD3+/4+).**
- No significant change to CD4/8 ratio.
- LDH, HSST TNT, and Fibronegin remained high throughout, though range observed decreased over time.
- **Consolidation and mixed GGO observed in longitudinal CT imaging.**
- **Patients showed different extents of inflammatory exudation in lungs, with overall tendency for improvement (except 2/7 patients that were readmitted after discharge with re-positive test).**

Patients Readmitted with PCR+ test

- **Seven patients repeated positive RT-PCR test and were readmitted to the hospital (9 to 17 day after initial discharge).**
- Improvement during readmission in 4 patients and observation of segmental progression CT in 2 patients (2/18 or 11.1% - re-positive 9 and 10 days post-discharge).
- Two patients showed new GGO, while others improved greatly.
- **Follow up CT necessary to monitor improvement during recovery and patients with lesion progression should be given more attention.**
- **Dynamic CT in addition to negative test essential in clinical diagnosis due to nasal swab PCR sampling bias (false-negatives).**
- **Increase in CRP occurred in 2 readmitted patients (and decr. in lymphocytes in one patient), but was not correlated with new lesions or disease progression vs. improvement (very low N).**

14.73.6 Significance

Study tracked key clinical features associated with disease progression, recovery, and determinants of clinical diagnosis/management of COVID-19 patients.

14.73.7 Credit

This review was undertaken by Natalie Vaninov as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.74 An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 and multiple 2 endemic, epidemic and bat coronavirus

Sheahan et al. *bioRxiv.* ([2253](#))

14.74.1 Keywords

- Treatment
- Antiviral
- Broad spectrum antiviral
- ribonucleoside analog β -D-N4 30 hydroxycytidine (NHC)
- Remdesivir

14.74.2 Main Findings

β -D-N4 30 -hydroxycytidine (NHC, EIDD-1931) is an orally bioavailable ribonucleoside with antiviral activity against various RNA viruses including Ebola, Influenza and CoV. NHC activity introduced *ds* mutations in the viral (but not cellular) RNA in a dose dependent *manner* that directly correlated with a decrease in viral titers. Authors show that NHC inhibited multiple genetically distinct Bat-CoV viruses in human primary epithelial cells *without affecting cell viability even at high concentrations (100 μ M)*. Prophylactic oral administration of NHC in C57BL/6 mice reduce lung titers of SARS-CoV and prevented weight loss and hemorrhage. Therapeutic administration of NHC in C57BL/6 mice 12 hours post infected with SARS-CoV reduced acute lung injury, viral titer, and lung hemorrhage. The degree of clinical benefit was dependent on the time of treatment initiation post infection. The authors also demonstrate that NHC reduces MERS-CoV infection titers, pathogenesis, and viral RNA in prophylactic and therapeutic settings.

14.74.3 Limitations

Most of the experiments were conducted using MERS-CoV, and SARS-CoV and a few experiments were conducted using other strains of CoV as opposed to SARS-CoV-2. The authors note the core residues that make up the RNA interaction sites (which constitutes the NHC interaction sites) are highly conserved among CoV and because of this conservation their understanding is that NHC can inhibit a broad-spectrum of CoV including SARS-CoV-2.

The increased viral mutation rates associated with NHC activity may have adverse effects if mutations cause the virus to become drug resistant, more infectious or speed-up immune evasion. *In addition, the temporal diminishing effectiveness of NHC on clinical outcome when NHC was used therapeutically is concerning. However, the longer window (7-10 days) for clinical disease onset in human patients from the time of infection compared to that of mice (24-48 hours), may associate with increased NHC effectiveness in the clinic.*

14.74.4 Significance

Prophylactic or therapeutic oral administration of NHC reduces lung titers and prevents acute lung failure in C57BL/6 mice infected with CoV. Given its *broad-spectrum antiviral activity, NHC could turn out to be a useful drug for treating current, emerging and future corona virus outbreaks.* ##### Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.75 Identification of antiviral drug candidates against SARS-CoV-2 from FDA-approved drugs

Sangeun Jeon et al. ([2254](#))

14.75.1 Keywords

- COVID-19
- SARS CoV-2
- antiviral drugs
- niclosamide
- ciclesonide

14.75.2 Main Findings

A panel of ~3,000 FDA- and IND-approved antiviral drugs were previously screened for inhibitory efficacy against SARS CoV, a coronavirus related to the novel coronavirus SARS CoV-2 (79.5%) homology. 35 of these drugs along with another 15 (suggested by infectious disease specialists) were tested in vitro for their ability to inhibit SARS CoV-2 infectivity of Vero cells while preserving cell viability. The infected cells were scored by immunofluorescence analysis using an antibody against the N protein of SARS CoV-2. Chloroquine, lopinavir and remdesivir were used as reference drugs.

Twenty four out of 50 drugs exhibited antiviral activity with IC₅₀ values ranging from 0.1-10 μ M. Among these, two stood out: 1) the anti-helminthic drug niclosamide which exhibited potent antiviral activity against SARS CoV-2 (IC₅₀=0.28 μ M). The broad-spectrum antiviral effect of niclosamide against SARS and MERS-CoV have been previously documented and recent evidence suggests that it may inhibit autophagy and reduce MERS CoV replication. 2) Ciclesonide, a corticosteroid used to treat asthma and allergic rhinitis, also exhibited antiviral efficacy but with a lower IC₅₀ (4.33 μ M) compared to niclosamide. The antiviral effects of ciclesonide were directed against NSP15, a viral ribonuclease which is the molecular target of this drug.

14.75.3 Limitations

The drugs were tested against SARS CoV-2 infectivity in vitro only, therefore preclinical studies in animals and clinical trials in patients will be need for validation of these drugs as therapeutic agents for COVID-19. In addition, niclosamide exhibits low adsorption pharmatokinetically which could be alleviated with further development of drug formulation to increase effective delivery of this drug to target tissues. Nonetheless, niclosamide and ciclesonide represent promising therapeutic agents against SARS CoV-2 given that other compounds tested in the same study including favipiravir (currently used in clinical trials) and atazanavir (predicted as the most potent antiviral drug by AI-inference modeling) did not exhibit antiviral activity in the current study.

14.75.4 Credit

This review was undertaken by K Alexandropoulos as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.76 Respiratory disease and virus shedding in rhesus macaques inoculated with SARS-CoV-2

Munster et al. *bioRxiv.* ([2255](#))

14.76.1 Keywords

animal model, pulmonary infiltrates, dynamic of antibody response, cytokine

14.76.2 Main Findings

Inoculation of 8 Resus macaques with SARS-Cov-2 , which all showed clinical signs of infection (respiratory pattern, reduced appetite, weight loss, elevated body temperature) resulting in moderate, transient disease. Four animals were euthanized at 3dpi, the 4 others at 21 dpi. Study of viral loads in different organs showed that nose swab and throat swabs were the most sensitive, with bronchio-alveolar lavage. Interstitial pneumonia was visible in radiographies and at the histological scale too. Clinically, the macaques had similar symptoms as described in human patients with moderate disease.

Viral shedding was consistently detected in nose swabs and throat swabs immediately after infection but less consistent thereafter which could reflect virus administration route (intranasal, oral). Bronchoalveolar lavages performed as a measure of virus replication in the lower respiratory tract on animals maintained for 21 days, contained high viral loads in 1 and 3dpi. The majority of the animals exhibited pulmonary edema and mild to moderate interstitial pneumonia on terminal bronchioles. In addition to the lung, viral RNA could also be detected throughout the respiratory track where viral replication mainly occurred.

Immunologic responses included leukocytosis, neutrophilia, monocytosis and lymphopenia in the majority of the animals at 1dpi. Lymphocytes and monocytes re-normalized at 2dpi. Neutrophils declined after 3dpi and

through 10dpi after which they started to recover. After infection, serum analysis revealed significant increases in **IL1ra, IL6, IL10, IL15, MCP-1, MIP-1b, but quick normalization** (3dpi). **Antibody response started around 7dpi, and the antibody titers stayed elevated until 21dpi** (day of animal euthanasia).

14.76.3 Limitations

The macaques were inoculated via a combination of intratracheal, intranasal, ocular and oral routes, which might not reproduce how humans get infected. Maybe this can lead to different dynamics in the host immune response. Also, the authors noted that the seroconversion was not directly followed by a decline in viral loads, as observed in covid19 patients.

14.76.4 Significance

This work confirms that rhesus macaques can be a good model to study Covid-19, as it has been shown by other groups ([2229](#), [2243](#), [2256](#)). While these experiments recapitulate moderate COVID-19 in humans, the mode of inoculation via a combination of intratracheal, intranasal, ocular and oral routes, might not reproduce how humans get infected and may lead to different dynamics in the host immune response. For example, the authors noted that the seroconversion was not directly followed by a decline in viral loads, as observed in COVID-19 patients. Therefore, it will be interesting to follow their antibody titers longer and further assess the possibility/effect of reinfection in these macaques. It is essential to be able to understand the dynamic of the disease and associated immune responses, and to work on vaccine development and antiviral drug testing.

14.76.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.77 ACE2 Expression is Increased in the Lungs of Patients with Comorbidities Associated with Severe COVID-19

([2257](#))

14.77.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- lung
- comorbidities
- histone
- epigenetics

14.77.2 Main Findings

- Transcriptomic analysis using systems-level meta-analysis and network analysis of existing literature to determine ACE2 regulation in patients who have frequent COVID-19 comorbidities [eg- cardiovascular diseases, familial pulmonary hypertension, cancer].
- Enrichment analyses indicated pathways associated with inflammation, metabolism, macrophage autophagy, and ER stress.
- ACE2 higher in adenocarcinoma compared to adjacent normal lung; ACE2 higher in COPD patients compared to normal.
- Co-expression analysis identified genes important to viral entry such as RAB1A, ADAM10, HMGBs, and TLR3 to be associated with ACE2 in diseased lungs.
- ACE2 expression could be potentially regulated by enzymes that modify histones, including HAT1, HDAC2, and KDM5B.

14.77.3 Limitations:

- Not actual CoVID-19 patients with co-morbidities, so interpretations in this study need to be confirmed by analyzing upcoming transcriptomics from CoVID-19 patients having co-morbidity metadata.
- As mentioned by authors, study does not look at diabetes and autoimmunity as risk factors in CoVID-19 patients due to lack of data; would be useful to extend such analyses to those datasets when available.
- Co-expression analysis is perfunctory and needs validation-experiments especially in CoVID-19 lung samples to mean anything.
- Epigenomic analyses are intriguing but incomplete, as existence of histone marks does not necessarily mean occupancy. Would be pertinent to check cell-line data (CCLE) or actual CoVID-19 patient samples to confirm ACE2 epigenetic control.

14.77.4 Significance

- Study implies vulnerable populations have ACE2 upregulation that could promote CoVID-19 severity. Shows important data-mining strategy to find gene-networks associated with ACE2 upregulation in co-morbid patients.
- Several of the genes co-upregulated with ACE2 in diseased lung might play an important role in CoVID-19 and can be preliminary targets for therapeutics
- If in silico findings hold true, epigenetic control of ACE2 expression could be a new target for CoVID-19 therapy with strategies such as KDM5 demethylases.

14.77.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.78 Meplazumab treats COVID-19 pneumonia: an open-labelled, concurrent controlled add-on clinical trial

Bian et al. *medRxiv*. ([2258](#))

14.78.1 Keywords

- Meplazumab
- CD147
- humanized antibody
- clinical trial

14.78.2 Main Findings

This work is based on previous work by the same group that demonstrated that SARS-CoV-2 can also enter host cells via CD147 (also called Basigin, part of the immunoglobulin superfamily, is expressed by many cell types) consistent with their previous work with SARS-CoV-1. ¹ A prospective clinical trial was conducted with 17 patients receiving Meplazumab, a humanized anti-CD147 antibody, in addition to all other treatments. 11 patients were included as a control group (non-randomized).

They observed a faster overall improvement rate in the Meplazumab group (e.g. at day 14 47% vs 17% improvement rate) compared to the control patients. Also, virological clearance was more rapid with median of 3 days in the Meplazumab group vs 13 days in control group. In laboratory values, a faster normalization of lymphocyte counts in the Meplazumab group was observed, but no clear difference was observed for CRP levels.

14.78.3 Limitations

While the results from the study are encouraging, this study was non-randomized, open-label and on a small number of patients, all from the same hospital. It offers evidence to perform a larger scale study. Selection bias as well as differences between treatment groups (e.g. age 51yo vs 64yo) may have contributed to results. The authors mention that there was no toxic effect to Meplazumab injection but more patient and longer-term studies are necessary to assess this.

14.78.4 Significance

These results seem promising as for now there are limited treatments for Covid-19 patients, but a larger cohort of patient is needed. CD147 has already been described to facilitate HIV ([2259](#)), measles virus ([2260](#)), and malaria ([2261](#)) entry into host cells. This group was the first to describe the CD147-spike route of SARS-CoV-2 entry in host cells ([2232](#)) p147. Indeed, they had previously shown in 2005 that SARS-CoV could enter host cells via this transmembrane protein ([2262](#)). Further biological understanding of how SARS-CoV-2 can enter host cells and how this integrates with ACE2R route of entry is needed. Also, the specific cellular targets of the anti-CD147 antibody need to be assessed, as this protein can be expressed by many cell types and has been shown to involved in leukocytes aggregation ([2263](#)). Lastly, Meplazumab is not a commercially-available drug and requires significant health resources to generate and administer which might prevent rapid development and use.

14.78.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.79 Potent human neutralizing antibodies elicited 1 by SARS-CoV-2 infection

Ju et al. *bioRxiv.* ([186](#))

14.79.1 Keywords

- monoclonal antibodies
- neutralization
- antibody cross-reactivity
- Receptor Binding Domain

14.79.2 Main Findings

In this study the authors report the affinity, cross reactivity (with SARS-CoV and MERS-CoV virus) and viral neutralization capacity of 206 monoclonal antibodies engineered from isolated IgG memory B cells of patients suffering from SARS-CoV-2 infection in Wuhan, China. All patients but one recovered from disease. Interestingly, the patient that did not recover had less SARS-CoV-2 specific B cells circulating compared to other patients.

Plasma from all patients reacted to trimeric Spike proteins from SARS-CoV-2, SARS-CoV and MERS-CoV but no HIV BG505 trimer. Furthermore, plasma from patients recognized the receptor binding domain (RBD) from SARS-CoV-2 but had little to no cross-reactivity against the RBD of related viruses SARS-CoV and MERS-CoV, suggesting significant differences between the RBDs of the different viruses. Negligible levels of cross-neutralization using pseudoviruses bearing Spike proteins of SARS-CoV-2, SARS-CoV or MERS-CoV, were observed, corroborating the ELISA cross-reactivity assays on the RBDs.

SARS-CoV-2 RBD specific B cells constituted 0.005-0.065% of the total B cell population and 0.023-0.329% of the memory subpopulation. SARS-CoV specific IgG memory B cells were single cell sorted to sequence the antibody genes that were subsequently expressed as recombinant IgG1 antibodies. From this library, 206 antibodies with different binding capacities were obtained. No discernible patterns of VH usage were found in the 206 antibodies suggesting immunologically distinct responses to the infection. Nevertheless, most high-binding antibodies were derived by clonal expansion. Further analyses in one of the patient derived clones, showed that the antibodies from three different timepoints did not group together in phylogenetic analysis, suggesting selection during early infection.

Using surface plasmon resonance (SPR) 13 antibodies were found to have 10^{-8} to 10^{-9} dissociation constants (Kd). Of the 13 antibodies, two showed 98-99% blocking of SARS-CoV-2 RBD-ACE2 receptor binding in competition assays. Thus, low Kd values alone did not predict ACE2 competing capacities. Consistent with competition assays the two antibodies that show high ACE2 blocking (P2C-2F6 and P2C-1F11) were the most capable of neutralizing pseudoviruses bearing SARS-CoV-2 spike protein (IC₅₀ of 0.06 and 0.03 µg/mL, respectively). Finally, using SPR the neutralizing antibodies were found to recognize both overlapping and distinct epitopes of the RBD of SARS-CoV-2.

14.79.3 Limitations

1. Relatively low number of patients
 - a. No significant conclusion can be drawn about the possible > correlation between humoral response and disease severity
2. *In vitro* Cytopathic Effect Assay (CPE) for neutralization activity
 - a. Huh7 cells were used in neutralization assays with > pseudoviruses, and they may not entirely mimic what happens in > the upper respiratory tract
 - b. CPE assay is not quantitative
3. Duplicated panel in Figure 4C reported (has been fixed in version 2)

14.79.4 Significance

This paper offers an explanation as to why previously isolated antibodies against SARS-CoV do not effectively block SARS-CoV-2. Also, it offers important insight into the development of humoral responses at various time points during the first weeks of the disease in small but clinically diverse group of patients. Furthermore, it provides valuable information and well characterized antibody candidates for the development of a recombinant antibody treatment for SARS-CoV-2. Nevertheless, it also shows that plasmapheresis might have variability in its effectiveness, depending on the donor's antibody repertoire at the time of donation.

14.79.5 Credit

Review by Jovani Catalan-Dibene as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.80 Characterisation of the transcriptome and proteome of SARS-CoV-2 using direct RNA sequencing and tandem mass spectrometry reveals evidence for a cell passage induced in-frame deletion in the spike glycoprotein that removes the furin-like cleavage site.

Davidson et al. ([2264](#))

14.80.1 Keywords

- Transcription
- RNA-seq
- proteomics
- mass spec
- furin cleavage site
- mutation
- pathogenicity

14.80.2 Main Findings

The authors performed long read RNA sequencing using an Oxford Nanopore MinION as well as tandem mass spec (MS) on Vero cells (a cell line derived from kidney cells of the African green monkey that is deficient in interferon) infected with SARS-CoV-2.

The authors found that passage of the virus in Vero cells gave rise to a spontaneous 9 amino acid deletion (679-NSPRRARSV-687 to I) in the spike (S) protein. The deleted sequence overlaps a predicted furin cleavage site at the S1 / S2 domain boundary that is present in SARS-CoV-2 but not SARS-CoV or the closely related bat coronavirus RaTG13, which are cleaved at S1 / S2 by other proteases ([16](#)). Furin cleavage sites at similar positions in other viruses have been linked to increased pathogenicity and greater cell tropism ([2265](#)). Loss of this site in SARS-CoV-2 has also already been shown to increase viral entry into Vero but not BHK cells (which are also interferon deficient) ([25](#)). The authors therefore make an important contribution in demonstrating that passage in Vero cells may lead to spontaneous loss of a key pathogenicity-conferring element in SARS-CoV-2.

14.80.3 Limitations

As the authors note, a similar study posted earlier by Kim et al., which also passaged SARS-CoV-2 in Vero cells, did not identify any loss in the S protein furin cleavage site ([2266](#)). It therefore remains to be determined how likely it is that this mutation spontaneously arises. A quantitative investigation using multiple experimental replicas to understand the spontaneous viral mutation rate at this site and elsewhere would be informative. Also, the mechanistic basis for the higher viral fitness conferred by loss of the furin cleavage site in Vero cells – but, evidently, not in vivo in humans, as this site is maintained in all currently sequenced circulating isolates – remains to be understood.

Due to the high base-call error rate of MinION sequencing, the authors' bioinformatic pipeline required aligning transcripts to a reference to correct sequencing artifacts. This presumably made it difficult or impossible to identify other kinds of mutations, such as single nucleotide substitutions, which may occur even more frequently than the deletions identified in this work. Pairing long read sequencing with higher-accuracy short-read sequencing may be one approach to overcome this issue.

14.80.4 Significance

As the authors suggest, animal studies using live virus challenge may need to periodically verify the genomic integrity of the virus, or potentially risk unknowingly using a likely less-pathogenic variant of the virus.

More broadly, the results emphasize the complexity and plasticity of the SARS-CoV-2 viral transcriptome and proteome. For example, the authors found multiple versions of transcripts encoding the nucleocapsid (N) protein, each with different small internal deletions, some of which were verified for translation by MS. A number of peptides arising from translation of unexpected rearrangements of transcripts were also detected. Additionally, the authors identified phosphorylation of a number of viral proteins (N, M, ORF 3a, nsp3, nsp9, nsp12 and S). For any cases where these have functional consequences, targeting the kinases responsible could be an avenue for drug development. Understanding the functional consequences of the mutations, transcript variations, and post translational modifications identified in this study will be important future work.

14.80.5 Credit

This review was undertaken by Tim O'Donnell, Maria Kuksin as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.81 A SARS-CoV-2-Human Protein-Protein Interaction Map Reveals Drug Targets and Potential Drug- Repurposing

Gordon et al. *bioRxiv* ([2267](#))

14.81.1 Keywords

- protein-protein interactions
- mass spectrometry
- drug targets

14.81.2 Main Findings

Gordon et al cloned, tagged and expressed 26 of the 29 SARS-CoV-2 proteins individually in HEK293T cells and used mass spectrometry to identify protein-protein interactions. They identified 332 viral-host protein-protein interactions. Furthermore, they used these interactions to identify 66 existing drugs known to target host proteins or host pathways (eg SARS-CoV-2 N and Orf8 proteins interact with proteins regulated by the mTOR pathway, so mTOR inhibitors Silmitasertib and Rapamycin are possible drug candidates).

14.81.3 Limitations

The main limitation of the study stems from the reductionist model: overexpression of plasmids encoding individual viral proteins in HEK293T cells. This precludes any interactions between the viral proteins, or the combined effects of multiple proteins on the host, as they are expressed individually. Moreover, HEK293T cells come from primary embryonic kidney and therefore might not reflect how SARS-CoV-2 interacts with its primary target, the lung. However, the authors found that the proteins found to interact with viral proteins in their experiments are enriched in lung tissue compared to HEK293Ts.

14.81.4 Significance

The authors provide a “SARS-CoV-2 interaction map,” which may provide potential hypotheses as to how the virus interacts with the host. Further, they identified existing drugs that could disrupt these host-viral interactions and curb SARS-CoV-2 infection. Although these interactions have not been validated, this paper acts as a valuable resource.

14.81.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.82 First Clinical Study Using HCV Protease Inhibitor Danoprevir to Treat Naïve and Experienced COVID-19 Patients

Chen et al. *medRxiv*. ([2268](#))

14.82.1 Keywords

- Clinical study
- HCV protease inhibitor

- Danoprevir
- Ritonavir
- Covid19 treatment

14.82.2 Main Findings

The authors treated 11 Covid-19 patients with Danoprevir, a commercialized HCV protease inhibitor ([2269](#))^(p4), boosted by ritonavir ([2270](#)), a CYP3A4 inhibitor (which enhances the plasma concentration and bioavailability of Danoprevir). Two patients had never received anti-viral therapy before (=naïve), whereas nine patients were on Lopinavir/Ritonavir treatment before switching to Danoprevir/Ritonavir (=experienced). The age ranged from 18 to 66yo.

Naïve patients that received Danoprevir/Ritonavir treatment had a decreased hospitalization time. Patients treated with Lopinavir/Ritonavir did not have a negative PCR test, while after switching to Danoprevir/Ritonavir treatment, the first negative PCR test occurred at a median of two days.

14.82.3 Limitations

The results of the study are very hard to interpret as there is no control group not receiving Danoprevir/Ritonavir treatment. This was especially true in naïve patients who seemed to have more mild symptoms before the start of the study and were younger (18 and 44yo) compared to the experienced patients (18 to 66yo). The possibility that the patients would have recovered without Danoprevir/Ritonavir treatment cannot be excluded.

14.82.4 Significance

The authors of the study treated patients with Danoprevir, with the rationale that this is an approved and well tolerated drug for HCV patients ([2270](#)), and that it could also target the protease from SARS-CoV-2 (essential for viral replication and transcription). Indeed, homology modelling data indicated that HCV protease inhibitors have the highest binding affinity to Sars-Cov2 protease among other approved drugs ([2271](#)).

While this study shows that the combination of Danoprevir and Ritonavir might be beneficial for Covid-19 patients, additional clinical trials with more patients and with better methodology (randomization and control group) are needed to make further conclusions.

14.82.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.83 Efficacy of hydroxychloroquine in patients with COVID-19: results of a randomized clinical trial

(1043)

14.83.1 Keywords

- hydroxychloroquine

14.83.2 Study Description

This is a randomized clinical trial of hydroxychloroquine (HCQ) efficacy in the treatment of COVID-19. From February 4 – February 28, 2020 142 COVID-19 positive patients were admitted to Renmin Hospital of Wuhan University. 62 patients met inclusion criteria and were enrolled in a double blind, randomized control trial, with 31 patients in each arm.

Inclusion criteria:

1. Age ≥ 18 years
2. Positive diagnosis COVID-19 by detection of SARS-CoV-2 by RT-PCR
3. Diagnosis of pneumonia on chest CT
4. Mild respiratory illness, defined by $\text{SaO}_2/\text{SPO}_2$ ratio $> 93\%$ or $\text{PaO}_2/\text{FIO}_2$ ratio > 300 mmHg in hospital room conditions (Note: relevant clinical references described below.)
 - a. Hypoxia is defined as an SpO_2 of 85-94%; severe hypoxia $< 85\%$.
 - b. The $\text{PaO}_2/\text{FIO}_2$ (ratio of arterial oxygen tension to fraction of inspired oxygen) is used to classify the severity of acute respiratory distress syndrome (ARDS). Mild ARDS has a $\text{PaO}_2/\text{FIO}_2$ of 200-300 mmHg, moderate is 100-200, and severe < 100 .
5. Willing to receive a random assignment to any designated treatment group; not participating in another study at the same time

Exclusion criteria:

1. Severe or critical respiratory illness (not explicitly defined, presumed to be respiratory function worse than outlined in inclusion criteria); or participation in trial does not meet patient's maximum benefit or safe follow up criteria
2. Retinopathy or other retinal diseases
3. Conduction block or other arrhythmias

4. Severe liver disease, defined by Child-Pugh score $\geq C$ or AST $>$ twice the upper limit
5. Pregnant or breastfeeding
6. Severe renal failure, defined by $\text{eGFR} \leq 30 \text{ mL/min/1.73m}^2$, or on dialysis
7. Potential transfer to another hospital within 72h of enrollment
8. Received any trial treatment for COVID-19 within 30 days before the current study

All patients received the standard of care: oxygen therapy, antiviral agents, antibacterial agents, and immunoglobulin, with or without corticosteroids. Patients in the HCQ treatment group received additional oral HCQ 400 mg/day, given as 200 mg 2x/day. HCQ was administered from days 1-5 of the trial. The primary endpoint was 5 days post enrollment or a severe adverse reaction to HCQ. The primary outcome evaluated was time to clinical recovery (TTCR), defined as return to normal body temperature and cough cessation for $> 72\text{h}$. Chest CT were imaged on days 0 and 6 of the trial for both groups; body temperature and patient reports of cough were collected 3x/day from day 0 – 6. The mean age and sex distribution between the HCQ and control arms were comparable.

14.83.3 Main Findings

There were 2 patients showing mild secondary effects of HCQ treatment. More importantly, while 4 patients in the control group progressed to severe disease, none progressed in the treatment group.

TTCR was significantly decreased in the HCQ treatment arm; recovery from fever was shortened by one day (3.2 days control vs. 2.2 days HCQ, $p = 0.0008$); time to cessation of cough was similarly reduced (3.1 days control vs. 2.0 days HCQ, $p = 0.0016$).

Overall, it appears that HCQ treatment of patients with mild COVID-19 has a modest effect on clinical recovery (symptom relief on average 1 day earlier) but may be more potent in reducing the progression from mild to severe disease.

14.83.4 Limitations

This study is limited in its inclusion of only patients with mild disease, and exclusion of those on any treatment other than the standard of care. It would also have been important to include the laboratory values of positive RT-PCR detection of SARS-CoV-2 to compare the baseline and evolution of the patients' viral load.

14.83.5 Limitations

Despite its limitations, the study design has good rigor as a double blind RCT and consistent symptom checks on each day of the trial. Now that the FDA has approved HCQ for treatment of COVID-19 in the USA, this study supports the efficacy of HCQ use early in treatment of patients showing mild symptoms, to improve time to clinical recovery, and possibly reduce disease progression. However, most of the current applications of HCQ have been in patients with severe disease and for compassionate use, which are out of the scope of the findings presented in this trial. Several additional clinical trials to examine [hydroxychloroquine](#) are now undergoing; their results will be critical to further validate these findings.

14.83.6 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

Structure-based modeling of SARS-CoV-2 peptide/HLA-A02 antigens

<https://doi.org/10.1101/2020.03.23.004176>

Immunology keywords:

CoVID-19, 2019-nCoV, SARS-CoV-2, comparative, homology, peptide, modeling, simulation, HLA-A, antigen

Summary of Findings:

- The authors utilize homology modeling to identify peptides from the SARS-CoV-2 proteome that potentially bind HLA-A*02:01.
- They utilize high-resolution X-ray structures of peptide/MHC complexes on Protein Data Bank, substitute homologous peptides with SARS-CoV-2 peptides, and calculate MHC/SARS-CoV-2 peptide Rosetta binding energy.
- They select MHC/SARS-CoV-2 complex models with highest binding energy for further study and publish models in an online database (<https://rosettamhc.chemistry.ucsc.edu>).

Limitations:

- The authors only utilize computational methods and predicted SARS-CoV-2 peptides must be validated experimentally for immunogenicity and clinical response.
- Due to computational burden and limited availability of high resolution X-ray structures on PDB, authors only simulate 9-mer and 10-mer peptide binding to HLA-A*02:01.
- Since the authors compare select existing X-ray structures as a starting point, backbone conformations that deviate significantly between test and template peptides are not captured. Furthermore, Rosetta modeling

protocols do not capture all possible structures and binding energy scoring does not fully recapitulate fundamental forces.^{1,2}

Importance/Relevance:

- The authors identify and publish high-scoring SARS-CoV-2 peptides that may direct a targeted, experimental validation approach toward a COVID-19 vaccine.
- The authors utilize Rosetta simulation to further filter results from NetMHCpan 4.0, supporting machine learning prediction with structural analysis.
- The authors develop RosettaMHC, a computationally efficient method of leveraging existing X-ray structures for identification of immunogenic peptides.

References:

1. Bender, B. J., Cisneros, A., 3rd, Duran, A. M., Finn, J. A., Fu, D., Lokits, A. D., . . . Moretti, R. (2016). Protocols for Molecular Modeling with Rosetta3 and RosettaScripts. *Biochemistry*, 55(34), 4748-4763. doi:10.1021/acs.biochem.6b00444
2. Alford, R. F., Leaver-Fay, A., Jeliazkov, J. R., O'Meara, M. J., DiMaio, F. P., Park, H., . . . Gray, J. J. (2017). The Rosetta All-Atom Energy Function for Macromolecular Modeling and Design. *J Chem Theory Comput*, 13(6), 3031-3048. doi:10.1021/acs.jctc.7b00125

Review by Jonathan Chung as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn school of medicine, Mount Sinai.

14.84 Serology characteristics of SARS-CoV-2 infection since the exposure and post symptoms onset

Lou et al. *medRxiv*. ([2272](#))

14.84.1 Keywords

- Seroconversion rate
- Total Antibody
- Ab
- IgG and IgM
- antibody

14.84.2 Main Findings

Currently, the diagnosis of SARS-CoV-2 infection entirely depends on the detection of viral RNA using polymerase chain reaction (PCR) assays. False negative results are common, particularly when the samples are collected from upper respiratory. Serological detection may be useful as an additional testing strategy. In this study the authors reported that a typical acute antibody response was induced during the SARS-CoV-2 infection, which was discussed earlier¹. The seroconversion rate for Ab, IgM and IgG in COVID-19 patients was 98.8% (79/80), 93.8% (75/80) and 93.8% (75/80), respectively. The first detectable serology marker was total antibody followed by IgM and IgG, with a median seroconversion time of 15, 18 and 20 days-post exposure (d.p.e) or 9, 10- and 12-days post-onset (d.p.o). Seroconversion was first detected at day 7d.p.e in 98.9% of the patients. Interestingly they found that viral load declined as antibody levels increased. This was in contrast to a previous study ([2212](#)), showing that increased antibody titers did not always correlate with RNA clearance (low number of patient sample).

14.84.3 Limitations

Current knowledge of the antibody response to SAR-CoV-2 infection and its mechanism is not yet well elucidated. Similar to the RNA test, the absence of antibody titers in the early stage of illness could not exclude the possibility of infection. A diagnostic test, which is the aim of the authors, would not be useful at the early time points of infection but it could be used to screen asymptomatic patients or patients with mild disease at later times after exposure.

14.84.4 Significance

Understanding the antibody responses against SARS-CoV2 is useful in the development of a serological test for the diagnosis of COVID-19. This manuscript discussed acute antibody responses which can be deducted in plasma for diagnostic as well as prognostic purposes. Thus, patient-derived plasma with known antibody titers may be used therapeutically for treating COVID-19 patients with severe illness.

14.84.5 Credit

This review was undertaken and edited by Konstantina A as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.85 SARS-CoV-2 launches a unique transcriptional signature from in vitro, ex vivo, and in vivo systems

Blanco-Melo et al. *bioRxiv.* ([2273](#))

14.85.1 Keywords

- host cellular response

- host-pathogen interaction
- type I interferon
- type III interferon
- inflammation
- RNA-seq
- comparative analysis

14.85.2 Main Findings

Given the high mortality rate of SARS-CoV-2 relative to other respiratory viruses such as seasonal IAV and RSV, there may be underlying host-pathogen interactions specific to SARS-CoV-2 that predispose to a worse clinical outcome. Using *in vivo*, *ex vivo*, and *in vitro* systems, the authors profiled host cell transcriptional responses to SARS-CoV-2 and to other common respiratory viruses (seasonal IAV and RSV). SARS-CoV-2 infection *in vitro* led to an induction of type I interferon response signaling and the upregulation of cytokine/chemokine transcripts. In comparison with IAV and RSV infection, SARS-CoV-2 *in vitro* appears to uniquely induce less type I and type III interferon expression and higher levels of two cytokines previously implicated in respiratory inflammation. Lastly, *in vivo* data from ferrets showed a reduced induction of cytokines and chemokines by SARS-CoV-2 infection relative to IAV infection.

14.85.3 Limitations

While these results are promising, there are several key weaknesses of this paper. 1) As the authors point out, there is an undetectable level of SARS-CoV-2 putative receptor (ACE2) and protease (TMPRSS2) expression in the lung epithelial cell line used for the *in vitro* studies. This raises the important question of whether viral replication actually occurs in any of the models used, which may explain the lack of interferon production observed *in vitro* in SARS-CoV-2 treated cells. Further studies characterizing viral titers across timepoints are needed. 2) Furthermore, these studies only characterize the host response at a single dose and timepoint per virus, and it is unclear why these doses/timepoints were chosen. This leaves open the possibility that the observed differences between viruses could be due to differences in dose, timing, host response, or a combination of all of these. 3) It is unclear whether ferrets are productively infected, which cell types are infected, and the extent/timing of the clinical course of infection. Moreover, the *in vitro* and *in vivo* data do not strongly correlate and the reasons for this are unclear.

14.85.4 Significance

This paper describes potentially unique transcriptional signatures of host cells exposed to SARS-CoV-2. If validated, these findings may help explain clinical outcomes and could be targeted in future therapeutic interventions.

14.85.5 Potential Conflicts of Interest Disclosure

The reviewers are also researchers at the Icahn School of Medicine at Mount Sinai.

14.85.6 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.86 A New Predictor of Disease Severity in Patients with COVID-19 in Wuhan, China

Zhou et al. *bioRxiv.* ([2274](#))

14.86.1 Keywords

- disease severity
- clinical data
- Neutrophils/lymphocytes ratio
- CRP
- D-dimer

14.86.2 Main Findings

377 hospitalized patients were divided into two groups: severe and non-severe pneumonia. The laboratory results of their first day of admission were retrospectively analyzed to identify predictors of disease severity.

After adjusting for confounding factors from chronic comorbidities (such as high blood pressure, type 2 diabetes, coronary heart disease, and chronic obstructive pulmonary disease), the independent risk factors identified for severe pneumonia were **age**, the **ratio of neutrophil/lymphocytes counts**, **CRP** and **D-dimer** levels.

To further increase the specificity and sensibility of these markers, they showed that their multiplication **[(Neutrophil/lymphocyte count) * CRP * D-dimer]** was a better predictor of disease severity, with higher sensitivity (95.7%) and specificity (63.3%), with a cutoff value of 2.68.

14.86.3 Limitations

This study included 377 hospitalized patients. Among them, 45.6% patients tested positive for SARS-Cov-2 nucleic acid test results, and others were included in the study based on clinically diagnosis even if the molecular diagnosis was negative. Thus, additional studies are needed to verify this on a larger number of covid-19 certified patients and the cutoff value might be adjusted. Also, all the patients that did not have the clinical characteristics of severe pneumonia were included in the non-severe pneumonia group, but usually patients are also divided into moderate and mild disease.

Also, studying different subset of lymphocytes could lead to a more specific predictor. Another study showed that the neutrophils to CD8+ T cells ratio was a strong predictor of disease severity ([2182](#)). Another more precise study showed that the percentage of helper T cells and regulatory T cells decrease

but the percentage of naïve helper T cells increases in severe cases ([2175](#)). Taking these subpopulations into account might make the predictor more powerful.

Other studies also noted an inverse correlation between disease severity and LDH ([2216](#)) or IL6 ([2225](#)) levels, but the authors here do not discuss LDH nor IL6 levels, although this could help to strengthen the predictor.

The study is based on the results obtained on the first day of admission, studying the dynamic of the changes in patients might also be interesting to better predict disease severity.

14.86.4 Significance

This study confirms that the neutrophil to lymphocyte ratio can be a predictor of disease severity as shown by many others ([2174](#), [2175](#), [2188](#)). The novelty here is that they show that a combination with other markers can enhance the specificity and sensibility of the predictor, although the study could be improved by taking into account sub-populations of lymphocytes and more biological factors from patients such as LDH and IL6.

14.86.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.87 Metabolic disturbances and inflammatory dysfunction predict severity of coronavirus disease 2019 (COVID-19): a retrospective study

Shuke Nie et al. *medRxiv.* ([2275](#))

14.87.1 Keywords

- metabolism
- fasting blood glucose
- serum total protein
- albumin
- blood lipid
- HDL-C
- APOA1
- lymphocytopenia
- IL-6
- CRP
- severity prediction of COVID19

14.87.2 Main Findings

Retrospective Study on 97 COVID-19 hospitalized patients (25 severe and 72 non-severe) analyzing clinical and laboratory parameter to predict transition from mild to severe disease based on more accessible indicators (such as fasting blood glucose, serum protein or blood lipid) than inflammatory indicators. In accordance with other studies, age and hypertension were risk factors for disease severity, and lymphopenia and increased IL-6 was observed in severe patients. The authors show that fasting blood glucose (FBG) was altered and patients with severe disease were often hyperglycemic. Data presented support that hypoproteinaemia, hypoalbuminemia, and reduction in high-densitylipoprotein (HDL-C) and ApoA1 were associated with disease severity.

14.87.3 Limitations

In this study non-severe patients were divided in two groups based on average course of the disease: mild group1 (14 days, n=28) and mild group 2 (30 days, n=44). However mild patients with a longer disease course did not show an intermediate phenotype (between mild patients with shorter disease course and severe patients), hence it is unclear whether this was a useful and how it impacted the analysis. Furthermore, the non-exclusion of co-morbidity factors in the analysis may bias the results (e.g. diabetic patients and glucose tests) It is not clear at what point in time the laboratory parameters are sampled. In table 3, it would have been interesting to explore a multivariate multiple regression. The correlation lacks of positive control to assess the specificity of the correlation to the disease vs. correlation in any inflammatory case. The dynamic study assessing the predictability of the laboratory parameter is limited to 2 patients. Hence there are several associations with disease severity, but larger studies are necessary to test the independent predictive value of these potential biomarkers.

14.87.4 Significance

As hospital are getting overwhelmed a set of easily accessible laboratory indicators (such as serum total protein) would potentially provide a triage methodology between potentially severe cases and mild ones. This paper also opens the question regarding metabolic deregulation and COVID-19 severity.

14.87.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.88 Viral Kinetics and Antibody Responses in Patients with COVID-19

([2276](#))

14.88.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- IgG
- IgM
- clinical
- kinetics
- antibodies

14.88.2 Main Findings

- Prospective cohort of 67 patients, clinical specimens taken and follow-up conducted.
- Viral shedding, serum IgM, IgG antibody against NP evaluated and correlated to disease severity and clinical outcome
- Viral RNA levels peaked at 1 week from febrile/cough symptom onset in sputum, nasal swabs, and stool samples. Shedding ranged from 12-19 days (median ranges) and was longer in severe patients.
- IgM and IgG titers stratified patients into three archetypes as 'strong vs weak vs non-responders'. Strong responders (with higher IgM/IgG titers) were significantly higher in severe patients.

14.88.3 Limitations

Specific for immune monitoring.

- Not clear if stool RNA captured from live infection in intestine/liver or from swallowed sputum. Transmission electron microscopy (TEM) carried out on sputum samples as proof of concept, but not stools. TEM unreasonable for actual clinical diagnosis.
- Several patients had co-morbidities (such as pulmonary and liver disease) that were not accounted for when tracking antibody responses. Viral kinetics and IgM/IgG titers in subsets of patients with underlying conditions/undergoing certain medication would be informative.

14.88.4 Significance

- Three archetypes of antibody response to SARS-CoV-2 with different disease progression and kinetics is useful to stratify patients, and for future serological tests.
- Strong spike-IgG levels often correlate with lymphopenia and CoVID-19 disease severity ([2277](#)), similar to macaque studies in SARS ([2278](#)). It would be critical to see if anti-NP or anti-Spike IgG antibodies for SARS-CoV-2 also elicit similar detrimental effects before clinical use.

14.88.5 Credit

14.89 COVID-19 infection induces readily detectable morphological and inflammation-related phenotypic changes in peripheral blood monocytes, the severity of which correlate with patient outcome

([2279](#))

14.89.1 Keywords

- Monocytes
- FSC-high
- PBMC
- ACE2
- inflammatory cytokines

14.89.2 Main Findings

This study is based on flow cytometry immunophenotyping of PBMCs from 28 patients diagnosed positive for SARS-Cov2 (COVID19). The authors identify a population of abnormally large (FSC-hi) monocytes, present in COVID19 patients, but absent in PBMCs of healthy volunteers (n=16) or patients with different infections (AIDS, malaria, TB). This FSC-hi monocytic population contains classical, intermediate and non-classical (monocytes (based on CD14 and CD16 expression) that produce inflammatory cytokines (IL-6, TNF and IL-10). The authors suggest an association of FSC-hi monocytes with poor outcome and correlate a high percentage of FSC-low monocytes, or higher ratio of FSC-low/hi monocytes, with faster hospital discharge.

14.89.3 Limitations

While identification of the monocytic population based on FSC is rather robust, the characterization of these cells remains weak. A comprehensive comparison of FSC-hi monocytes with FSC-low monocytes from patients and healthy controls would be of high value. It is unclear if percentages in blood are among CD45+ cells. Furthermore, it would have been important to include absolute numbers of different monocytic populations (in table 1 there are not enough samples and it is unclear what the authors show).

The authors show expression of the ACE2 receptor on the surface of the monocytes, and highlight these cells as potential targets of SARS-Cov2. However, appropriate controls are needed. CD16 has high affinity to rabbit IgG and it is unclear whether the authors considered unspecific binding of rabbit anti-ACE2 to Fc receptors. Gene expression of ACE-2 on monocytes

needs to be assessed. Furthermore, it would be important to confirm infection of monocytes by presence of viral proteins or viral particles by microscopy.

Considering the predictive role of FSC-hi monocytes on the development of the disease and its severity, some data expected at this level are neither present nor addressed. Although the cohort is small, it does include 3 ICU patients. What about their ratio of FSC-low vs FSC-hi monocytes in comparison to other patients? Was this apparent early in the disease course? Does this population of FSC-hi monocytes differ between ICU patients and others in terms of frequency, phenotype or cytokine secretion?

In general, figures need to be revised to make the data clear. For example, in Fig. 5, according to the legend it seems that patients with FSC-high monocytes are discharged faster from the hospital. However according to description in the text, patients were grouped in high or low levels of FSC-low monocytes.

14.89.4 Significance

Despite the limitations of this study, the discovery of a FSC-high monocyte population in COVID-19 patients is of great interest. With similar implication, a recent study by Zhou et al. ([2180](#)) identified a connection between an inflammatory CD14+CD16+ monocyte population and pulmonary immunopathology leading to deleterious clinical manifestations and even acute mortality after SARS-CoV-2 infections. Although the presence of these monocytes in the lungs has yet to be demonstrated, such results support the importance of monocytes in the critical inflammation observed in some COVID19 patients.

14.89.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.90 Correlation between universal BCG vaccination policy and reduced morbidity and mortality for COVID-19: an epidemiological study

Miller et al. *medRxiv*. ([2280](#))

14.90.1 Keywords

- BCG vaccine
- epidemiology
- vaccination policy

14.90.2 Main Findings

The authors compared middle and high income countries that never had a universal BCG vaccination policy (Italy, Lebanon, Nederland, Belgium) and countries with a current policy (low income countries were excluded from the analysis as their number of cases and deaths might be underreported for the moment). **Countries that never implement BCG vaccination have a higher mortality rate than countries which have a BCG vaccination policy** (16.38 deaths per million people vs 0.78). Next, **the authors show that an earlier start of vaccination correlates with a lower number of deaths per million inhabitants**. They interpret this as the vaccine protecting a larger fraction of elderly people, which are usually more affected by COVID-19. Moreover, higher number of COVID-19 **cases** were presented in countries that never implemented a universal BCG vaccination policy.

14.90.3 Limitations

While this study aims to test an intriguing hypothesis unfortunately, the data is not sufficient at this time to accurately make any determinations. Several caveats must be noted including: not all countries are in the same stage of the pandemic, the number of cases/deaths is still changing very rapidly in a lot of countries and thus the association may only reflect exposure to the virus. This analysis would need to be re-evaluated when all the countries are passed the pandemic and more accurate numbers are available. Additionally, very few middle and high-income countries ever implemented universal BCG vaccination, which can be a source of bias (5 countries, vs 55 that have a BCG vaccine policy). Effective screening and social isolation policies also varied considerable across the countries tested and may reflect another important confounder. The authors could consider analyzing the Case Fatality Rate (CFR, % of patients with COVID-19 that die), to more correct for exposure although testing availability will still bias this result. Variability in mortality within countries or cities with variable vaccination and similar exposure could also be appropriate although confounders will still be present.

14.90.4 Significance

BCG vaccine is a live attenuated strain derived from *Mycobacterium bovis* and used for a vaccine for tuberculosis (TB). This vaccine has been proven to be efficient in preventing childhood meningitis TB, but doesn't prevent adult TB as efficiently. For this reason, several countries are now only recommending this vaccine for at-risk population only.

This study shows that there is a correlation between BCG vaccination policy and reduced mortality for Covid-19. Indeed, BCG vaccine has been shown to protect against several viruses and enhance innate immunity ([2281](#)), which could explain why it could protect against SARS-CoV-2 infection, but the exact mechanism is still unknown. **Moreover, the efficiency of adult/older people vaccination and protection against Covid-19 still needs to be assessed.** Regarding this, Australian researchers are starting a clinical trial of BCG vaccine for healthcare workers ([2282](#)), to assess if it can protect them against Covid-19.

14.90.5 Credit

This review was undertaken as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.91 Non-neural expression of SARS-CoV-2 entry genes in the olfactory epithelium

([2283](#))

14.91.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- anosmia
- olfaction
- scRNAseq

14.91.2 Main Findings

- Study analyzed bulk and scRNAseq data of olfactory cell types from publicly-available mouse, nonhuman primate and human datasets.
- show that *ACE2* and *TMPRSS2* (genes involved in SARS-CoV-2 entry) are expressed in olfactory epithelial (OE) cells, basal stem cells and respiratory epithelium (RE), but not sensory neurons.
- Comparison of human RE and OE datasets (Deprez et al. 2019; Durante et al. 2020) revealed that *ACE2* and *TMPRSS2* expression in OE sustentacular cells was similar to expression in the remainder of the non-nasal respiratory tract.

14.91.3 Limitations

- Transcript data alone from healthy respiratory/olfactory cells is not sufficient to confirm infectivity of nasal passage, or to indicate damage to epithelia.
- No mechanism defined for anosmia; it is not clear if epithelial injury leads to reduced sensitivity or increased inflammation and altered immune contexture drives neural/epithelial dysfunction. Will be critical to test this in CoVID-19 patient samples or mouse models.

14.91.4 Significance

- Study provides possible rationale for anosmia observed in several CoVID-19 patients.
- Raises possibility that nasal respiratory goblet, ciliated cells, and olfactory epithelia may serve as a viral reservoir after initial SARS-CoV-2 infection.

- Human olfactory sensory neurons express several other molecules important to CoV (not CoV-19) entry such as *FURIN*, *ST6GAL1*, *ST3GAL4*; this suggests wider mechanism of neuronal infectivity in other coronaviruses.

14.91.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

Title:

SARS-CoV-2 proteome microarray for mapping COVID-19 antibody interactions at amino acid resolution

Immunology keywords: SARS-CoV-2, COVID-19, high throughput, peptide microarray, antibody epitope screening

The main finding of the article:

This study screened the viral protein epitopes recognized by antibodies in the serum of 10 COVID-19 patients using a new SARS-CoV-2 proteome peptide microarray. The peptide library was constructed with 966 linear peptides, each 15 amino acids long with a 5 amino acid overlap, based on the protein sequences encoded by the genome of the Wuhan-Hu-1 strain.

To investigate crossreactivity between SARS-CoV-1 and SARS-CoV-2, they tested rabbit monoclonal and polyclonal antibodies against SARS-CoV-1 nucleocapsid (N) in the microarray. Antibodies against SARS-CoV-1 N displayed binding to the SARS-CoV-2 nucleocapsid (N) peptides. Polyclonal antibodies showed some crossreactivity to other epitopes from membrane (M), spike (S), ORF1ab and ORF8. This suggests that previous exposure to SARS-CoV-1 may induced antibodies recognizing both viruses.

Screening of IgM and IgG antibodies from 10 COVID-19 patients showed that many antibodies targeted peptides on M, N, S, Orf1ab, Orf3a, Orf7a, and Orf8 from SARS-CoV-2, while immunodominant epitopes with antibodies in more than 80 % COVID-19 patients were present in N, S and Orf3. It is shown that the receptor binding domain (RBD) resides on S protein and RBD is important for SARS-CoV-2 to enter the host cells via ACE2. Among six epitopes on S protein, structural analysis predicted that three epitopes were located at the surface and three epitopes were located inside of the protein. Furthermore, some IgM antibodies from 1 patient and IgG antibodies from 2 patients bound to the same epitope (residue 456-460, FRKSN) which resided within the RBD, and structural analysis determined that this epitope was located in the region of the RBD loop that engages with ACE2.

Critical analysis of the study:

In addition to the limitations mentioned in the manuscript, it would have been informative to do the analysis over the course of the disease. The pattern of antibody recognition, especially on S protein, and the course of

antibodies of different isotypes recognizing the same peptide might correlate to the clinical course in these patients. It would also have been informative to analyze the presence of cross-reactive antibodies from patients previously exposed to SARS-CoV-1.

The importance and implications for the current epidemics:

This study identified linear immunodominant epitopes on SARS-CoV-2, Wuhan-Hu-1 strain. This is a valuable information to design vaccines that will elicit desirable immune responses.

The Novel Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Directly Decimates Human Spleens and Lymph Nodes

Review by Matthew D. Park

Revised by Miriam Merad

Keywords: COVID-19, SARS-CoV-2, spleen, lymph node, ACE2, macrophage

Main findings

It has been previously reported that COVID-19 patients exhibit severe lymphocytopenia, but the mechanism through which this depletion occurs has not been described. In order to characterize the cause and process of lymphocyte depletion in COVID-19 patients, the authors performed gross anatomical and *in situ* immune-histochemical analyses of spleens and lymph nodes (hilar and subscapular) obtained from post-mortem autopsies of 6 patients with confirmed positive viremia and 3 healthy controls (deceased due to vehicle accidents).

Primary gross observations noted significant splenic and LN atrophy, hemorrhaging, and necrosis with congestion of interstitial blood vessels and large accumulation of mononuclear cells and massive lymphocyte death. They found that CD68⁺ CD169⁺ cells in the spleens, hilar and subscapular LN, and capillaries of these secondary lymphoid organs expressed the ACE2 receptor and stain positive for the SARS-CoV-2 nucleoprotein (NP) antigen, while CD3⁺ T cells and B220⁺ B cells lacked both the ACE2 receptor and SARS-CoV-2 NP antigen. ACE2⁺ NP⁺ CD169⁺ macrophages were positioned in the splenic marginal zone (MZ) and in the marginal sinuses of LN, which suggests that these macrophages were positioned to encounter invading pathogens first and may contribute to virus dissemination.

Since SARS-CoV-2 does not directly infect lymphocytes, the authors hypothesized that the NP⁺ CD169⁺ macrophages are responsible for persistent activation of lymphocytes via Fas::FasL interactions that would mediate activation-induced cell death (AICD). Indeed, the expression of Fas was significantly higher in virus-infected tissue than that of healthy controls, and TUNEL staining showed significant lymphocytic apoptosis. Since pro-inflammatory cytokines like IL-6 and TNF- α can also engage cellular apoptosis and necrosis, the authors interrogated the cytokine expression of the secondary lymphoid organs from COVID-19 patients; IL-6, not TNF- α , was elevated in virus-infected splenic and lymph node tissues, compared to those

of healthy controls, and immunofluorescent staining showed that IL-6 is primarily produced by the infected macrophages. *In vitro* infection of THP1 cells with SARS-CoV-2 spike protein resulted in selectively increased *Il6* expression, as opposed to *Il1b* and *Tnfa* transcription. Collectively, the authors concluded that a combination of Fas up-regulation and IL-6 production by NP⁺ CD169⁺ macrophages induce AICD in lymphocytes in secondary lymphoid organs, resulting in lymphocytopenia.

In summary, this study reports that CD169⁺ macrophages in the splenic MZ, subscapular LN, and the lining capillaries of the secondary lymphoid tissues express ACE2 and are susceptible to SARS-CoV-2 infection. The findings point to the potential role of these macrophages in viral dissemination, immunopathology of these secondary lymphoid organs, hyperinflammation and lymphopenia.

Limitations

Technical

A notable technical limitation is the small number of samples (n=6); moreover, the analysis of these samples using multiplexed immunohistochemistry and immunofluorescence do not necessarily provide the depth of unbiased interrogation needed to better identify the cell types involved.

Biological

The available literature and ongoing unpublished studies, including single-cell experiments of spleen and LN from organ donors, do not indicate that ACE2 is expressed by macrophages; however, it remains possible that ACE2 expression may be triggered by type I IFN in COVID-19 patients. Importantly, the SARS-CoV-2 NP staining of the macrophages does not necessarily reflect direct infection of these macrophages; instead, positive staining only indicates that these macrophages carry SARS-CoV-2 NP as antigen cargo, which may have been phagocytosed. Direct viral culture of macrophages isolated from the secondary lymphoid organs with SARS-CoV-2 is required to confirm the potential for direct infection of macrophages by SARS-CoV-2. Additionally, it is important to note that the low to negligible viremia reported in COVID-19 patients to-date does not favor a dissemination route via the blood, as suggested by this study, which would be necessary to explain the presence of virally infected cells in the spleen.

Relevance

Excess inflammation in response to SARS-CoV-2 infection is characterized by cytokine storm in many COVID-19 patients. The contribution of this pathology to the overall fatality rate due to COVID-19, not even necessarily directly due to SARS-CoV-2 infection, is significant. A better understanding of the full effect and source of some of these major cytokines, like IL-6, as well as the deficient immune responses, like lymphocytopenia, is urgently needed. In this study, the authors report severe tissue damage in spleens and lymph nodes of COVID-19 patients and identify the role that CD169⁺ macrophages may play in the hyperinflammation and lymphocytopenia that are both

characteristic of the disease. It may, therefore, be important to note the effects that IL-6 inhibitors like Tocilizumab and Sarilumab may specifically have on splenic and LN function. It is important to note that similar observations of severe splenic and LN necrosis and inflammation in patients infected with SARS-CoV-1 further support the potential importance and relevance of this study.

14.92 Cigarette smoke triggers the expansion of a subpopulation of respiratory epithelial cells that express the SARS-CoV-2 receptor ACE2

(2284)

14.92.1 Keywords

- CoVID-19
- 2019-nCoV
- SARS-CoV-2
- respiration
- cigarette
- ACE2
- lung

14.92.2 Main Findings

- Study uses scRNAseq, bulk seq data and air-liquid interface culture experiments to show that cigarette smoke causes a dose-dependent upregulation of ACE2 in mouse and human lungs (transplantation, tumor resection, or IPF datasets).
- ACE2 was not up-regulated in patients with asthma or lung-sarcoidosis or in mouse models of cystic fibrosis or carcinogen exposure.
- Cathepsin B (alternate protease involved in viral entry) is increased in smoke-exposed mouse or human lungs.
- Smoke triggers a protective expansion of mucus-secreting MUC5AC+ goblet and SCGB1A1+ club cells; ACE2 presence in these cells is increased upon smoke exposure.

14.92.3 Limitations:

- Long-term smokers usually have several co-morbidities including immune dysfunction, which can affect interpretation of CoV-2 susceptibility in these datasets. Ideally, analyses can control for major co-morbidities across smokers and non-smokers (immune suppression, cardiovascular disease and atherosclerosis).
- Hyperplasia of ACE2+ goblet cells upon smoking needs to be separated from ACE2 upregulation in existing goblet cells.

- ACE2 expression increase alone does not confirm increased viral entry into goblet cells; future studies with air-liquid interface cultures testing CoV-2 infectivity in *ex vivo* epithelial cells from human epithelial lines, *ex vivo* samples or hACE2 mice will be very informative.

14.92.4 Significance

- This study may partially explain why smokers are more likely to develop severe SARS-CoV-2 infections. Also, the reversibility of ACE2 expression upon smoking cessation suggests that quitting smoking could lessen CoV-2 susceptibility.
- Absence of ACE2 upregulation in other lung inflammation pathologies implies CoV-2 susceptibility might be smoking-specific, and not fibrosis-specific.
- Another preprint showed ACE2 expression increases in lung of patients with CoV-2 co-morbidities such as hypertension ([2257](#)); these studies collectively paint a better picture of CoV-2 susceptibility before actual experiments can be carried out.

14.92.5 Credit

Review by Samarth Hegde as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

14.93 The comparative superiority of IgM-IgG antibody test to real-time reverse transcriptase PCR detection for SARS-CoV-2 infection diagnosis

Liu et al. *medRxiv.* ([2285](#))

14.93.1 Keywords

- IgM/IgG antibody test
- nucleic acid test
- SARS-CoV-2 detection

14.93.2 Main Findings

The study compares IgM and IgG antibody testing to RT-PCR detection of SARS-CoV-2 infection. 133 patients diagnosed with SARS-CoV-2 in Renmin Hospital (Wuhan University, China) were analyzed. The positive ratio was 78.95% (105/133) in IgM antibody test (SARS-CoV-2 antibody detection kit from YHLO Biotech) and 68.42% (91/133) in RT-PCR (SARS-CoV-2 ORF1ab/N qPCR detection kit). There were no differences in the sensitivity of SARS-CoV-2 diagnosis in patients grouped according to disease severity. For example, IgG responses were detected in 93.18% of moderate cases, 100% of severe

cases and 97.3% of critical cases. In sum, positive ratios were higher in antibody testing compared to RT-PCR detection, demonstrating a higher detection sensitivity of IgM-IgG testing for patients hospitalized with COVID-19 symptoms.

14.93.3 Limitations

This analysis only included one-time point of 133 hospitalized patients, and the time from symptom onset was not described. There was no discussion about specificity of the tests and no healthy controls were included. It would be important to perform similar studies with more patients, including younger age groups and patients with mild symptoms as well as asymptomatic individuals. It is critical to determine how early after infection/symptom onset antibodies can be detected and the duration of this immune response.

14.93.4 Significance

The IgM-IgG combined testing is important to improve clinical sensitivity and diagnose COVID-19 patients. The combined antibody test shows higher sensitivity than individual IgM and IgG tests or nucleic acid-based methods, at least in patients hospitalized with symptoms.

14.93.5 Credit

Review by Erica Dalla as part of a project by students, postdocs and faculty at the Immunology Institute of the Icahn School of Medicine, Mount Sinai.

Title: Lectin-like Intestinal Defensin Inhibits 2019-nCoV Spike binding to ACE2

Immunology keywords: defensins, spike protein, intestinal Paneth cells, ACE2 binding

Main Findings:

Human ACE2 was previously identified as the host receptor for SARS-CoV-2. Despite ACE2 being expressed in both lung alveolar epithelial cells and small intestine enterocytes, respiratory problems are the most common symptom after viral infection while intestinal symptoms are much less frequent. Thus, the authors here investigate the biology behind the observed protection of the intestinal epithelium from SARS-CoV-2. Human defensin 5 (HD5), produced by Paneth cells in the small intestine, was shown to interact with human ACE2, with a binding affinity of 39.3 nM by biolayer interferometry (BLI). A blocking experiment using different doses of HD5 coating ACE2 showed that HD5 lowered viral spike protein S1 binding to ACE2. Further, a molecular dynamic simulation demonstrated a strong intermolecular interaction between HD5 and the ACE2 ligand binding domain. To test HD5 inhibitory effect on S1 binding to ACE2, human intestinal epithelium Caco-2 cells were preincubated with HD5. Preincubation strongly reduced adherence of S1 to surface of cells. HD5 was effective at a concentration as low as 10 µg/mL, comparable to the concentration found in the intestinal fluid.

Limitations:

The study focuses exclusively on intestinal cells. However, HD5 could have been tested to block ACE2-S1 binding in human lung epithelial cells as a potential treatment strategy. It would be useful to know whether HD5 could also prevent viral entry in lung cells.

Relevance:

This work provides the first understanding of the different efficiency of viral entry and infection among ACE2-expressing cells and tissues. Specifically, the authors show that human defensin 5 produced in the small intestine is able to block binding between S1 and ACE2 necessary for viral entry into cells. The study provides a plausible explanation on why few patients show intestinal symptoms and suggests that patients with intestinal disease that decrease defensins' production may be more susceptible to SARS-CoV-2. It also indicates that HD5 could be used as a molecule to be exogenously administered to patients to prevent viral infection in lung epithelial cells.

Title:**Susceptibility of ferrets, cats, dogs and different domestic animals to SARS-coronavirus-2**

Immunology keywords: SARS-CoV-2, ferret, cat, laboratory animal, domestic animals

The main finding of the article:

This study evaluated the susceptibility of different model laboratory animals (ferrets), as well as companion (cats and dogs), and domestic animals (pigs, chickens and ducks) to SARS-CoV-2. They tested infection with two SARS-CoV2 isolates, one from an environmental sample collected in the Huanan Seafood Market in Wuhan (F13-E) and the other from a human patient in Wuhan (CTan-H).

Ferrets were inoculated with either of the two viruses by intranasal route with 10^5 pfu, and the viral replication was evaluated. Two ferrets from each group were euthanized on day 4 post infection (p.i.). At day 4 p.i., viral RNA and infectious viruses were detected only in upper respiratory tract (nasal turbinate, upper palate, tonsils, but not in the trachea, lungs or other tissues. Viral RNA and virus titer in the remaining ferrets were monitored in nasal washes and rectal swabs on days 2, 4, 6, 8 and 10 p.i. Viral RNA and infectious viruses were detected in nasal washes until day 8 p.i. One ferret in each group developed fever and loss of appetite on days 10 and 12 p.i., however, viral RNA was practically undetectable. These two ferrets showed severe lymphoplasmacytic perivascularitis and vasculitis in the lungs and lower antibody titers compared to other 4 ferrets.

Cats. Five subadult 8-month-old domestic cats were inoculated with CTan-h virus and three uninfected cats were placed in a cage adjacent to each of the infected cats to monitor respiratory droplet transmission. Viral RNA was detected in the upper respiratory organs from all infected cats and in one out

of three exposed cats. All infected (inoculated and exposed) cats developed elevated antibodies against SARS-CoV2. Viral replication studies with juvenile cats (70-100 days) revealed massive lesions in the nasal and tracheal mucosa epithelium and lungs of two inoculated cats which died or were euthanized on day 3 p.i., and infection in one out of three exposed cats. These results indicated SARS-CoV2 could replicate in cats, that juvenile cats were more susceptible than adults, and that SARS-CoV2 could be transmitted via respiratory droplets between cats.

Dogs and others. Five 3-month-old beagle dogs were inoculated and housed with two uninoculated beagles in a room. Two virus inoculated dogs seroconverted, but others including two contact dogs were all seronegative for SARS-CoV2 and infectious virus was not detected in any swabs collected. Viral RNA was not detected in swabs from pigs, chickens, and ducks inoculated or contacted. These results indicated that dogs, pigs, chickens, and ducks might have low or no susceptibility to SARS-CoV2.

Critical analysis of the study:

This manuscript describes the viral replication and clinical symptoms of SARS-CoV2 infection in ferrets, and the SARS-CoV2 infection and transmission in cats. Clinical and pathological analysis was not performed in cats, therefore the correlation of virus titer with symptoms severity in the adult and juvenile cats could not be determined.

The importance and implications for the current epidemics:

SARS-CoV-2 transmission to tigers, cats and dogs has been previously reported. It should be noted that this manuscript did not evaluate the transmission from cats to human. Nevertheless, it clearly showed higher susceptibility of ferrets and domestic cats to SARS-CoV-2. This data strongly indicates the need for surveillance of possible infection and transmission of SARS-CoV-2 by domestic cats.

14.94 Virus-host interactome and proteomic survey of PMBCs from COVID-19 patients reveal potential virulence factors influencing SARS-CoV-2 pathogenesis

Li et al. *bioRxiv.* ([195](#))

14.94.1 Keywords

- PBMC
- virulence factors – interaction network – nsp9
- nsp10 – NKRF

14.94.2 Main findings

The authors identified **intra-viral protein-protein interactions** (PPI) with two different approaches: genome wide yeast-two hybrid (Y2H) and co-immunoprecipitation (co-IP). A total of 58 distinct PPI were characterized. A screen of **viral-host PPI** was also established by overexpressing all the SARS-CoV-2 genes with a Flag epitope into HEK293 cells and purifying each protein complex. Interacting host proteins were then identified by liquid chromatography and tandem mass spectrometry. 251 cellular proteins were identified, such as subunits of ATPase, 40S ribosomal proteins, T complex proteins and proteasome related proteins, for a total of 631 viral-host PPI. Several interactions suggesting protein-mediated modulation of the immune response were identified, highlighting the multiple ways SARS-CoV-2 might reprogram infected cells.

Subsequently, the authors compared global proteome profiles of PBMCs from healthy donors (n=6) with PBMC from COVID-19 patients with mild (n=22) or severe (n=13) symptoms. 220 proteins were found to be differentially expressed between *healthy donors and mild COVID-19 patients*, and a pathway analysis showed **a general activation of the innate immune response**. 553 proteins were differentially expressed between the PBMC of *mild and severe COVID-19 patients*, most of them (95%) being downregulated in severe patients. Functional pathway analysis indicated a defect of T cell activation and function in severe COVID-19. There was also evidence suggesting reduced antibody secretion by B cells. Together, these results suggest a **functional decline of adaptive immunity**. A FACS analysis of PBMC from severe patients indicated higher levels of IL6 and IL8 but not IL17 compared to mild patients.

Finally, the authors focused on NKRF, an endogenous repressor of IL8/IL6 synthesis that was previously identified as interacting with SARS-Cov-2 nsp9,10,12,13 and 15. Individually expressed nsp9 and nsp10 (but not nsp12, nsp13, nsp15) induced both IL6 and IL8 in lung epithelial A459 cells, indicating that nsp9 and nsp10 may be directly involved in the induction of these pro-inflammatory cytokines. The authors finally argue that nsp9 and nsp10 represent potential drug targets to prevent over-production of IL6 and IL8 in infected cells, and reducing the over-activation of neutrophils.

14.94.3 Limitations

First, the authors seem to have forgotten to include the extended data in the manuscript, and their proteomic data does not seem to be publicly available for the moment, which limits greatly our analysis of their results.

While this work provides important data on host and viral PPI, only 19 interactions were identified by Y2H system but 52 with co-IP. The authors do not comment about what could lead to such differences between the two techniques and they don't specify whether they detected the same interactions using the two techniques.

Moreover, the PBMC protein quantification was performed comparing bulk PBMC. Consequently, protein differences likely reflect differences in cell populations rather than cell-intrinsic differences in protein expression. While

this analysis is still interesting, a similar experiment performed on pre-sorted specific cell populations would allow measuring proteome dynamics at a higher resolution.

Finally, the authors did not discuss their results in regards to another SARS-CoV-2 interactome of host-viral PPI that had been published previously¹. This study reported 332 host-virus PPI, but no interaction of viral proteins with NKRFP was found. Some interactions were found in both studies (eg. N and G3BP1, Orf6 and RAE1). However, the time point used to lyse the cells were different (40h previously vs 72h here), which could explain some of the differences.

14.94.4 Relevance

The identification of many interactions between intra-viral and host-virus PPI provides an overview of host protein and pathways that are modulated by SARS-CoV-2, which can lead to the identification of potential targets for drug development.

In the model proposed by the authors, nsp9 and nsp10 from SARS-CoV-2 induce an over-expression of IL6 and IL8 by lung epithelial cells, which recruits neutrophils and could lead to an excess in lung infiltration. Nsp9 has been shown to be essential for viral replication for SARS-CoV-1², and shares a 97% homology with nsp9 from SARS-CoV-2³. Further, nsp9 crystal structure was recently solved³, which can help to develop drug inhibitors if this protein is further confirmed as being important for the virulence of SARS-CoV-2.

1. Gordon DE, Jang GM, Bouhaddou M, et al. A SARS-CoV-2-Human Protein-Protein Interaction Map Reveals Drug Targets and Potential Drug-Repurposing. *bioRxiv*. March 2020:2020.03.22.002386. doi:10.1101/2020.03.22.002386

2. Miknis ZJ, Donaldson EF, Umland TC, Rimmer RA, Baric RS, Schultz LW. Severe acute respiratory syndrome coronavirus nsp9 dimerization is essential for efficient viral growth. *J Virol*. 2009;83(7):3007-3018. doi:10.1128/JVI.01505-08

3. Littler DR, Gully BS, Colson RN, Rossjohn J. *Crystal Structure of the SARS-CoV-2 Non-Structural Protein 9, Nsp9*. Molecular Biology; 2020. doi:10.1101/2020.03.28.013920

Title: Prediction and Evolution of B Cell Epitopes of Surface Protein in SARS-CoV-2

Keywords: SARS-CoV-2; Epitopes; Bioinformatics; Evolution

Summary/Main findings:

Lon et al. used a bioinformatic analysis of the published SARS-CoV-2 genomes in order to identify conserved linear and conformational B cell epitopes found on the spike (S), envelope (E), and membrane (M) proteins. The characterization of the surface proteins in this study began with an assessment of the peptide sequences in order to identify hydrophilicity

indices and protein instability indices using the Port-Param tool in ExPASy. All three surface proteins were calculated to have an instability score under 40 indicating that they were stable. Linear epitopes were identified on the basis of surface probability and antigenicity, excluding regions of glycosylation. Using BepiPred 2.0 (with a cutoff value of 0.35) and ABCpred (with a cutoff value of 0.51), 4 linear B cell epitopes were predicted for the S protein, 1 epitope for the E protein, and 1 epitope for the M protein. For structural analysis, SARS-CoV assemblies published in the Protein Data Bank (PDB) acting as scaffolds for the SARS-CoV-2 S and E amino acid sequences were used for input into the SWISS-MODEL server in order to generate three-dimensional structural models for the assessment of conformational epitopes. Using Ellipro (cutoff value of 0.063) and SEPPA (cutoff value of 0.5), 1 conformational epitope was identified for the S protein and 1 epitope was identified for the E protein, both of which are accessible on the surface of the virus. Finally, the Consurf Server was used to assess the conservation of these epitopes. All epitopes were conserved across the published SARS-CoV-2 genomes and one epitope of the spike protein was predicted to be the most stable across coronavirus phylogeny.

Critical Analysis/Limitations:

While this study provides a preliminary identification of potential linear and conformational B cell epitopes, the translational value of the epitopes described still needs extensive experimental validation to ascertain whether these elicit a humoral immune response. The conformational epitope analyses are also limited by the fact that they are based off of predicted 3D structure from homology comparisons and not direct crystal structures of the proteins themselves. Additionally, since there was not a published M protein with a high homology to SARS-CoV-2, no conformational epitopes were assessed for this protein. Finally, while evolutionary conservation is an important consideration in understanding the biology of the virus, conservation does not necessarily imply that these sites neutralize the virus or aid in non-neutralizing *in vivo* protection.

Relevance/Implications:

With further experimental validation that confirms that these epitopes induce effective antibody responses to the virus, the epitopes described can be used for the development of treatments and vaccines as well as better characterize the viral structure to more deeply understand pathogenesis.

15 Appendix B

Contributors were asked to complete this template to summarize and evaluate new papers related to diagnostics.

Title: Please edit the title to add the name of the paper after the colon

Please paste a link to the paper or a citation here:

Link:

What is the paper's [Manubot-style citation](#)?

Citation:

Please list some keywords (3-10) that help identify the relevance of this paper to COVID-19

- keyword 1 (replace me, copy and paste more than three if needed)
- keyword 2 (replace me, copy and paste more than three if needed)
- keyword 3 (replace me, copy and paste more than three if needed)

Please note the publication / review status

- ☐ Pre-print
- ☐ New Peer-Reviewed Paper
- ☐ Peer-Reviewed Paper Pre-2020

Which areas of expertise are particularly relevant to the paper?

- ☐ virology
- ☐ epidemiology
- ☐ biostatistics
- ☐ immunology
- ☐ pharmacology

Questions to answer about each paper:

Please provide 1-2 sentences introducing the study and its main findings

Study question(s) being investigated:

What type of testing scenario is being considered?

Is it a screening test (used for individuals with no symptoms), diagnostic test (used for individuals with symptoms), or definitive test (used for individuals who have had previous positive test results on diagnostic or screening tests)?

Study population:

What is the model system (e.g., human study, animal model, cell line study)?

What is the sample size?

What is the “pre-test” probability of disease in the study population (i.e., what is the anticipated prevalence of the disease?)

For human studies, the following are related to the pre-test probability:

What countries/regions are considered?

What is the age range, gender, other relevant characteristics?

What is the setting of the study (e.g., random sample of school children, retirement communities, etc.)?

What other specific inclusion-exclusion criteria are considered?

Reference test:

What reference test is considered as a “gold standard” comparator for the test under investigation?

Test assignment:

How are the new and reference tests assigned?

Examples of assignment could include: Recruited individuals have initially undergone neither the new nor the reference test; individuals tested as positive or negative by the reference test undergo the new test; individuals who have undertaken the new test are assessed by the standard test.

Are there any other relevant details about the study design?

Depending on how individuals are chosen, the test may be biasing towards more sick or less sick individuals or very clear-cut positive/negative cases. Any factors that would influence this bias should be included here.

Test conduct:

How were tests performed?

Describe technical details of assays used, when measurements were taken and by whom, etc. for both the new and standard tests.

Test Assessment

Describe how individuals are classified as positive or negative, e.g. if a threshold is used.

Is there evidence that the test is precise/reproducible when repeated more than once?

Are measurements complete?

For example: Do some participants undergo just one test (the new or the reference test)? Are there individuals with inconclusive results?

Results summary:

What are the estimated sensitivity, specificity, positive predictive value (PPV), and negative predicted value (NPV)?

Note that the PPV and NPV represent “post-test” probabilities of disease and are generally more meaningful than sensitivity and specificity. Sometimes the post-test odds will be given instead.

What are the confidence bounds around these intervals?

Interpretation of results for study population:

How good is the test at ruling in or ruling out a disease based on the post-test probabilities?

Are there identified side affects of the test?

Is patient adherence to the test likely to be an issue?

Extrapolation of conclusions to other groups of individuals

How well is the test likely to work in populations with different pretest odds?

For example, if the prevalence is lower, then the PPV will also be lower, but the NPV will be higher.

How costly is the test?

How difficult is it to perform the test in different settings?

Could the test be combined with other existing tests?

Summary of reliability

1-2 sentences on concluding remarks, including summary of strengths, weaknesses, limitations.

Progress

Check off the components as they are completed. If the component is not applicable, check the box as well.

- ☐ 1-2 sentences introducing the study and its main findings
- ☐ Describe testing scenario
- ☐ Describe model system
- ☐ Sample size
- ☐ Describe prevalence of disease
- ☐ Describe countries/regions are considered
- ☐ Describe age range, gender, other relevant characteristics

- ❑ Describe setting of the study
- ❑ Describe other specific inclusion-exclusion criteria
- ❑ Describe "gold standard"
- ❑ Describe how the new and reference tests assigned
- ❑ Describe other relevant details about the study design
- ❑ Describe how the tests were performed
- ❑ Describe how individuals are classified as positive or negative
- ❑ Describe if test is precise/reproducible
- ❑ Describe whether measurements are complete
- ❑ What are the estimated sensitivity, specificity, positive predictive value (PPV), and negative predicted value (NPV)?
- ❑ What are the confidence bounds around these intervals?
- ❑ Describe post-test probabilities
- ❑ Describe side affects of the test
- ❑ Describe patient adherence
- ❑ Describe how it will extrapolate
- ❑ How costly is the test?
- ❑ How difficult is it to perform the test in different settings?
- ❑ Could the test be combined with other existing tests?
- ❑ Summary of reliability

16 Appendix C

Contributors were asked to complete this template to summarize and evaluate new papers related to therapeutics.

Title: Please edit the title to add the name of the paper after the colon

Please paste a link to the paper or a citation here:

Link:

What is the paper's [Manubot-style citation](#)?

Citation:

Please list some keywords (3-10) that help identify the relevance of this paper to COVID-19

- keyword 1 (replace me, copy and paste more than three if needed)
- keyword 2 (replace me, copy and paste more than three if needed)
- keyword 3 (replace me, copy and paste more than three if needed)

Please note the publication / review status

- ☐ Pre-print
- ☐ New Peer-Reviewed Paper
- ☐ Peer-Reviewed Paper Pre-2020

Which areas of expertise are particularly relevant to the paper?

- ☐ virology
- ☐ epidemiology
- ☐ biostatistics
- ☐ immunology
- ☐ pharmacology

Questions to answer about each paper:

Please provide 1-2 sentences introducing the study and its main findings

Study question(s) being investigated:

How many/what drugs/combinations are being considered?

What are the main hypotheses being tested?

Study population:

What is the model system (e.g., human study, animal model, cell line study)?

What is the sample size? If multiple groups are considered, give sample size for each group (including controls).

- number treated with treatment A
- number treated with treatment B

For human studies:

What countries/regions are considered?

What is the age range, gender, other relevant characteristics?

What is the setting of the study (random sample of school children, inpatient, outpatient, etc)?

What other specific inclusion-exclusion criteria are considered?

For example, do the investigators exclude patients with diagnosed neoplasms or patients over/under a certain age?

Treatment assignment:

How are treatments assigned?

For example, is it an interventional or an observational study?

Is the study randomized?

A study can be interventional but not randomized (e.g., a phase I or II clinical trial is interventional but often not randomized).

Provide other relevant details about the design.

This includes possible treatment stratification (e.g., within litters for animal studies, within hospitals for human studies), possible confounding variables (e.g., having a large age range of individuals), possible risks of bias and how they are addressed (e.g., is there masking in a clinical trial? how are individuals chosen in an observational study?).

Outcome Assessment:

Describe the outcome that is assessed and whether it is appropriate.

For example: Is the outcome assessed by a clinician or is it self-reported? Is the outcome based on viral load or a functional measurement (e.g., respiratory function, discharge from hospital)? What method is used to measure the outcome? How long after a treatment is the outcome measured?

Are outcome measurements complete?

For example, are there individuals lost to follow up?

Are outcome measurements subject to various kinds of bias?

For example, a lack of masking in randomized clinical trials.

Statistical Methods Assessment:

What methods are used for inference?

For example, logistic regression, nonparametric methods.

Are the methods appropriate for the study?

For example, are clustered data treated independently or are clusters adjusted for, such as different hospitals or litters?

Are adjustments made for possible confounders?

For example, adjustment for age, sex, or comorbidities.

Results Summary:

What is the estimated association?

For example, is it an estimated odds ratio, a median difference in detected cases, etc?

What measures of confidence or statistical significance are provided?

For example, confidence intervals, p-values, and/or Bayes factors.

Interpretation of results for study population:

Can we make a causal interpretation for the individuals in the study of drug -> outcome, such as "taking drug A improves likelihood of survival twofold over taking drug B."

For example, with a well-performed animal study or randomized trial it is often possible to infer causality. If is an observational study, does it match up with some of the Bradford Hill criteria?

<https://www.edwardtufte.com/tufte/hill>

https://en.wikipedia.org/wiki/Bradford_Hill_criteria

Are there identified side effects or interactions with other drugs?

For example, is the treatment known to cause liver damage or to not be prescribed for individuals with certain comorbidities?

Are there specific subgroups with different findings?

For example, do individuals with a specific baseline seem to do particularly well? Be particularly cautious with respect to multiple testing here.

Extrapolation of conclusions to other groups of individuals not specifically included in the study:

If the study is an animal study, which animal and how relevant is that model?

Is the model system appropriate? Is there evidence from past use that it's highly-relevant to therapeutic design in this context?

If it is a human study, what characteristics of the study population may support/limit extrapolation?

- Can results extrapolate easily to other similar groups? (e.g., same country, similar age groups)
- What would happen if conditions are extended in terms of dose or duration?
- Can results be extrapolated to other populations or in very different settings? (e.g., different age group, primary care setting vs emergency department etc)

Summary of reliability

1-2 sentences on concluding remarks, including summary of strengths, weaknesses, limitations.

Progress

Check off the components as they are completed. If the component is not applicable, check the box as well.

- ☐ 1-2 sentences introducing the study and its main findings
- ☐ Describe How many/what drugs/combinations are being considered
- ☐ Describe the model system
- ☐ What is the sample size?
- ☐ What countries/regions are considered
- ☐ What is the age range, gender, other relevant characteristics
- ☐ Describe study setting
- ☐ Describe other specific inclusion-exclusion criteria
- ☐ Describe how treatments are assigned
- ☐ Describe randomization (or not) and other relevant details about the design
- ☐ Describe the outcome that is assessed and whether it is appropriate.
- ☐ Describe whether the outcome measurements are complete
- ☐ Are outcome measurements subject to various kinds of bias?
- ☐ Describe methods used for inference
- ☐ Describe whether the methods are appropriate for the study
- ☐ Are adjustments made for possible confounders?
- ☐ Describe the estimated association
- ☐ What measures of confidence or statistical significance are provided?
- ☐ Describe whether a causal interpretation can be made
- ☐ Are there identified side effects or interactions with other drugs?
- ☐ Are there specific subgroups with different findings?
- ☐ If the study is an animal study, which animal and how relevant is that model?

- ❑ If it is a human study, what characteristics of the study population may support/limit extrapolation?
- ❑ Summary of reliability

17 Appendix D

Contributors were asked to complete this template to summarize and evaluate new papers related to topics besides therapeutics and diagnostics.

Title: Please edit the title to add the name of the paper after the colon.

General Information Please paste a link to the paper or a citation here:

Link:

What is the paper's [Manubot-style citation](#)?

Citation:

Is this paper primarily relevant to Background or Pathogenesis?

- ☐ Background
- ☐ Pathogenesis
- ☐ Methods

Please list some keywords (3-10) that help identify the relevance of this paper to COVID-19

- keyword 1 (replace me, copy and paste more than three if needed)
- keyword 2 (replace me, copy and paste more than three if needed)
- keyword 3 (replace me, copy and paste more than three if needed)

Please note the publication / review status

- ☐ Pre-print
- ☐ New Peer-Reviewed Paper
- ☐ Peer-Reviewed Paper Pre-2020

Which areas of expertise are particularly relevant to the paper?

- ☐ virology
- ☐ epidemiology
- ☐ biostatistics
- ☐ immunology
- ☐ pharmacology
- ☐ other:

Summary

Suggested questions to answer about each paper: - What did they analyze? - What methods did they use? - Does this paper study COVID-19, SARS-CoV-2, or a related disease and/or virus? - What is the main finding (or a few main takeaways)? - What does this paper tell us about the background and/or

diagnostics/therapeutics for COVID-19 / SARS-CoV-2? - Do you have any concerns about methodology or the interpretation of these results beyond this analysis?

Any comments or notes?

1. <https://asapbio.org/preprints-and-covid-19> as well as <https://retractionwatch.com/retracted-coronavirus-covid-19-papers>↵
2. <https://depts.washington.edu/pandemicalliance/covid-19-literature-report/latest-reports>↵
3. <https://outbreaksci.prereview.org>↵
4. <https://asapbio.org/preprints-and-covid-19>↵
5. <https://disqus.com/by/sinaiimmunologyreviewproject>↵
6. <https://rapidreviewscovid19.mitpress.mit.edu>↵
7. <https://greenelab.github.io/covid19-review>↵
8. <https://casrai.org/credit>↵
9. <https://www.gitter.im>↵
10. <https://greenelab.github.io/covid19-review>↵
11. <https://greenelab.github.io/covid19-review/manuscript.pdf>↵
12. Vaccines: <https://github.com/owid/covid-19-data>; Clinical Trials: https://github.com/ebmdatalab/covid_trials_tracker-covid; Cases and Deaths: <https://github.com/CSSEGISandData/COVID-19>↵
13. <https://github.com/greenelab/covid19-review/blob/master/.github/workflows/update-external-resources.yaml>↵
14. <https://github.com/greenelab/covid19-review/tree/external-resources>↵
15. <https://github.com/greenelab/covid19-review/blob/external-resources/environment.yml>↵
16. <https://forums.zotero.org/discussion/74933/import-from-clinical-trials-registry> and <https://forums.zotero.org/discussion/77721/add-reference-from-clinical-trials-org>↵
17. <https://www.zotero.org> and <https://github.com/zotero/translation-server>↵
18. <https://github.com/zotero/translators/pull/2153>↵
19. <https://identifiers.org>↵

20. <https://pandoc.org>↵
21. <http://aspell.net>↵
22. <https://github.com/pandoc/lua-filters/tree/master/spellcheck>↵
23. https://twitter.com/j_perkel/status/1245454628235309057↵
24. CONTRIBUTING.md and INSTRUCTIONS.md within the repository↵
25. https://github.com/ismms-himc/covid-19_sinai_reviews↵
26. https://github.com/CSSEGISandData/COVID-19/tree/master/csse_covid_19_data/csse_covid_19_time_series↵
27. <https://github.com/owid/covid-19-data>↵